1 Melatonin content of pepper and tomato fruits: effects of cultivar

2 and solar radiation

```
4 Patrick Riga<sup>*1</sup>, Sonia Medina<sup>2</sup>, Libia Alejandra García-Flores<sup>2</sup> and Ángel Gil-
```

- 5 Izquierdo²
- 6 ¹ Department of Plant Production and Protection, NEIKER-Basque Institute of Agricultural Research
- 7 and Development, Parque Tecnológico de Bizkaia P. 812, E-48160 Derio, Spain.
- 8 ² Department of Food Science and Technology, CEBAS-CSIC, P.O. Box 164, E-30100 Espinardo,
- 9 Murcia, Spain.
- 10
- 11 * Corresponding author. Address: Department of Plant Production and Protection,
- 12 NEIKER-Basque Institute of Agricultural Research and Development, Parque
- 13 Tecnológico de Bizkaia P. 812, E-48160 Derio, Spain.
- 14 Phone: +34 94 403 43 00, Fax: +34 94 403 43 10, E-mail: priga@neiker.net
- 15
- 16 **Running title:** Melatonin content of pepper and tomato fruits
- 17
- 18

ABSTRACT 19

20 We evaluated the effect of cultivar and solar radiation on the melatonin content of 21 Capsicum annuum (pepper) and Solanum lycopersicum (tomato) fruits. The melatonin content of red pepper fruits ranged from 31 to 93 ng g⁻¹ (dry weight). The 22 melatonin content of tomato ranged from 7.5 to 250 ng g⁻¹ (dry weight). We also 23 studied the effect of ripeness on melatonin content and identified one group of 24 pepper cultivars in which the melatonin content increased as the fruit ripened and 25 26 another in which it decreased as the fruit ripened. Under shade conditions, the melatonin content in most of tomato cultivars tended to increase (up to 135%), 27 whereas that of most pepper cultivars decreased (to 64%). Overall, the results also 28 demonstrated that the melatonin content of the fruits was not related to carbon fluxes 29 from leaves. 30

- 31
- 32

33	Keywords:	N-acetyl-5-methoxytryptamine,	Capsicum	annuum,	Solanum
34	lycopersicum	, UHPLC-MS/MS.			
35	C				

36 1. Introduction

37

Plants can synthesize melatonin (N-acetyl-5-methoxytryptamine), a methoxylated 38 indolamine (Murch, KrishnaRaj, and Saxena, 2000). Although the physiological 39 40 functions of melatonin (MEL) in plants remain to be definitively established, some 41 functional roles have recently been proposed (Tan, Hardeland, Manchester, 42 Korkmaz, Ma, Rosales-Corral, and Reiter, 2012). Studies have documented that MEL is as a potent free-radical scavenger in plants (Paredes, Korkmaz, Manchester, 43 Tan, and Reiter, 2009; Tan et al., 2012; Wang, Yin, Liang, Li, Ma, and Yue, 2012) 44 45 and in animals (Gitto, Pellegrino, Gitto, Barberi, and Reiter, 2009; Melchiorri, Reiter, Sewerynek, Hara, Chen, and Nistico, 1996). In addition, MEL promotes 46 growth of roots (Arnao and Hernández-Ruiz, 2007; Sarropoulou, Therios, and 47 48 Dimassi-Theriou, 2012) and leaves (Okazaki, Higuchi, Aouini, and Ezura, 2010; Wang, Sun, Li, Wei, Liang, and Ma, 2013), and it may be involved in the regulation 49 of circadian rhythms and photoperiodic reactions (Kolar and Machakova, 2005), 50 among other plant functions. 51

52 The increasing interest in plant MEL expressed by the scientific community is partly due to beneficial effects of MEL on human health. When plants containing 53 54 melatonin are eaten, the melatonin is absorbed and the antioxidant capacity of the 55 blood is elevated (Iriti, Varoni, and Vitalini, 2010; Reiter, Manchester, and Tan, 56 2005). In addition to its antioxidant properties, MEL can modulate the immune 57 system (Carrillo-Vico, Lardone, Alvarez-Sanchez, Rodriguez-Rodriguez, and Guerrero, 2013; Mauriz, Collado, Veneroso, Reiter, and Gonzalez-Gallego, 2013), 58 59 exert both direct and indirect anticancer effects (Alvarez-Garcia, Gonzalez, Alonso-60 Gonzalez, Martinez-Campa, and Cos, 2012; Uguz, Cig, Espino, Bejarano, Naziroglu,

61 Rodriguez, and Pariente, 2012) act as a neurotransmitter, thus alleviating 62 neurodegenerative disorders such as Alzheimer's disease, and modulate sleep and circadian regulating systems in humans (Rosales-Corral, Acuña-Castroviejo, Coto-63 64 Montes, Boga, Manchester, Fuentes-Broto, Korkmaz, Ma, Tan, and Reiter, 2012; Slats, Claassen, Verbeek, and Overeem, 2013). Melatonin also plays other 65 physiological roles in mammals and is involved in, e.g., regulation of body 66 67 temperature, sexual maturation, mood and cardiovascular functions (Pandi-Perumal, 68 Zisapel, Srinivasan, and Cardinali, 2005). Healthy subjects synthesize MEL not only in the pineal gland (Stehle, Saade, Rawashdeh, Ackermann, Jilg, Sebestény, and 69 Maronde, 2011), but also in a wide range of other organs, i.e., gastrointestinal tract, 70 71 airway epithelium, pancreas, adrenal glands, thyroid gland, thymus, urogenital tract 72 and placenta (Kvetnoy, 1999). However, individuals who are deficient in MEL must 73 ingest it from chemical or natural sources. In a recent human study, it has been demonstrated that concentrations of urinary aMT6 (a marker of circulating MEL in 74 the body) increased significantly after consumption of some fruits (Johns, Johns, 75 76 Porasuphatana, Plaimee, and Sae-Teaw, 2012). In addition, ingestion of a tart cherry 77 juice concentrate increased the urinary MEL levels and was beneficial in improving sleep duration and quality in adults (Howatson, Bell, Tallent, Middleton, McHugh, 78 79 and Ellis, 2012). The uptake of MEL from vegetables and fruits as an alternative to 80 the intake of synthetic MEL is therefore of increasing interest (Korkmaz, 2011).

MEL occurs widely in higher plants, and it has been identified and quantified in several families of both monocotyledons and dicotyledons (Paredes et al., 2009). Nevertheless, available information on the natural effects of cultivar and solar radiation on the MEL contents of edible plants is rather scarce. Some data have shown intraspecific variations in the MEL content in tomatoes (Dubbels, Reiter,

Klenke, Goebel, Schnakenberg, Ehlers, Schiwara, and Schloot, 1995; Sturtz, Cerezo, 86 Cantos-Villar, and Garcia-Parrilla, 2011), strawberries (Sturtz et al., 2011) and 87 88 grapes (Iriti and Faoro, 2006). The environmental conditions under which plants grow also greatly influences their MEL content, e.g., its content increased from 22 to 89 142 ng g f.w.⁻¹ in the leaves of tomato plants grown in a controlled growing chamber 90 and in the open field respectively (Arnao and Hernández-Ruiz, 2013). Water 91 92 hyacinth (Eichhornia crassipes (Mart) Solms) plants grown under sunlight contain 93 more MEL than plants grown under artificial light (Arnao and Hernández-Ruiz, 94 2013; Tan, Manchester, Di Mascio, Martinez, Prado, and Reiter, 2007).

95 The objectives of the present study were to explore the following: the MEL 96 content of pepper fruits (*Capsicum annuum* L.), the effect of the ripeness of pepper 97 fruits on the MEL content, the effect of cultivar and solar radiation on MEL content 98 in pepper and tomato fruits, and the relationship between the MEL content and dry 99 weight (d.w.) of the fruits, to elucidate whether the MEL content is associated with 100 the carbon flux from leaves to fruits.

101

A COR

103

104 2.1. Reagents

105

N-acetyl-5-methoxytryptamine (MEL) standard was purchased from Fluka (Neu-106 Ulm, Germany). All LC-MS grade solvents were obtained from J. T. Baker 107 (Phillipsburg, NJ) and formic acid was purchased from Panreac Química S.A. 108 ANU 109 (Barcelona, Spain).

110

111 2.2. Experimental conditions

112

Six varieties of pepper (Capsicum annuum L.) and seven varieties of tomato 113 (Solanum lycopersicum Mill.) were chosen from commercial hybrids and local non-114 hybrid cultivars, all of which displayed different morphological types (Table 1). 115 Seeds of the local cultivars were obtained from the germplasm stored by NEIKER. 116 All cultivars were grown in soil, in the same unheated soft polyethylene-covered 117 greenhouse in Derio (Basque Country, northern Spain) (latitude, 43° 17' N; 118 longitude, 2° 52' W; altitude, 65 m above sea-level). The climate in the region is 119 Atlantic temperate. Measured air temperature (T, °C) and total cumulated radiation 120 121 (Rad, Wh m⁻²) in the greenhouse corresponding to the harvest periods were: min. T =122 7.9, max. T = 49, mean T = 23.8; min Rad = 390, max. Rad = 4546 and mean Rad = 123 2779.

Tomato and pepper plants with four true leaves were planted at a density of 1.7 124 plants per m^2 . Plants were drip irrigated with a nutrient solution (meg l^{-1}) containing 125

126 1.0 Mg^{2+} , 1.7 K⁺, 3.4 NO₃⁻, and 1.0 SO₄²⁻. The electrical conductivity of the nutrient 127 solution was 0.5 dS m⁻¹ and the pH varied between 5.0 and 5.5.

128 When the fruits were 0.5-1 cm thick, some of them were shaded with aluminium foil. The foil was placed loosely around the fruit as a skirt, which was closed at the 129 130 peduncle and open at the bottom to enable air circulation and thus minimize 131 differences in air humidity and temperature between the shaded and non-shaded fruit. However, the temperature regime experienced by the non-shaded fruits obviously 132 133 differed slightly because these fruits were sometimes directly exposed to solar 134 radiation. It is very difficult to manipulate the amount of incident solar radiation that reaches fruit without inducing changes in the microclimate. Nevertheless, we 135 considered that the main effect on shaded fruits was the large reduction in solar 136 137 radiation.

138

139 *2.3. Sample preparation*

140

141 Batches of mature unshaded green, shaded and unshaded light-red pepper fruits 142 and batches of shaded and unshaded light red tomato fruits were harvested from the same plants and washed with deionised water. Each batch of fruit weighed at least 143 144 1.5 kg, except the batches of cherry tomatoes, which weighed 350 g. Four batches 145 per treatment (unshaded green, unshaded red and shaded red) were processed. Non-146 edible parts (peduncle, calyx, placenta and seeds in pepper fruits and peduncle and 147 columella in tomato fruits) were discarded. The edible parts were homogenized in a 148 conventional food blender (Type 4184 Braun, Barcelona, Spain) at speed 5 for 1 149 minute. Two subsamples of the pure thus obtained (from 40 to 50 g each) were dried at 65 °C for 7 days (to constant weight) to determine the d.w. 150

151 For each batch, 30 g of puree was mixed with 30 ml of absolute methanol (HPLC 152 grade, Lab-Scan) and homogenized in a bullet blender 50 (Next Advance, USA) with 153 1.4 mm stainless beads, at speed 8 for 12 min. The homogenate was filtered 154 (Whatman paper No. 4) and centrifuged (Sorvall Legend XTR, Thermo Fischer 155 Scientific, Madrid, Spain) at 7600 x g for 15 minutes. The pellet was discarded and 156 the supernatant was evaporated to dryness under vacuum. The residue was resuspended in a mixture of methanol and water (1:1, v:v) before analysis of MEL. 157 158 All steps were carried out under dim light. NU

159

160 2.4. UHPLC-QqQ-MS/MS analysis

161

MEL determination and quantification was analyzed using a UHPLC-MS/MS 162 163 (UHPLC-1290 Series and a 6460 QqQ-MS/MS; Agilent Technologies, Waldbronn Germany) with an ACQUITY BEH C18 column (2.1 x 150 mm; 1.7 µm; Waters, 164 165 Milford, MA). Chromatographic separation was achieved using a binary gradient 166 consisting of (A) water and (B) methanol as LC grade solvents, both containing 0.1% 167 formic acid (v/v). The flow rate was 0.30 ml min-1 using a linear gradient (t; %B): 168 (0.00; 40), (1.50; 40), (1.51; 90), (3.50; 90), (3.51; 40). The volume injection was 20 169 ul. Multiple reaction monitoring mode (MRM) in positive mode was carried out and 170 MEL quantification was based on the 233/216 MRM transition (Mena, Gil-171 Izquierdo, Moreno, Martín, and García-Viguera, 2012; Rodriguez-Naranjo, Gil-172 Izquierdo, Troncoso, Cantos, and Garcia-Parrilla, 2011; Rodriguez-Naranjo, Gil-173 Izquierdo, Troncoso, Cantos-Villar, and Garcia-Parrilla, 2011). This MRM transition 174 was selected because of its specificity and better signal-to-noise ratio. Nitrogen was 175 used as the collision gas for the fragmentation by collision-induced dissociation of

176 the compounds at the collision cell of the triple quadrupole mass spectrometer. Mass 177 spectrometer parameters were set as follows: drying-gas flow: 8 min-1; sheath gas 178 flow: 12 min-1; sheath gas temperature: 350 °C; nebulizer pressure: 30 psi; capillary 179 voltage: 4000 V and nozzle voltage: 1000 V. MassHunter Software version B 04.00 180 was used for MS control and data gathering and MassHunter. Software version B 181 03.01 was used for data processing, peak integration and linear regression. 182 183 2.5. Statistical analysis 184 We used an independent-samples t test to compare means, and Duncan's test to 185

identify homogeneous subsets of means that are not different from each other (at a
significant level of 0.05). We used SSPS 10 software for all analyses.

189 3. Results and discussion

190

191 3.1. Effect of pepper and tomato cultivars on MEL contents

192

All six pepper cultivars analysed contained quantifiable amounts of MEL. The 193 content of this compound in red fruits ranged from 4.48 ng g^{-1} f.w. in the Barranca 194 cultivar to 11.90 ng g⁻¹ f.w. in the F26 cultivar (Fig. 1A). One-way analysis of 195 196 variance revealed a significant effect of cultivar on MEL content (p=0.000). Duncan's test identified three distinct groups of cultivars according to their contents: 197 the group with the lowest MEL contents (4.48-6.23 ng g^{-1} f.w.) included the 198 Barranca, NC9, Derio and Velero cultivars; the group containing intermediate 199 amounts of MEL (6.23-7.72 ng g⁻¹ f.w.) included Velero and Cristal and the group 200 201 containing the highest amounts of MEL comprised the F26 cultivar. On a d.w. basis, its content ranged from 31.01 ng g^{-1} d.w. to 93.40 ng g^{-1} d.w. in Barranca and F26 202 respectively (Fig. 1B). 203

MEL contents in samples of the seven cultivars of red tomato fruits ranged from 0.64 ng g⁻¹ f.w. (Ciliegia) to 14.77 ng g⁻¹ f.w. (Optima) (Fig. 2A). The one-way analysis of variance revealed a significant effect of these cultivars on MEL content (p=0.000). On a d.w. basis, its contents ranged from 7.47 ng g⁻¹ d.w. (Ciliegia) to 249.98 ng g⁻¹ d.w. (Optima) (Fig. 2B).

Although a lower MEL content has been reported for mature red tomato fruit (3-17 pg g⁻¹ f.w.) (Van Tassel, Roberts, Lewy, and O'Neill, 2001) and for pepper fruits, from 25.5 to 581 pg g⁻¹ f.w. (Huang and Mazza, 2011), the content in pepper and tomato fruits reported here is consistent with the values recently found for eleven varieties of tomato (4.1-114.5 ng g⁻¹ f.w.) (Sturtz et al., 2011). These authors

214 attributed the differences to the newly developed analytical method used. The latter 215 data revealed intraspecific differences of MEL contents in fruit among members of L. 216 esculentum, as previously suggested by Dubbels et al. (1995) and among members of 217 Prunus cerasus (Burkhardt, Tan, Manchester, Hardeland, and Reiter, 2001). 218 Nevertheless, our data confirm the important effect of cultivars on the MEL content 219 per unit of d.w. of pepper and tomato fruits. Dry weight is a more reliable measure 220 than the fresh weight (f.w.) because the water content of a plant or plant part will 221 obviously depend on several different factors (variety, the amount of water available 222 in soils, climatic conditions, time elapsed between harvest and analysis, and position 223 of fruit within the plant). Choice of the unit can also considerably modify the results. For example, on a f.w. basis, the Optima cultivar contained 23 times more MEL than 224 Ciliegia, but on a d.w. basis, the difference was 33.5-fold, due to the differences on 225 226 their d.w. values. However, from a nutritional point of view, data are usually presented on a f.w. basis, as this enables calculation of the contribution of the edible 227 228 part of a fruit or vegetable to the amount of this neurohormone in the human diet.

229 To explore the possible relationships between biomass allocation to the fruit and 230 MEL content, the pepper and tomato cultivars were chosen to yield a wide range of fruit d.w. (%), from about 10 to 16% in pepper and from about 6 to 12% in tomato. 231 232 There was no correlation between MEL content and fruit d.w. in either tomatoes or 233 peppers (Fig. 3). Therefore, its allocation in fruit does not appear to be related to the 234 carbon fluxes from leaves. Like other metabolites, the MEL content in fruit depends 235 on its uptake from phloem vessels, the extent to which it is degraded and how it is 236 metabolized. As far as we know, the possibility that MEL could be synthesized in 237 fruit has not previously been considered. However, the following have led us to 238 suggest that tomato fruit may biosynthesize MEL: the content in tomatoes harvested

239 at the mature green stage and allowed to ripen under controlled conditions increased 240 (Van Tassel et al., 2001) and there is some evidence that mitochondria and 241 chloroplasts (intracellular organelles present in green fruits) have the capacity to synthesize MEL in situ (Tan, Manchester, Liu, Rosales-Corral, Acuna-Castroviejo, 242 243 and Reiter, 2013).

- 244
- 245

3.2. Effect of the stage of ripeness of pepper fruit on MEL contents

246

247 Except for the Velero pepper cultivar, the MEL content per fresh or d.w. units tended to increase from mature green to red fruits. However, these differences were 248 only statistically significant in Barranca and F26 (Fig. 4 A and B), and there were no 249 250 such significant differences in the Derio and NC9 cultivars. The opposite pattern was 251 observed in Velero cultivar, in which the MEL contents in red fruits were lower than those of mature green fruits. Therefore, the stage of ripeness in Capsicum annuum 252 253 fruit plays an important role in the final MEL content, which is consistent with 254 previous data observed in tomatoes (Okazaki and Ezura, 2009; Van Tassel et al., 255 2001), i.e. that mature green fruits contained the lowest amounts and red tissue the 256 highest amounts of MEL. In the current study, we identified two types of pepper 257 cultivars: those in which the MEL content increased concomitantly with the 258 maturation degree (Barranca, F26 and NC9 although not significant) and in others 259 two cultivars where its content decreased (Velero and Derio although not 260 significant). The present results therefore demonstrate that the effect of the stage of 261 ripeness on MEL contents in pepper fruit is not straightforward and strongly depends 262 on the genotype.

264 3.3. Effect of the shading treatment on MEL contents in pepper and tomato fruits

265

266 Shaded tomato fruits contained more MEL than non-shaded fruits, except in the 267 Optima and Ciliega cultivars. The content in the shaded fruits of Optima was lower than that of control fruits, whereas that of the shaded fruit of Ciliega did not differ 268 269 from the control fruit (Fig. 5A). In contrast, in most pepper cultivars, the MEL 270 content of shaded fruit was lower than that shown for non-shaded fruit, whereas the 271 shading treatment did not affect the content in Barranca fruits and increased it in 272 Velero fruits (Fig. 5B). Therefore, it appears that the shading treatment had different 273 effects in these plants, leading to an increase of the MEL content in the tomatoes 274 cultivars and decrease in the pepper cultivars.

275 The differences in MEL contents between shaded and control fruits did not 276 correlate with the differences in fruit d.w. (Table 2). For most of the pepper and tomato cultivars, the shading treatment induced a decrease in the d.w. of the fruits, 277 278 which is consistent with previous findings that the d.w. tomato fruit tends to decrease 279 linearly with decreasing incident solar radiation (Riga, Anza, and Garbisu, 2008). 280 The present results demonstrate that the MEL contents of the fruits were not related to the carbon fluxes from leaves, as shown above from the analysis of the 281 282 relationship between the MEL content in red fruit of each cultivar and respect to its d.w. (Fig. 3). 283

Regarding the tomato cultivars, Jack was the most sensitive to the shading treatment as the MEL content increased by 135%, whereas Ciliegia was not sensitive to the reduction in incident radiation (Table 2). Of the pepper cultivars, California and F26 were the most sensitive, but showed a different pattern of response; the

shading treatment led to an increase in the MEL content of 64% in the Californiacultivar and to a decrease of the same rate per cent in the F26 cultivar.

290 Light is known to affect MEL metabolism in plants, as Murch et al., (2000) 291 demonstrated that light intensity regulates the incorporation of radio-labelled 292 serotonin into MEL. Under low light conditions, plantlets of Hypericum perforatum 293 cv. Anthos metabolized less MEL than under higher light intensities. Melatonin 294 content of the leaves of Eichhornia crassipes (Mart.) Solms plants grown under 295 sunlight are about 16 times higher than those of plants grown under artificial light 296 (Tan et al., 2007). In a recent study, the MEL content measured in leaves of tomatoes grown in an open field was found to be about 6.5 fold higher than in plants grown in 297 a controlled growing chamber and the difference between the plants grown in field 298 299 and in vitro was about 9.4-fold (Arnao and Hernández-Ruiz, 2013). The phenomenon 300 whereby light intensity induces the metabolism and accumulation of MEL appears to 301 be related to the photoprotection against oxidative stresses induced by free radicals 302 or reactive oxygen species, produced during the process of photosynthesis (Arnao 303 and Hernández-Ruiz, 2009; Paredes et al., 2009; Tan et al., 2007). However, caution 304 should be taken in extrapolating the role of MEL as a scavenger from photosynthetic 305 tissues to non-photosynthetic organs such as a mature fruit.

Data on the effects of light on MEL content in fruit are rather scarce. Under field conditions, grapes (*Vitis vinifera*) from bunches naturally shaded by leaves retained higher MEL contents, about 10 fold higher than in grapes those exposed to solar radiation (Boccalandro, González, Wunderlin, and Silva, 2011). It has been suggested that under intense radiation, larger amounts of MEL are consumed (because of its role as a free radical scavenger) than its quantity produced or imported from leaves, thus leading to low MEL contents in the fruit. In contrast,

313 under low levels of radiation, less MEL is consumed and the content in fruits will be 314 therefore higher. The response of most tomato cultivars and only one pepper cultivar 315 to the shading treatment was similar to that of V. vinifera. However, the opposite 316 response was observed in most pepper cultivars and two tomato cultivars, i.e. the 317 shaded fruits contained less MEL and others were not sensitive to the treatment. 318 These findings demonstrated at least two important aspects: there were intraspecific 319 differences in the response of the plants to reduced incident light in both C. annum 320 and S. lycopersicum, and the MEL content in shaded fruit did not always increase as 321 expected under the concept that the steady-state of this compound in fruit is directly 322 affected by the light (Boccalandro et al., 2011).

323 It has been suggested that the decrease in MEL contents observed in fruit exposed to intense light (such as sunlight) may be a regulatory response of plants to maintain 324 325 a high level of MEL in leaves exposed to light stress conditions and thereby to provide efficient protection against free radicals derived from photosynthetic 326 327 processes, so that the allocation of MEL to a sink organ like a fruit will be reduced 328 (Arnao and Hernández-Ruiz, 2013). However, in the present study, leaves were not 329 shaded and all plants were grown under the same conditions, and therefore the 330 differences in MEL contents between shaded and control fruit were not due to a 331 higher or lower stress levels in leaves.

To conclude, the melatonin content of red pepper fruits ranged from 31.0 to 93.4 ng g⁻¹ (d.w.) and tomato fruits from 7.47 to 249.98 ng g⁻¹ (d.w.). The effect of the stage of ripeness on MEL contents in pepper fruit is not straightforward and strongly depends on the genotype. Under shading treatment, the content of this hormone in most of tomato cultivars tended to increase (up to 135%), whereas that of most pepper cultivars decreased (to 64%). Overall, the results also demonstrated that the

338 melatonin content of the fruits was not related to carbon fluxes from leaves. Further

339 studies should be performed in order to elucidate whether the MEL content of fruit is

340 derived only from leaves or is also biosynthesized in the fruit, to identify the factors

341 that affect MEL transport from leaves and/or roots, and to clarify the physiological

342 functions of MEL in fruit.

References

346	Alvarez-Garcia, V., Gonzalez, A., Alonso-Gonzalez, C., Martinez-Campa, C., &
347	Cos, S. (2012). Melatonin interferes in the desmoplastic reaction in breast
348	cancer by regulating cytokine production. Journal of Pineal Research, 52,
349	282-290.
350	Arnao, M. B. & Hernández-Ruiz, J. (2009). Protective effect of melatonin against
351	chlorophyll degradation during the senescence of barley leaves. Journal of
352	Pineal Research, 46, 58-63.
353	Arnao, M. B. & Hernández-Ruiz, J. (2007). Melatonin promotes adventitious- and
354	lateral root regeneration in etiolated hypocotyls of Lupinus albus L. Journal
355	of Pineal Research, 42, 147-152.
356	Arnao, M. B. & Hernández-Ruiz, J. (2013). Growth conditions influence the
357	melatonin content of tomato plants. Food Chemistry, 138, 1212-1214.
358	Boccalandro, H. E., González, C. V., Wunderlin, D. A., & Silva, M. F. (2011).
359	Melatonin levels, determined by LC-ESI-MS/MS, fluctuate during the
360	day/night cycle in Vitis vinifera cv Malbec: evidence of its antioxidant role in
361	fruits. Journal of Pineal Research, 51, 226-232.
362	Burkhardt, S., Tan, D. X., Manchester, L. C., Hardeland, R., & Reiter, R. J. (2001).
363	Detection and quantification of the antioxidant melatonin in montmorency
364	and balaton tart cherries (Prunus cerasus). Journal of Agricultural and Food
365	Chemistry, 49, 4898-4902.

366	Carrillo-Vico,	A., Lardone	e, P., Alva	arez-Sanchez,	N., R	Rodriguez-Ro	driguez, A	۸., &	z

- Guerrero, J. (2013). Melatonin: buffering the immune system. *International Journal of Molecular Sciences*, 14, 8638-8683.
- 369 Dubbels, R., Reiter, R. J., Klenke, E., Goebel, A., Schnakenberg, E., Ehlers, C.,
- 370 Schiwara, H. W., & Schloot, W. (1995). Melatonin in edible plants identified
- by radioimmunoassay and by high performance liquid chromatography-mass
 spectrometry. *Journal of Pineal Research*, 18, 28-31.
- 373 Gitto, E., Pellegrino, S., Gitto, P., Barberi, I., & Reiter, R. J. (2009). Oxidative stress
- of the newborn in the pre- and postnatal period and the clinical utility of
 melatonin. *Journal of Pineal Research*, 46, 128-139.
- 376 Howatson, G., Bell, P., Tallent, J., Middleton, B., McHugh, M., & Ellis, J. (2012).

377 Effect of tart cherry juice (*Prunus cerasus*) on melatonin levels and enhanced
378 sleep quality. *European Journal of Nutrition*, 51, 909-916.

- Huang, X. & Mazza, G. (2011). Simultaneous analysis of serotonin, melatonin,
 piceid and resveratrol in fruits using liquid chromatography tandem mass
 spectrometry. *Journal of Chromatography A*, 1218, 3890–3899.
- 382 Iriti, M. & Faoro, F. (2006). Grape phytochemicals: A bouquet of old and new
 383 nutraceuticals for human health. *Medical Hypotheses*, 67, 833-838.
- 384 Iriti, M., Varoni, E. M., & Vitalini, S. (2010). Melatonin in traditional Mediterranean
 385 diets. *Journal of Pineal Research*, 49, 101-105.
- Johns, N. P., Johns, J., Porasuphatana, S., Plaimee, P., & Sae-Teaw, M. (2012).
 Dietary intake of melatonin from tropical fruit altered urinary excretion of 6-

388	sulfatoxymelatonin in healthy volunteers. Journal of Agricultural and Food
389	Chemistry, 61, 913-919.
390	Kolar, J. & Machakova, I. (2005). Melatonin in higher plants: occurrence and
391	possible functions. Journal of Pineal Research, 39, 333-341.
392	Korkmaz, A. (2011). Melatonin; from pineal gland to healthy foods. Spatula DD -
393	Peer Reviewed Journal on Complementary Medicine and Drug Discovery, 1,
394	33-36.
395	Kvetnoy, I. (1999). Extrapineal melatonin: location and role within diffuse
396	neuroendocrine system. Histochemical Journal, 31, 1-12.
397	Mauriz, J. L., Collado, P. S., Veneroso, C., Reiter, R. J., & Gonzalez-Gallego, J.
398	(2013). A review of the molecular aspects of melatonin's anti-inflammatory
399	actions: recent insights and new perspectives. Journal of Pineal Research, 54,
400	1-14.
401	Melchiorri, D., Reiter, R. J., Sewerynek, E., Hara, M., Chen, L., & Nistico, G.
402	(1996). Paraquat toxicity and oxidative damage: Reduction by melatonin.

403 *Biochemical Pharmacology*, 51, 1095-1099.

404 Mena, P., Gil-Izquierdo, Á., Moreno, D. A., Martín, N., & García-Viguera, C.
405 (2012). Assessment of the melatonin production in pomegranate wines. *LWT*406 - *Food Science and Technology*, 47, 13-18.

407 Murch, S. J., KrishnaRaj, S., & Saxena, P. K. (2000). Tryptophan is a precursor for
408 melatonin and serotonin biosynthesis in in vitro regenerated St. John's wort

409	(Hypericum perforatum L. cv. Anthos) plants. Plant Cell Reports, 19, 698-
410	704.

- 411 Okazaki, M. & Ezura, H. (2009). Profiling of melatonin in the model tomato
 412 (*Solanum lycopersicum* L.) cultivar Micro-Tom. *Journal of Pineal Research*,
 413 46, 338-343.
- Okazaki, M., Higuchi, K., Aouini, A., & Ezura, H. (2010). Lowering intercellular
 melatonin levels by transgenic analysis of indoleamine 2,3-dioxygenase from
 rice in tomato plants. *Journal of Pineal Research*, 49, 239-247.
- 417 Pandi-Perumal, S. R., Zisapel, N., Srinivasan, V., & Cardinali, D. P. (2005).
 418 Melatonin and sleep in aging population. *Experimental Gerontology*, 40, 911419 925.
- 420 Paredes, S. D., Korkmaz, A., Manchester, L. C., Tan, D. X., & Reiter, R. J. (2009).
 421 Phytomelatonin: a review. *Journal of Experimental Botany*, 60, 57-69.
- 422 Reiter, R. J., Manchester, L. C., & Tan, D. X. (2005). Melatonin in walnuts:
 423 Influence on levels of melatonin and total antioxidant capacity of blood.
 424 *Nutrition (Burbank, Los Angeles County, Calif)*, 21, 920-924.
- 425 Riga, P., Anza, M., & Garbisu, C. (2008). Tomato quality is more dependent on
 426 temperature than on photosynthetically active radiation. *Journal of the*427 *Science of Food and Agriculture*, 88, 158-166.
- Rodriguez-Naranjo, M. I., Gil-Izquierdo, A., Troncoso, A. M., Cantos, E., & GarciaParrilla, M. C. (2011). Melatonin: A new bioactive compound in wine. *Journal of Food Composition and Analysis*, 24, 603-608.

431	Rodriguez-Naranjo, M. I., Gil-Izquierdo, A., Troncoso, A. M., Cantos-Villar, E., &
432	Garcia-Parrilla, M. C. (2011). Melatonin is synthesised by yeast during
433	alcoholic fermentation in wines. Food Chemistry, 126, 1608-1613.
434	Rosales-Corral, S. A., Acuña-Castroviejo, D., Coto-Montes, A., Boga, J. A.,
435	Manchester, L. C., Fuentes-Broto, L., Korkmaz, A., Ma, S., Tan, D., &
436	Reiter, R. J. (2012). Alzheimer's disease: pathological mechanisms and the
437	beneficial role of melatonin. Journal of Pineal Research, 52, 167-202.
438	Sarropoulou, V. N., Therios, I. N., & Dimassi-Theriou, K. N. (2012). Melatonin
439	promotes adventitious root regeneration in in vitro shoot tip explants of the
440	commercial sweet cherry rootstocks CAB-6P (Prunus cerasus L.), Gisela 6
441	(<i>P. cerasus</i> \times <i>P. canescens</i>), and MxM 60 (<i>P. avium</i> \times <i>P. mahaleb</i>). Journal
442	of Pineal Research, 52, 38-46.
443	Slats, D., Claassen, J. A. H. R., Verbeek, M. M., & Overeem, S. (2013). Reciprocal
444	interactions between sleep, circadian rhythms and Alzheimer's disease: Focus
445	on the role of hypocretin and melatonin. Ageing Research Reviews, 12, 188-
446	200.
4 4 7	Stable L. H. Saada A. Damarkidak O. Ashammann K. Ella A. Sahartéan T. S

- Stehle, J. H., Saade, A., Rawashdeh, O., Ackermann, K., Jilg, A., Sebestény, T., &
 Maronde, E. (2011). A survey of molecular details in the human pineal gland
 in the light of phylogeny, structure, function and chronobiological diseases. *Journal of Pineal Research*, 51, 17-43.
- 451 Sturtz, M., Cerezo, A. B., Cantos-Villar, E., & Garcia-Parrilla, M. C. (2011).
 452 Determination of the melatonin content of different varieties of tomatoes

453	(Lycopersicon	esculentum)	and	strawberries	(Fragaria	ananassa).	Food
454	Chemistry, 127	, 1329-1334.					

455 Tan, D. X., Hardeland, R., Manchester, L. C., Korkmaz, A., Ma, S., Rosales-Corral,

S., & Reiter, R. J. (2012). Functional roles of melatonin in plants, and
perspectives in nutritional and agricultural science. *Journal of Experimental Botany*, 63, 577-597.

- Tan, D. X., Manchester, L. C., Di Mascio, P., Martinez, G. R., Prado, F. M., &
 Reiter, R. J. (2007). Novel rhythms of N₁-acetyl-N₂-formyl-5methoxykynuramine and its precursor melatonin in water hyacinth:
 importance for phytoremediation. *The FASEB Journal*, 21, 1724-1729.
- Tan, D. X., Manchester, L. C., Liu, X., Rosales-Corral, S. A., Acuna-Castroviejo, D.,
 & Reiter, R. J. (2013). Mitochondria and chloroplasts as the original sites of
 melatonin synthesis: a hypothesis related to melatonin's primary function and
 evolution in eukaryotes. *Journal of Pineal Research*, 54, 127-138.
- 467 Uguz, A. C., Cig, B., Espino, J., Bejarano, I., Naziroglu, M., Rodriguez, A. B., &
 468 Pariente, J. A. (2012). Melatonin potentiates chemotherapy-induced
 469 cytotoxicity and apoptosis in rat pancreatic tumor cells. *Journal of Pineal*470 *Research*, 53, 91-98.
- 471 Van Tassel, D. L., Roberts, N., Lewy, A., & O'Neill, S. D. (2001). Melatonin in plant
 472 organs. *Journal of Pineal Research*, 31, 8-15.
- Wang, P., Sun, X., Li, C., Wei, Z., Liang, D., & Ma, F. (2013). Long-term exogenous
 application of melatonin delays drought-induced leaf senescence in apple. *Journal of Pineal Research*, 54, 292-302.

- 476 Wang, P., Yin, L., Liang, D., Li, C., Ma, F., & Yue, Z. (2012). Delayed senescence
- 477 of apple leaves by exogenous melatonin treatment: toward regulating the
- Accepted ascorbate-glutathione cycle. Journal of Pineal Research, 53, 11-20. 478
- 479
- 480

Table 1. Origin and phenotypic characteristics of pepper and tomato cultivars. sd: standard deviation, n: number of samples.

Cultivars	Origin		Туре	Fruit weig	gth (g)	Fruit width	n (mm)	Fruit length	(mm)	length/	width	n
				mean	sd	mean	sd	mean	sd	mean	sd	
Pepper												
Barranca	Local cultivar	NEIKER	Triangular	59.1	10.3	51.5	13.9	110	20.4	2.30	0.67	16
Cristal	Comercial hybrid	Ramiro Arnedo	Elongate	67.6	11.5	44.5	4.52	115	14.0	2.61	0.36	26
Derio	Local cultivar	NEIKER	Elongate	35.3	7.51	33.4	5.90	125	12.2	3.88	0.93	28
F26	Local cultivar	NEIKER	Elongate	61.2	0.98	36.7	0.80	207	2.20	5.66	0.70	12
NC9	Local cultivar	NEIKER	Elongate	48.3	4.24	35.1	3.35	166	7.22	4.77	0.81	24
Velero	Comercial hybrid	Seminis	Blocky	175	24.1	86.8	6.01	78.6	6.20	0.91	0.11	24
Tomato									/			
Ciliegia	Comercial hybrid	Vilmorin	Cherry	7.78	2.38	24.2	1.73	23.8	1.84	0.98	0.04	16
Isis	Local cultivar	NEIKER	Cherry	16.4	3.37	30.6	2.25	28.8	2.58	0.94	0.04	25
Jack	Comercial hybrid	Seminis	Beef	280	78.4	84.7	9.64	68.9	5.57	0.82	0.05	12
Jesus	Local cultivar	NEIKER	Beef	462	167	110.7	13.2	79.5	10.8	0.72	0.07	16
NKT072	Local cultivar	NEIKER	Elongate	152	50.7	51.9	7.49	120	16.9	2.36	0.43	17
Optima	Comercial hybrid	Royal Sluis	Beef	221	35.0	77.4	-7.22	65.6	3.95	0.86	0.13	12
Prico	Local cultivar	NEIKER	Beef	228	85.9	84.4	10.5	61.3	7.76	0.73	0.06	15
		6	0									
		6										
		6										

Table 2. Effect of the shading treatment on melatonin content pepper and tomato fruits of. Values were calculated using the following formula: $Mel(_{shaded fruit})*100/Mel(_{control fruit})-100$, where Mel is the melatonin content per d.w.. ns: differences between shaded and control fruit are not significant.

Cultivar		nelatonin Differences in d.w. (%)
	content (%)	
	Peppers	
Velero	64.4	-20.1
Derio	-51.2	-5.16
F26	-64.0	ns
NC9	-41.4	-13.6
Barranca	ns	-9.41
Cristal	-40.8	ns
	Tomatoes	
Optima	-36.6	-3.54
Jack	136	-7.00
Ciliegia	ns	10.7
Isis	73.5	-13.0
NKT072	95.9	-6.59
Prico	104	ns
Jesus	ns	-12.9

Figure 1.

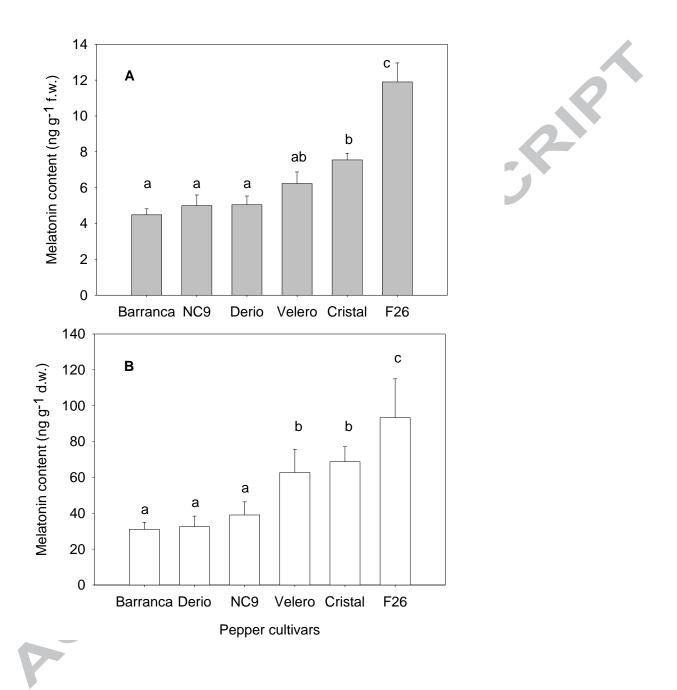


Fig. 1. Effect of cultivar on the melatonin content of light-red pepper fruits. The melatonin contents are expressed on a fresh eight (A) and dry weight (B) basis. All cultivars were grown under the same greenhouse conditions. Each column represents the mean \pm standard deviation from 4 replicates. Means followed by the same letters are not significantly different at *p*<0.05 (Duncan test).

Figure 2.

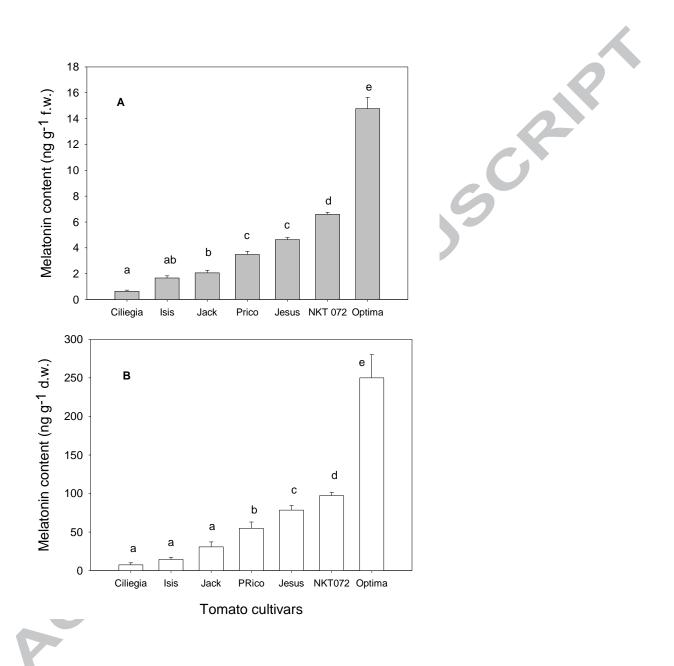


Fig. 2. Effect of cultivar on the melatonin content of light-red tomato fruits. The melatonin contents are expressed on a f.w. (A) and d.w. (B) basis. All cultivars were grown under the same greenhouse conditions. Each column represents the mean \pm standard deviation from 4 replicates. Means followed by the same letters are not significantly different at *p*<0.05 (Duncan test).

Figure 3.

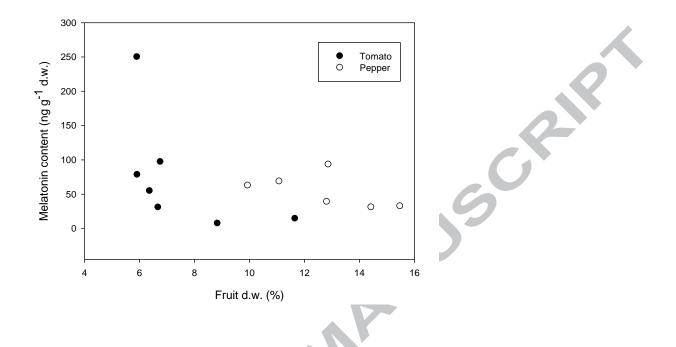


Fig. 3. Relationships between melatonin content and fruit d.w. for all tomato and pepper cultivars. Each point represents the mean value from 4 replicates for melatonin values and from 8 replicates for d.w. values.

Figure 4.

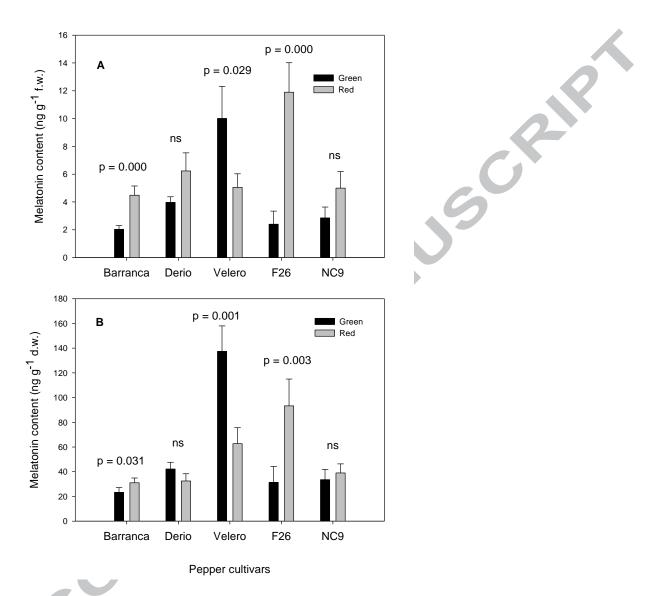


Fig. 4. Effect of stage of ripeness on melatonin content in mature green and light-red pepper fruits. Data are expressed on a f.w. (A) and a d.w. (B) basis. All cultivars were grown under the same greenhouse conditions. Each column represents the mean \pm standard deviation from 4 replicates. *p*-values from the independent-samples t test (Student's test) are shown. ns: not significant.

Figure 5.

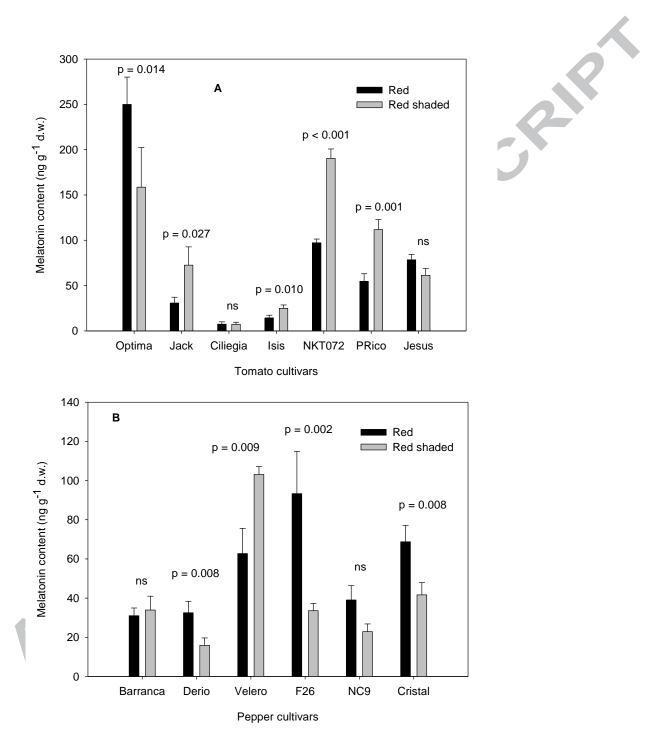


Fig. 5. Effect of solar radiation on melatonin content in tomato (A) and pepper (B) cultivars. All cultivars were grown under the same greenhouse conditions and some fruits were shaded with an aluminium foil. Each column represents the mean \pm standard deviation from 4

replicates. *p*-values from the independent samples t test (Student's test) are shown. ns: not Acctebric significant.

Highlights

Melatonin in pepper and tomato fruits is strongly affected by incident solar radiation.

All cultivars did not show the same pattern of response to the shading treatment.

Melatonin accumulation in the fruits does not appear to depend on carbon flux from n Recorded to the second secon leaves.