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Advanced sample preparation to analyse selected emerging pollutants in Anguilla anguilla by liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS)

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INTRODUCTION

Samples of liver and muscle of Anguilla anguilla (Linnaeus, 1758) species were chosen as environmental matrices. This species is a catadromous fish, spending the majority of their life cycle in fresh water or estuaries and returning to the sea to spawn traveling thousands of kilometers to the Sargasso Sea. It is present in the Albufera lake, located in the Mediterranean coast of Valencia, Spain, where the rice farming and eels fishing are closely related.

Anguilla anguilla species has been selected because of their environmental and cultural interest in the área of Valencia. Furthermore, their high content in proteins and lipids (7-15% and 5-20% respectively in wet weight), pose a challenge for organic contaminants extraction.



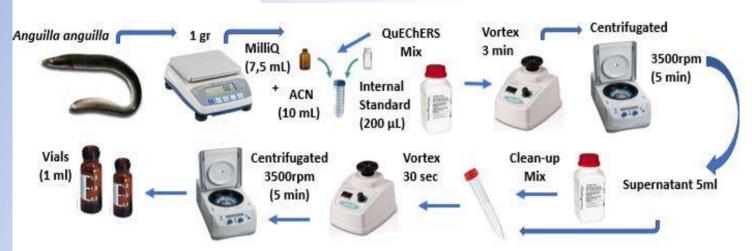
MATERIAL AND METHODS

Multiresidue method

Vildagliptin

108,9

61,4







ExionLC AD coupled to a Sciex QTRAP 6500⁺ mass spectrometer (Sciex, Concord, Ontario, Canada). **Target analytes:** -5 Pesticides Perfluoroalkyl -5 substances (PFASs)

- -9 Pharmaceuticals
- -2 Illicit drugs
- 2 transitions "lon precursor \rightarrow lon product" per compound.

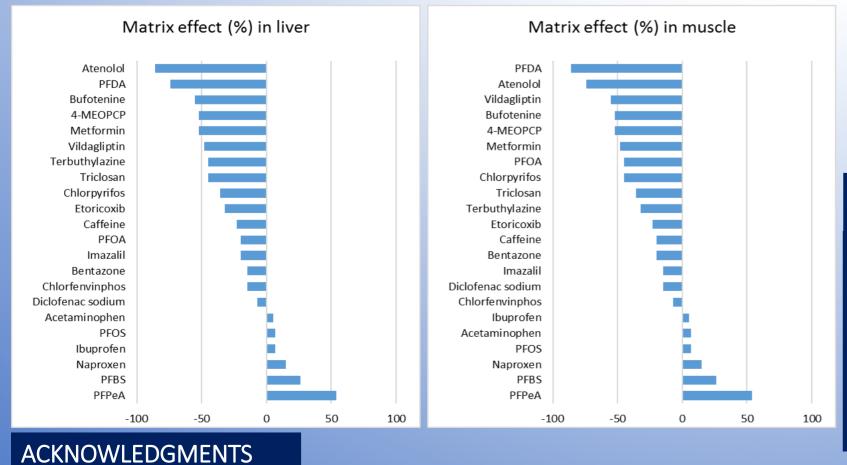
Extraction was based on QuEChERs. The dispersive solid phase extraction (dSPE) Enhanced Matrix Removal (EMR-lipid), especially developed for lipid removal was applied for clean-up.

RESULTS AND DISCUSSION

Recoveries	s (%) of the targe	et compounds		Differences between compounds accumulated in muscle and liver				
Recoveries (%)	Liver 50 ng/g 500 ng/g	Muscle 50 ng/g 500 ng/g	Chlorphyrifos STANDARD	*16.911 * ¹⁰ ³ , Chlorphyrifos LIVER	* 16.911	1 x10 ⁴ Chlorphyrifos MUSCLE		

					8.
4MeO-PCP	9,0	2,6	15,7	8,6	3- 7-
Acetaminophen	115,6	92,2	110,3	85,1	2,5- 6- 0,8-
Atenolol	123,2	120,4	111,2	96,3	2- 5- 0,7- 0,6-
Bentazone	-	60,4	-	53,4	1,5- 3- 0,4-
Bufotenine	42,3	90,7	46,1	88,6	2- 0,5- 1-
Caffeine	148,6	123,6	45,6	51,0	
Chlorfenvinphos	95,3	81,1	123,1	116,4	1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 1 Counts vs. Acquisition Time (min) Counts vs. Acquisition Time (min)
Chlorpyrifos	100,3	81,6	132,4	144,6	
Diclofenac	132,5	89,9	130,6	92,3 [×]	x10 ⁴ PFDA STANDARD
Etoricoxib	91,0	61,0	85,1	70,6	2,4- 2,2- 1,2-
Ibuprofen	124,5	112,9	100,5	98,6	2- 1,8- 1-
Imazalil	99,9	89,5	89,4	81,3	1,6- 1,4- 0,8- 0,7-
Naproxen	86,7	90,8	80,1	85,3	1,2- 1- 0,6-
PFBS	79,9	111,9	84,3	99,6	0,8- 0,6- 0,4-
PFDA	132,4	100,9	142,3	136,4	0,4- 0,2- 0,1-
PFOA	139,5	116,8	135,4	129,2	0
PFOS	120,8	98,8	110,3	99,4	Counts vs. Acquisition Time (min) Counts vs. Acquisition Time (min) Counts vs. Acquisition Time (min)
PFPeA	89,9	98,7	90,3	81,2	Results of samples from a bioacummulation study, showed differences between the compo
Terbuthylazine	110,2	95,9	100,3	92,1	accumulated in muscle and liver. While compounds such us chlorphyrifos, vildagliptin, PFDA, F
Triclosan	68,5	87,9	88,3	81,4	accumulated in mascle and iven. While compounds such as emorphymos, viaughptin, r bry,

were present in both matrices.



129,9

132,6

The extraction procedure achieved satisfactory recoveries for all the compounds except 4-MeO-PCP. For both liver and muscle, repeatability was satisfactory (<22% RSDs) for most compounds, as well as reproducibility (<30% RSDs) and LODs ranged 0,55-25 ng g^{-1} w.w. Strong matrix effect was present in several compounds with values of <±50% for the 72% of the them.

1,3- ¹ 1.2-

CONCLUSIONS

QuEChERS along with EMR-Lipid dSPE is a promising extraction procedure for multi-residue approaches.

The validated method was successfully applied to the analysis incurred in liver and muscle of Anguilla anguilla samples, despite the complexity of the matrices, further demonstrating the utility of the method to detect emerging pollutants and for implementation in regulatory and commercial laboratories.

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