1 2 3 4 5	This is the accepted version of the following article: Morales J (2020) Eggshell Biliverdin as an Antioxidant Maternal Effect: Biliverdin as an Antioxidant Resource in Oviparous Animals. BioEssays 42: 2000010, which has been published in final form at: https://onlinelibrary.wiley.com/doi/abs/10.1002/bies.202000010. This article may be used for non-commercial purposes in accordance with the Wiley Self-Archiving Policy
6	[http://www.wileyauthors.com/self-archiving].
7	
8	
9	Eggshell biliverdin as an antioxidant maternal effect
10	
11	Judith Morales
12	
13	National Museum of Natural Sciences, Spanish National Research Council (CSIC), c/ José
14	Gutiérrez Abascal 2, 28006 Madrid, Spain. Correspondence: jmorales@mncn.csic.es; Tel.: +34
15	914 111 328 (ext. 981341).
16	
17	
18	Subtitle: Eggshell biliverdin as an antioxidant resource in oviparous animals
19	Article type: Hypotheses
20	Author contribution: JM conceived the research and wrote the paper.
21	Data availability statement: No data was used.
22	
23	

## 24 Abstract

25	In this essay, I propose the hypothesis that biliverdin pigment plays an antioxidant role in the
26	avian eggshell. Due to its ability to scavenge free radical species and to reduce mutation,
27	biliverdin potentially counteracts the oxidative action of pathogens that penetrate the eggshell
28	and/or protects the shell membrane from oxidation, thus promoting the proven antioxidant
29	and antimicrobial capacities of the shell membrane itself. Additionally, biliverdin may be able
30	to inhibit viral replication in the eggshell due to its ascribed antiviral properties. Moreover,
31	previous results in other taxa leave open the question of whether biliverdin can be absorbed
32	by the embryo from the eggshell and play a role in embryogenesis. These mechanisms of
33	antioxidant action of eggshell biliverdin remain totally unexplored in birds and in other
34	oviparous animals. I develop the main assumptions and predictions of the antioxidant
35	hypothesis, and propose directions for future research.
36	
37	Keywords: antioxidants, biliverdin pigmentation, eggshell colouration, eggshell membrane,
38	maternal effects, oxidative stress
39	
40	

# 42 **1. Introduction**

43 The colour of avian eggs has long attracted evolutionary ecologists. In the nineteenth century, renowned naturalists such as Darwin, Wallace and Hewitson already observed its variation 44 across species and suggested that it was related to nesting habits.<sup>[1]</sup> Since then, the function of 45 46 avian eggshell colouration has remained a major topic in the field (for some reviews and discussions in the past two decades, see<sup>[1-6]</sup>). Recently, attention has also been directed to 47 48 understanding the evolutionary origin of eggshell colouration. Indeed, the two main pigments 49 present in avian eggs, protoporphyrin (responsible for brown hues) and biliverdin IXα (for blue and green hues), have even been studied and identified in dinosaur eggshells.<sup>[7]</sup> 50 51 Adaptive explanations proposed for variation in eggshell pigmentation can be included 52 in two main, non-mutually exclusive sets of hypotheses. The first one focuses on eggshell 53 colour as an external trait that can be visually perceived by a diverse array of observers. It can favour camouflage from predators<sup>[8]</sup> and avian brood parasites,<sup>[9]</sup> or the opposite, it can 54 55 increase conspicuousness to signal egg unpalatability to predators.<sup>[10]</sup> In species with bi-56 parental care, shell colour has been proposed as a post-mating signal of female quality to encourage greater paternal investment ("sexual signalling hypothesis"),<sup>[3]</sup> or to force it 57 ("blackmail hypothesis").<sup>[11]</sup> The second set of hypotheses is based on the direct action of 58 59 protoporphyrin and biliverdin, whose chemical properties can adaptively modulate the 60 structure and physiology of the shell and thus influence embryonic survival. Specifically, protoporphyrin has been suggested to enhance eggshell strength<sup>[4, 12]</sup> and to promote a 61 photoactive antimicrobial defence.<sup>[13]</sup> Biliverdin, through interactions with the light 62 63 environment, has been proposed to accelerate embryonic development and to favour the light-dependent repair of DNA lesions caused by specific UV radiation.<sup>[14]</sup> This interaction 64 would only apply to open-nesting species; however, the evidence in these species is mixed.<sup>[15,</sup> 65 66 <sup>16]</sup> On the other hand, it has been reported that both pigments reduce near-infrared 67 absorbance and solar overheating.<sup>[17]</sup> Again, these actions would be associated mainly with

68 open-nesting species exposed to high levels of harmful radiation. In contrast, in a comparative 69 analysis performed at a global scale, Wisocki and co-authors concluded that brown eggshell 70 colouration is more frequent in open-nesting species living in cold and humid climates.<sup>[6]</sup> 71 Moreover, they found that darker eggs heated more rapidly than lighter ones when exposed to 72 solar radiation, supporting a thermoregulatory role for both eggshell pigments (biliverdin and 73 protoporphyrin) combined. Nonetheless, although the analysis yielded an impressive dataset, 74 in which the eggshell colour of more than 600 species belonging to most avian orders was 75 measured, the interspecific patterns of eggshell blue-green chroma were not explained by the 76 environmental factors studied.<sup>[6]</sup>

77 An often overlooked property of biliverdin that has the potential to directly promote 78 embryo survival is its antioxidant capacity. Traditionally, biliverdin and its derivative in 79 humans, bilirubin, have been considered as toxic, but this is mainly evident when the concentration of bile pigments is unusually high.<sup>[18]</sup> Nowadays, biliverdin's ability to scavenge 80 free radical species and reduce mutation is also clearly recognized<sup>[19-22]</sup> (in birds, see<sup>[23]</sup>). 81 82 Certainly, the sexual signalling hypothesis of blue-green eggshell colouration is grounded in 83 biliverdin's antioxidant properties: females that lay more colourful blue-green eggs signal their 84 low oxidative status to males.<sup>[3]</sup> In the present essay, however, I argue that these antioxidant 85 properties also have the potential to directly promote embryo survival. This "antioxidant 86 hypothesis" is motivated by the facts that i) biliverdin's molecular structure (a tetrapyrrole 87 with an extended system of conjugated double bonds and reactive hydrogen atoms) is 88 responsible for its antioxidant properties<sup>[21]</sup> and ii) the biliverdin molecule in the eggshell is exactly the same as in other organismal tissues in which it plays an antioxidant role.<sup>[21, 24]</sup> 89 90 According to the antioxidant hypothesis, biliverdin counteracts the oxidative action of 91 pathogens that penetrate the eggshell and/or protects the shell membrane from oxidation, 92 thus promoting the proven antioxidant and antimicrobial capacities of the eggshell membrane 93 itself.<sup>[25]</sup> Biliverdin might also directly inhibit viral replication in the eggshell, due to its ascribed

antiviral properties.<sup>[26]</sup> Moreover, as previously speculated,<sup>[27]</sup> it is not implausible to 94 95 hypothesize that the avian embryo gradually absorbs biliverdin traces from the eggshell, as it does with calcium. Biliverdin IX $\alpha$ , the same molecule used by female birds to pigment their 96 97 eggs, is also present in the oocyte, egg and embryonic cytoplasm of the amphibian Xenopus *laevis*, and plays a crucial role in dorsal axis development.<sup>[28]</sup> Although amphibian eggs differ 98 99 significantly from avian eggs, mainly because they lack a shell and pigments mostly accumulate 100 in the yolk, the study in X. laevis leaves open to question a direct function of eggshell biliverdin 101 during avian embryonic development. Similarly, in insects, CV-bilin, a biliverdin isomer present 102 in eggs has been proposed to function in cellular regulation.<sup>[29]</sup> 103 To my knowledge, these mechanisms of antioxidant action of eggshell biliverdin (Table

104 1) remain totally unexplored. In the following two sections, I develop the assumptions and 105 predictions of the antioxidant hypothesis of eggshell biliverdin, the previous observations that 106 support them, and possible ways to test the hypothesis.

107

# **2. Conceptual framework of the antioxidant hypothesis**

### 109 **2.1.** Assumptions: inherent antioxidant capacity and high permeability of biliverdin

110 There are two main assumptions underlying the antioxidant hypothesis of eggshell biliverdin. 111 The first one is that the biliverdin molecule possesses the ability to scavenge free radical 112 species and to reduce mutation, which, as mentioned above, has been consistently demonstrated (e.g.,<sup>[19-22]</sup>). By reducing oxidative damage and mitigating infections, eggshell 113 114 biliverdin potentially protects the integrity of the thin organic membrane that envelops the 115 albumen and connects the true shell with the egg contents (Figure 1). Thus, the second 116 assumption is that biliverdin extends deep into the calcified layers of the eggshell and can 117 come in direct contact with the eggshell membrane. Indeed, small amounts of the pigment 118 (less than 0.01 absorbance/g) have been detected in the innermost calcified eggshell layers in various bird species<sup>[30, 31]</sup> (see also<sup>[7]</sup> in dinosaur eggs), which can be visually perceived in some 119

cases (Figure 2a). This high permeability also implies that the molecule surrounds eggshell
 pores and cracks, gateways for microbes. Intriguingly, pore density has been positively
 associated with biliverdin-based eggshell pigmentation in a passerine species at a wide
 geographic scale.<sup>[32]</sup>

124 Additionally, in the case that the embryo absorbs eggshell biliverdin as a resource 125 during development, a third assumption should be that the pigment permeates the shell 126 membrane itself. This assumption is supported by three arguments. First, the shell membrane penetrates the basal calcified layer, <sup>[33, 34]</sup> also called the cone or mammillary layer, which can 127 128 be permeated by biliverdin (Figure 1). Second, protoporphyrin, which is less permeable than 129 biliverdin,<sup>[35]</sup> has been identified in the shell membrane of various bird species.<sup>[36]</sup> Third, 130 biliverdin shows high affinity to carrier proteins that are abundant both in the shell membrane and in the albumen,<sup>[33, 37]</sup> like albumin, globulin and vitellogenin,<sup>[33, 38]</sup> the major yolk precursor 131 protein associated with embryonic development. Biliverdin also interacts with cysteine-rich 132 proteins, <sup>[39]</sup> which are abundant in the shell membrane, and forms soluble complexes with egg 133 134 white lysozyme, an antioxidant enzyme that is present both in the shell membrane and in the mammillary layer.<sup>[40]</sup> 135

136 If biliverdin is absorbed by the embryo during development, one question to address is 137 why do females not allocate it directly inside the egg. One possible reason is that biliverdin 138 assimilation is similar to that of eggshell calcium, which is gradually absorbed within fine physiological limits and whose excess inside the egg can be highly toxic for the embryo. <sup>[41]</sup> If 139 140 true, I would expect biliverdin traces to be more easily detected in the shell membrane at 141 advanced stages of embryo development, coinciding with a higher exposure to oxidative 142 stress.<sup>[42]</sup> Suggestive of this, I have observed that the shell membrane of the blue-footed booby 143 (Sula nebouxi) partially has a greenish tint at hatching (Figure 2a, arrow); nonetheless, the 144 pigment composition of this membrane needs to be confirmed analytically. In this context, one 145 may ask how biliverdin could be transported inside the egg. On the one hand, eggshell

membrane proteins represent a potential mechanism, given the high affinity of biliverdin for
them. On the other hand, calcium is implicated in biliverdin synthesis in the shell gland and its
transport to uterine fluid,<sup>[43]</sup> suggesting the possibility that the embryo gradually absorbs
biliverdin together with calcium. The latter mechanism would depend, first, on the possibility
that biliverdin-calcium complexes can be formed, and then on the relative amount of eggshell
protoporphyrin pigment, which shows high affinity for calcium.<sup>[24]</sup>

152

#### 153 **2.2.** Predictions: highly conserved but only abundant under high oxidative stress

154 If biliverdin plays a beneficial antioxidant role during development, the main prediction should 155 be that most bird species allocate it in the eggshell. However, there are many birds that do not lay blue-green eggshells,<sup>[1, 6]</sup> at least apparently. One possibility is that biliverdin traces are 156 157 indeed present in the eggshells of most species and that they are enough to elicit an antioxidant effect. In fact, it has been claimed that unpigmented eggs are rare in nature and 158 that most white eggs contain pigment traces,<sup>[50]</sup> but this needs to be demonstrated. 159 160 As most studies on biliverdin focus on signalling mechanisms or eggshell colour as a 161 proxy for pigment concentration, the prevalence of biliverdin traces is unknown in most cases. 162 However, apparently unpigmented eggs of some species have been shown to contain 163 biliverdin (e.g., the eggs of the black-footed and the Humboldt's penguin, the Mandarin duck 164 and the wood pigeon, respectively, Spheniscus demersus, S. humboldti, Aix galerlculata and *Columba palumbus*;<sup>[51]</sup> see also<sup>[52]</sup> in white Leghorns, *Gallus gallus domesticus*), and brown 165 166 eggs can contain it in very high amounts.<sup>[16, 51]</sup> In all the previous cases, the presence of 167 biliverdin would go unnoticed if only blue-green eggshell colouration is relied on as a measure. 168 Note also that the eggs of many marine and raptor species are blue only when fresh and become white or grey almost immediately after laying (see Figure 2b)<sup>[53]</sup>. Nonetheless, it is 169 170 possible to observe that the inner shell layers of such eggs at hatching contain high amounts of 171 biliverdin (Figure 2a). Colour fading of the outermost eggshell layers also occurs in

songbirds.<sup>[54]</sup> Thus, the antioxidant hypothesis predicts, in the first place, that most species
contain biliverdin traces in the inner eggshell layers, regardless of eggshell external
appearance. Only in species in which biliverdin-based pigmentation has evolved as a signal
(directed at predators, brood parasites or males) or as a photo-protective agent should it be
present in high amounts in the outermost shell layers as well.

177 Although the physiology of closely related species is not expected to differ to the 178 extent that biliverdin is either allocated or not in the eggshell, it is plausible that the associated 179 costs for females limit the amount allocated. In fact, the allocation of biliverdin to the eggshell 180 has been shown to decrease antioxidant levels in the defence system of laying females, [55] a 181 cost that is alleviated when the females are supplemented with carotenoids and antioxidant vitamins prior to egg laying.<sup>[27, 46, 56]</sup> Thus, a second prediction of the hypothesis is that large 182 183 amounts of the pigment are allocated to the eggshell only in those species in which the 184 antioxidant benefits for the embryo compensate the oxidative costs for females. Due to the 185 ability of biliverdin to scavenge free radical species, thus reducing DNA damage, its role should 186 be more prominent when embryos are particularly exposed to oxidation during development. 187 First, life-history strategies characterized by rapid embryonic growth and small body 188 mass and brain size have been consistently linked to increased oxidative damage across divergent taxonomic groups.<sup>[57-59]</sup> Second, high amounts of biliverdin in the inner shell layers 189 190 are predicted to be more relevant in the case of plastic increases in growth rate, for example, 191 catch-up growth strategies in late-hatching animals, which can lead to higher oxidative damage.<sup>[60]</sup> Third, biliverdin traces are predicted to be more easily detected in species or 192 193 populations that are more exposed to external sources of oxidative stress, such as pathogens 194 and pollutants. Previous results seem to support these predictions. For instance, biliverdin eggshell colouration has been reported to be more frequent in smaller species<sup>[61, 62, 15]</sup> (but 195 see<sup>[1]</sup>) and in passerine lineages with shorter incubation periods.<sup>[15]</sup> Intraspecific studies have 196 197 also reported that biliverdin-based pigmentation is associated with higher haemolytic bacterial

loads in the eggshell,<sup>[47]</sup> and is related to environmental contamination.<sup>[63, 64]</sup> The latter 198 199 relationships could merely be due to changes in haem biosynthesis following high exposure to pathogens or contaminants.<sup>[65]</sup> However, it is also possible that eggshell pigmentation is in part 200 201 adaptively upregulated to protect the embryo against external sources of oxidation. 202 Finally, a third prediction of the antioxidant hypothesis is that the shell membrane 203 shows stronger antioxidant and antimicrobial properties when biliverdin is present, especially 204 in the inner shell layers. Furthermore, by enhancing the properties of the shell membrane, 205 which is the last barrier between the true shell and the albumen, biliverdin would also be 206 protecting the egg contents. Then, we may likewise predict that the yolk and the albumen 207 show greater antioxidant capacity when eggshell biliverdin is present in the inner shell layers. 208 In agreement with the latter prediction, in various bird species, biliverdin-based pigmentation reflects the antioxidant quality of egg contents<sup>[44-46]</sup> (but see<sup>[47]</sup>), including yolk vitamin E and 209 210 carotenoids, both of which protect against oxidation. Moreover, in poultry species, blueshelled egg yolks possess higher radical scavenging activity than white-shelled egg yolks, [48] one 211 of the reasons commercial blue eggs are more valued in Korea<sup>[48]</sup>. Although none of the above 212 213 studies demonstrated a direct physiological function of biliverdin in the shell, they have 214 revealed that its allocation is narrowly linked to the antioxidant quality of bird mothers<sup>[49]</sup> and their eggs.<sup>[44-46]</sup> 215

216

# 217 **3. Testing the hypothesis**

The antioxidant hypothesis assumes that biliverdin is in direct contact with the shell
membrane and, if absorbed by the embryo, permeates the shell membrane itself (Table 1).
Further studies on the antioxidant role of eggshell biliverdin should thus aim to detect
biliverdin traces/concentration directly in the innermost calcified layers and in the shell
membrane. This has been achieved by means of layer-by-layer dissolution methods combined
with spectrophotometric measurements of the supernatant.<sup>[30]</sup> The shell membrane can be

224 easily detached from the calcified layers and be analysed separately. Additionally, the use of Raman spectroscopy has been successfully used in eggshell fragments<sup>[7, 66]</sup> and can map 225 226 pigments across vertical egg sections.<sup>[7]</sup> Given that the external eggshell colour do not 227 accurately reflect the concentration of pigments in the inner layers (Fig. 2b),<sup>[67]</sup> 228 spectrophotometric methods should focus on the inner eggshell surface. This may limit the 229 scope for studying, for instance, specimens from Museum egg collections. However, in natural 230 populations, broken eggshells can be easily collected at hatching. Actually, if biliverdin is 231 gradually absorbed by the embryo, I would expect that it is more easily detected in the shell 232 membrane at later stages of embryo development.

233 The antioxidant capacity of the shell membrane and the egg contents (yolk and 234 albumen) needs to be compared between species that lay eggs pigmented with biliverdin and those that do not, [48] and also among species that allocate biliverdin in different amounts. The 235 236 use of a battery of tests (i.e., assessment of total antioxidant capacity, as well as radical 237 scavenging activity, yolk lipid peroxidation and the amount of specific antioxidants like 238 lysozyme, vitamin E, carotenoids, superoxide dismutase and glutathione peroxidase) would 239 contribute to a better characterization of the antioxidant capacity of different egg 240 components.<sup>[48]</sup> Intriguingly, the eggshells of certain bird species also contain biliverdin 241 reductase (Hanley, D., personal communication), which is the enzyme that catalyses the reduction of biliverdin to bilirubin in other vertebrates.<sup>[20]</sup> Future studies could also explore the 242 243 occurrence of this molecule in the different shell layers and test whether it increases the 244 antioxidant potential of biliverdin.

However, to reveal causal relationships between the presence of biliverdin in the eggshell and the antioxidant potential of the shell membrane/egg components, experimental manipulation of biliverdin levels at laying is needed. Experiments of this kind have been performed by exogenous injection of the pigment into the shell gland of ducks, *Anas platyrhynchos*,<sup>[31]</sup> although this approach would entail more difficulty in smaller species. A

250 complementary manipulation would be to induce an external oxidative challenge during embryo development (e.g., elevated oxygen concentration during incubation),<sup>[42]</sup> and analyse 251 252 the resulting relationship between eggshell biliverdin concentration and the oxidative status of 253 the shell membrane and egg components. In addition, it would be extremely interesting to 254 assess biliverdin concentration in the plasma of hatchlings in response to the above 255 experimental approaches. Increased pigment concentration in plasma after experimental 256 enhancement of the pigment in the eggshell would support absorption by the embryo during 257 development.

258 The hypothesis also predicts that eggshell biliverdin should be more important when 259 the embryo is more exposed to oxidative stress (Table 1). Inter- and intra-specific analyses 260 could test this prediction by exploring the occurrence of eggshell biliverdin traces in relation to 261 species/populations growth strategies and to exposure to pathogens and contaminants. Inter-262 specific studies should control for potential selective forces known to favour biliverdin-based 263 pigmentation (at least, in the outermost eggshell layers), like nesting habits, male parental care, predation risk and light exposure.<sup>[1, 3, 8, 68]</sup> As mentioned in the preceding section, the 264 265 relationship with microbes and contaminants could merely reveal changes in haem 266 biosynthesis pathway following high exposure to these external oxidative agents or could also 267 be due to an adaptive upregulation of biliverdin synthesis. Experimental manipulation of the 268 exposure to pathogens and contaminants could help to distinguish between these two 269 possibilities. An adaptive response would be supported if: i) eggshell biliverdin is upregulated 270 after the experimental challenge and ii) is accompanied by a decrease in oxidative stress.

271

### **4. Conclusions and outlook**

In this essay, I propose the hypothesis that biliverdin pigment functions as an antioxidant in
the eggshell, and potentially in the shell membrane and the embryo. Due to the ability of
biliverdin to scavenge free radical species, thus reducing DNA damage, the hypothesis predicts

276 it to act as an antioxidant in most bird species, but its role should be more prominent when 277 embryos are particularly exposed to oxidation. Future research should focus on studying the 278 presence of biliverdin traces in the inner eggshell layers and in the link of these traces with 279 embryonic exposure to oxidative damage, mediated, for instance, by fast growth strategies, 280 exposure to pollution or microbial infections. Experimental studies manipulating eggshell 281 biliverdin to explore its effect on the eggshell membrane and on embryonic development are 282 also promising lines of research. The antioxidant hypothesis of biliverdin in the eggshell (and 283 beyond it) is compatible with other explanations proposed for blue-green eggshell 284 pigmentation, for example, the idea that biliverdin can accelerate embryonic development due 285 to a photo-active effect, predicted to be more important in species with faster growth rates.<sup>[14,</sup> 286 <sup>68]</sup> Biliverdin-based pigmentation might have been shaped by a combination of selective forces that vary according to both the life history of a species and the environment,<sup>[6]</sup> thus making it 287 288 difficult to elucidate its adaptive function. This essay does not pretend to offer a unique 289 explanation for the observable patterns of blue-green eggshell colouration. Rather, it aims to 290 reopen the debate about these unexplained patterns with a different perspective, one based 291 on the antioxidant potential of biliverdin in the eggshell, an ancient biomolecule that was 292 already present in dinosaur eggshells, well before the modern bird radiation.<sup>[7]</sup> This and other 293 hypotheses on the role of biliverdin during embryogenesis need to be further explored in birds 294 and in other taxa.

295

ACKNOWLEDGEMENTS: I thank Juan Moreno and Manuel Jiménez-Tenorio for insightful
 discussions on the ideas presented here. I am also thankful to Daniel Hanley for invaluable
 comments and to Melinda Modrell for English correction of a previous version of this paper.
 This work was supported by a Ramón y Cajal by the Ministerio de Economía, Industria y
 Competitividad MINECO (CGL2016-79390-P) and the European Regional Development Fund
 (FEDER).

302

### 303 CONFLICT OF INTEREST

- 304 The author declares no conflict of interest.
- 305

#### 306 **REFERENCES**

- 307 [1] R. M. Kilner, *Biol. Rev.* **2006**, 81, 383.
- 308 [2] T.J. Underwood, S.G. Sealy, in Avian Incubation, Behaviour, Environment and Evolution (Ed:
- 309 D.C. Deeming) Oxford University Press, Oxford **2002**, pp. 280-298.
- 310 [3] J. Moreno, J.L. Osorno, *Ecol. Lett.* **2003**, 6, 803.
- 311 [4] A.G. Gosler, J.P. Higham, S.J. Reynolds, *Ecol. Lett.* **2005**, *8*, 1105.
- 312 [5] P. Cassey, *Curr. Biol.* **2009**, 19, 1083.
- 313 [6] P.A. Wisocki, P. Kennelly, I. Rojas-Rivera, P. Cassey, D. Hanley, *Nature Ecol. Evol.* 2020, 4,
- 314 148.
- 315 [7] J. Wiemann, T-R. Yang, M.A. Norell, *Nature* **2018**, 563, 555.
- 316 [8] M. C. Stoddard, K.L.A. Marshall, R.M. Kilner, Avian Biol. Res. 2011, 4, 196.
- 317 [9] N.B. Davies, M.De L. Brooke, J. Anim. Ecol. 1989, 58, 207.
- 318 [10] H.B. Cott, *Nature* **1948**, 161, 8.
- 319 [11] D. Hanley, S. Doucet, D.C. Dearborn, *The Auk* **2010**, 127, 453.
- 320 [12] S.E. Solomon, *Egg and Eggshell Quality*, Iowa State University Press, Ames, IA **1997**.
- 321 [13] S.I. Ishikawa, K. Suzuki, E. Fukuda, K. Arihara, Y. Yamamoto, T. Mukai et al., FEBS Lett.
- **2010,** 584, 770.
- 323 [14] G. Maurer, S.J. Portugal, P. Cassey, J. Avian Biol. 2011, 42, 494.
- 324 [15] J.J. Soler, J. Moreno, J.M. Avilés, A.P. Møller, *Evolution* **2005**, 59, 636.
- 325 [16] P. Cassey, G.H. Thomas, S.J. Portugal, G. Maurer, M.E. Hauber, T. Grim, et al., Biol. J. Linn.
- *Soc.* **2012**, 106, 657.
- 327 [17] G.S. Bakken, V.C. Vanderbilt, W.A. Buttemer, W.R. Dawson, *Science* **1978**, 200, 321.

- 328 [18] S.F. Asad, S. Singh, A. Ahmad, N.U. Khan, S.M. Hadi, *Chem. Biol. Interact.* 2001, 137, 59.
- 329 [19] R. Stocker, Proc. Natl. Acad. Sci. USA **1987**, 84, 5918.
- 330 [20] A.F. McDonagh, *Nat. Struct. Biol.* **2001**, *8*, 198.
- 331 [21] H. Kaur, M.N. Hughes, C.J. Green, P. Naughton, R. Foresti, R. Motterlini, FEBS Lett. 2003,
- 332 543, 113.
- 333 [22] A.C. Bulmer, K. Ried, J.T. Blanchfield, K.-H. Wagner, *Mut. Res.* **2008**, 658, 28.
- 334 [23] J.L. Baylor, M.W. Butler, J. Exp. Biol. 2019, 222, 1.
- 335 [24] G.Y. Kennedy, H.G. Vevers, Comp. Biochem. Physiol. 1973, 44B, 11.
- 336 [25] Y. Shi, J. Kovacs-Nolan, B. Jiang, R. Tsao, Y. Mine, J. Funct. Foods **2014**, 10, 35.
- 337 [26] Z. Zhu, A.T. Wilson, B.A. Luxon, K.E. Brown, M.M. Mathahs, S. Bandyopadhyay, et al.,
- 338 *Hepatology* **2010**, 52, 1897.
- 339 [27] J. Morales, A. Velando, R. Torres, *Behav. Ecol. Sociobiol.* **2011**, 65, 197.
- 340 [28] K.H. Falchuk, J.M. Contin, T.S. Dziedzic, Z.L. Feng, T.C. French, G.J. Heffron, et al., Proc. Natl
- 341 *Acad. Sci. USA* **2002**, 99, 251.
- 342 [29] H. Kayser, V. Wray, M. Nimtz, FEBS J. 2014, 281, 2366.
- 343 [30] X.-T. Wang, X.-M. Deng, C.-J. Zhao, J.-Y. Li, G.-Y. Xu, L.-S. Lian, C.-X. Wu, Poult. Sci. 2007,
- 344 86*,* 2236.
- 345 [31] H.C. Liu, M.C. Hsiao, Y.H. Hu, S.R. Lee, W.T.K. Cheng, Asian-Aust. J. Anim. Sci. 2010, 23,
- 346 162.
- 347 [32] J. Morales, S. Ruuskanen, T. Laaksonen, T. Eeva, R. Mateo, E. Belskii, E. et al., J. Avian Biol.
- **2013**, 44, 111.
- [33] J. Gautron, Y. Nys, in: *Bioactive egg compounds*, Springer, Berlin **2007**, pp. 103-108.
- 350 [34] S.G. Tullet, *Comp. Biochem. Physiol.* **1984**, 78A, 5.
- 351 [35] L. R. Milgrom, M.J. Warren, in *The Colours of Life: An Introduction to the Chemistry of*
- 352 Porphyrins and Related Compounds (Ed: L.R. Milgrom), Oxford University Press, Oxford
- 353 **1997**, pp. 1-175.

- 354 [36] M.R. Lang, J.W. Wells, World. Poultry Sci. J. 1967, 43, 238.
- 355 [37] V.K. Kodali, S.A. Gannon, S. Paramasivam, S. Raje, T. Polenova, C. Thorpe, *PLoS One* 2011,
  356 6, 1.
- 357 [38] G.V. Marinetti, J.T. Bagnara, *Science* **1983**, 219, 985.
- 358 [39] J. Wiemann, T-R. Yang, P.N. Sander, M. Schneider, M. Engeser, S. Kath-Schorr, C.E. Müller,
- 359 P.M. Sander, *PeerJ* **2017**, 5, e3706.
- 360 [40] M.T. Hincke, J. Gautron, M. Panhéleux, J.M. Garcia-Ruiz, M.D. McKee, Y. Nys, Mater. Biol.
- **2000**, 19, 443.
- 362 [41] B.M. Freeman, M.A. Vince, in: *Development of the avian embryo*. Springer, Dordrecht
- **1974**, pp. 237-248.
- 364 [42] H. Watson, P. Salmón, C. Isaksson, J. Exp. Biol. 2018, 221, 1.
- 365 [43] Z. Wang, G. Meng, Y. Bai, R. Liu, Y. Du, L. Su, *BMC genomics* **2017**, 18, 725.
- 366 [44] R. Hargitai, C. Moskát, M. Bán, D. Gil, I. López-Rull, E. Solymos, J. Avian Biol. 2010, 41, 177.
- 367 [45] C. Navarro, T. Pérez-Contreras, J.M. Avilés, K.J. McGraw, J.J. Soler, J. Avian Biol. 2011, 42,
- 368 538.
- 369 [46] M.W. Butler, K.J. McGraw, Funct. Ecol. 2013, 27, 1176.
- 370 [47] H. Hoi, A. Darolová, J. Krištofík, M. Poláček, J. Majtán, M. Zeman, et al., Ethol. Ecol. Evol.
- **2019**, 31, 249.
- 372 [48] J. Sujiwo, D. Kim, J.-Y. Yoon, H. Kim, J.-S. Kim, S.-K. Lee, et al., *Korean J. Food Sci. An.* **2017**,
- 373 37, 181.
- 374 [49] D. Hanley, G. Heiber, D.C. Dearborn, *The Condor* **2008**, 110, 767.
- 375 [50] M.D. Shawkey, L. D'Alba, *Nature* **2019**, 570, E43.
- 376 [51] G.Y. Kennedy, H.G. Vevers, Comp. Biochem. Physiol. 1976, 55B, 117.
- 377 [52] T. Tamura, S. Fujii, J. Fac. Fish. Anim. Husb. **1967**, 7, 35.
- 378 [53] J. Morales, R. Torres, A. Velando, *Naturwissenschaften* **2010**, 97, 173.
- 379 [54] J. Moreno, E. Lobato, J. Morales, Ornis Fenn **2011**, 88, 51.

- 380 [55] J. Morales, A. Velando, J. Moreno, *Behav. Ecol. Sociobiol.* 2008, 63, 227.
- 381 [56] R. Hargitai, N. Boross, Z. Nyiri, Z. Eke, *Behav. Ecol. Sociobiol.* **2016**, 70, 2093.
- 382 [57] A.P. Martin, S. Palumbi, Proc. Natl. Acad. Sci. USA 1993, 90, 4087.
- 383 [58] I. Galván, J. Erritzøe, F. Karadaş, A.P. Møller, J. Comp. Physiol. B 2012, 182, 947.
- 384 [59] P. Monaghan, N.B. Metcalfe, R. Torres, *Ecol. Lett.* **2009**, 12, 75.
- 385 [60] N. Metcalfe, C. Alonso-Álvarez, *Funct. Ecol.* **2010**, 24, 984.
- 386 [61] D. Lack, Ibis **1958**, 100, 145.
- 387 [62] M. Schönwetter, *Handbuch der Oologie*, Akademie Verlag, **1960 1992**.
- 388 [63] A. Jagannath, R.F. Shore, L.A. Walker, P.N. Ferns, A.G. Gosler, J. Apppl. Ecol. 2008, 45, 133.
- 389 [64] D. Hanley, S. Doucet, J. Appl. Ecol. **2012**, 49, 1055.
- 390 [65] R. Mateo, G. Castells, A.J. Green, C. Godoy, C. Cristòfolb, J. Chromat. 2004, 810, 305.
- 391 [66] D.B. Thomas, M.E. Hauber, D. Hanley, G.I.N. Waterhouse, S. Fraser, K.C. Gordon, J. Exp.
- *Biol.* **2015**, 218, 2670.
- 393 [67] D. Hanley, T. Grim, P. Cassey, M.E. Hauber, *Biol. Lett.* **2015**, 11, 20150087.
- 394 [68] G. Maurer, S.J. Portugal, M.E. Hauber, I. Mikšík, D.G.D. Russell, P. Cassey, *Funct. Ecol*.
- **2015**, 29, 209.

**Table 1**. The two main mechanisms of action of eggshell biliverdin (BLV) according to the antioxidant hypothesis, the central assumptions and predictions of

the hypothesis and possible ways to test them.

Proposed specific action	Assumptions/predictions	Ways to test assumptions/predictions
BLV protects the shell membrane against oxidation	<ul> <li>BLV traces present in most bird species</li> <li>High permeability of BLV through the inner calcified layers</li> </ul>	<ul> <li>Apply analytical methods (e.g., layer-by-layer dissolution; Raman spectroscopy) to different shell layers<sup>[7, 30, 66]</sup></li> </ul>
-	• BLV more abundant when the embryo is more exposed to oxidation	<ul> <li>Explore BLV traces/concentration in species /populations that differ in growth strategies and exposure to pathogens and contaminants</li> </ul>
	<ul> <li>Higher antioxidant capacity of the shell membrane/egg contents when BLV is present/more abundant</li> </ul>	• Experimental manipulation of BLV at laying or of oxidative stress during embryo development; <sup>[31, 42]</sup> investigate differences in shell membrane/egg antioxidant capacity in relation to BVL <sup>[48]</sup>
Biliverdin traces are absorbed by the embryo as an antioxidant resource	<ul> <li>Additional to the above:</li> <li>BLV permeates the shell membrane</li> <li>BLV more abundant at later stages of embryo development</li> </ul>	<ul> <li>Analytical methods (see above)<sup>[7, 30, 66]</sup> applied to inner shell layers at different stages of development; assessment of biliverdin levels in the plasma of hatchlings after BLV manipulation at laying</li> </ul>

400 Legend to Figures

401 Figure 1. Structure of a typical avian eggshell showing both the previously reported extent of

402 biliverdin in the innermost calcified layers<sup>[30, 31]</sup> and the potential reach in the shell membrane

403 proposed in this essay. Reprinted from *World's Poultry Science Journal*, M.R. Lang & J.W. Wells,

- 404 A Review of eggshell pigmentation, page 240, Copyright (1987), and minimally modified with
- 405 permission from Taylor & Francis Ltd (<u>http://www.tandfonline.com</u>; license: 4792060147918)
- 406 and from Elsevier
- 407 (<u>https://www.sciencedirect.com/science/article/abs/pii/0300962984900835</u>; license number:

408 4784730822058), publisher of the original figure in *Comparative Biochemistry and Physiology* 

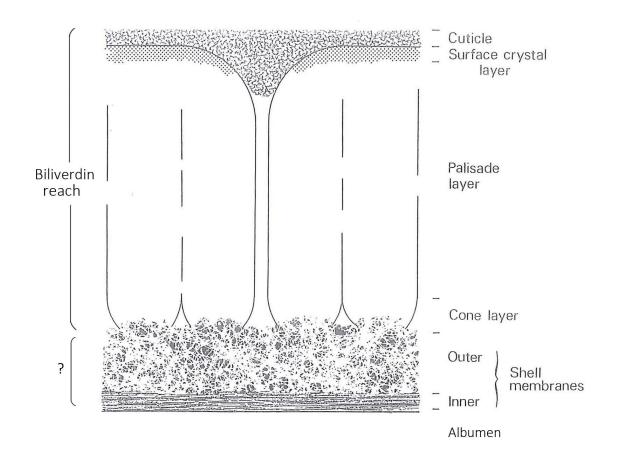
- 409 Part A: Physiology, 78 (1), S.G. Tullet, The porosity of avian eggshells, page 5, Copyright (1984)
- 410

411 Figure 2. a) Inner surface of a blue-footed booby (*Sula nebouxii*) eggshell on the hatching day.

412 Note the intense biliverdin-based blue-green colouration. The arrow points to remaining shell

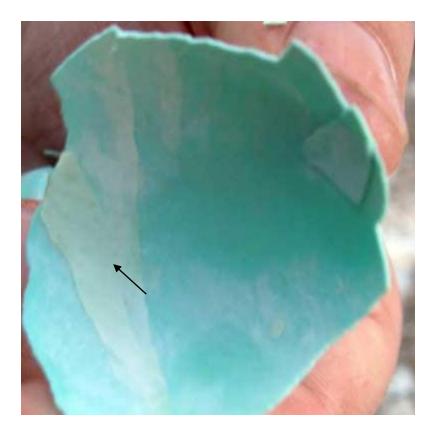
413 membrane that has a greenish tint; b) external surface of a blue-footed booby eggshell close to

414 hatching. Credit: J. Morales.





416 Fig. 1





418 Fig. 2a



- 421 Fig. 2b