Genetic epidemiology of *Sarcoptes scabiei* in the Iberian wolf in Asturias, Spain

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A B S T R A C T

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Background: During the last decades, attempts have been made to understand the molecular epidemiology of *Sarcoptes scabiei*, and to detect and clarify the differences between isolates from different hosts and geographic regions. Two main phenomena have been described: (i) host-taxon derived-*Sarcoptes* mite infection in European wild animals (revealing the presence of three separate clusters, namely herbivore-, carnivore- and omnivore-derived *Sarcoptes* populations in Europe) and (ii) prey-to-predator *Sarcoptes* mite infection in the Masai Mara ecosystem.

Results: Using one multiplex of 9 microsatellite markers and *Sarcoptes* mite samples from sympatric Pyrenean chamois, red deer, red fox and Iberian wolf, different population structure analyses revealed concordance with the host-taxon law described for wild animals in Europe, with two main host-derived *Sarcoptes* mite populations, herbivore- and carnivore-derived. Surprisingly, Iberian wolf derived *Sarcoptes* populations had the highest genetic diversity among the other populations, including two different subpopulations: one similar to the herbivore-derived *Sarcoptes* populations, and another similar to carnivore (fox)-derived *Sarcoptes* mite population.

Conclusions: The host-taxon effect in wild animals is still supported with the maintenance of carnivore- and herbivore-derived *Sarcoptes* clusters' separation in analyzed mites. However, this phenomenon could be modified with the inclusion of a large predator as wolf in the present work, revealing prey-to-predator *Sarcoptes* mite infection between the studied host-taxa and suggesting the importance of wolf's immune system for explaining the high variability reported in *C. lupus* derived mites. Further studies of host diet, behavior and movement, and regarding the role played by its immune system, would be of great help to clarify interactions between the two hypotheses, host-taxon and prey-to-predator.

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1. Introduction

Despite representing the first human illness with a known etiologic agent (Montesu et al., 1991; Gakuya et al., 2012a) and affecting more than 100 mammal species

all over the world (Bornstein et al., 2001; Pence and Ueckerman, 2002), the taxonomical status of the ectoparasitic burrowing mite *Sarcoptes scabiei* has been the subject of continuous debate for decades (Pence et al., 1975; Fain, 1978; Andrews, 1983). The development of molecular techniques has provided new approaches and novel data on this subject (Zahler et al., 1999; Walton et al., 2004a; Gu and Yang, 2008); in particular, microsatellites are a valuable technique for the analysis of *S. scabiei* and the relationship between hosts. Microsatellite analysis supports the standing of *S. scabiei* as a single highly variable species with different strains manifesting physiological host-specificity (Walton et al., 2004b; Alasaad et al., 2008b, 2011b, 2012).

A mange epizootic first detected in 1993 in the southern border of Asturias severely affected Southern chamois (*Rupicapra pyrenaica parva*) from the area (Fernández-Moran et al., 1997), leading to an overall 61.3% reduction of their population during the first years after the outbreak (González-Quirós et al., 2002). Although the origin of the outbreak could not be demonstrated, it has been attributed to many domestic goats sharing pastures with chamois in the study area, with subsequent cross-infection (Lavín et al., 2000; Menzano et al., 2007). The epizootic is still expanding eastwards today with sporadic cases of sarcoptic mange in chamois, red deer (*Cervus elaphus*) and roe deer (*Capreolus capreolus* – Oleaga et al., 2008a,b) from the area. Red foxes (*Vulpes vulpes*) and wolves (*Canis lupus*) have also been diagnosed with sarcoptic mange in Western, Central and Eastern Regions of Asturias, and disease is now considered endemic in red foxes.

Three main groups of *S. scabiei* mites have been genetically identified in European wildlife studied so far, namely herbivore, carnivore and omnivore-derived *Sarcoptes* clusters (Rasero et al., 2010). These mite “strains” occur in sympatric as well as geographically distant populations, including Southern chamois, red deer, roe deer and red fox in our study area in Asturias, Northern Spain (Alasaad et al., 2011b). In sympatric mammals, differentiation of mites into “strains” may result from null or limited interspecific transmission, due to low frequency of direct contacts among hosts, low densities of one or more hosts, or different immunological response to the invading mites, limiting their population size, hence their number on the skin surface and their transmissibility to other individuals of the same or a different species. On the other hand, genetically based preference for a particular host (leading to host-associated mating) or the evolution of higher performance or viability on that host (usually entailing a worse performance on the others, Kassen, 2002) have been proposed as possible mite-dependent mechanisms limiting gene flow (Magalhães et al., 2007). The apparent lack of gene flow or recent admixture between carnivore-, herbivore-, and omnivore-derived *Sarcoptes* populations reported in European studied species, including sympatric animals, led to the so called “host-taxon law” formulation (Rasero et al., 2010; Alasaad et al., 2011b).

Parasites are of pivotal importance in food webs. Moreover food webs are very incomplete without parasites (Lafferty et al., 2006), with endo-parasites (Sukhdeo, 2012) as the most frequently described trophically transmitted

parasites (Luong et al., 2013). The only report, to our knowledge, about ecto-parasites transmission was on *Sarcoptes* mite transmission in Masai Mara ecosystem (Gakuya et al., 2011). Theoretically, the higher frequency of direct contacts among hosts within trophic chains (especially when acting as predator, considering that prey are not a “long-lasting available habitat” after predation for ectoparasitic mites) should favor a more efficient gene flow between different mite “strains”. Nevertheless, the study of *S. scabiei* molecular features in a predator/prey ecosystem has not been carried out in any European model, so far. Recently, molecular and epidemiological studies including wild felids and ungulates in Africa have signaled the existence of a potential predator/prey bond in sarcoptic mange transmission, the so called prey-to-predator parasitic infection (Gakuya et al., 2011, 2012b).

The recent isolation of *S. scabiei* mites in 9 out of 12 wolves bearing skin lesion in Asturias (Northern Spain) suggested an increase in morbidity of sarcoptic mange in this wild canid population (Domínguez et al., 2008; Oleaga et al., 2011) and allowed a genetic study of mites in this host for the first time. This work aims to study the genetic structure of *S. scabiei* mites affecting wolves in Northern Spain, and to genetically compare them with mites originating from sympatric wild herbivores (red deer and Southern chamois) and carnivores (red fox). Both wild and domestic ungulates are common prey of the wolf in the study area (Meriggi and Lovari, 1996; Barja, 2009), whereas predation and scavenging on many foxes have been suspected as a source of infection in this top chain carnivore (Bornstein et al., 1995; Mörner et al., 2005; Domínguez et al., 2008). Such molecular analyses could help in determining the extent to which sarcoptic mites may be interspecifically transmitted in one of the few available large predator-prey food chains in Europe.

2. Materials and methods

2.1. Study area

The study was carried out in the Principality of Asturias, a 10,603 km² autonomous region located in North-Western Spain. The area, with a mixture of open pastures and meadows with deciduous and mixed forests, is home of many different wildlife species, including carnivores like the endangered brown bear (*Ursus arctos*), wolf and red fox, and several wild ungulates (red deer, roe deer, wild boar – *Sus scrofa* – and Cantabrian chamois) that represent a high percentage of the diet of wolves (Llaneza et al., 1996).

2.2. Sampling and mite isolation

The study of wolves submitted for necropsy in Asturias (Northern Spain) as part of a wildlife diseases surveillance program allowed the confirmation of sarcoptic mange by mite isolation in 9 out of 47 wolves from 2008 to 2010 (Oleaga et al., 2011), and the extraction of genetic material from mites belonging to 8 of these 9 mange wolves (seven of them were collected in population control hunts carried out by wildlife officers, while one wolf died as a consequence of vehicle collision). Acari were collected after

incubation of 8 skin pieces from each animal on Petri dishes for 24 h at 37 °C and meticulous examination for the detection and identification of ectoparasites using an Olympus SZX9 (10–57×) magnifier (Alasaad et al., 2009). Collected mites were identified as *S. scabiei* according to Wall and Shearer (1997) and preserved in 70% ethanol until DNA extraction. DNA was extracted from 19 mites belonging to 8 mangy confirmed wolves (seven adults and two 6 month old pups).

Mites obtained from 7 chamois ($n=14$), 8 red deer ($n=13$) and 6 red foxes ($n=12$) with confirmed sarcoptic mange were also collected in Asturias from 2006 to 2010, and analyzed in order to compare molecular features with those reported in Iberian wolves.

2.3. DNA extraction and fluorescent-based polymerase chain reaction (PCR) analysis of microsatellite DNA

The DNA of individual *Sarcoptes* mites was extracted using the HotSHOT Plus ThermalSHOCK technique (Alasaad et al., 2008a). Two blanks (reagents only) were included in each extraction to monitor for contamination.

As described by Alasaad et al. (2008b), nine specific *Sarcoptes* mite microsatellites (Sarms 33–38, 40, 41, and 44) were used with one 9× multiplex PCR. One primer from each set was 5' labeled with 6-FAM, VIC, NED or PET® fluorescent dye tag (Applied Biosystems, Foster City, CA, USA). Each 15 µl PCR reaction mixture consisted of 3 µl of the single mite DNA, together with the PCR mixture containing all primer pairs (ranging from 0.04 to 0.1 µM per primer), 200 µM of each dNTP, 1.5 µl of 10× PCR buffer (200 mM KCl and 100 mM Tris–HCl, pH 8.0), 1.5 mM MgCl₂ and 0.15 µl (0.5 U/reaction) HotStar Taq (QIAGEN, Milano, Italy). The thermal profile in a 2720 thermal cycler (Applied Biosystems, Foster City, CA, USA) was as follows: 15 min at 95 °C (initial denaturing), followed by 37 cycles of three steps of 30 s at 94 °C (denaturation), 45 s at 55 °C (annealing) and 1.5 min at 72 °C (extension), before a final elongation of 7 min at 72 °C. Fluorescent PCR amplification products were analyzed using formamide with Size Standard 500 Liz (Applied Biosystems, Foster City, CA, USA) by ABI PRISM 310 Genetic Analyser with pop4. Allele calling was performed using the GeneMapper v. 4.0 software (Applied Biosystems, Foster City, CA, USA). Possible genotyping mistakes (scoring error due to stuttering, large allele dropout) were estimated using MICROCHECKER (Oosterhout et al., 2004).

2.4. Molecular analyses

Expected (H_E) and observed (H_O) heterozygosity, linkage disequilibria (LD), and HWE tests were calculated using GENEPOP (v.3.4 [Raymond and Rousset, 1995]). Deviations from HWE and tests for LD were evaluated using Fisher's exact tests and sequential Bonferroni corrections.

The heterogeneity of genetic diversity among the different *Sarcoptes* mite populations was estimated by the partition of variance components (AMOVA) applying conventional F_{ST} statistics using allele's frequencies as implemented in Arlequin 3.11 (Excoffier, 2006). The analysis of relationships between mites was carried out by the

Bayesian assignment test of the software STRUCTURE (v.2.3.3 [Pritchard et al., 2000]). Burn-in and run lengths of Markov chains were both 100,000. We ran 20 independent runs for each K (for $K=1–10$). The most likely number of clusters was determined using two approaches; by estimating the posterior probability for each K , using the method of Evanno et al. (2005). Finally, each of the inferred clusters was associated with the component populations of its mites.

The degree of genetic relationship among populations was further investigated with FCA as implemented in Genetix v.4.05.2 (Belkhir, 1999).

3. Results

Twenty-eight alleles were detected in the 9 microsatellite loci studied in wolf derived-*Sarcoptes* mites, including one private allele (present only in the wolf population, Sarms 44). The number of alleles for each locus ranged in the wolf population from two (Sarms40 and Sarms 41) to four (Sarms35, Sarms37 and Sarms38). All the alleles detected in studied ungulate species ($n=12$, with 11, 12

Table1

Comparison of allele size and total number of detected alleles in the four sympatric species-derived mite populations studied in Asturias at the 9 microsatellite loci. Private alleles are shown in black, while alleles present both in ungulates and red fox (that are also present in wolf) are underlined and in italics.

Locus#	Chamois	Red deer	Red fox	Wolf
Sarms33	226	226	232 240	226 232 240
Sarms34	176 198	176 198	174	176 198 174
Sarms35	162	162	148 152 156	162 148 152 156
Sarms36	279	279 281	279 281 283	<u>279</u> <u>281</u> 283
Sarms37	172	172	164 170 178	172 164 170 178
Sarms38	213 215	213 215	209 211	213 215 209 211
Sarms40	215	215	217 243	215 217
Sarms41	236	236	234	236 234
Sarms44	262	262	262 270 272	<u>262</u> 270 268
Total number of alleles	11	12	20	28

Fig.1. Results ofSTRUCTURE analysis showing ΔK as proposed by Evanno et al. (2005) method. The best fit of the data was two clusters.

and 9 alleles for chamois, red deer and roe deer, respectively, Alasaad et al., 2011b) and 18 out of the 20 reported in red foxes in Asturias were also present in the wolf population, whereas the only two alleles previously detected in Asturian wildlife but absent in the wolf were two private alleles from red fox (Sarms40 and Sarms44, Table 1).

While mites isolated from wolves contained 27 of the 29 alleles previously detected in *S. scabiei* mites derived from sympatric ungulates and red fox from Asturias, only 3 of these 29 alleles were present both in red fox and ungulate

derived mites. This fact allowed the consideration of 9 (out of 12) and 17 (out of 20) alleles as “exclusive” for ungulate species and red fox derived mites respectively, before including *C. lupus* mites in the study.

There was no evidence of linkage disequilibrium for any of the loci examined ($P > 0.05$). The analysis of molecular data showed deviation from Hardy-Weinberg equilibrium (HWE) in all the 9 microsatellite loci studied in wolf mites, the highest value from studied wildlife species in Asturias (HWE had been detected in two and six loci from Pyrenean chamois and red fox respectively, Alasaad et al., 2011b).

Several mite specimens were collected from each of 4 wolves, with intra-host variation detected in three of them. Variation was reported in all loci with the exception of Sarms44. Sarms36 showed variation in the 3 wolves where intra-host variation was detected, and Sarms34, Sarms37 and Sarms38 presented variation in two wolves.

Mites isolated from wolves offered the highest mean number of alleles (3.111 ± 0.782) from the four studied wild mammal species in Asturias, while herbivore-derived populations showed a lower mean number of alleles (0.444 ± 0.882 in Pyrenean chamois and 0.667 ± 1.000 in red deer) than red fox-derived mites (2.000 ± 1.225) (Table 2).

Mean expected heterozygosity was 0.049 ± 0.097 for chamois, 0.117 ± 0.181 for red deer, 0.416 ± 0.232 for red fox and 0.571 ± 0.140 for wolf.

The Bayesian assignment test of the software STRUCTURE, $\ln \Pr(X|K)$ for the likely number of populations, indicated $K=2$ as the uppermost cluster value (Fig. 1). This differentiation of two clusters as the best fit grouped

Table2

Number of alleles detected in mites belonging to the four sympatric species studied in Asturias.

Locus #	Rupic. 2010	Cervus	Vulpes	Canis	Mean	s.d.	Total number
Sarms33	1	1	2	3	1.750	0.957	3
Sarms34	2	2	1	3	2.000	0.816	3
Sarms35	1	1	3	4	2.250	1.500	4
Sarms36	1	2	3	3	2.250	0.957	3
Sarms37	1	1	3	4	2.250	1.500	4
Sarms38	2	2	2	4	2.500	1.000	4
Sarms40	1	1	2	2	1.500	0.577	3
Sarms41	1	1	1	2	1.250	0.500	2
Sarms44	1	1	3	3	2.000	1.555	4
Mean	0.444	0.667	2.000	3.111	1.556	1.244	3.333
s.d.	0.882	1.000	1.225	0.782	0.972	0.191	0.667

Fig.2. Bar plot of the degree of individual variation between 58 *S. scabiei* from different host species in Asturias (Spain) assigned to given genetic clusters in STRUCTURE, when two ($K=2$) populations are assumed in the dataset. Each cluster is represented by a different color. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

Table 3

Matrix of fixation index (FST) significant *P* values, with significance level *P*=0.05 (above diagonal), and population pairwise FST (below diagonal) for each pairwise comparison of four *Sarcoptes* mite populations from Asturias, Spain.

	<i>Rupicapra pyrenaica</i>	<i>Cervus elaphus</i>	<i>Vulpes vulpes</i>	<i>Canis lupus</i>
<i>Rupicapra pyrenaica</i>		0.081	<0.001*	<0.001*
<i>Cervus elaphus</i>	0.118		<0.001*	<0.001*
<i>Vulpes vulpes</i>	0.742	0.709		<0.001*
<i>Canis lupus</i>	0.374	0.332	0.236	

* Significance level *p*=0.05.

Sarcoptes mites from all herbivore hosts in one cluster, while *Sarcoptes* mites from red fox formed another cluster and wolves showed mites similar to one or the other cluster (Fig. 2).

AMOVA analysis showed differentiation among populations ($F_{ST}=0.48352$; $P<0.001$), signaling that the mite component populations differed greatly. The wolf-derived *Sarcoptes* mite population was statistically ($P<0.001$) different from those living in the two studied herbivore species (chamois and red deer) and from red-fox derived *Sarcoptes* mite populations. Herbivore-derived mite populations formed only one group ($P=0.081$). There were statistically supported ($P<0.001$) genetic differentiations between the other species-derived *Sarcoptes* mite populations. FST between wolf derived- and herbivore-derived *Sarcoptes* mite populations was lower than that between fox derived- and herbivore-derived *Sarcoptes* mite populations (Table 3).

The scatter plot of the Factorial Component Analysis (FCA) for the individual mites collected on chamois, red deer, red fox and wolf, agreed with the results obtained by the Bayesian assignment test: while red deer and chamois derived mites were similar to each other and appeared grouped, red fox mites gathered separately, and mites collected on wolves were grouped in part with the ungulate-derived mites and in part with the red fox-derived mites (Fig. 3).

4. Discussion

The present study reveals that wolf mites show the highest number of different alleles ($n=28$) out of the four wild species surveyed ($n=20$ alleles in red fox, with none of the ungulate species housing mites with more than 12 detected alleles [Alasaad et al., 2011b – Table 1]). Wolf mites contained 27 of the 29 alleles previously detected in *S. scabiei* originating from sympatric ungulates and red fox in Asturias, thus explaining the low number of private alleles reported in this work and suggesting limited genetic separation and relatively high gene flow between *Sarcoptes* mite populations when including wolf in the analysis. The higher variability reported in *C. lupus* mites agrees with results dealing with the mean number of alleles, deviation from Hardy-Weinberg equilibrium (HWE), intra-host variation and mean expected heterozygosity, all of them showing in wolf mites the highest values found in the four analyzed host species. Regarding the genetic diversity among the different *Sarcoptes* mite populations, the obtained FST values are to be considered high, but they are similar to those obtained in previous studies on *S. scabiei* (e.g. Gakuya et al., 2011). We have no clear explanation of such high FST values, however the questionable taxonomic status of this parasite could be behind this high FST values, and hence further molecular studies with wider range of molecular markers are needed.

Fig. 3. Factorial Component Analysis (FCA) of the proportion of variation of *Sarcoptes* mite populations of red deer, chamois, red fox and wolf from Asturias (Northern Spain) assigned to given genetic clusters in Genetix. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

In the Masai Mara (Kenya) ecosystem (Gakuya et al., 2011), the genetic study of mites belonging to different species revealed “prey-to-predator” *Sarcoptes* gene flow leading to deviations from the host taxon phenomenon formulated by Rasero et al. (2010). This work is the first one, in Europe, in which a large predator and the corresponding main sylvatic prey are included in a study aimed to investigate the molecular diversity and gene flow of *Sarcoptes* mites. As in the case of other large predators in Masai Mara, the inclusion of wolf permitted to investigate the outcome of the enhanced opportunities of *Sarcoptes* mite transmission between host taxa within a consolidated “predator/prey” chain, with the eventual evasion of the genetic separation of host-associated mites. In Gakuya’s work (2011) the “favorite prey” effect led to the identification of four different clusters and highlighted inconsistencies with the host-taxon law. As an interesting difference, in our case data showed the maintenance of the two previously described host-associated clusters (red fox vs. herbivore derived mite populations, Figs. 1–3, Alasaad et al., 2011b) and an apparently milder “prey-to-predator” effect associated with the presence of wolf, whose mites belonged to both clusters with scarce variations. The molecular results in the present work can be better assessed when considering FCA information (showing mites distributed within those from red fox or ungulates, Fig. 3) and the Bayesian assignment test for the likely number of populations K (with $K=2$ as the uppermost cluster value, Fig. 1). These data confirm a substantial compliance with the “host taxon law”, with maintenance of a differentiation of mites in two populations (ungulate and red fox derived), and the appearance in wolf of mites belonging to one of these two recognized clusters in Asturias (thus revealing certain “prey to predator” effect). The bar plot representing the degree of individual variation between *S. scabiei* from the different studied host species (Fig. 2) agrees with this view of wolf being able to “collect” mites both from red fox and from ungulates.

In the “prey-to-predator” *Sarcoptes* gene flow reported in wolves in Asturias, two main differences with the Masai Mara ecosystem results can be found: (i) a more evident “carnivore to carnivore” (red fox-wolf) gene flow than observed between lion and cheetah and (ii) an apparent lack of real gene flow, “evolution” or mixture between red fox- and herbivore-derived mite populations, even though wolves may harbor mites belonging to both. The first difference is probably related to the different ecological role that the red fox plays compared to the wolf, lion or cheetah, implying a different nature and frequency of contacts between wolf and red fox in comparison with the two African predators. On the other hand, further research is necessary to clarify the reasons for the apparent lack of gene admixture and scarce “variation-evolution” suggested by molecular data in wolf derived mites.

It is the Authors’ opinion that peculiarities of the immune response developed by the wolf against invading *Sarcoptes* mites may be a key in the understanding of reported molecular data. As shown by Oleaga et al. (2012), in scabietic wolves the hypersensitivity-associated alopecic form is prevalent on the anergy-associated hyperkeratotic form which is usually observed in ungulates

and red fox. Demographic and necropsy data suggest that this predator is able to exert a certain control over infestation with *S. scabiei* (Oleaga et al., 2011), in contrast with the usually deadly development of scabies in the other studied sympatric species in Asturias (Oleaga et al., 2012). The apparent efficiency of the immune system of the wolf against mite proliferation may suggest (i) the development of a limited number of mite generations on it and (ii) the possibility of repeated infestations with mites from any of the sympatric species this canid may get into contact (as suggested by the detection, in the present work, of one wolf infested by both “ungulate-like” and “red-fox-like” mites). These two features may hamper host adaptation and facilitate gene flow among mite populations, thus preventing genetic selection and favoring the high variability reported in *C. lupus* derived mites. A similar situation has been suggested in wild boars, which show greater resistance to these parasites than other mammalian species and whose mites showed high variability and heterozygosity parameters (Rasero et al., 2010). In the case of wolf, the wide range of geographical movement and the large number of species this canid can get into contact (both as predator and as scavenger) are additional arguments favoring repeated infestations with genetically distinct mites from different species.

To complement this type of study it would be interesting to obtain mites isolated from sympatric domestic animals, such as dogs or goats as a possible source of sarcoptic mange for ungulates and even wolf in Asturias. The implementation of tools like the *Sarcoptes*-World Molecular Network (*Sarcoptes*- WMN, Alasaad et al., 2011a) can improve collaboration between researchers and allow more profound studies in the future.

Taking into account these and previous results dealing with hosts (namely wild boars) which have not been considered in this study (Rasero et al., 2010), it seems necessary focus on the pathobiology of the mites and the efficiency of the immune response developed by the different host species, as key aspects for a better interpretation and tuning of the “host-taxon law” when studying gene flow and evolution of parasitic mite populations. On the other hand, the role that host species play in studied ecosystems, especially when including susceptible carnivores displaying preying behavior on mangy prey, must be also considered for a correct evaluation of a possible “prey-to-predator” effect. The consideration of mites and their hosts as an ecosystem in constant change and adaptation process requires due attention not only to the biology, ecology and behavior of host species, but also to their immune system performance and the possibly host species population-specific variability of clinical presentations of sarcoptic mange.

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