- Ecological niche segregation among five toothed whale species off the NW Iberian
- 2 Peninsula using ecological tracers as multi-approach.
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Abstract This study aims to assess niche segregation among the five main toothed whales that frequent the NW Iberian Peninsula waters: the common dolphin, the harbour porpoise, the bottlenose dolphin, the striped dolphin and the long-finned pilot whale. We used cadmium (Cd) and stable isotope ratios (δ^{13} C and δ^{15} N) as ecological tracers to assess degree of segregation in diet/trophic level and in foraging habitat, over various time-scales. δ^{13} C values highlighted different habitat while Cd concentrations highlighted feeding differences between oceanic and neritic species. Moreover, δ^{15} N values suggest different trophic levels of prey targeted within oceanic and neritic species. Hence, results revealed long-term ecological segregation among five toothed whales that coexist in the NWIP and demonstrated the ability of ecological tracers to discriminate ecological niches among closely related species.

Introduction

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Hutchinson (1957) defined the central concept of ecological niche as a set of biotic and 40 abiotic conditions in which a species is able to persist and maintain stable population sizes. 41 These conditions can be structured along three main dimensions: trophic (e.g. diet 42 composition, trophic level and prey quality), spatial (horizontally and vertically) and temporal 43 (from diel activity patterns to yearly migratory cycles); temporal and spatial dimensions of the 44 niche include various environmental influences on distribution. Description of these different 45 dimensions allows estimation of inter-specific overlap *versus* segregation within a community 46 or a guild (Pianka 1974). The first mechanism that allows for niche differentiation is resource 47 48 partitioning (trophic dimension). In this case, different predators living in the same habitat exploit the resources differently, minimizing the overlap in terms of prey composition 49 (Ballance 2002). The second mechanism involves the spatial and/or temporal dimensions of 50 51 the availability of different resources (Huisman and Weissing 2001). The study of the ecological niche as well as of the segregation processes can help to advance 52 our understanding of ecosystem functioning (Leibold 1995) although this objective presents 53 added difficulties since the ecological niche in marine ecosystems is hard to measure 54 (Newsome et al. 2007). Renewed interest in this concept has arisen due to technological 55 56 advances such as the use of chemical or ecological tracers in marine organisms, e.g. stable isotopes, in particular of carbon (C) and nitrogen (N), analysis of which has become a 57 powerful tool in the study of ecological niches (Jardine et al. 2006; Newsome et al. 2007), 58 notably because δ^{13} C and δ^{15} N provide information on trophic level and foraging area 59 (DeNiro and Epstein 1978; Cabana and Rasmussen 1996; Hobson 1999; Fry 2006). Isotopic 60 results are especially relevant in combination with others approaches such as stomach content 61 analysis, habitat modeling or other potential ecological tracers (e.g. trace elements, persistent 62 organic pollutants or fatty acids) to investigated the ecological niches of marine top predators 63

(e.g. Zhao et al. 2004; Herman et al. 2005; Krahn et al. 2007; Cherel et al. 2008; Kiszka et al. 64 2011). 65 Together with stable isotopes, ecological and chemical tracers' elements offer an integrated 66 measure of prey assimilation over different periods of time. This contrast with conventional 67 dietary analyses (e.g. faeces or stomach contents) which provide information on food ingested 68 over a very short-time scale (several days, see e.g. Santos et al. 2001). Depending on the 69 combination of elements and tissues used, the period of integration represented by the specific 70 signature or concentration will differ, relating biological half-life of tracers and tissue 71 turnover rates. Thus, ecological tracers can provide information from a medium-time scale 72 73 (from several weeks to months, i.e. fatty acids in blubber or stable isotopes in liver and muscle), to long-term time scales (from several years to the life span of the animals) when 74 ecological tracers are analyzed in hard tissues with no turnover such as teeth (Hobson and 75 76 Clark 1992; Hobson and Sease 1998; Nordstrom et al. 2008, Caurant et al. 2009). Moreover, ecological tracers can be viewed as proxies of habitat, feeding habits and trophic 77 position. Thus the relative abundance of heavy to light carbon isotopes (δ^{13} C) has been used 78 to discriminate between habitats where phytoplankton is the only source of organic carbon 79 (pelagic and offshore habitats) and those where macrophytes are a significant source of 80 organic carbon (vegetated inshore and benthic habitats) (DeNiro and Epstein 1978; Fry 2006). 81 The relative abundance of heavy to light nitrogen isotopes (δ^{15} N) is commonly taken as an 82 indicator of trophic level (Cabana and Rasmussen 1996) because $\delta^{15}N$ increases from prey to 83 predator due to the preferential excretion of the light isotope (Hobson 1999). However, $\delta^{15}N$ 84 also vary strongly by habitat: inshore and offshore systems, latitudes and between oceanic 85 basins. Thus, Ruiz-Cooley et al. (2012) recently showed that the $\delta^{15}N$ values from 86 zooplankton and/or prey such as squid from similar sizes exhibited large differences in $\delta^{15}N$ 87 (> 4 %) between inshore and offshore systems due to biochemical properties of the habitat. In 88

the same way, Chouvelon et al. (2012) showed marked decreasing $\delta^{15}N$ values from inshore to offshore into different marine species from the Bay of Biscay, highlighting the role of stable isotopes of N as an indicator of feeding area. Considering trace elements, elevated concentrations of cadmium (Cd) in marine mammals are usually associated to a predominance of cephalopods or crustaceans in the diet (Honda et al. 1983; Das et al. 2000) due to the bioaccumulation of this element in their digestive gland where most of the detoxification processes take place (Bustamante et al. 1998b) and the highest concentrations of Cd are found in oceanic species (Bustamante et al. 1998a; Lahaye et al. 2005). The high bioavailability of Cd induces a high potential for its trophic transfer to upper level predators such as marine mammals (Bustamante et al. 2002a, 2002b). In addition, cephalopods show large differences in Cd concentrations depending upon their origin (specifically latitude, with highest Cd enrichment found in sub-polar food webs (Bryan 1976; Miramand and Bentley 1992; Bustamante et al. 1998a). Consequently, marine top predators from polar and sub-polar areas feeding on cephalopods should exhibit high Cd concentrations in their tissues. The Northwest of the Iberian Peninsula (NWIP) is at the northern limit of the East Central Atlantic upwelling system, characterized by high biodiversity and productive fisheries. NWIP represents an important habitat for cetaceans, at least 20 species of marine mammals (16 cetaceans and 4 pinnipeds) have been recorded and the rate of stranding is one of the highest in Europe (López et al. 2002). The most frequent species in the area are toothed whales, specifically common dolphin (Delphinus delphis), bottlenose dolphin (Tursiops truncatus), harbour porpoise (Phocoena phocoena), striped dolphin (Stenella coeruleoalba) and longfinned pilot whale (Globicephala melas) (Cendrero 1993; López et al. 2002, 2004). The ongoing occurrence of cetaceans in the area has permitted, for many years, systematic boatand land-based surveys, as well as several studies on diet, distribution, habitat preferences and

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interactions with human activities including fisheries among others (López et al. 2003, 2004; Santos et al. 2007a, b, In press; Pierce et al. 2010; Spyrakos et al. 2011). Several studies have tried to relate coastal and at-sea sightings with environmental parameters (e.g. depth, SST and chlorophyll-a) to improve understanding of spatio-temporal trends in cetacean distribution in the NWIP, and results suggest the existence of mechanisms allowing a fine-scale partitioning of habitats and/or resources. For instance, stables isotopes of C and N were analyzed in muscle tissues (i.e. medium-time scale information) of these five species and in their prey in order to better understand their position in the NWIP food web (Mèndez-Fernandez et al. 2012). Toothed whales occupy high trophic positions (TPs from 4.3 to 5.3) and no significantly between-species differences have been detected. However, an overlap in TP does not necessarily imply an overlap in diet, since animals may feed on different prey species with similar TPs. Thus, the present study aims to provide further insight into habitat and resource partitioning among these toothed whale species, extending previous stable isotopes of C and N results in muscle (from Mèndez-Fernandez et al. 2012) with new analyses on C and N isotopes in teeth as well as Cd in liver and kidney tissues, to cover a wide range of temporal scales. We hypothesized that each toothed whale species occupies its own ecological niche in the area, defined by differences in at least one of the ecological tracers used. Cd and $\delta^{15}N$ values will determine the trophic dimension of the ecological niche over different time-scales (Bustamante et al. 1998a; Hobson 1999), whereas, information on the spatial dimension (offshore vs inshore, pelagic vs benthic) will be provided by δ^{13} C and also δ^{15} N isotope values (Hansson et al. 1997; Hobson 1999; Sherwood and Rose 2005; Ménard et al. 2007; Chouvelon et al. 2012). An important aspect of the temporal dimension is the possibility that habitat use and trophic position may change over time. Consequently, effects of seasons and

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years were studied on common dolphin, the species for which the biggest sample size was available.

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Methods

Study area and sampling

Fieldwork was carried out in the North West of the Iberian Peninsula (NWIP), from Northern limit of the Galician coast (43° 31' N, 7° 2' W) to Nazaré on the Portuguese coast (39° 36' N, 9° 3' W; Fig. 1). Stranded and by-caught cetaceans were attended by experienced personnel of the Galician (Coordinadora para o Estudo dos Mamiferos Mariños, CEMMA) or Portuguese (Sociedade Portuguesa de Vida Salvagem, SPVS) stranding networks. Animals were identified to species, measured (total length and several other standard measurements are routinely taken), sexed and, if the state of preservation of the carcass allowed it, full necropsies were performed and samples collected following a standard protocol defined by the European Cetacean Society (after Kuiken and Garcia Hartmann 1991). Samples collected between 2004 and 2008 were used for this study, and muscle, teeth, liver and kidney tissues were sampled, whenever possible, from stranded and by-caught individuals of five toothed whales species: common dolphin, bottlenose dolphin, harbour porpoise, striped dolphin and long-finned pilot whale (see Table 1). The common dolphin was the best represented species, as a result of the high number of individuals by-caught in the Spanish and Portuguese fisheries. After the necropsies, all the tissue samples were stored at -20°C until being processed in the laboratory. Since decomposition state can affect results from stable isotope analysis of muscle tissue, individuals selected for analysis were those with condition code 1 to 3, i.e. originally stranded alive, freshly dead or moderately decomposed (after Kuiken and Garcia Hartmann 1991).

Age determination

At least 5 teeth were collected from the mid-section of the left mandible of each sampled individual, selecting the least worn/damaged and least curved teeth, to ensure sufficient material for replicate preparations. Teeth were preserved frozen and age was determined by analysing growth layer groups (GLGs) in the dentine of teeth, following adapted methods based on Lockyer (1993), Hohn and Lockyer (1995) and Rogan et al. (2004). Teeth were decalcified and sectioned using a freezing microtome. The most central and complete sections (including the whole pulp cavity) were selected from each tooth, stained with Mayer's haematoxylin (modified by Grue) and 'blued' in a weak ammonia solution, mounted on glass slides, and allowed to dry. GLGs were counted under a binocular microscope. All readings were initially made blind (without access to individual biological data) and replicate counts were made by at least two readers. If the age estimates obtained by the two readers differed by more than 1 year, readings were repeated. If the increments were difficult to count, both readers discussed the interpretation and either reached an agreed age or judged the tooth to be unreadable.

Stable isotope analyses

Muscle samples were previously analyzed for $\delta^{13}C$ and $\delta^{15}N$ (see Mèndez-Fernandez et al. 2012) and, as for whole tooth samples, they were freeze-dried and then ground into a fine powder before isotopic analyses. Since lipids are highly depleted in ^{13}C relative to other tissue components (DeNiro and Epstein 1978), they were extracted from muscle tissues using cyclohexane. To this end, an aliquot of approximately 100 mg of muscle powder was agitated with 4 mL of cyclohexane for 1h. Next, the sample was centrifuged for 5 min at 4000 g, and the supernatant containing the lipids was discarded. Then, the sample was dried in an oven at $45^{\circ}C$ for 48 h, and 0.35 ± 0.05 mg subsamples of lipid-free dry powder were finally weighed in tin capsules for stable isotope analyses. Any sample for $\delta^{13}C$ analysis that may contain biogenic carbonates (i.e. inorganic carbon) must be acidified and carbonates can be removed

in the dry powder leaving only the organic portion of the sample, composed of collagen. 188 Thus, carbonates were removed from powdered whole tooth samples by digesting with 189 approximately 1 mL of a 4 M-hydrochloric acid solution at 45°C for 48 h. The digested 190 contents were taken up in milli-Q ultrapure quality water, homogenized and freeze-dried at 191 -80° C for 2 days. Finally, 1.45 \pm 0.05 mg subsamples of each homogenized carbonate-free 192 dried dental sample were weighed into tin capsules. 193 194 These analyses were performed with an elemental analyzer coupled to an Isoprime (Micromass) continuous-flow isotope-ratio mass spectrometer (CF IR-MS). The results are 195 presented in the usual δ notation relative to Pee Dee belemnite marine fossil limestone for 196 δ^{13} C and atmospheric N₂ for δ^{15} N, in parts per thousand (‰). Replicate measurements of 197 internal laboratory standards (acetanilide) during each run indicate measurements errors < 0.2 198 % for both δ^{13} C and δ^{15} N. 199 200 Trace element analyses The equipment used in the sample processing was cleaned and subsequently decontaminated 201 for 24 h in a solution composed of 35 ml HNO₃ (65%) and 50 mL HCl (36%) for 1 L of Milli-202 Ro quality water. Fresh samples were freeze-dried, ground to powder and then each sample 203 was treated in duplicate. 204 205 For Cd analysis, aliquots of the liver and kidney samples (i.e. the main storage tissues; 0.1-0.3 g) were digested with 6mL 67-70% HNO₃ and 2 mL 34-37% HCl. Acid digestion was 206 performed overnight under ambient temperature before been heated in a microwave 207 (ThermoFisher Scientific Milestone Start D) during 30 min with increasing temperature up to 208 105°C, and 15 min at 105°C (1200 W). After the mineralization process, each sample was 209 diluted to 30 or 50 mL with milli-Q quality water, according to the volume of acid added to 210 the mineralization (3.0 mL or 4.5 mL). Thus, for samples with a weight < 0.1g the mixture 211

was 3mL 67-70% HNO₃/2mL 34-37% HCl and the milli-Q water complement was 25mL.

Elements were analysed using a Varian Vista-Pro ICP-OES and a Thermo Fisher Scientific XSeries 2 ICP-MS. Three control samples (two CRMs and one blank) treated and analysed in the same way as the samples were included in each analytical batch. CRMs were DOLT-4 (dogfish liver; NRCC) and TORT-2 (lobster hepatopancreas; NRCC) and the detection limit was 0.15 μg g⁻¹ dry weight (d.wt). Finally, this protocol was validated by an international intercalibration method (Christopher et al. 2007).

Data treatment

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Cadmium concentration data showed a marked departure from normality which was substantially improved by log transformation. The distribution of δ^{13} C values (at least for model residuals) was normal and no transformation was needed. Although $\delta^{15}N$ values were not exactly normally distributed, transformation did not improve the distribution and untransformed data were therefore used in analysis. To investigate the possible role of confounding factors, notably age and season but also interannual differences, on niche segregation, generalized additive models (GAMs; Hastie and Tibshirani 1990) were used. GAM is a semi-parametric extension of generalized linear modelling (McCullagh and Nelder 1989). GAMs are more flexible; they permit both linear and complex non-linear relationships between species and environmental predictors, as well as combinations of the two within the same model (Wood and Augustin 2002), without the problematic step of postulating response curve shape or parametric response function. GAMs estimate response curves with a non-parametric smoothing function ("smoothers") instead of parametric terms, and as such are said to be data-driven methods. Differences between the sexes were also tested but since there was no effect for any species and tracers we removed this parameter from the final GAMs.

For common dolphin, Gaussian GAMs with an identity link function were fitted to stable

isotope values and Cd concentrations (log-transformed) (response variables) with age, season

and year as potential explanatory variables. In these GAMs, age was considered as a continuous explanatory variable, while the year and the season of sampling of individuals were treated as categorical explanatory variables. Season was defined as a categorical variable with 4 values: spring (March, April and May), summer (June, July and August), autumn (September, October and November) and winter (December, January and February). The age smoother was constrained to take a relatively simple form, avoiding overfitting, by setting to the number of knots to k=4. Overfitting occurs when the model learns the training data extremely well but is not able to generalize well. To test for significant interactions between age-season and age-year effects, models were fitted both with single age smoothers and with separate age smoothers for each season or year and, in both cases, the two versions were compared using a F test. If the seasonal effect was significant, we also explored using month instead of season as an explanatory variable, fitting it as a smoother, to provide greater detail on the form of the seasonal pattern. The general form of the model performed for each tracer was then:

Tracer value ~ s(age, k = 4) + s(month, k = 4) + year [+ interaction terms]

In principle this approach could be extended to species comparisons but sample sizes for all species except common dolphin were too small to allow fitting a common model (with relevant interaction terms) for all species, testing species differences using models such as the following, taking into account the fact that age effects and seasonal patterns are likely to differ between species (i.e. that there are species-age and species-season interaction):

Tracer value ~ species + s(age,
$$k = 4$$
, by = as.factor(species))

+ s(month, k = 4, by = as.factor(species)) + year [+ other interaction terms]

In practice, because sample sizes were small for most species, full models could not be tested.

However, relationships between tracers and explanatory variables found in common dolphins

dolphin model contained an age effect, this structure was used in the all species model. 263 For each GAM performed, a stepwise procedure was used to select the most parsimonious 264 model using the Akaike Information Criterion (AIC; Akaike 1981). The best model was found 265 using backwards selection (successive eliminations), removing the least important non-266 significant terms sequentially until all remaining effects were significant. Stepwise procedures 267 have been criticized by several authors (e.g. Whittingham et al. 2006), for example because 268 they effectively update the hypothesis tested at each step. However, resulting models can be 269 viewed more as a refinement of the original hypothesis than a hypothesis test per se. Thus all 270 271 such models require tests with independent data sets and we therefore argue that stepwise procedures remain a useful tool to guide future work. The percentage of deviance explained 272 by the final model was calculated as follows: 273 Explained deviance = ((Null model deviance – final model residual deviance)/Null model 274 deviance)*100, with the null model that only contains the intercept terms. 275 Final models were checked for normality and trends in residuals and for outliers and other 276 influential data points (Zuur et al. 2007). The models performed for common dolphin, 277 including respective AIC values, are summarized in Table 2. 278 To provide a quick and easily understandable summary of between species differences in 279 accumulation/assimilation of trace elements and stable isotopes, ANOVA (analysis of 280 variance) with Tukey tests was also performed, specifically to test for differences in Cd 281 concentrations (ug g⁻¹ wet weight, w.wt) and stable isotope values (δ^{13} C and δ^{15} N) (log-282 transformed) between species. 283 Statistical analyses mentioned above were performed using R version 2.11.1 (R Development 284 Core Team 2010) and BRODGAR 2.5.1 statistical software. 285

were used as a guide for the construction of species comparison models, e.g. if the common

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Results

- The number of samples available by tissue, species and year are summarized in Table 1.
- 289 Cadmium concentration in kidney
- 290 Concentrations of Cd were higher in the kidney than in the liver for all species; ranges and
- values (mean \pm SD) are summarized in Table 3.
- 292 ANOVA confirmed that Cd concentrations (µg g⁻¹ w.wt, log-transformed) in kidney varied
- significantly between species (ANOVA, $F_{(4)} = 5.31$, P < 0.001), being significantly higher in
- striped dolphin than in common dolphin and harbour porpoise and significantly higher in pilot
- 295 whale than in harbour porpoise (Tukey tests). However, no significant differences were found
- between pilot whale and striped dolphin (P > 0.05, Tukey test). The final GAM for Cd
- 297 concentrations (µg g⁻¹ w.wt, log-transformed) in the kidney of common dolphins explained
- 298 37.9% of deviance (N = 96, AIC = 133.6; Table 2) and included only a significant effect of
- age (P < 0.0001, d.f. = 2.81). Cd concentrations increased with increasing age, although
- leveling off after around 10 years (Fig. 2a).
- A GAM model for all species which included separate age-smoothers for each species
- performed better than one assuming a common age smoother across all species (F test, P <
- 303 0.0001) and explained 47.7% of deviance. The age effect was significant and showed
- different trends among species (Fig. 3), except for bottlenose dolphin (P = 0.060). This was
- probably a result of the low number of samples and high proportion of young (immature)
- animals of this species. Significant between-species variation is confirmed, with pilot whale,
- striped dolphin and common dolphins all showing higher age-corrected accumulation of Cd in
- the kidney than harbour porpoise and striped dolphin having a higher concentration than
- common dolphin (P < 0.0001 in all cases).
- 310 Cadmium concentration in liver

In liver, Cd concentrations also varied significantly between species (ANOVA, $F_{(4)} = 13.85$, 311 P < 0.0001), being significantly higher in striped dolphin than in all the other species except 312 pilot whale, significantly lower for harbour porpoise than in all the other species except 313 bottlenose dolphin and significantly higher in pilot whale than in common dolphin (Tukey 314 tests). The final model for Cd in liver of common dolphin explained 47.4% of deviance (N = 315 95, AIC = 110.3; Table 2). Cd concentrations in liver increased with increasing age (d.f. = 316 2.99, P < 0.0001) up to around age 7 (Fig. 2b). As for Cd in kidney, seasonal and between-317 year differences were non-significant. 318 The final GAM for Cd concentrations in liver for all species included separate age smoothers 319 for each species and explained 52.4% of deviance (N = 143, AIC = 253.9). The age effects 320 were significant for all the species except for striped dolphin and pilot whale. Again, 321 differences between species were highly significant. Age-corrected concentrations were 322 higher in pilot whale and striped dolphin than in common dolphin or harbour porpoise, and 323 higher in common dolphin than in harbour porpoise (P < 0.0001 in all cases). 324 δ^{13} C values in muscle and teeth 325 A summary of δ^{13} C and δ^{15} N values in the different tissues (muscle and tooth) of the five 326 toothed whales from the NWIP is presented in Table 3. Results for muscle are based on data 327 from Mèndez-Fernandez et al. (2012) 328 δ^{13} C values (‰) in muscle and teeth varied significantly between species, as shown by 329 ANOVA analysis (ANOVA, $F_{(4)} = 11.1$ and 10.91, respectively, P < 0.0001). Striped dolphin 330

showed significantly lower $\delta^{13}C$ values in both muscle and teeth (Tukey tests) compared to all other species, except for common dolphin in teeth. Common dolphin also showed significantly lower $\delta^{13}C$ values than harbour porpoise in both muscle and teeth, and lower $\delta^{13}C$ values than pilot whale and bottlenose dolphin in teeth (Tukey tests) (Fig. 4).

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The final GAM for δ^{13} C values (‰) in the muscle of common dolphin explained 34.9% of

deviance (N = 104, AIC = 119.8; Table 2). There was no effect of age but the effect of season

was highly significant (P < 0.0001, d.f. = 2.3), with the highest values in April and the lowest

in September (Fig. 5a).

- GAM results for δ^{13} C in muscle of all species confirmed clear differences between species
- and that the effect of season also differed between species, with the seasonal trends for
- common dolphin, harbour porpoise and bottlenose dolphin all being statistically significant
- 342 (Figs. 5a-c). The model explained 45.7% of deviance. Seasonally adjusted δ¹³C in muscle was
- 343 higher in pilot whale, porpoise and bottlenose dolphin than in common or striped dolphin, and
- higher in common dolphin than in striped dolphin. The GAM for δ^{13} C in teeth gave similar
- results, although deviance explained was only 26.5% (Table 2).
- δ^{15} N values in muscle and teeth
- 347 ANOVA confirmed that $\delta^{15}N$ values (‰) in muscle and teeth also varied significantly
- between species (ANOVA, $F_{(4)} = 25.92$ and 40.61, respectively, P < 0.0001), being
- significantly lower in striped dolphin than in all the other species, except in teeth of pilot
- whale, and significantly higher in teeth of harbour porpoise than in teeth of all the other
- species (Tukey tests). Finally, $\delta^{15}N$ values in muscle of common dolphin were significantly
- lower than in muscle of bottlenose dolphin (Tukey tests) (Fig. 4).
- The final GAM for δ^{15} N values in muscle of common dolphin explained 19.7% of deviance
- (N = 97, AIC = 151.9; Table 2). There was no effect of season or year but a strong negative
- and linear effect of age (P < 0.001, d.f. = 1.0) where $\delta^{15}N$ values decreased with increasing
- 356 age (Fig. 6).
- 357 GAM results for the all-species analysis of $\delta^{15}N$ in muscle indicated that a model with
- separate age-smoothers for each species was not a significant improvement on a model with a
- 359 common smoother for age. Although there were some influential outliers ("hat" values

approaching 1.0), using separate smoothers and/or transformation of $\delta^{15}N$ data produced no marked improvement in fit. The final model therefore contained a significant age effect (P < 0.0001, d.f. = 2.8), depicting a decline in $\delta^{15}N$, at least until age 7 and also a significant species effect (P < 0.0001). It explained 52.3% of deviance. Harbour porpoise, pilot whale and bottlenose dolphin all had higher (age-corrected) $\delta^{15}N$ values than common dolphin and striped dolphin, while values for common dolphin were also higher than for striped dolphin. Finally, for $\delta^{15}N$ in teeth of all species, the best model included separate age effects for each species (with these being significant for all species except the pilot whale) and a species effect (P < 0.0001), explaining 77.3% of deviance. Although there were some influential data points, the distribution of residuals appeared to be satisfactory. $\delta^{15}N$ values were higher in porpoise than in pilot whales and common dolphins, while all three of these species had higher values than striped dolphin.

Discussion

General comments

We have shown how the application of a multi-tracer approach has allowed the evaluation of the foraging niche of the five most common toothed whales in the NWIP. This approach has permitted the investigation of the foraging niche in two dimensions (i.e. trophic and spatial) covering a range of different time spans. Moreover, this approach also provided complementary information to the studies using conventional methods previously carried out in the area (e.g. based on stomach contents analysis or direct observations). Overall, the main finding in this work is that none of the individual ecological tracers examined alone reveals complete ecological segregation amongst the five species studied whereas the integration of results from all these tracers indicates segregation of all five species in the long-term (conceptually represented in Fig. 7). The NWIP represents only a part of the range of these

species within the NE Atlantic and each individual may not spend a substantial proportion of its time within this area. This is why a possible segregation found by tracers may attest to a different distribution of the species on a broader geographic scale out of the NWIP. A further and important consideration is that all the ecological tracers are mainly transmitted *via* food with only differences in tracer signatures are really informative (similar signatures could arise from different combinations of different prey).

Identifying the limitations of the study is necessary to delineate its range of validity. Sampling from stranded individuals does not often allow examination of inter-annual or seasonal

consecutive years are available. However, and despite the potential biases of the sampling

variation since it is very unlikely that sufficient individuals covering all seasons over several

regime, the stranded and by-caught animals used in this study reflects the ecology of the

different species in the study area and provide evidence of their distinct ecological niches.

Resource partitioning inferred from Cd concentrations

It is well-known that diet, size, weight, longevity and metabolic rates (i.e. ingestions and assimilation) have an influence on Cd bioaccumulation and variations of concentrations among species (Caurant et al. 1994; Aguilar et al. 1999; Das et al. 2003a). Thus, as shown (Figs. 2 and 3), Cd is a metal which accumulates in cetacean tissues, but with different accumulation rates according to the species. In common dolphin, the species for which we had the largest sample size, concentrations increased with age up to around 10 years old, after which concentrations reached a plateau (Fig. 2a). Similar results are well-documented in the literature for several marine mammal species (Caurant et al. 1994; Bustamante et al. 2004; Lahaye et al. 2006) and this pattern seems to be the result of an equilibrium between absorption and excretion in the animals and not due to a shift in diet with time. These results emphasize the need to take age into account when carrying out comparison between species.

Differences in Cd concentrations between areas have already been reported in several marine mammal species. As an example, in common dolphin from the Bay of Biscay, the consumption of cephalopods appeared to be a major route of exposure to Cd and the use of Cd as a tracer confirmed the existence of dietary segregation between oceanic and neritic common dolphins over a long time scale (Lahaye et al. 2005). In the present study, Cd concentrations in both liver and kidney differentiated pilot whale and striped dolphin from the other three species (Table 3). High Cd concentrations in tissues of both species are consistent with their teuthophagous feeding habits. In the NWIP, pilot whales mainly feed on neritic and/or benthic cephalopods such as common and curled octopuses Octopus vulgaris and Eledone cirrhosa (González et al. 1994; Pierce et al. 2007; Santos et al. In press). However, striped dolphin feeds on both cephalopods (mainly Gonatus sp. and ommastrephids) and fish (including blue whiting *Micromesistius poutassou*, scads *Trachurus trachurus*, sand smelt Atherina presbyter and silvery lightfish Maurolicus muelleri, Santos et al. 2007b) as well as on crustaceans (specifically pelagic shrimp species, Jose Cedeira pers comm). Therefore, their fish consumption could explain their lower Cd concentrations (Table 3) in comparison with pilot whales. Conversely, bottlenose dolphins and harbour porpoises are mainly fish feeders (Santos et al. 2007a, b) and in this area, harbour porpoise is mainly a coastal species (López et al. 2002; Pierce et al. 2010). This can explain the significantly lower Cd concentrations found in harbour porpoise compared to the main teuthophagous species over a medium and long timescale, and the absence of differences between porpoises and bottlenose dolphin at any timescale. This last result suggests that trophic competition can occur between these two species as reported in adjacent areas of the Northeast Atlantic by Spitz et al. (2006). However, consumption of different prey species, revealed by stomach contents analyses carried out on

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stranded Iberian Peninsula porpoises and bottlenose dolphins (Santos et al. 2001, 2007a, b), 433 would mitigate the competition in this area (discussed below). 434 The low hepatic and renal Cd concentrations recorded for common dolphin suggests a lower 435 proportion of cephalopods in its diet than expected (Evans 1994; Santos et al. In press; Table 436 3). Significant differences were found between common dolphin and pilot whale, over a 437 medium time-scale (hepatic concentrations), and with striped dolphin, over both medium 438 (hepatic) and long (renal) time-scales, indicating that segregation occur among these species 439 even though common dolphin also frequents oceanic habitats. 440 Spatial and trophic segregation inferred from stable isotopes 441 Results on δ^{13} C values obtained in the present study are generally in agreement with the 442 habitat preferences inferred from direct observations and/or from habitat modeling carried out 443 in the study area for some of the study species (López et al. 2004; Pierce et al. 2010; Spyrakos 444 445 et al. 2011). The δ^{13} C values in the tissue analyzed (i.e. muscle and teeth) allowed a good differentiation of 446 the most oceanic from the most coastal species. Thus, the significantly lower δ^{13} C values 447 exhibited in striped and common dolphins compared to the other species confirm, over 448 medium and long time-scales, the mostly oceanic distribution observed for both species in this 449 area. Striped dolphin is most frequently seen over depths of 100-500 m along both the western 450 and northern coasts (López et al. 2002; Spyrakos et al. 2011). Common dolphin is the most 451 abundant cetacean in the NWIP (Aguilar 1997; López et al. 2004) and mainly found in waters 452 deeper than 200m depth (Spyrakos et al. 2011). Despite this, common dolphin is the second 453 most frequently sighted species from the coast, albeit seen most often in areas where the shelf 454 is narrower (Pierce et al. 2010). This characteristic would explain their low and intermediate 455 δ^{13} C values, between those of striped dolphin and the other toothed whales studied (Fig. 4). 456 This information could suggest that common dolphins move following pelagic schooling fish 457

in the area (such as sardine and blue whiting; Moura et al. 2012; Santos et al. In press) and, as a consequence, seasonal overlap and/or competition or habitat with striped dolphins may occur. Based on δ^{13} C values, bottlenose dolphin, harbour porpoise and pilot whale did not show significant differences from each other over medium and long time-scales but showed significantly high δ^{13} C values than common and striped dolphins (Fig. 4). Coastal sightings suggest that bottlenose dolphin is associated with coastal areas where the continental shelf is relatively wide and productivity relatively high (Pierce et al. 2010) but this species is also seen in offshore waters (López et al. 2004). Recent studies revealed two ecologically and genetically distinct groups of bottlenose dolphins among strandings in Galicia (northern part of the NWIP), which is consistent with the existence of both inshore and offshore groups of bottlenose dolphin (Fernández et al. 2011a, b). Thus the high variability obtained in our isotopic δ^{13} C values for bottlenose dolphin may be due to the presence of some offshore individuals in our sampling (Fig. 4). Again based on coastal sightings, harbour porpoise tend to be seen where the shelf is narrower while boat-based sightings studies suggest association with water depths of 100-200m (Spyrakos et al. 2011). Although the pilot whale is essentially known as oceanic and deep water feeder, it shows some degree of dietary plasticity and can apparently forage successfully in both oceanic and neritic habitats (Spitz et al. 2011). This fact, together with the age/maturity of sampled animals, might explain the high muscular and dental δ^{13} C values obtained in this study. Nitrogen isotopic signatures are the main source of information on feeding habitat and on resource utilization and trophic positions (TP) of the marine mammals in the food web. Here we used the TP obtained in Mèndez-Fernandez et al. (2012) complementing $\delta^{15}N$ values obtained from hard tissues, i.e. long-term information, to compare species showing overlap in

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 δ^{13} C values and/or Cd concentrations, i.e. living in the same habitat and feeding in the same 482 483 type of prey. The age effect on muscular $\delta^{15}N$ values was previously tested for common dolphin (Fig. 6) 484 and showed a negative relationship. Several studies found the same negative effect of age (or 485 body length) on δ^{15} N values. In younger animals, this is consistent with individuals switching 486 from primarily feeding on milk to individuals primarily feeding on living prey (Hobson et al. 487 1997; Das et al. 2003b; Knoff et al. 2008; Fernández et al. 2011a; Meissner et al. 2011). 488 Additionally, it was demonstrated that the consumption of blue whiting, one of the main prey 489 of common dolphin in the Iberian Peninsula, increased with (post-weaning) dolphin body 490 length (Fernández et al. 2011a, Santos et al. In press). Since blue whiting have low δ^{15} N 491 values (Mèndez-Fernandez et al. 2012), the decrease of δ^{15} N values with common dolphin age 492 makes sense (Fig. 6). 493 The comparison between species showed that striped dolphins exhibited the lowest $\delta^{15}N$ 494 values in muscle and teeth, revealing the low TP of this species and its oceanic habitat (Table 495 3). The presence of crustaceans and oceanic fish species (which usually showed low TP in the 496 food web) may be the main reason for their low TP. Therefore, striped dolphin is well 497 segregated from the other four species in the trophic dimension as well as in the spatial 498 dimension (Fig. 4). The higher $\delta^{15}N$ values in common dolphin compared to striped dolphin, 499 over medium and long time-scales, are consistent with its more strongly piscivorous feeding 500 habits as its higher presence in coastal waters. However, these values were still low in 501 comparison with the other toothed whale species (Table 3), probably as the consequence of 502 feeding on fish with a low TP such as blue whiting and sardine Sardina pilchardus (Mèndez-503 Fernandez et al. 2012), the most important prey of common dolphin in Galician waters 504 (Santos et al. In press). According to its $\delta^{15}N$ values, common dolphin is not significantly 505 different from pilot whale and one might conclude that there is a large overlap between their 506

diets, if Cd concentrations (this study) did not contradict this, by confirming the higher occurrence of cephalopods in pilot whale diet (Pierce et al. 2007; Spitz et al. 2011; Santos et al. In press). Furthermore and as discussed above, pilot whales exhibit some degree of dietary plasticity allowing them to forage successfully in both oceanic and neritic habitats (Spitz et al. 2011). This dietary plasticity can also explain their relatively high $\delta^{15}N$ values and the variability obtained, and might result from a distribution which shifts seasonally between oceanic and neritic habitats. In the present study, we have insufficient samples of this species to confirm a seasonal trend in isotopic composition. Data from coastal sightings are also limited but 5 out of 6 sightings of the species during 2003-07 were during the months September to November, which would be consistent with seasonal use of coastal waters (Pierce et al. 2010; unpubl data). Bottlenose dolphin and harbour porpoise exhibited elevated δ¹⁵N values, with harbour porpoise showing significantly higher values in teeth compared to the other species (Table 3), indicating that they feed on prey with a high TP and located at more coastal areas on a longterm basis. Thus, scad, pouting Trisopterus luscus and garfish Belone belone were identified as the most important prey in their diet (Santos et al. 2004, 2007b; Pierce et al. 2010) and these prey also exhibit high δ^{15} N values (Mèndez-Fernandez et al. 2012). Bottlenose dolphin is slightly more generalist and its main prey, in terms of both biomass and number, are blue whiting and hake *Merluccius merluccius* of less than 30 cm length (Mahe et al. 2007; Santos et al. 2007a), which exhibit lower δ^{15} N values than harbour porpoise prey (Mèndez-Fernandez et al. 2012). Time integration of ecological tracers

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Ecological tracers were analyzed in tissues with different rates of turnover and results thus gave us access to different periods of integration. However, sampling did not give us the opportunity to study segregation over the whole year, except for the common dolphin, and the

variations which may occur between seasons. Foraging niche segregation probably depends on the availability of resources in the area. The NWIP is an upwelling area which is characterized by high biological productivity (Fraga 1981) and where primary production, biomass and phytoplankton species composition are influenced by this hydrographic perturbation, which can cause large variations in nutrient availability (González et al. 1979; Tenore et al. 1995). Therefore we expected to find differences in the segregation of these species according to the time of year. Despite the possible seasonal movements demonstrated for common dolphin (Fig. 5a), this species is well-separated from striped dolphin by its δ^{13} C isotopic composition over a medium time-scale, $\delta^{15}N$ isotopic composition and Cd concentrations. Thus, over a long time-scale both species mostly share the same habitat but feed on different resources. The other species for which results were highly influenced by the time period of integration is the harbour porpoise. Only dental $\delta^{15}N$ isotopic composition discriminate harbour porpoise from bottlenose dolphin, the most similar species in terms of ecological niche, which means that although both species can share the same coastal habitat and type of prey, they feed on resources at different TP in the long-term. Such foraging segregation might allow their co-existence in NWIP waters.

Conclusions

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In this work we studied the three main dimensions of the ecological niche of five toothed whale species through the analyses of different ecological tracers in different soft and hard tissues, to test whether there is niche segregation to avoid competition among these five toothed whales which frequent the NWIP waters. Over a medium time-scale period of integration, ecological tracers did not reveal a complete segregation among the five species. Thus, the harbour porpoise and bottlenose dolphin could face competition for habitat in waters off the NW Iberian Peninsula. However, integrating information given by long-term

- ecological tracers showed complete ecological niche segregation among all species (see Fig.
- 7). We hypothesize the following conceptual scheme to sum up our results:
- 1 Harbour porpoise has a coastal foraging niche with the highest trophic position among the
- 559 five species.
- 560 2 Bottlenose dolphin has the most similar foraging niche to harbour porpoises but its lower
- trophic position suggests different targeted prey and/or the use of offshore areas.
- 3 Common dolphin and pilot whale can forage in both oceanic and neritic waters as shown
- by their $\delta^{15}N$ and $\delta^{13}C$ stable isotope ratios but the renal cadmium concentrations revealed a
- dietary difference (fish-based vs cephalopod-based, respectively).
- 4 The striped dolphin showed the most oceanic ecological niche.
- Consequently, the present work provides useful indications on trophic and habitat preferences
- in toothed whales that frequent the NWIP and demonstrates that ecological tracers provide a
- relatively simple way to quantify ecological niche differences over different time-scales,
- especially when looking at closely related species within a common area. However, there is a
- 570 need for a larger sample sizes and a stronger representation of older animals to strengthen
- these conclusions.

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References

592	Aguilar A (1997) Inventario de los cetáceos de las aguas atlánticas peninsulares: aplicación de
593	la directiva 92/43/CEE. Memoria Final del Departamento de Biología Animal,
594	University of Barcelona, Barcelona
595	Aguilar A, Borrell A, Pastor T (1999) Biological factors affecting variability of persistent
596	pollutant levels in cetaceans [special issue]. J Cetac Res Manage 1:83-116
597	Akaike H (1981) Likelihood of a model and information criteria. J econom 16:3-14
598	Ballance LT (2002) Cetacean ecology. In: Perrin WF, Wiirsig B, Thewissen JGM (eds)
599	Encyclopedia of Marine Mammals. Academic Press, San Diego, pp 196–201
600	Bryan GW (1976) Heavy metal contamination in the sea. In: Johnston R (ed) Marine
601	pollution, ch 3. Academic Press, London, pp 185–302
602	Bustamante P, Caurant F, Fowler SW, Miramand P (1998a) Cephalopods as a vector for the
603	transfer of cadmium to top marine predators in the north-east Atlantic Ocean. Sci Total
604	Environ 220:71–80
605	Bustamante P, Cherel Y, Caurant F, Miramand P (1998b) Cadmium, copper and zinc in
606	octopuses from Kerguelen Islands, Southern Indian Ocean. Pol Biol 19:264-271
607	Bustamante P, Teyssié JL, Fowler SW, Cotret O, Danis B, Miramand P, Warnau M (2002a)
608	Biokinetics of zinc and cadmium accumulation and depuration at different stages in
609	the life cycle of the cuttlefish Sepia officinalis. Mar Ecol Progr Ser 231:167–177
610	Bustamante P, Cosson RP, Gallien I, Caurant F, Miramand P (2002b) Cadmium
611	detoxification processes in the digestive gland of cephalopods in relation to
612	accumulated cadmium concentrations. Mar Environ Res 53:227-241
613	Bustamante P, Morales CF, Mikkelsen B, Dam M, Caurant F (2004) Trace element
614	bioaccumulation in grey seals Halichoerus grypus from the Faroe Islands. Mar Ecol
615	Progr Ser 267:291–301

616	Cabana G, Rasmussen JB (1996) Comparison of aquatic food chains using introgen isotopes.
617	Proc Nat Acad Sci USA 93:10844-10847
618	Caurant F, Amiard JC, Amiard-Triquet C, Sauriau PG (1994) Ecological and biological
619	factors controlling the concentrations of trace elements (As, Cd, Cu, Hg, Se, Zn) in
620	delphinids Globicephala melas from the North Atlantic Ocean. Mar Ecol Progr Ser
621	103:207–219
622	Caurant F, Chouvelon T, Lahaye V, Méndez-Fernandez P, Rogan E, Spitz J, Ridoux V (2009)
623	The use of ecological tracers for discriminating populations: the case of the short-
624	beaked common dolphin Delphinus delphis in the European Atlantic waters. RIWC
625	Madeira.
626	Cendrero O (1993) Nota sobre los hallazgos de cetáceos en el norte de España. Bol Inst Esp
627	Oceanogr 9:251–255
628	Cherel Y, Le Corre M, Jaquemet S, Ménard F, Richard P, Weimerskirch H (2008) Resource
629	partitioning within a tropical seabird community: new information from stable
630	isotopes. Mar Ecol Prog Ser 366:281–291
631	Chouvelon T, Spitz J, Caurant F, Mèndez-Fernandez P, Chappuis A, Laugier F, Le Goff E,
632	Bustamante P (2012) Revisiting the use of $\delta^{15}N$ in meso-scale studies of marine food
633	webs by considering spatio-temporal variations in stable isotopic signatures - The case
634	of an open ecosystem: The Bay of Biscay (North-East Atlantic). Progr Oceanogr 101:
635	92–105
636	Christopher SJ, Pugh RS, Ellisor MB, Mackey EA, Spatz RO, Porter BJ, Bealer KJ, Kucklick
637	JR, Rowles TK, Becker PR (2007) Description and results of the NIST/NOAA 2005
638	Interlaboratory Comparison Exercise for Trace Elements in Marine Mammals. Accred
639	Qual Assur 12:175–187

640	Das K, Lepoint G, Loizeau V, Debacker V, Dauby P, Bouquegneau J-M (2000) Tuna and
641	dolphin associations in the North-east Atlantic: Evidence of different ecological niches
642	from stable isotope and heavy metal measurements. Mar Poll Bull 40:102-109
643	Das K, Debacker V, Pillet S, Bouquegneau J-M (2003a) Heavy metals in marine mammals.
644	In: Vos JG, Bossart GD, Fournier M, O'shea TJ (eds) Toxicology of Marine Mammals
645	Volume 3 - Systems. Taylor & Francis Inc, New York, pp 135–167
646	Das K, Lepoint G, Leroy Y, Bouquegneau J-M (2003b) Marine mammals from the southern
647	North Sea: feeding ecology data from $\delta^{13}C$ and $\delta^{15}N$ measurements. Mar Ecol Progr
648	Ser 263:287–298
649	DeNiro MJ, Epstein S (1978) Influence of diet on the distribution of carbon isotopes in
650	animals. Geochim Cosmochim Acta 42:495–506
651	Evans WE (1994) Common dolphin, white-bellied porpoise Delphinus delphis Linnaeus,
652	1758. In: Ridgway S, Harrison R (eds) Handbook of marine mammals, Volume 5.
653	Academic Press, London, pp 191–224
654	Fernández R, García-Tiscar S, Santos MB, López A, Martínez-Cedeira JA, Newton J, Pierce
655	GJ (2011a) Stable isotope analysis in two sympatric populations of bottlenose
656	dolphins Tursiops truncatus: evidence of resource partitioning? Mar Biol 158:1043-
657	1055
658	Fernández R, Santos MB, Pierce GJ, Llavona A, López A, Silva MA, Ferreira M, Carrillo M,
659	Cermeño P, Lens S, Piertney SB (2011b) Fine-scale genetic structure of bottlenose
660	dolphins, Tursiops truncatus, in Atlantic coastal waters of the Iberian Peninsula.
661	Hydrobiologia 670:111–125
662	Fraga F (1981) Upwelling off the Galician Coast, Northwest Spain. In: Richards FA (ed)
663	Coastal Upwelling. American Geophysical Union, Washington DC, pp 176–182
664	Fry B (2006) Stable isotope ecology. Springer, New York

665	González AF, López A, Guerra A, Barreiro A (1994) Diets of marine mammals stranded on
666	the northwestern Spanish Atlantic coast with special reference to Cephalopoda. Fish
667	Res 21:179 –191
668	González N, González JJ, García C, Cabanas JM (1979) Dinámica de nutrientes en las Rías de
669	Arosa y Muros. Bol Inst Esp Oceanogr 1:51-81
670	Hansson S, Hobbie JE, Elmgren R, Larsson U, Fry B, Johansson S (1997) The stable nitrogen
671	isotope ratio as a marker of food-web interactions and fish migration. Ecology 78:
672	2249–2257
673	Hastie TJ, Tibshirani RJ (1990) Generalized additive models. Chapman & Hall, New-York
674	Herman DP, Burrows DG, Wade PR, Durban JW, Matkin CO, LeDuc RG, Barrett-Lennard
675	LG, Krahn, MM (2005) Feeding ecology of eastern North Pacific killer whales
676	Orcinus orca from fatty acid, stable isotope, and organochlorine analyses of blubber
677	biopsies. Mar Ecol Progr Ser 302:275-291
678	Hobson KA (1999) Tracing origins and migration of wildlife using stable isotopes: a review.
679	Oecologia 120:314–326
680	Hobson KA, Clark RG (1992) Assessing avian diets using stable isotopes: I. Turnover of
681	carbon-13. Condor 94:181–188
682	Hobson KA, Sease JL (1998) Stable isotope analyses of tooth annuli reveal temporal dietary
683	records: an example using steller sea lions. Mar Mam Sci 14:116–129
684	Hobson KA, Sease JL, Merrick RL, Piatt JF (1997) Investigating trophic relationships of
685	pinnipeds in Alaska and Washington using stable isotope ratios of nitrogen and
686	carbon. Mar Mam Sci 13:114–132
687	Hohn AA, Lockyer C (1995) Protocol for obtaining age estimates from harbour porpoise
688	teeth. Appendix 3, Report of the harbour porpoise age determination workshop. In:
689	Bjørge A, Donovan GP (eds) Biology of phocoenids. RIWC Cambridge

690	Honda K, Tatsukawa R, Itano K (1983) Heavy metal concentrations in muscle, liver and
691	kidney tissue of striped dolphin, Stenella coeruleoalba, and their variations with body
692	length, weight, age and sex. Agric Biol Chem 47:1219-1228
693	Huisman J, Weissing FJ (2001) Biological conditions for oscillations generated by
694	multispecies oscillations. Ecology 82:2682–2695
695	Hutchinson GE (1957) A Treatise on Limnology. Wiley & Sons, New York
696	Jardine TD, Kidd KA, Fisk AT (2006) Applications, Considerations, and Sources of
697	Uncertainity When Using Stable Stable Isotope Analysis in Ecotoxicology. Env Sci
698	Technol 40:7501–7511
699	Kiszka J, Simon-Bouhet B, Martínez L, Pusineri C, Richard P, Ridoux V (2011) Ecological
700	niche segregation within a community of sympatric dolphins around a tropical island.
701	Mar Ecol Progr Ser 433:273–288
702	Knoff A, Hohn A, Macko S (2008) Ontogenetic diet changes in bottlenose dolphins (Tursiops
703	truncatus) reflected through stable isotopes. Mar Mam Sci 24:128-137
704	Krahn MM, Hanson MB, Baird RW, Boyer RH, Burrows DG, Emmons CK, Ford JKB, Jones
705	LL, Noren DP, Ross PS, Schorr GS, Collier TK (2007) Persistent organic pollutants
706	and stable isotopes in biopsy samples (2004/2006) from Southern Resident killer
707	whales. Mar Pollut Bull 54:1903–1911
708	Kuiken T, Garcia Hartmann M (1991) Proceedings of the first European Cetacean Society
709	workshop on 'Cetacean pathology: dissection techniques and tissue sampling'. ECS
710	Newslett 17:1–39
711	Lahaye V, Bustamante P, Spitz J, Dabin W, Das K, Pierce GJ, Caurant F (2005) Long-term
712	dietary segregation of common dolphins Delphinus delphis in the Bay of Biscay,
713	determined using cadmium as an ecological tracer. Mar Ecol Progr Ser 305:275–285

Lahaye V, Bustamante P, Dabin W, Van Canneyt O, Dhermain F, Cesarini C, Pierce GJ, 714 715 Caurant F (2006) New insights from age determination on toxic element accumulation in striped and bottlenose dolphins from Atlantic and Mediterranean waters. Mar Pollut 716 Bull 52:1219-1230 717 Leibold MA (1995) The niche concept revisited: Mechanistic models and community context. 718 Ecology 76:1371-1382 719 Lockyer C (1993) A report on patterns of deposition of dentine and cement in teeth of pilot 720 whales, genus Globicephala. RIWC Special Issue 14:138–161 721 López A (2003) Estatus dos Pequenos cetaceos da Plataforma de Galicia. Dissertation, 722 723 University of Santiago de compostela, Santiago de compostela López A, Santos MB, Pierce GJ, González AF, Valeiras X, Guerra A (2002) Trends in 724 strandings of cetaceans on the Galician coast, NW Spain, during the 1990s. J Mar Biol 725 Assoc UK 82:513-521 726 López A, Pierce GJ, Valeiras X, Santos MB, Guerra A (2004) Distribution patterns of small 727 cetaceans in Galician waters. J Mar Biol Assoc UK 84:283-294 728 Mahe K, Amara R, Bryckaert T, Kacher M, Brylinski JM (2007) Ontogenetic and spatial 729 variation in the diet of hake (Merluccius merluccius) in the Bay of Biscay and the 730 731 Celtic Sea. ICES J Mar Sci 64:1210–1219 McCullagh P, Nelder JA (1989) Generalized linear models. Chapman & Hall, London 732 Meissner AM, MacLeod CD, Richard P, Ridoux V, Pierce G (2011) Feeding ecology of 733 striped dolphins, Stenella coeruleoalba, in the north-western Mediterranean Sea based 734 on stable isotope analyses. J Mar Biol Assoc UK 735 Ménard F, Lorrain A, Potier M, Marsac F (2007) Isotopic evidence of distinct feeding 736 ecologies and movement patterns in two migratory predators (yellowfin tuna and 737 swordfish) of the western Indian Ocean. Mar Biol 153:141–152 738

739	Mèndez-Fernandez P, Bustamante P, Bode A, Chouvelon T, Ferreira M, López A, Pierce GJ,
740	Santos MB, Spitz J, Vingada JV, Caurant F (2012) Foraging ecology of five toothed
741	whale species in the Northwest Iberian Peninsula, inferred using $\delta^{13}C$ and $\delta^{15}N$
742	isotopic signatures. J Exp Mar Biol Ecol 413:150–158
743	Miramand P, Bentley D (1992) Concentration and distribution of heavy metals in tissues of
744	two cephalopods, Eledone cirrhosa and Sepia officinalis, from the French coast of the
745	English Channel. Mar Biol 114:407–414
746	Moura AE, Sillero N, Rodrigues A (2012) Common dolphin (Delphinus delphis) habitat
747	preferences using data from two platforms of opportunity. Acta Oecol 38:24-32
748	Newsome SD, Martínez del Rio C, Bearhop S, Phillips DL (2007) A niche for isotopic
749	ecology. Front Ecol Environ 5:429–436
750	Nordstrom CA, Wilson LJ, Sara IJ, Tollit DJ (2008) Evaluating quantitative fatty acid
751	signature analysis (QFASA) using harbour seals Phoca vitulina richardsi in captive
752	feeding studies. Mar Ecol Prog Ser 360:245–263
753	Pianka ER (1974) Niche overlap and diffuse competition. Proc Nat Acad Sci USA 71:2141-
754	2145
755	Pierce GJ, Santos MB, Ross HM, Reid RJ, Patterson AIP, López A, Cedeira J (2007) Diets of
756	teuthophagous small cetaceans from the Scottish and Galician coasts. Poster, European
757	Cetacean Society, San Sebastian, Spain
758	Pierce GJ, Caldas M, Cedeira J, Santos MB, Llavona Á, Covelo P, Martínez G, Torres J,
759	Sacau M, López A (2010) Trends in cetacean sightings along the Galician coast,
760	north-west Spain, 2003-2007, and inferences about cetacean habitat preferences. J Mar
761	Biol Assoc UK 90:1547–1560
762	R Development Core Team (2010) R: a language and environment for statistical computing.
763	R Foundation for Statistical Computing, Vienna, http://www.R-project.org

764	Rogan E, Murphy S, Learmonth JA, González A, Dabin W (2004) Age determination in small
765	cetaceans from the NE Atlantic. Appendix 4. In: Pierce GJ, Santos MB, Learmonth
766	JA, Smeenk, C and others (eds) Bioaccumulation of persistent organic pollutants in
767	small cetaceans in European waters: transport pathways and impact on reproduction.
768	Final Report to the European Commission's Directorate General for Research on
769	Project EVK3-2000-00027, Aberdeen
770	Ruiz-Cooley RI, Engelhaupt DT, Ortega-Ortiz JG (2012) Contrasting C and N isotope ratios
771	from sperm whale skin and squid between the Gulf of Mexico and Gulf of California:
772	Effect of habitat. Mar Biol 159:151–164
773	Santos MB, Clarke MR, Pierce DGJ (2001) Assessing the importance of cephalopods in the
774	diets of marine mammals and other top predators: problems and solutions. Fish Res
775	52:121–139
776	Santos MB, Pierce GJ, Learmonth JA, Reid RJ, Ross HM, Patterson IAP, Reid DG, Beare D
777	(2004) Variability in the Diet of Harbor porpoises (Phocoena phocoena) in scottish
778	waters 1992-2003. Mar Mam Sci 20:1-27
779	Santos MB, Fernández R, López A, Martínez JA, Pierce GJ (2007a) Variability in the diet of
780	bottlenose dolphin, Tursiops truncatus, in Galician waters, north-western Spain, 1990-
781	2005. J Mar Biol Assoc UK 87:231–241
782	Santos MB, Pierce GJ, Ross HM, Reid RJ, Patterson AIP, López A, Cedeira J (2007b) Diets
783	of piscivorous small cetaceans from the Scottish and Galician coasts. Poster, European
784	Cetacean Society, San Sebastian, Spain
785	Santos MB, Monteiro SS, Vingada, JV, Ferreira M, López A, Cedeira JAM, Reid RJ,
786	Brownlow A, Pierce GJ (In press) Patterns and trends in diet of long-finned pilot
787	whales (Globicephala melas) based on the analysis of stomachs contents of animals
788	stranded on Northeast Atlantic coasts. Mar Mam Sci

789	Santos MB, Pierce GJ, López A, Martínez JA, Fernández MT, Ieno E, Mente E, Porteiro C
790	Carrera P, Meixide M (In press) Variability in the diet of common dolphins
791	(Delphinus delphis) in Galician waters 1991-2003 and relationship with prey
792	abundance. Mar Ecol Progr Ser
793	Sherwood GD, Rose GA (2005) Stable isotope analysis of some representative fish and
794	invertebrates of the Newfoundland and Labrador continental shelf food web. Estuar
795	Coast Shelf Sci 63:537–549
796	Spitz J, Rousseau Y, Ridoux V (2006) Diet overlap between harbour porpoise and bottlenose
797	dolphin: An argument in favour of interference competition for food? Estuar Coast
798	Shelf Sci 70:259–270
799	Spitz J, Cherel Y, Bertin S, Kiszka J, Dewez A, Ridoux V (2011) Prey preferences among the
800	community of deep-diving odontocetes from the Bay of Biscay, Northeast Atlantic
801	Deep-Sea Res Pt I 58:273–282
802	Spyrakos E, Santos-Diniz TC, Martínez-Iglesias G, Torres-Palenzuela JM, Pierce GJ (2011)
803	Spatiotemporal patterns of marine mammal distribution in coastal waters of Galicia,
804	NW Spain. Hydrobiologia 670:87–109
805	Tenore KR, Alonso-Noval M, Álvarez-Ossorio M, Atkinson LP, Cabanas JM, Cal RM,
806	Campos HJ, Castillejo F, Chesney EJ, González N, Hanson RB, Mcclain CR, Miranda
807	A, Roman MR, Sanchez J, Santiago G, Valdes L, Varela M, Yoder J (1995) Fisheries
808	and oceanography off Galicia, NW Spain: Mesoscale spatial and temporal changes in
809	physical processes and resultant patterns of biological productivity. J Geophys Res
810	100:10943–10966
811	Whittingham MJ, Stephens PA, Bradburry RB, Freckleton RP (2006) Why do we still use
812	stepwise modelling in ecology and behavior. J Anim Ecol 75:1182-1189

813	Wood S, Augustin N (2002) GAMs with integrated model selection using penalized
814	regression splines and applications to environmental modelling. Ecol Model 157:157-
815	177
816	Zhao L, Castellini MA, Mau TL, Trumble SJ (2004) Trophic interactions of Antarctic seals as
817	determined by stable isotope signatures. Polar Biol 27:368–373
818	Zuur AF, Ieno EN, Smith GM (2007) Analysing Ecological Data. Springer, New York
819	

Table 1 Summary of numbers of samples collected per species, year and tissue from cetaceans stranded and by-caught in the North West of the Iberian Peninsula.

Species	Number of samples				
	Muscle	Teeth	Kidney	Liver	
Common dolphin Delphinus delphis	103	98	101	101	
2004	4	3	4	4	
2005	16	17	16	16	
2006	12	12	11	12	
2007	41	39	41	40	
2008	30	27	29	29	
Harbour porpoise <i>Phocoena phocoena</i>	17	15	15	15	
2004	5	5	5	4	
2005	1	1	1	1	
2006	3	3	3	3	
2007	5	3	3	4	
2008	3	3	3	3	
Bottlenose dolphin Tursiops truncatus	8	8	8	8	
2004	1	1	1	1	
2005	2	2	2	2	
2006	3	3	3	3	
2007	2	2	2	2	
Striped dolphin Stenella coeruleoalba	18	20	18	18	
2004	2	1	2	2	
2005	4	4	4	4	
2006	3	3	3	3	

2007	6	9	6	6
2008	3	3	3	3
Long-finned pilot whale Globicephala melas	9	3	9	8
2004	4	0	4	3
2006	1	1	1	1
2007	2	2	2	2
2008	2	0	2	2

Table 2 Results of the GAM models for explaining common dolphin ($Delphinus\ delphis$) tracers of the North West of the Iberian Peninsula. Explanatory variables, deviance explained and Akaike Information Criterion (AIC) value for each model are given. Significance level of variables is indicated by asterixes: *P < 0.05; **P < 0.01; ***P < 0.001; ns: not significance. Final models chosen are highlighted in bold.

Tracer	Variables	Significance	AIC	Deviance explained
Renal Cd	CdK \sim s(Age, k = 4) + s(Month, k = 4) + Year	ns	135.2	39.1%
	CdK \sim s(Age, k = 4) + s(Month, k = 4)	ns	133.9	38.7%
	CdK ~ s(Age, k = 4)	***	133.6	37.9%
Hepatic Cd (CdL)	CdL \sim s(Age, k = 4) + s(Month, k = 4) + Year	ns	114.2	47.5
	CdL \sim s(Age, k = 4) + s(Month, k = 4)	ns	112.2	47.5%
	CdL ~ s(Age, k = 4)	***	110.3	47.4%
δ^{13} C Muscle (δ^{13} CM)	δ^{13} CM ~ s(Age, k = 4) + s(Month, k = 4) + Year	ns	119.8	34.9%
	δ^{13} CM ~ s(Age, k = 4) + s(Month, k = 4)	ns	121.1	32.4%
	δ^{13} CM \sim s(Month, k = 4)	***	122.8	29.8%
δ^{13} C Teeth (δ^{13} CT)	δ^{13} CT ~ s(Age, k = 4) + s(Month, k = 4) + Year	ns	111	26.9%
	δ^{13} CT ~ s(Age, k = 4) + s(Month, k = 4)	ns	109.5	26.5%
	$\delta^{13}CT \sim s(Month, k = 4)$	***	115.8	20%
δ^{15} N Muscle (δ^{15} NM)	δ^{15} NM ~ s(Age, k = 4) + s(Month, k = 4) + Year	ns	151.5	26%
	δ^{15} NM ~ s(Age, k = 4) + s(Month, k = 4)	ns	149.8	25.9%
	δ^{15} NM ~ s(Age, k = 4)	***	151.9	19.7%
δ^{15} N Teeth (δ^{15} NT)	δ^{15} NT ~ s(Age, k = 4) + s(Month, k = 4) + Year	ns	162	52.4%
	δ^{15} NT ~ s(Age, k = 4) + s(Month, k = 4)	ns	163.1	50.8%
	δ^{15} NT ~ s(Age, k = 4)	***	161.3	50.7%

Table 3 Age (mean \pm SD, in years), carbon (δ^{13} C) and nitrogen (δ^{15} N) stable isotope values (mean \pm SD, ‰) and cadmium (Cd) concentrations (mean \pm SD, μg g⁻¹ wet weight, w.wt) in different tissues of the toothed whale species collected along the North West of the Iberian Peninsula. Ranges (minimum, maximum) are presented in parentheses. δ^{13} C and δ^{15} N stable isotope values in muscle tissue are from Mèndez-Fernandez et al. (2012).

Species	Age	δ^{13} C		δ^{15} N		Cd	
		Muscle	Teeth	Muscle	Teeth	Liver	Kidney
Common dolphin	6.1 ± 5.3	-17.0 ± 0.5	-14.3 ± 0.5	11.7 ± 0.6	13.0 ± 0.8	0.4 ± 0.5	2.3 ± 2.7
Delphinus delphis	(0, 20)	(-18.1, -16.0)	(-15.6, -13.4)	(10.1, 13.6)	(11.3, 15.2)	(< 0.01, 2.9)	(< 0.01, 16.4)
Harbour porpoise Phocoena phocoena	5.4 ± 6.1	-16.5 ± 0.7	-13.7 ± 0.7	13.0 ± 1.2	15.5 ± 0.6	0.1 ± 0.1	2.2 ± 5.3
	(0, 18)	(-17.6, -15.1)	(-14.7, -12.4)	(11.5, 17.1)	(14.4, 16.2)	(< 0.01, 0.4)	(0.09, 19.0)
Bottlenose dolphin Tursiops truncatus	4.3 ± 2.0	-16.5 ± 0.8	-13.8 ± 0.8	12.6 ± 0.9	13.7 ± 1.0	1.2 ± 2.8	5.7 ± 13.8
	(0.5, 7)	(-17.3, -15.2)	(-14.8, -12.3)	(11.3, 14.0)	(11.6, 15.1)	(0.01, 8.2)	(0.15, 39.8)
Striped dolphin 4.1	4.1 ± 5.0	-17.6 ± 0.6	-14.6 ± 0.4	10.8 ± 0.7	12.4 ± 0.8	3.9 ± 3.8	10.3 ± 11.0
Stenella coeruleoalba	enella coeruleoalba (0, 15)	(-18.5, -16.5)	(-15.5, -13.8)	(9.8, 12.4)	(10.8, 14.9)	(0.04, 13.2)	(0.1, 40.7)
C I	7.4 ± 5.0	-16.5 ± 0.5	-13.5 ± 0.8	12.2 ± 0.6	13.6 ± 0.5	8.3 ± 8.4	30.0 ± 26.9
	(0, 14)	(-17.3, -15.7)	(-14.4, -13.0)	(11.0, 13.3)	(13.3, 14.3)	(0.01, 22.2)	(< 0.01, 79.3)

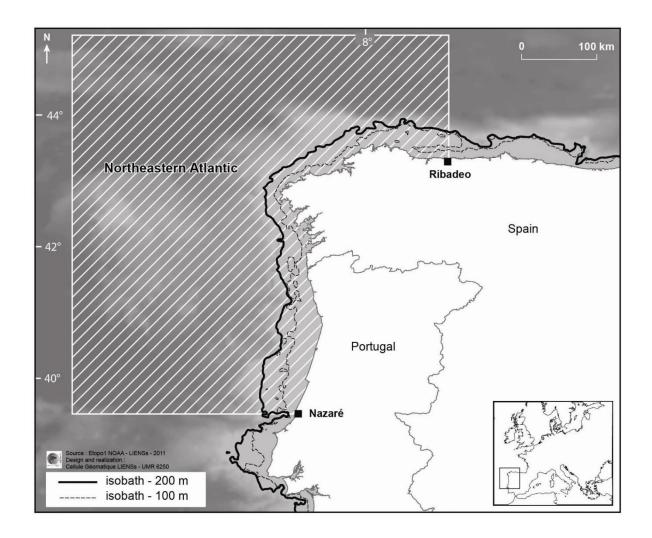


Figure 1 Map of the sampling area with 100 and 200 m isobaths. The 200 m isobaths is taken as the limit for the shelf-break. The sampling area is framed in white, representing the north and south limits for the strandings and the western latitude of the by-catches.

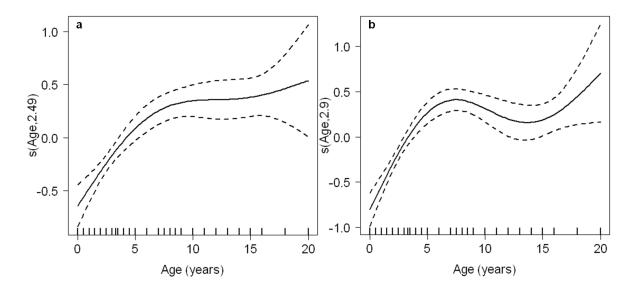


Figure 2 Smoothers for effects of age on Cd concentrations ($\mu g g^{-1} w.wt$) in kidney (a) and in liver (b) of common dolphin *Delphinus delphis*. The *y*-axis shows the contribution of the smoother to the predictor function (in arbitrary units). Smoothers illustrate the partial effect of age, i.e. the effect of age once the effects of all other explanatory variables in the model have been taken into account. Tick marks in *x*-axis represent sampled individuals. Dashed lines represent 95% confidence bands for the smoothers.

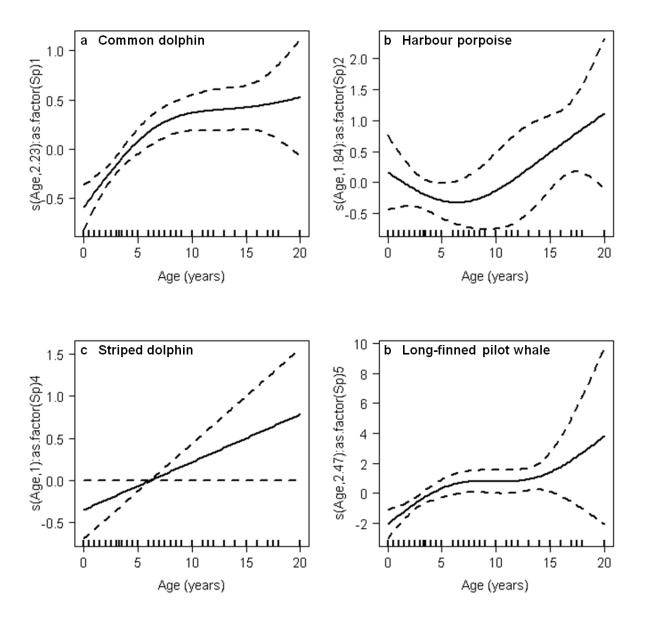


Figure 3 Smoothers for between species differences of age-corrected accumulation of Cd concentrations in kidney (μg g⁻¹ w.wt) of common dolphin *Delphinus delphis* (a) harbour porpoise *Phocoena phocoena* (b) striped dolphin *Stenella coeruleoalba* (c) and pilot whale *Globicephala melas* (d). The *y*-axis shows the contribution of the smoother to the predictor function (in arbitrary units). Smoothers illustrate the partial effect of age, i.e. the effect of age once the effects of all other explanatory variables in the model have been taken into account. Tick marks in *x*-axis represent sampled individuals. Dashed lines represent 95% confidence bands for the smoothers.

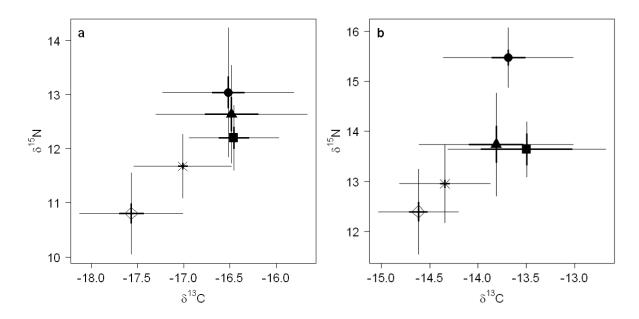


Figure 4 Carbon (δ^{13} C) and nitrogen (δ^{15} N) stable isotope values in muscle tissue (a) and in teeth (b) of common dolphin *Delphinus delphis* (*), harbour porpoise *Phocoena phocoena* (•), bottlenose dolphin *Tursiops truncatus* (\blacktriangle), Striped dolphin *Stenella coeruleoalba* (\diamond) and long-finned pilot whale *Globicephala melas* (\blacksquare). Data are mean (∞) \pm SD and SEM (bold). δ^{13} C and δ^{15} N stable isotope values in muscle from Mèndez-Fernandez et al. (2012).

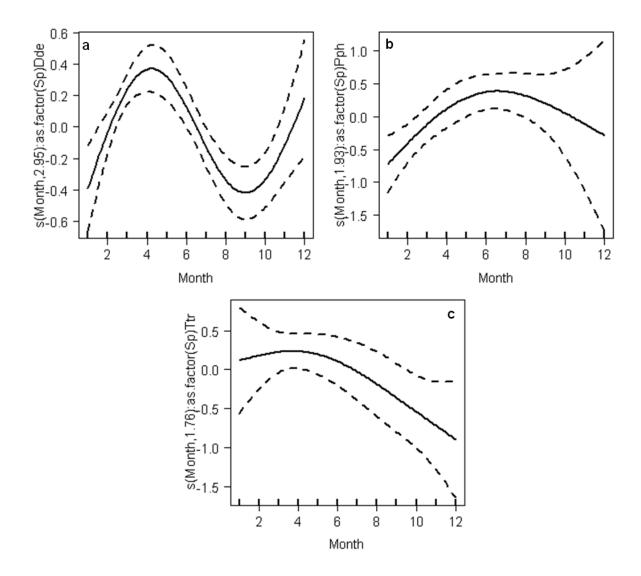


Figure 5 Smoothers for between species differences of season-adjusted effects on δ^{13} C values in muscle of common dolphin *Delphinus delphis* (a) harbour porpoise *Phocoena phocoena* (b) and bottlenose dolphin *Tursiops truncatus* (c). The *y*-axis shows the contribution of the smoother to the predictor function (in arbitrary units). Smoothers illustrate the partial effect of season once the effects of all other explanatory variables in the model have been taken into account. Tick marks in *x*-axis represent sampled individuals. Dashed lines represent 95% confidence bands for the smoothers.

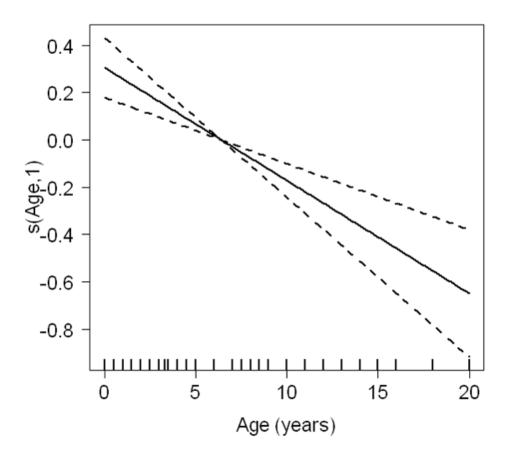


Figure 6 Smoothers for effects of age on $\delta^{15}N$ values in muscle of common dolphin *Delphinus delphis*. The y-axis shows the contribution of the smoother to the predictor function (in arbitrary units). Smoothers illustrate the partial effect of age once the effects of all other explanatory variables in the model have been taken into account. Tick marks in x-axis represent sampled individuals. Dashed lines represent 95% confidence bands for the smoothers.

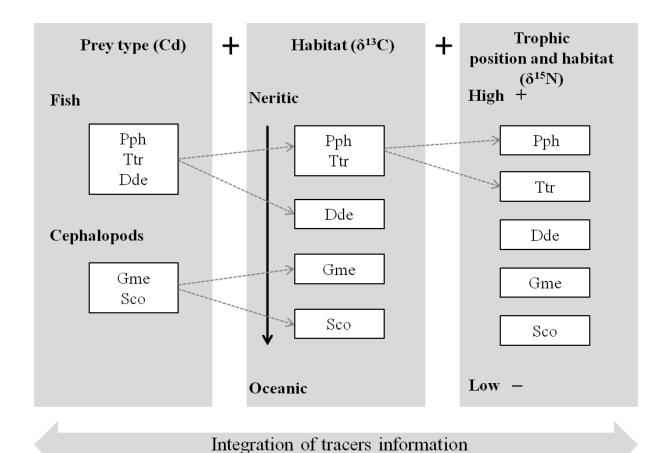


Figure 7 Conceptual scheme of foraging niche segregations obtained by the sum of the information given by ecological tracers' in the long-term (i.e. several years) and in the five toothed whale species. Dde: common dolphin *Delphinus delphis*, Pph: harbour porpoise *Phocoena phocoena*, Ttr: bottlenose dolphin *Tursiops truncatus*, Sco: striped dolphin *Stenella coeruleoalba* and Gme: pilot whale *Globicephala melas*. Ecological tracer values of species lumped together were no significantly different (Tukey test, P > 0.05).