DrosOmics: A Browser to Explore -omics Variation Across High-Quality Reference Genomes From Natural Populations of *Drosophila melanogaster*

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Abstract

The advent of long-read sequencing technologies has allowed the generation of multiple high-quality de novo genome assemblies for multiple species, including well-known model species such as *Drosophila melanogaster*. Genome assemblies for multiple individuals of the same species are key to discover the genetic diversity present in natural populations, especially the one generated by transposable elements, the most common type of structural variant. Despite the availability of multiple genomic data sets for *D. melanogaster* populations, we lack an efficient visual tool to display different genome assemblies simultaneously. In this work, we present DrosOmics, a population genomic-oriented browser currently containing 52 high-quality reference genomes of *D. melanogaster*, including annotations from a highly reliable set of transposable elements, and functional transcriptomics and epigenomics data for 26 genomes. DrosOmics is based on JBrowse 2, a highly scalable platform, which allows the visualization of multiple assemblies at once, key to unraveling structural and functional features of *D. melanogaster* natural populations. DrosOmics is an open access browser and is freely available at http://gonzalezlab.eu/drosomics.

Key words: Drosophila, transposable elements, epigenomics, transcriptomics, histone marks, ATAC-seq.

Introduction

The advent of technological developments in DNA sequencing has led to a high number of whole-genome sequences for multiple individuals of the same species, revolutionizing the field of population genomics (Mitsuhashi and Matsumoto 2020; Sakamoto et al. 2020). Among these developments, long-read sequencing technologies allow to improve the quality and completeness of reference genomes because of their ability to span repetitive regions, leading to a better detection and annotation of structural variants such as transposable elements (TEs; Solares et al. 2018; Du and Liang 2019; Miga et al. 2020; Rech et al. 2022). *Drosophila melanogaster*, one of the central model species for population genomics studies (Casillas and Barbadilla 2017) and for studying TEs (Mccullers and Steiniger 2017), has benefited from these technological developments and currently has several de novo reference genomes representative of natural populations sampled worldwide (Long et al. 2018; Chakraborty et al. 2019; Rech et al. 2022).

Complementary to these population genome sequence data, there has been a large increase in the generation of other functional -omics data, such as transcriptomics and epigenomics. While the genome sequence represents a static component of the organism, functional -omics data vary across different body parts or tissues (spatial), life stages (temporal), and conditions (e.g., different environments and experimental treatments). Capturing this information is relevant for understanding, for example, how organisms adapt to changing environments, plant and animal breeding, and medical genetics.

One challenge faced in genomic studies is to visualize and retrieve large amounts of genome and functional -omics data, to, for example, perform a manual inspection of regions of interest to validate the presence of a genetic variant, or to retrieve the sequence of a particular genomic region. For that, web-based genome browsers are a suitable platform that allows browsing, visualizing, and retrieving data through an efficient and user-friendly graphical interface (Schattner 2008; Buels et al. 2016). Current population genomics-oriented browsers are based on the first available JBrowse version, which allows only a single genome to be displayed. These browsers, for example, PopHuman (https://pophuman.uab.cat; Casillas et al. 2018), were designed for visualizing and querying population genetic statistics for each of the populations included. For *D. melanogaster*, there are two such population genomics-oriented browsers available: PopFly (https://popfly.uab.cat; Hervas et al. 2017) and DEST (https://dest.bio; Kapun et al. 2021). However, until recently, JBrowse-based browsers precluded a comparative analysis of sequence and structural variants across genomes.

Open Access

New Approaches

In this work, we present DrosOmics (http://gonzalezlab.eu/drosomics), a population genomics-oriented browser based on the new version of JBrowse, JBrowse 2 (Diesh et al. 2022). DrosOmics allows browsing, visualizing, and retrieving genome sequences and functional -omics data for 52 genomes of *D. melanogaster* collected in 32 locations worldwide (table 1). Based on JBrowse 2, DrosOmics allows one to explore the sequence and structural variation of several populations simultaneously and to quickly navigate through the different genome views. The genomes included in DrosOmics have been annotated with a high-quality and manually curated library of TEs, which includes the sequence variability present in several natural populations (Rech et al. 2022). DrosOmics also compiles functional annotations for 26 of these genomes, including transcriptomics (RNA-seq) and epigenomic data sets (ChIP-seq and ATAC-seq), some of them provided here for the first time (table 2). This allows genetic variants, for example, single-nucleotide polymorphisms (SNPs) and structural variants, such as TEs, to be associated with variation in gene expression and epigenetic modifications. JBrowse 2-based browsers are highly scalable, and we provide guidelines to the research community on how to contribute to DrosOmics (http://gonzalezlab.eu/drosomics/contribution.html).

DrosOmics Browser Content Overview

DrosOmics compiles 52 high-quality genome assemblies that have been obtained using long-read sequencing technologies (table 1 and supplementary table S1, Supplementary Material online).

DrosOmics contains gene and TE annotations for the aforementioned 52 *D. melanogaster* reference genomes. Note that the current version of DrosOmics provides gene annotations that were transferred to each genome from the reference release 6 of *D. melanogaster* (Larkin et al. 2021; see supplementary file 1, Supplementary Material online). Functional -omics data for 26 genomes were retrieved from previous research (Everett et al. 2020; Salces-Ortiz et al. 2020; Green et al. 2022; Horvath et al. 2023; table 2 and supplementary table S1, Supplementary Material online) or are reported in this work for the first time (table 2, supplementary table S2, and supplementary file 1, Supplementary Material online Genome and functional annotations can be graphically displayed along the chromosome arms for multiple genomes simultaneously (table 2). DrosOmics is built on JBrowse 2 (release 1.6.9; Diesh et al. 2022) and is currently running under a Docker container using Apache on a CentOS 7.9.2009 Linux x64 server with 4 vCores processors and 16 Gb RAM.

DrosOmics has a Help section that includes documentation about all the data compiled in the browser, such as detailed data descriptions, data sources, and citations, as well as details about the browser tracks available. Importantly, it also provides an exhaustive tutorial with step-by-step worked examples introducing the usage of the browser. All the compiled data and support resources provided by DrosOmics are open access and available at http://gonzalezlab.eu/drosomics.

Table 1. The 52 *Drosophila melanogaster* Reference Genome Assemblies Compiled in DrosOmics.

Genomes	Source Locations	Data Source
32 genomes	12 localities from 8 countries: America (1), Europe (7)	Rech et al. (2022)
14 genomes	14 localities from 11 countries: Africa (2), Europe (2), North America (1), North Atlantic Ocean (1), South America (2), Asia (3)	Chakraborty et al. (2019)
5 genomes	5 localities from 5 countries: Africa (1), Europe (1), America (1), Asia (1), Oceania (1)	Long et al. (2018)
1 genome (ISO-1)	America	Larkin et al. (2021)

Table 2. Available Tracks (>500) in DrosOmics.

Track Type	Track Name	Description	Genomes	Data Sets
Genome sequence	Reference sequence	Displayed by the chromosome arm (2R, 2L, 3R, 3L, and X)	All	52
Genome annotations	TE annotations	• *De novo* TE annotations	All	104
	Gene annotations	• Gene annotations transferred from *D. melanogaster* release 6 annotation		
RNA-seq	RNA-seq coverage	• Basal conditions: head, gut, ovary	◆ 5 genomes	15 (3 repl)
		• Basal conditions: whole body	■ 8 genomes	8 (2 repl)
		• Basal and malathion treatment: gut	▲ 4 genomes	8 (3 repl)
		• Basal and copper treatment: whole body ♀	● 6 genomes	12 (3 repl)
		• Basal and desiccation treatment: whole body ♀	▶ 6 genomes	12 (3 repl)
ChIP-seq	ChIP-seq FC signal, ChIP-seq peak calling	• H3K9me3 (repressive mark): head, gut, ovary	◆ 5 genomes	15 (3 repl)
		• H3K27ac (active mark): head, gut, ovary	◆ 5 genomes	15 (3 repl)
ATAC-seq	ATAC-seq FC signal, ATAC-seq peak calling	• Basal and malathion treatment whole body ♀	▲ 4 genomes	8 (3 repl)

NOTE.—Further details on raw data number accessions, data processing, and associated publications are available on supplementary file 1 and supplementary table S1, Supplementary Material online. The symbols indicate the publication source for the functional data: ◆ this publication; ■ Everett et al. (2020); ▲ Salces-Ortiz et al. (2020); ● Green et al. (2022); ▶ Horvath et al. (2023). The number of biological replicates (repl) for each data set is given. FC, fold change.
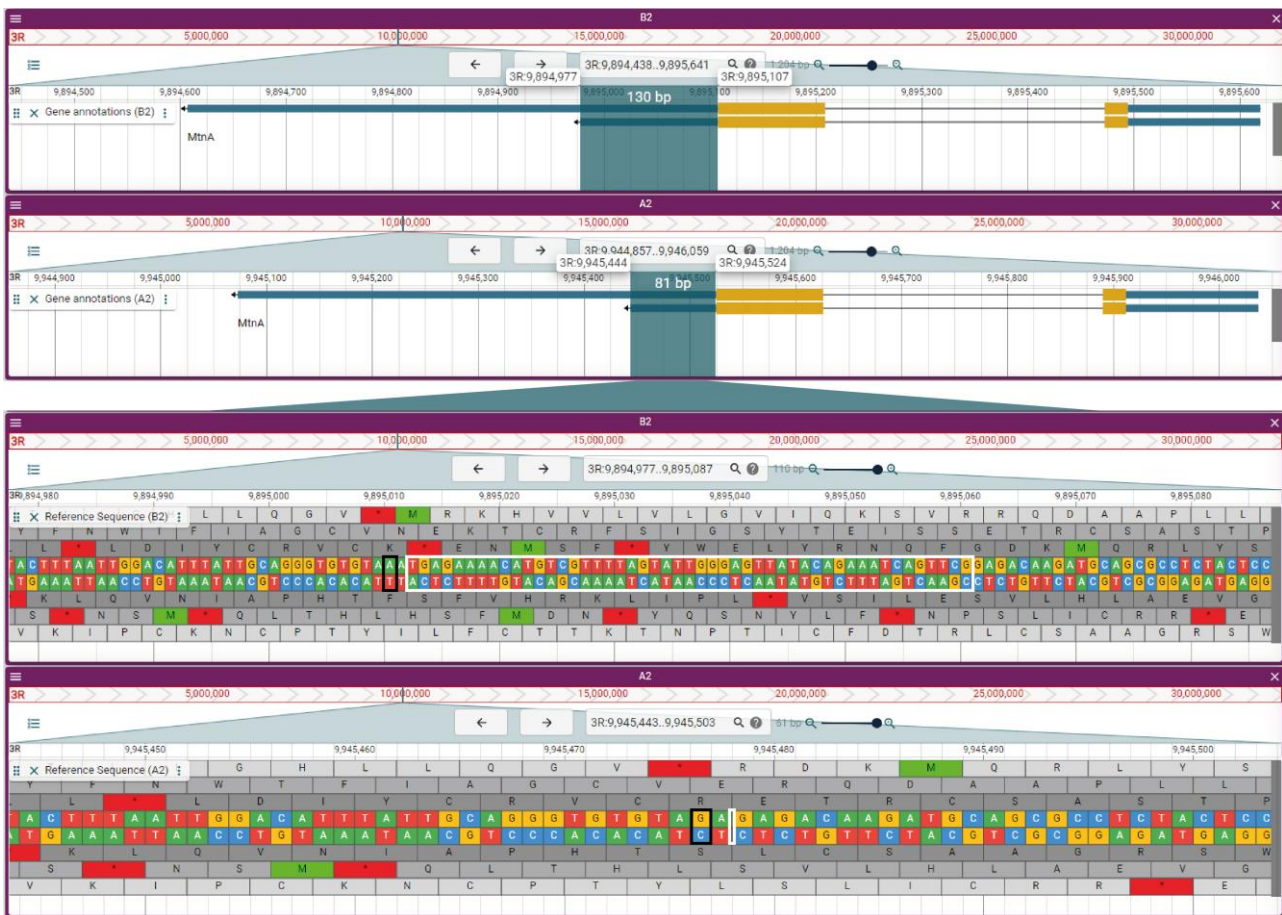
Fig. 1. DrosOmics snapshot showing the *MtnA* gene and its 3′UTR sequence in two genomes. Genome view of the *MtnA* gene from genome *B2* (top; Cape Town [South Africa]; Chakraborty et al. 2019) and genome *A2* (bottom; Bogota [Colombia]; Chakraborty et al. 2019). The transcript model of *MtnA* is displayed with separated features: UTRs (narrow blue rectangles), coding exons (wide golden rectangles), and introns (black lines). By zooming to the sequence level in the 3′UTR of *MtnA* gene, it is possible to visualize the 49-bp indel (highlighted in white) present in *B2* and absent in *A2*, and the linked SNP (highlighted in black) described in Catalán et al. (2016).

DrosOmics Case Studies

We have selected three case studies to demonstrate the usability of the browser: (1) analysis of a region of interest in multiple genomes; (2) analysis of structural variability between genomes; and (3) study of the association between structural, epigenetic, and expression data.

Case Study 1

With DrosOmics, the user is able to visualize and retrieve the sequence of a region of interest in any of the 52 genomes available. This is of special interest for the study of the presence and/or absence of genetic variants in natural populations. One example is the analysis of the presence of a 49-bp indel polymorphism, and a linked SNP, in the 3′UTR region of the *metallothionein A* (*MtnA*) gene in some natural populations. This indel has been associated with higher expression of *MtnA*, linked, in turn, to oxidative stress tolerance (Catalán et al. 2016). With DrosOmics, the user can navigate to the *MtnA* gene region and visualize the sequence

of the 3′UTR from any of the genomes to determine if they carry the indel or not (fig. 1). Additionally, the nucleotide sequence of the selected region of a genome can be retrieved using the "Get sequence" option. This case study exemplifies the utility of the browser in visualizing a sequence from multiple genomes for validation and further characterization.

Case Study 2

One of the main advantages of JBrowse 2 is that it allows the study of structural variation across genomes. To exemplify this, we will focus on the genome region surrounding the *Cyp6a17* gene, which was found to be associated with increased cold preference when knocked down (Kang et al. 2011). Chakraborty et al. (2019) described that *Cyp6a17* was not present in a high proportion of African and North American flies, suggesting an adaptive role of this mutation on temperature regulation in the wild. With DrosOmics, it is possible to identify and visualize genomes with present and absent genotypes. Figure 2 shows four genomes

FIG. 2. DrosOmics snapshot showing the region surrounding *Cyp6a17* gene in four genomes from North American *Drosophila melanogaster*. RAL-176 and RAL-375 (top) have *Cyp6a17* and RAL-059 and RAL-177 (bottom) have the absent genotype. RNA-seq profiles are shown in turquoise.

from North American *D. melanogaster*: two (RAL-176 and RAL-375) have the *Cyp6a17* gene and the other two (RAL-059 and RAL-177) do not. By including RNA-seq coverage tracks, it is possible to visualize the expression differences of the genes in the two genotypes: when present, *Cyp6a17* and its neighboring paralogs – *Cyp6a22* and *Cyp6a23*– exhibit similar expression levels. When *Cyp6a17* is absent, the expression level of *Cyp6a23* is notably higher than that of *Cyp6a22*. With this example, we show that DrosOmics can be a tool to easily explore the structural differences between genomes and their possible functional consequences.

Case Study 3

This example links gene expression changes with the epigenetic effects of a TE. TEs have been associated with gene downregulation due to the spread of repressive histone marks targeting TEs (Sienski et al. 2012; Choi and Lee 2020). Figure 3 shows the region surrounding the

gene *Myo31DF* in two genomes: TOM-007 and JUT-011 (Rech et al. 2022). TOM-007 has the insertion of a TE that is not present in the JUT-011 genome. By activating ChIP-seq tracks for the repressive histone mark H3K9me3 (fold enrichment and histone peaks), one can observe the enrichment and presence of peaks surrounding the TE insertion that are not present when the TE is absent. Because the TE is in the promoter region of *Myo31DF*, this epigenetic effect can have an impact on the gene expression level. Indeed, the RNA-seq levels of the gene with the insertion are lower than when the insertion is absent (z-score $= -25.4$, $P < 0.001$; see supplementary file 1, Supplementary Material online; Coronado-Zamora M, González J, in preparation).

Finally, we would like to highlight that DrosOmics is already being used to facilitate primer design in a protocol to experimentally validate by PCR TE insertions, aimed at high school teachers (Merenciano et al. 2023).
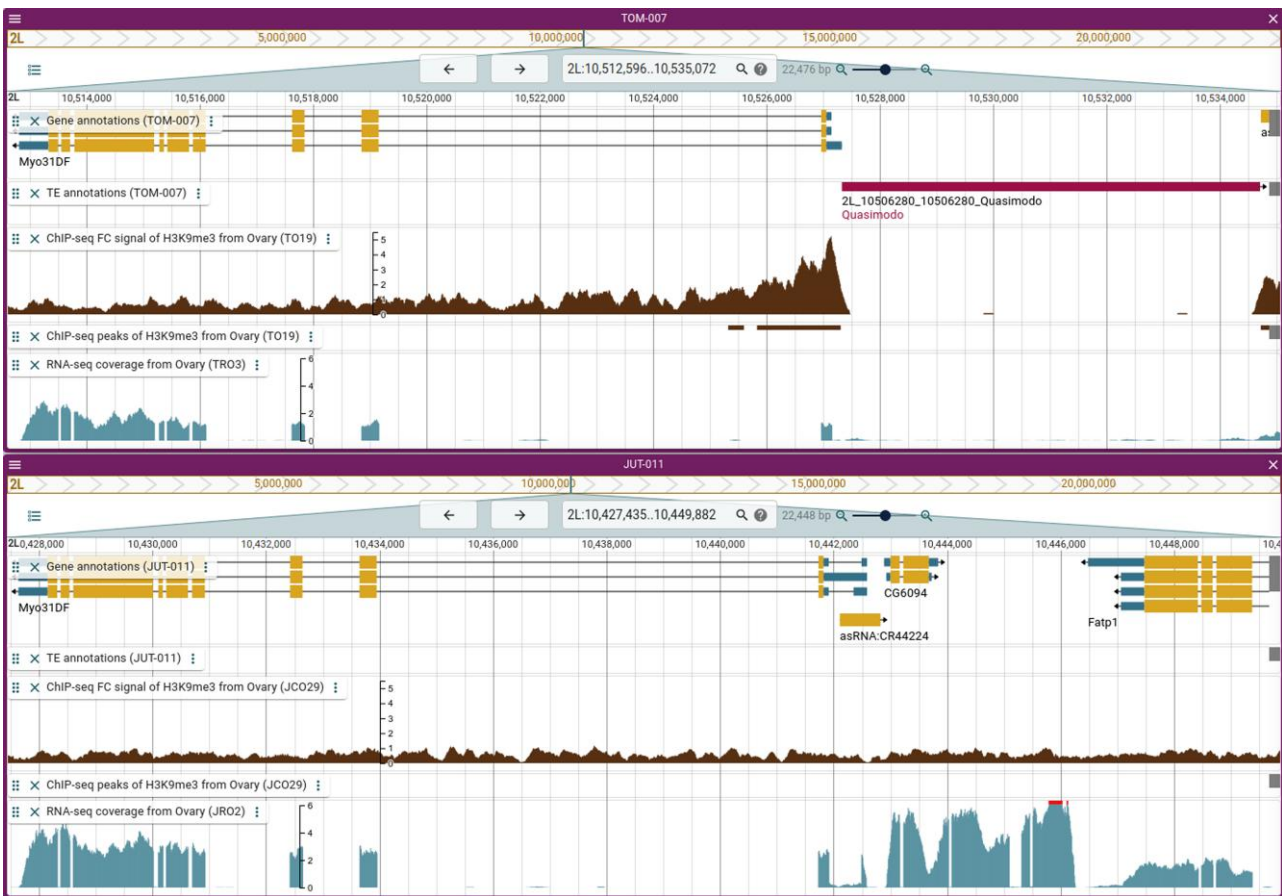
FIG. 3. DrosOmics snapshot showing the region surrounding *Myo31DF* gene in two genomes: TOM-007 (with a TE insertion in the promoter region) and JUT-11 (no TE insertion). The brown track represents the ChIP-seq fold-change signal of H3K9me3 and the turquoise track the RNA-seq coverage levels, as indicated in the corresponding left labels. The presence of the TE induces the enrichment of H3K9me3, affecting the promoter region of *Myo31DF*. This leads to the downregulation of the gene.

Conclusions

The DrosOmics browser, based on JBrowse 2, compiles genome sequences, TEs and gene annotations, as well as functional -omics data for 52 genomes from *D. melanogaster* natural populations. Furthermore, as JBrowse 2 is a highly scalable platform, it allows a continuous update by the addition of new genomes, annotations, and functional data sets. We believe that DrosOmics will facilitate the unraveling of structural and functional features of *D. melanogaster* natural populations, thus providing a great opportunity to explore and test new evolutionary biology hypotheses. DrosOmics is an open access browser and is freely available at http://gonzalezlab.eu/drosomics.

Supplementary Material

Supplementary data are available at *Molecular Biology and Evolution* online.

Acknowledgments

Data Availability

All processed data tracks, annotations, and genomes are available to download from http://gonzalezlab.eu/drosomics. Genome sequences from a particular region can be retrieved directly from the browser using the "Get sequence" dialog available when selecting a genome region. The Docker container and scripts used to create DrosOmics are available at the GitHub repository: https://github.com/

GonzalezLab/DrosOmics. The RNA-seq and ChIP-seq raw data for five genomes that are reported here for the first time are available in the NCBI Sequence Read Archive database under BioProject PRJNA643665.

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