

Taxonomical considerations and molecular phylogeny of the closely related genera *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus* (Nematoda: Telotylenchinae), with one new and four known species from Iran

Manouchehr Hosseinvand¹, Ali Eskandari¹, Shahin Ganjkhanloo¹, Reza Ghaderi², Pablo Castillo³ and Juan E. Palomares-Rius³

¹ Department of Plant Protection, Faculty of Agriculture, University of Zanjan, 45371-38791, Zanjan, Iran

² Department of Plant Protection, School of Agriculture, Shiraz University, 71441-65186, Shiraz, Iran

³ Instituto de Agricultura Sostenible (IAS), Consejo Superior de Investigaciones Científicas (CSIC), Avenida Menéndez Pidal s/n, 14004 Córdoba, Campus de Excelencia Internacional Agroalimentario, ceiA3, Spain

Correspondence

Ali Eskandari, Department of Plant Protection, Faculty of Agriculture, University of Zanjan, 45371-38791, Zanjan, Iran

Email: eskandari.a@znu.ac.ir

Contributing authors

Manouchehr Hosseinvand (m.houseinvand@gmail.com); Shahin Ganjkhanloo (Sh.ganjkhanloo69@gmail.com); Reza Ghaderi (rghaderi@shirazu.ac.ir); Pablo Castillo (p.castillo@csic.es); Juan E. Palomares-Rius (palomaresje@ias.csic.es)

Running title: Review of *Tylenchorhynchus* and related genera

Keywords: 18S rRNA, 28S rRNA, *Bitylenchus parvulus* n. sp., Identification key, *Sauertylenchus maximus*, synonymy, *Tylenchorhynchus*.

Abstract

During several nematological surveys in cultivated and natural habitats in Khuzestan and Zanzan provinces of Iran, a new species, *Bitylenchus parvulus* n. sp., two new records for Iran namely *Tylenchorhynchus agri* and *T. graciliformis*, and a population of *Bitylenchus parvus* and one of *Sauertylenchus maximus* were recovered and characterised based upon morphological and molecular approaches. The new species is characterized by lip region with five to seven annuli, stylet 17.7 (17.0-18.5) μm long, sub-cylindrical tail narrowing abruptly near terminus, cuticle near anterior part of vulva wrinkled and post-rectal sac occupied whole of tail cavity. The phylogenetic analyses were carried out using molecular data from D2-D3 expansion segments of 28S rDNA for all studied species and the partial 18S rDNA for the new species. The representatives of *Bitylenchus* and *Sauertylenchus* formed distinct clades from *Tylenchorhynchus* members supporting the hypothesis in which *Bitylenchus* and *Sauertylenchus* could be considered as valid genera but rejecting the 'large-genus' concept for *Tylenchorhynchus*. Also, *S. ibericus* was proposed as a junior synonym of *S. maximus* based on the results from morphological and phylogenetic analysis. Furthermore, an identification key for all known species included in three genera *Bitylenchus*, *Tylenchorhynchus* and *Sauertylenchus* is presented herein. The number of transverse annuli on the lip region and presence/absence of post-rectal sac were considered as main diagnostic characters for classifying of the species into seven groups, and other morphological and morphometric characters were subsequently used for distinguishing species in each group.

Introduction

Nematode members of the genus *Tylenchorhynchus* Cobb, 1913, known under the common name "stunt nematodes", are one of the largest groups of plant-parasitic nematodes which are migratory ectoparasites of the roots of many plants, including various agricultural crops and native plants (Siddiqi, 2000; Handoo *et al.*, 2014). *Bitylenchus* Filipjev, 1934 was proposed as a subgenus of the genus *Tylenchus* Bastian, 1865 by Filipjev (1934) with *Tylenchus* (*Bitylenchus*) *dubius* (Bütschli, 1873) Filipjev, 1934 as its type species; however, it was synonymized with *Tylenchorhynchus* by Filipjev (1936). Some nematologists (Fortuner & Luc, 1987; Handoo, 2000; Geraert, 2011) considered *Bitylenchus* as a synonym of *Tylenchorhynchus*, but others recognized both as valid genera (Gomez-Barcina *et al.*, 1992; Siddiqi, 2000; Andrásy, 2007; Handoo *et al.*, 2014; Ghaderi *et al.*, 2014). The genus *Bitylenchus* was differentiated from *Tylenchorhynchus* in having areolated outer bands of lateral fields, a large post-anal intestinal sac containing intestinal granules and fasciculi, relatively more thickened cuticle at the female tail tip, and gubernaculum lacking a crest (Gomez-Barcina *et al.*, 1992). The genus *Sauertylenchus* Sher, 1974 is also similar to *Bitylenchus* except for the difference in the structure of the lip region which has an offset labial disc and the first labial annulus divided into six sectors and a relatively longer stylet (Sher, 1974; Siddiqi, 1986, 2000; Gomez-Barcina *et al.*, 1992).

Siddiqi (2000) listed 105 valid species under *Tylenchorhynchus*, 29 valid species under *Bitylenchus* and further five species under *Sauertylenchus*. Geraert (2011) considered 133 species under *Tylenchorhynchus* (the genus *Bitylenchus* was considered as its synonym) and only one species under the genus *Sauertylenchus* (*S. labiodiscus* Sher, 1974). Subsequently, *Tylenchorhynchus bambusi* Singh, Lal, Rathour & Ganguly, 2010; *B. hispaniensis* Handoo, Palomares-Rius, Cantalapiedra-Navarrete, Liébanas, Subbotin & Castillo, 2014; *B. capsicum* Zarina & Akhtar, 2014; *T. mediterraneus* Handoo, Palomares-Rius, Cantalapiedra-Navarrete, Liébanas, Subbotin & Castillo, 2014 and *T. iranensis* Azimi, Mahdikhani-Moghadam, Rouhani & Rajabi Memari, 2016 were described from India, Spain, Pakistan, Spain, and Iran, respectively. Sturhan (2014) transferred *Paratrophurus striatus* Castillo, Siddiqi & Gomez-Barcina, 1989 to the genus *Tylenchorhynchus* and named it *T. casigo* (Castillo, Siddiqi & Gomez-Barcina, 1989) Sturhan, 2014.

Recently, Handoo *et al.* (2014) studied relationships between the genera *Bitylenchus* and *Tylenchorhynchus* using integrative taxonomy. In this sense, the use of integrative taxonomy combined with the phylogenetically test of alternative hypotheses confirmed the separation of

these two genera, rejecting the Fortuner & Luc's conceptual view of *Tylenchorhynchus* as a large genus (Fortuner & Luc, 1987). Because of the few and difficult characters to study in this group of nematodes, the use of molecular markers for species identification in the genus is of high importance. The use of nuclear ribosomal RNA genes [D2–D3 expansion segments of the large ribosomal subunit (28S), internal transcribed spacer (ITS), and partial small ribosomal subunit (18S)] have been proven useful in the latest phylogenetical studies of this group of nematodes (Handoo *et al.*, 2014; Azizi *et al.*, 2016; Hosseinvand *et al.*, 2019a; Gharakhani *et al.*, 2019; Hosseinvand *et al.*, 2020).

Taxonomic position of certain genera incorporated into the large genus *Tylenchorhynchus*, particularly *Bitylenchus* and *Sauertylenchus* is still controversial despite several phylogenetic and morphological studies. Sufficient morphological information is not available on several known species, which make their identification a challenging task. So far, several identification keys have been given for species included in these genera, mostly under *Tylenchorhynchus sensu lato* (Allen, 1955; Brzeski, 1998; Brzeski & Dolinski, 1998; Handoo, 2000; Geraert, 2011). In the present study, we provided a revised and new updated key for species included in the genera *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus*. Moreover, we described a new species from *Bitylenchus* and provided additional morphological and molecular information on four other known species namely *B. parvus* (Allen, 1955) Jairajpuri, 1982; *Sauertylenchus maximus* (Allen, 1955) Siddiqi, 2000; *Tylenchorhynchus agri* Ferris, 1963 and *T. graciliformis* Siddiqi & Siddiqui, 1983.

Materials and methods

Taxonomic review of Bitylenchus, Sauertylenchus and Tylenchorhynchus

The original and several relevant published descriptions of the species included in the genera *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus* were studied in detail. Based on obtained information from these descriptions, as well as those given in the monograph by Geraert (2011), we provided an updated and revised identification key for all known species in these genera. For practical reasons, we prefer to include all species in a single key due to close morphological affinity of the genera and the problems arise when assigning a correct taxon for a given population. Besides the characters implemented in the key by Geraert (2011), we used new additional characters such as the number of tail annuli, areolation of the lateral field, and the shape of gubernaculum for obtaining a better resolution to separate closely related species.

The taxonomic position of the valid species and their synonyms are mainly based on Siddiqi (2000).

Sampling and morphological identification

Soil samples were collected from the rhizosphere of different wild plants in Khuzestan and Zanjan provinces of Iran. Nematodes were extracted by the tray method (Whitehead & Hemming, 1965), then killed and fixed by hot FPG (4: 1: 1, formaldehyde: propionic acid: glycerol), and processed to anhydrous glycerol (De Grisse, 1969). The nematodes were transferred to a drop of glycerol and a surrounding ring of paraffin wax on permanent slides and studied using a light microscope, equipped with a Dino-eye microscope eye-piece camera in conjunction with its Dino Capture version 2.0 software. Drawings were made through a drawing tube attached to a light microscope. Specimens were identified at species level using available identification keys (Geraert, 2011).

Molecular characterization

For molecular analyses, and in order to avoid mistakes in case of mixed populations in the same sample, two specimens from each sample were temporarily mounted in a drop of 1M NaCl containing glass beads (to avoid nematode crushing/damaging) to ensure confirming specimens with the unidentified populations. All necessary morphological and morphometric data (pictures and measurements) of the studies specimens in temporary slides were recorded using the abovementioned camera-equipped microscope. This was followed by DNA extraction from single individuals as described by Subbotin *et al.* (2000). The D2-D3 region was amplified using the D2A (5'-ACAAGTACCGTGAGGGAAAGTTG-3') and D3B (5'-TCGGAAGGAACCAGCTACTA-3') primers (Nunn, 1992). Finally, the 18S rRNA was amplified using primers 988F (5'-CTCAAAGATTAAGCCATGC-3'), 1912R (5'-TTTACGGTCAGAACTAGGG-3'), 1813F (5'-CTGCGTGAGAGGTGAAAT-3') and 2646R (5'-GCTACCTTGTTACGACTTTT-3') (Holterman *et al.*, 2006).

All PCR assays were carried out according to the conditions described by Archidona-Yuste *et al.* (2016). Then, the amplified PCR products were purified using ExoSAP-IT (Affimetrix, USB products) and used for direct sequencing on a DNA multicapillary sequencer (Model 3130XL genetic analyser; Applied Biosystems, Foster City, CA, USA), using the BigDye Terminator Sequencing Kit V.3.1 (Applied Biosystems, Foster City, CA, USA), at the Stab Vida sequencing facilities (Caparica, Portugal). The newly obtained sequences were

submitted to the GenBank database under the accession numbers indicated on the phylogenetic trees. 153
154

Phylogenetic analyses 155 156

D2-D3 segments of 28S rRNA and 18S rRNA sequences of different genera of Telotylenchinae 157
Siddiqi, 1960 were obtained from GenBank and used for phylogenetic reconstruction. 158
Outgroup taxa for each dataset were chosen following previous published studies (Handoo *et* 159
al., 2014; Azizi *et al.*, 2016; Hosseinvand *et al.*, 2019a). The D2-D3 region alignment consisted 160
of 54 sequences as ingroups and two sequences of *Psilenchus hilarulus* de Man, 1921 161
(KP313831) and *Coslenchus paramaritus* Hosseinvand, Eskandari, & Ghaderi, 2019b 162
(MK542004) as outgroups. The 18S rRNA alignment consisted of 30 sequences as in groups 163
and *Geocenamus quadrifer* (Andrássy, 1954) Brzeski, 1991 (AY993997) and *P. hilarulus* 164
(KX798728) as outgroups. The newly obtained sequences were edited and aligned with other 165
relevant sequences available in GenBank using Muscle alignment tool implemented in 166
MEGA7 program (Kumar *et al.*, 2016). The ambiguously aligned parts and divergent regions 167
were removed from the alignments using the online version of Gblocks 0.91b (Castresana, 168
2000) (http://molevol.cmima.csic.es/castresana/Gblocks_server.html). The best-fit model of 169
nucleotide substitution used for the phylogenetic analysis was statistically selected using 170
jModelTest 2.1.10 (Darriba *et al.*, 2012) with the Akaike Information Criterion (AIC). 171
Phylogenetic trees were generated with Bayesian inference method using MrBayes 3.2.6 172
(Huelsenbeck & Ronquist, 2001; Ronquist *et al.*, 2012). The best-fit model, the base frequency, 173
the proportion of invariable sites, and the gamma distribution shape parameters and substitution 174
rates in the AIC were then used in MrBayes for the phylogenetic analyses. The general time- 175
reversible model with invariable sites and a gamma-shaped distribution (GTR + I + G), for the 176
D2-D3 segments of 28S rRNA and 18S rRNA genes, was run with four chains for 2×10^6 177
generations. A combined analysis of the two genes was not undertaken due to some sequences 178
not being available for all species. The topologies were used to generate a 50 % majority-rule 179
consensus tree. Posterior probabilities (PP) are given on appropriate clades. Trees from all 180
analyses were visualized and saved using FigTree software V.1.4.3 181
(<http://tree.bio.ed.ac.uk/software/figtree/>) (Rambaut, 2014) and edited with Adobe® Acrobat® 182
XI Pro 11.0.1. 183

Results	186
<i>Key to the species of Bitylenchus, Sauertylenchus and Tylenchorhynchus</i>	187
All known species of these three genera are treated in a single identification key here (see discussion for justification). For practical reasons, we separate this complex nematode group in several morphotype groups, based on the main diagnostic characters (mainly the number of lip region annuli and presence/absence of post-rectal sac) used for species delineation in these genera.	188 189 190 191 192 193 194
Group 1. Lip region without annuli, smooth; post-rectal sac absent or very slightly overlapping rectum (presented in the original description of <i>T. amgi</i> but not drawn)	195 196
Group 2. The number of transverse annuli on lip region (excluding labial disc) usually 2 or 3, rarely 4; post-rectal sac absent or very slightly overlapping rectum	197 198
Group 3. The number of transverse annuli on lip region (excluding labial disc) usually 2 or 3, rarely 4; post-rectal sac present	199 200
Group 4. The number of transverse annuli on lip region (excluding labial disc) usually 4 or 5, rarely 6; post-rectal sac absent or very slightly overlapping rectum	201 202
Group 5. The number of transverse annuli on lip region (excluding labial disc) usually 4 or 5, rarely 6; post-rectal sac present	203 204
Group 6. The number of transverse annuli on lip region (excluding labial disc) mostly 6-10, fine; post-rectal sac absent or very slightly overlapping rectum	205 206
Group 7. The number of transverse annuli on lip region (excluding labial disc) mostly 6-10, fine; post-rectal sac present	207 208 209
<i>Dichotomous keys for species identification in proposed morphospecies groups:</i>	210
Group 1.	211 212
1. Average of stylet length below 15 μm <i>T. cynodoni</i> Kumar, 1981	213
- Average of stylet length 17-22 μm 2	214
- Average of stylet length 23 μm <i>T. robustus</i> Thorne & Malek, 1968 (lip region described as having no visible transverse annuli; however, this species may have 6-10 mostly fine annuli, and belongs to the group 6)	215 216 217
2. Average value of c' ratio 2 to 3.5 3	218

- Average value of c' ratio nearly 4 or more	6	219
3. Average of cuticular annuli on the ventral side of tail below 30	4	220
- Average of cuticular annuli on the ventral side of tail more than 40	<i>T. sudanensis</i> (Decker, Yassin & El-Amin, 1975) Castillo, Siddiqi & Gomez-Barcina, 1989	221 222
4. Lip region distinctly offset; tail sub-cylindrical	<i>T. oryzae</i> Kaul & Waliullah, 1995	223
- Lip region continuous to slightly offset; tail cylindrical	5	224
5. Lip region hemispherical, basal bulb saccate; stylet knobs posteriorly or laterally directed	<i>T. parasudanensis</i> (Elbadri, Moon, Lee & Choo, 2010) Geraert, 2011	225 226
- Lip region conoid truncate, basal bulb pyriform; stylet knobs anteriorly directed	<i>T. tuberosus</i> Zarina & Maqbool, 1994	227 228
6. Cuticular annuli on the ventral side of tail below 25 in average	7	229
- Cuticular annuli on the ventral side of tail more than 35 in average	8	230
7. Stylet with anteriorly directed basal knobs; cuticular annuli 1.0-1.2 μm apart	<i>T. amgi</i> Kumar, 1981	231 232
- Stylet with posteriorly directed basal knobs; cuticular annuli 1.5-2.4 μm apart	<i>T. leviterminalis</i> Siddiqi, Mukherjee & Dasgupta, 1982	233 234
8. Lip region hemispherical, slightly offset, tail cylindrical, very slightly arcuate ventrally	<i>T. ooti</i> Siddiqi, 2008	235 236
- Lip region conoid-truncate, continuous, tail cylindrical to sub-clavate with characteristic shallow depression on its dorsal side.....	<i>T. microcephalus</i> Siddiqi & Patel, 1990	237 238
		239
Group 2.		240
		241
1. Average of stylet length below 18 μm	2	242
- Average of stylet length 18-22 μm	17	243
- Average of stylet length more than 23 μm	28	244
2. Cuticular annuli on the ventral side of tail more than 25 in average	3	245
- Cuticular annuli on the ventral side of tail below 25 in average	5	246
3. Average of body length more than 800 μm	<i>T. handooi</i> Khan, 2004	247
- Average of body length less than 700 μm	4	248
4. c' = 2.7-3.2; stylet 16-17 μm	<i>T. karnalensis</i> Saha, Singh, Lal & Kaushal, 2002	249
- c' = 4; stylet 14-16 μm	<i>T. delhiensis</i> Chawla, Bhamburkar, Khan & Prasad, 1968	250
5. Lip region distinctly offset	6	251

- Lip region continuous or slightly offset 7	252
6. Basal bulb saccate and offset from intestine; cuticular annuli 1.6-1.8 μm wide at mid-body; tail slightly ventrally curved; $c = 17-24$ <i>T. microconus</i> Siddiqi, Mukherjee & Dasgupta, 1982	253 254 255
- Basal bulb pyriform with slight overlapping, cuticular annuli 2.7 μm wide at mid-body; tail straight; $c = 14-17$ <i>T. aspericutis</i> Knobloch, 1975	256 257
7. Tail cylindrical 8	258
- Tail conical to sub-cylindrical 9	259
8. Lateral field marked by peculiar oblique striae <i>T. gossypii</i> Nasira & Maqbool, 1996	260
- Lateral field not marked by such striae <i>T. lucknowensis</i> Singh & Jain, 1983	261
9. Average of stylet length below 14 μm <i>T. tritici</i> Golden, Maqbool & Handoo, 1987	262
- Average of stylet length more than 14 μm 10	263
10. $c' = 3.9$ <i>T. elamini</i> Elbadri, Moon, Lee & Choo, 2010	264
- $c' = 2.0-3.5$ 11 (including 8 similar species)	265
11. Average of body length less than 600 μm 12	266
- Average of body length more than 600 μm 16	267
12. Stylet knobs anteriorly directed 13	268
- Stylet knobs posteriorly directed 14	269
13. Lip region with 2 transverse annuli; $c' = 2.3-2.8$; average of stylet length more than 17 μm <i>T. varicaudatus</i> Singh, 1971	270 271
- Lip region with 3-4 transverse annuli; $c' = 3.0-3.4$; average of stylet length less than 17 μm <i>T. qasimii</i> Ramzan, Handoo & Fayyaz, 2008	272 273
14. Tail annuli 20-24 <i>T. iarius</i> Saha, Gaur & Lal, 1998	274
- Tail annuli 11-20 15	275
15. $c' = 2.3-2.9$ <i>T. rudis</i> Siddiqi, 2008	276
- $c' = 3.2$ <i>T. haki</i> Fotedar & Mahajan, 1971	277
16. Tail annuli 13-16; cuticular annuli about 3 μm wide at mid-body; spicules with distinct terminal notch <i>T. mashhoodi</i> Siddiqi & Basir, 1959	278 279
- Tail annuli 17-21; cuticular annuli 2.6 μm wide at mid-body; spicules without terminal notch <i>T. bohrrensis</i> Gupta & Uma, 1980	280 281
- Tail annuli 23; cuticular annuli about 2.2 μm wide at mid-body; spicules without terminal notch <i>T. elegans</i> Siddiqi, 1961	282 283
17. Tail annuli more than 25 in average 18	284

- Tail annuli less than 25 in average	21	285
18. Tail clavate; c' = 4; tail with distinct hyaline near 10 µm thick	<i>T. clavicaudatus</i>	286
Seinhorst, 1963		287
- Tail conical to sub-cylindrical; c' below 3.5; tail without distinct hyaline	19	288
19. Lip region distinctly offset	<i>T. sanwali</i> Kumar, 1982	289
- Labial region continuous to slightly offset	20	290
20. Tail conical to almost funnel-shaped with narrow terminus	<i>T. paratriversus</i> Brzeski,	291
1992		292
- Tail regularly conical with hemispherical terminus	<i>T. thermophiles</i> Golden, Baldwin &	293
Mundo-Ocampo, 1995		294
21. Tail clavate	22	295
- Tail conical to sub-cylindrical	23 (two similar species)	296
22. Pharynx 115-160 µm; spicules 19-23 µm; tail hyaline 7-13 µm thick	<i>T. crassicaudatus</i>	297
Williams, 1960		298
- Pharynx 105-120 µm; spicules 21-26 µm; tail hyaline 5-8 µm thick	<i>T. kegasawai</i>	299
Minagawa, 1995		300
23. Tail terminus striated; longitudinal striae present on anterior end	<i>T. claytoni</i> Steiner,	301
1937		302
- Tail terminus smooth; longitudinal striae absent	24	303
24. Male not found, even in populations with very high number of females	<i>T. annulatus</i>	304
(Cassidy, 1930) Golden, 1971		305
- Males usually present	25	306
25. Lip framework refractive; stylet knobs concave, 5 µm across	<i>T. ancorastyletus</i>	307
Ivanova, 1983		308
- Lip framework non-refractive; stylet knobs usually not concave, 4 µm or less	26	309
26. c' = 2.0; stylet 19-23 µm; stylet knobs posteriorly directed	<i>T. mexicanus</i> Knobloch &	310
Laughlin, 1973		311
- c' = 2.7-3.0; stylet 17-20 µm; stylet knobs laterally or anteriorly directed	27 (three similar	312
species)		313
27. Lip region annuli three; body length 620-710 µm; spicules 20-22 µm	<i>T. badliensis</i>	314
Saha & Khan, 1982		315
- Lip region annuli two; body length 570-630 µm; spicules 23-24 µm	<i>T. coffeae</i> Siddiqi &	316
Basir, 1959		317

- Lip region annuli 2; body length 580-660 μm ; spicules 24-26 μm	<i>T. musae</i> Kumar, 1981	318
28. Body length 700-800 μm ; tail hyaline very thick (12-16 μm); spicules 21-25 μm	<i>T. clavus</i> Khan, 1990	319 320
- Body length 800-1000 μm ; tail hyaline normal (below 8 μm thick); spicules 28-33 μm	<i>T. silvaticus</i> Ferris, 1963	321 322 323
Group 3.		324
1. Average of stylet length less than 18 μm ; spicules 24-28 μm	<i>T. microcephaloides</i> (Zarina, Siddiqi & Shahina, 2004) Geraert, 2011	326 327
- Average of stylet length between 18 and 22 μm ; spicules shorter than 25 μm	2	328
- Average of stylet length more than 24 μm ; spicules shorter than 25 μm	<i>T. botrys</i> Siddiqi, 1985	329 330
2. Cuticular annuli on the ventral side of tail below 25 in average; tail terminus smooth (except in <i>T. kashmirensis</i>)	3	331 332
- Cuticular annuli on the ventral side of tail 25-35 in average; tail terminus striated	<i>B. equatorialis</i> Talavera & Siddiqi, 1995	333 334
- Cuticular annuli on the ventral side of tail more than 35 in average; tail terminus smooth	<i>T. neoclavicaudatus</i> Mathur, Sanwal & Lal, 1979	335 336
3. Lip region distinctly offset; tail terminus striated	<i>T. kashmirensis</i> Mahajan, 1974	337
- Lip region continuous or slightly offset; tail terminus smooth	4	338
4. Tail cylindrical to clavate	5 (two similar species)	339
- Tail conical to sub-cylindrical	6	340
5. Tail cylindrical to slightly clavate	<i>T. nudus</i> Allen, 1955	341
- Tail distinctly clavate.....	<i>T. bambusi</i> Singh, Lal, Rathour & Ganguly, 2010	342
6. $c' = 2.8-4.1$; post-rectal sac filling less than 20% of tail length	<i>T. alami</i> Shaw & Khan, 1996	343 344
- $c' = 2.1-2.6$; post-rectal sac filling about 45-70% of tail length	7	345
7. Spicules 21-25 μm ; stylet knobs with concave anterior surfaces	<i>T. agri</i> Ferris, 1963	346
- Spicules 18 μm ; stylet knobs rounded	<i>T. ewingi</i> Hopper, 1959	347 348
Group 4.		349
		350

1. Average of stylet length below 18 μm 2 (some species have a stylet near to 18 μm ; comparison with those bearing 18-22 μm stylet is recommended)	351
- Average of stylet length 18-22 μm 15	353
- Average of stylet length more than 22 μm 28	354
2. Cuticular annuli on the ventral side of tail below 30 in average 3	355
- Cuticular annuli on the ventral side of tail more than 30 in average 11	356
3. Lip region distinctly offset 4	357
- Lip region continuous or slightly offset 5	358
4. Tail terminus smooth; spicules 24-26 μm <i>T. variannulatus</i> Siddiqi, 2008	359
- Tail terminus striated; spicules 14 μm <i>T. leucaenus</i> Azmi, 1991	360
- Tail terminus striated; male unknown <i>T. crotoni</i> Pathak & Siddiqui, 1997	361
5. Body ventrally contracted behind vulva <i>T. contractus</i> Loof, 1964	362
- Body not contracted behind vulva 6	363
6. Tail with broadly rounded terminus; lateral field strongly areolated; gubernaculum hammer- shaped <i>T. aerolatus</i> Tobar Jiménez, 1970	364
- Tail with narrower terminus; lateral field not strongly areolated; gubernaculum simple 7	366
7. Spermatheca non-functional; males absent or very rare <i>T. clarus</i> Allen, 1955 (<i>T.</i> <i>variannus</i> Mavlyanov, 1978 is a possible synonym)	367
- Spermatheca functional; males usually present 8	369
8. Stylet length 11-13 μm <i>T. nordiensis</i> Khan & Nanjappa, 1974	370
- Stylet length 15-19 μm 9 (three similar species; also they should be compared with <i>T.</i> <i>zeae</i> and <i>T. chirchikensis</i> which have only slightly longer stylet)	371
9. Tail terminus flattened <i>T. shivanandi</i> Shaw & Khan, 1992	373
- Tail terminus hemispherical 10	374
10. Spicules 18-21 μm <i>T. brassicae</i> Siddiqi, 1961	375
- Spicules 21-26 μm <i>T. spinaceai</i> Singh, 1976	376
11. Tail terminus striated <i>T. mangiferae</i> Luqman & Khan, 1986	377
- Tail terminus smooth 12	378
12. Cuticle on either side of vulva irregular <i>B. usmanensis</i> (Khurma & Mahajan, 1988) Siddiqi, 2000	379
- Cuticle near vulva normal 13	381
13. Lip region hemispherical; vulva with lateral membrane <i>T. fatimae</i> Khan, Saeed & Akhter, 2004	382
	383

- Lip region sunken and dome-shaped; vulva without lateral membrane	14	384
14. Lip region with 5-6 transverse annuli; lateral field areolated	<i>B. quaidi</i> (Golden, Maqbool & Handoo, 1987) Siddiqi, 2000	385 386
- Lip region with 4 transverse annuli; lateral field apparently not areolated	<i>T. minutus</i> Karapetjan, 1979	387 388
15. Cuticular annuli on the ventral side of tail below 25 in average	16	389
- Cuticular annuli on the ventral side of tail between 25 and 35 in average	25	390
- Cuticular annuli on the ventral side of tail more than 35 in average	27	391
16. Tail cylindrical to clavate	17	392
- Tail conical to sub-cylindrical	19	393
17. Body length 730-830 μm ; $c' = 3.0-3.8$	<i>T. idahoensis</i> Siddiqi, 2008	394
- Body length 570-710 μm ; $c' = 2.5-3.0$	18	395
18. Tail clavate; tail annuli about 15	<i>T. dactylurus</i> Das, 1960	396
- Tail cylindrical; tail annuli 8-10	<i>T. georgiensis</i> Eliashvili, 1971	397
19. Lip region distinctly offset	20	398
- Lip region continuous or slightly offset	22	399
20. Tail with very narrow terminus	<i>T. aduncus</i> de Guiran, 1967	400
- Tail terminus not so narrow	21	401
21. Body length 460-660 μm ; tail terminus striated	<i>T. ismaili</i> Azmi & Ahmad, 1991	402
- Body length 670-830 μm ; tail terminus smooth	<i>T. graciliformis</i> Siddiqi & Siddiqi, 1983	403
22. Tail with very narrow terminus	<i>T. paulettae</i> Bloemers & Wanless, 1998	404
- Tail terminus not so narrow	23	405
23. Tail terminus striated; spicules 26-29 μm	<i>T. irregularis</i> Wu, 1969	406
- Tail terminus smooth; spicules less than 25 μm	24 (three similar species)	407
24. Body length 530-640 μm	<i>T. zaeae</i> Sethi & Swarup, 1968	408
- Body length 620-700 μm	<i>T. chirchikensis</i> Mavlyanov, 1978	409
- Body length 700-780 μm	<i>T. projectus</i> Khan, 1990	410
25. Body length 520-590 μm ; lip region with 5 annuli; stylet knobs rounded	<i>T. ebriensis</i> Seinhorst, 1963	411 412
- Body length 600-760 μm ; lip region with 4 annuli; stylet knobs with concave anterior end	26	413 414
26. Tail terminus narrow, striated	<i>T. eremicolus</i> Allen, 1955	415
- Tail terminus hemispherical, smooth	<i>T. digitatus</i> Das, 1960	416

27. Tail with finger-shaped outgrowth; $c' = 2.2$	<i>T. erevanicus</i> Karapetjan, 1979	417
- Tail with distinct hyaline (9-12 μm); $c' = 5$	<i>T. areoterminalis</i> Siddiqi, 2008	418
- Tail normal; $c' = 2.8$	<i>T. hordei</i> Khan, 1972	419
28. Tail with elongated-conical with narrow terminus; tail annuli more than 30	<i>T. antarcticus</i> Wouts & Sher, 1981	420
		421
- Tail bifurcated; tail annuli 26-27	<i>T. bicaudatus</i> Khakimov, 1973	422
- Tail suddenly constricted ventrally before middle, distal part hook-shaped with a bursa-like structure formed by lateral fields; tail annuli less than 25	<i>T. bursifer</i> Loof, 1960	423
		424
- Tail conical; tail annuli 15-22	<i>T. cylindricus</i> Cobb, 1913	425
- Tail almost cylindrical; tail annuli near 25	<i>S. velatus</i> (Sauer & Annells, 1981) Siddiqi, 2000	426
		427
- Tail almost cylindrical; tail annuli 14-15	<i>T. tarjani</i> Andr�ssy, 1969	428
		429
Group 5.		430
		431
1. Average of stylet length below 18 μm	2	432
- Average of stylet length more than 18 μm	7	433
2. Tail annuli more than 40	<i>T. paracanalisis</i> Khan, 1991	434
- Tail annuli less than 40	3	435
3. Average of stylet length below 15 μm ; vulva without distinct epiptygma	4	436
- Average of stylet length more than 15 μm ; vulva without epiptygma	<i>B. capsicumii</i> Zarina & Akhtar, 2014	437
		438
- Average of stylet length more than 15 μm ; vulva with distinct epiptygma	<i>B. queirozi</i> (Monteiro & Lordello, 1976) Jairajpuri, 1982	439
		440
4. Cuticle posterior to vulva with irregular undulations	<i>B. ventrosignatus</i> (Tobar Jim�nez, 1969) Jairajpuri, 1982	441
		442
- Cuticle posterior to vulva normal	5	443
5. Tail terminus smooth	<i>T. subcylindricus</i> Singh & Jain, 1982	444
- Tail terminus striated	6	445
6. Tail conical with narrow terminus; spicules 20-23 μm	<i>B. swarupi</i> (Singh & Khera, 1978) Jairajpuri, 1982	446
		447
- Tail almost cylindrical with hemispherical terminus; spicules 23-26 μm	<i>B. cuticaudatus</i> (Ray & Das, 1983) Siddiqi, 1986	448
		449

7. Tail annuli less than 25	<i>T. kamlae</i> Shaw & Khan, 1996	450
- Tail annuli 25-35	8	451
- Tail annuli 35-45	<i>B. bryobius</i> (Sturhan, 1966) Jairajpuri, 1982	452
8. Body length 480-800 µm; spicules 18-26 µm	9	453
- Body length 830-1300 µm; spicules 33-35 µm	<i>S. pratensis</i> (Gomez-Barcina, Siddiqi & Castillo, 1992) Siddiqi, 2000	454 455
9. Lip region usually with 4 transverse annuli	<i>B. colombianus</i> Siddiqi, 1985	456
- Lip region with 5 or 6 transverse annuli	10 (two similar species)	457
10. Stylet length 16-20 µm	<i>B. iphilus</i> Minagawa, 1995	458
- Stylet length 19-21 µm	<i>B. malinus</i> (Lin, 1992) n. comb.	459 460
Group 6.		461
1. Average of stylet length below 17 µm	2 (two similar species)	462 463
- Average of stylet length 18-22 µm	3	464
- Average of stylet length more than 22 µm	6	465
2. Body length 580-700 µm; stylet 16-17 µm	<i>T. latus</i> Allen, 1955	466
- Body length 710-730 µm; stylet 14-16 µm	<i>T. persicus</i> Sultan, Singh & Sakhuja, 1991	467
3. Tail annuli less than 20	<i>T. shimizui</i> Talavera, Watanabe & Mizukubo, 2002	468
- Tail annuli 20-35	4	469
- Tail annuli more than 35	5	470
4. Lip region continuous; c' = 1.7-2.3; spicules 29-34 µm	<i>T. casigo</i> (Castillo, Siddiqi & Gomez-Barcina, 1989) Sturhan, 2014	471 472
- Lip region offset; c' = 3.4-5.0; spicules 20-22 µm	<i>T. teres</i> (Khan & Darekar, 1979) Siddiqi, 1986	473 474
5. Lip region distinctly offset; body length 800-1200 µm; tail regularly sub-cylindrical	<i>T. mediterraneus</i> Handoo, Palomares-Rius, Cantalapiedra-Navarrete, Liébanas, Subbotin & Castillo, 2014	475 476 477
- Lip region continuous; body length 750 µm; tail contracted behind anus	<i>T. manubriatus</i> Litvinova, 1946	478 479
6. Tail annuli 16-30; vulva with protruding double epiptygma	<i>T. siccus</i> Nobbs, 1990	480
- Tail annuli more than 30; epiptygma absent	7	481

7. Tail conical ending to a very narrow terminus; stylet 23-26 μm	<i>T. tenuicauda</i> Wouts & Sher, 1981	482
- Tail cylindrical ending to a hemispherical terminus; stylet 26-31 μm	<i>T. kegenicus</i> Litvinova, 1946	483
		484
		485
		486
Group 7.		487
1. Average of stylet length 10-11 μm	<i>B. depressus</i> Jairajpuri, 1982	488
- Average of stylet length 12-15 μm	2	489
- Average of stylet length 15-19 μm	6	490
- Average of stylet length 19-22 μm	10	491
- Average of stylet length 22-30 μm	13	492
- Average of stylet length 35-40 μm	<i>S. labiodiscus</i> Sher, 1974	493
2. Cuticle near vulva irregular	3	494
- Cuticle near vulva normal	4	495
3. Longitudinal striae present on anterior end (pharyngeal region); spicules 23-27 μm	<i>B. brevilineatus</i> (Williams, 1960) Jairajpuri, 1982	496
		497
- Longitudinal striae absent; spicules 21-23 μm	<i>B. goffarti</i> (Sturhan, 1966) Jairajpuri, 1982	498
4. Lip region continuous	<i>B. mediocris</i> Talavera & Siddiqi, 1995	499
- Lip region offset	5	500
5. Lateral field not areolated; lip region with 6-7 transverse annuli; spermatheca with spheroid sperm	<i>B. vulgaris</i> (Upadhyay, Swamp & Sethi, 1972) Jairajpuri, 1982	501
		502
- Lateral field areolated; lip region with 7-9 transverse annuli; spermatheca lacking sperm	<i>B. singularis</i> Siddiqi & Sharma, 1994	503
		504
6. Lip region offset	7	505
- Lip region continuous	9	506
7. Lip region with 5-7 transverse annuli; epiptygma present; intestinal fasciculi absent; tail sub-cylindrical, abruptly narrowing near terminus	<i>B. parvulus n. sp.</i>	507
		508
- Lip region with 6-10 transverse annuli; epiptygma absent; intestinal fasciculi present; tail sub-cylindrical to cylindrical, with normal hemispherical terminus	8	509
		510
8. $c' = 2.2-3.7$; spermatheca filled with sperm, males common.....	<i>B. dubius</i> (Bütschli, 1873) Filipjev, 1934	511
		512
- $c' = 3.1-4.4$; spermatheca empty, males absent	<i>B. tobari</i> (Sauer & Annells, 1981) Siddiqi, 1986	513
		514

9. Gubernaculum with crest, distinctly curved proximally	<i>T. iranensis</i> Azimi, Mahdikhani- Moghadam, Rouhani & Rajabi Memari, 2016	515 516
- Gubernaculum lacking crest, simple	<i>B. parvus</i> (Allen, 1955) Jairajpuri, 1982 (three other species, <i>B. teeni</i> (Hashim, 1984) Siddiqi, 1986, <i>B. huesingi</i> (Paetzold, 1958) Jairajpuri, 1982 and <i>B. hispaniensis</i> Handoo, Palomares-Rius, Cantalapiedra-Navarrete, Liébanas, Subbotin & Castillo, 2014 are very similar)	517 518 519 520
10. Tail annuli less than 30; c' below 2	<i>B. kidwaii</i> (Rashid & Heyns, 1990) Siddiqi, 2000	521
- Tail annuli more than 30; c' more than 2	11	522
11. Post-rectal sac covers one-third of tail length at maximum	<i>B. serranus</i> Gomez-Barcina, Siddiqi & Castillo, 1992	523 524
- Post-rectal sac covers almost the total tail length	12	525
12. Stylet knobs anteriorly directed; c' = 2.7; Stylet knobs anteriorly directed	<i>B. canalis</i> (Thorne & Malek, 1968) Jairajpuri, 1982	526 527
- Stylet knobs posteriorly directed; c' = 3-4; Stylet knobs posteriorly directed	<i>B. natalensis</i> (Kleynhans, 1984) Siddiqi, 1986	528 529
13. Labial region distinctly offset; average of stylet length 25 µm; vulva with epiptygma	<i>S. pamiricus</i> (Ivanova, 1989) n. comb.	530 531
- Labial region slightly offset; average of stylet length below 25 µm; vulva without epiptygma	14	532 533
14. Tail terminus annuli finer than other tail annuli	<i>S. maximus</i> (Allen, 1955) Siddiqi, 2000	534
- Tail terminus annuli as wide as other tail annuli	<i>S. ibericus</i> (Mahajan & Nombela, 1987) Siddiqi, 2000	535 536 537 538
<i>Descriptions and observations on Iranian populations</i>		539
A total of 81 individuals including 62 female and 19 male specimens belonging to seven species were used for morphological analyses. Description of the new species and some species are given and observations are made on other species.		540 541 542 543
<i>Bitylenchus parvulus</i> n. sp.		544
ZooBank (zoobank.org) identifier: urn:lsid:zoobank.org:pub: 9089459F-7E88-4039-80EB- F0B65B600CFC.		545 546
(Figs 1, 2 & 3)		547

<i>Measurements</i>	548
The morphometrics of <i>B. parvulus</i> n. sp. are present in Table 1.	549
	550
<i>Description</i>	551
<i>Female</i> . Medium sized, ventrally arcuate to J-shaped after heat fixation. Lateral field with	552
four incisures, outer bands areolated, 29 (21-37)% of the corresponding body diameter. Cuticle	553
annuli delicate, 1.0 (0.5-1.5) μm wide at mid-body. Cuticle just anterior to vulva with irregular	554
undulations (wrinkling) at its ventral side. Labial framework slightly sclerotized; lip region	555
slightly offset from body contour by a distinct depression, with five to seven annuli, 3.6 (3.0-	556
4.0) μm high and 7.4 (6.5-8.5) μm wide. Amphidial apertures indistinct. Stylet well-developed,	557
conus 9.1 (8.5-9.5) μm , about one half of stylet length, stylet knobs rounded, slightly sloping	558
backward, 3.7 (3.0-4.5) μm in width. Dorsal pharyngeal gland orifice (DGO) 2.8 (2.0-3.5) μm	559
from stylet knobs. Procorpus cylindrical, as long as isthmus, median bulb well developed,	560
ovate, with distinct valve, 17.5 (15.0-19.5) μm long and 11.6 (10.0-13.5) μm wide, occupying	561
48 (57-66)% of the corresponding body diameter. Nerve ring at anterior half of isthmus, 91	562
(80-101) μm from anterior end. Excretory pore position varies from posterior end of isthmus	563
to anterior end of basal bulb. Hemizonid 5-6 annuli anterior to excretory pore, 109 (96-119)	564
μm from anterior end. Basal bulb pocket-shaped, offset from intestine, 27 (22-34) μm in length	565
and 13.5 (10.0-15.5) μm in width. Cardia small. Intestine generally without distinct fasciculi.	566
Post-rectal sac occupies almost entire tail cavity. Reproductive system didelphic-amphidelphic,	567
gonads straight, oocytes arranged in single row, spermatheca non-offset, usually without	568
sperm, uterus long, vagina perpendicular to the body axis, 8.7 (7.5-10.5) μm long, occupying	569
24 (22-26) % of the vulval body width. Vulva near mid-body, as a transverse slit, with small	570
epiptygma. Phasmids at 42 (31-50)% of tail length. Tail sub-cylindrical, often dorsally convex,	571
usually narrowing abruptly near terminus giving a bluntly digitate appearance to its striated	572
terminus.	573
<i>Male</i> . Body ventrally curved to J-shaped. Cuticle annuli 0.8 (0.5-1.0) μm at mid-body.	574
Lateral field occupying 29 (26-37)% of body width, not areolated. Labial framework not	575
sclerotised. Labial region slightly offset from body with five to six annuli, 6.8 (6.5-7.5) μm	576
wide and 3.5 (3.0-4.5) μm high. Stylet knobs well developed, slightly sloping backward, 3.4	577
(3.0-3.5) μm in width. Median bulb oval, 10.0 (9.0-11.0) μm \times 14.5 (13.5-16.5) μm . Basal bulb	578
pyriform, 11.4 (10.5-12.0) μm \times 21.5 (18.0-26.5) μm . Testis single, 430 (417-460) μm long,	579
occupying 65.5 (63.0-68.0)% of total body length. Spicules curved ventrally, with velum and	580

pointed tip. Gubernaculum simple, crescent-shaped, about one-third of spicules length. Bursa elongated, 58 (51-62) μm long. Tail conical, ending to a pointed terminus.

Type locality

The new species was recovered from the rhizosphere of milk vetch (*Astragalus* sp.) in Sendan mountain, Zanjan province, northwestern Iran (GPS coordinates: 49°07'13"N, 36°42'41"E).

Type material

Holotype, 16 paratype females and 6 paratype males were deposited in the nematode collection of the Department of Plant Protection, College of Agriculture, Shiraz University, Shiraz, Iran; and two paratype females and one paratype male at the USDA Nematode Collection (*Bitylenchus parvulus* T-7480p).

Etymology

The species epithet refers to the proximity of the new species with the other known species, *B. parvus*.

Diagnosis and relationships

The new species is characterized by lip region with five to seven annuli, stylet 17.7 (17.0-18.5) μm long, sub-cylindrical tail with digitate terminus, cuticle just anterior to vulva wrinkled, and post-rectal sac occupied whole of tail cavity.

Regarding general characters, lip region annuli, tail shape and post-rectal sac, *Bitylenchus parvulus* n. sp. is similar to a group of some closely related species namely: *B. dubius*; *B. tobari* (Sauer & Annells, 1981) Siddiqi, 1986; *B. hispaniensis*; *B. huesingi* (Paetzold, 1958) Jairajpuri, 1982; *B. parvus*; *B. serranus* Gomez-Barcina, Siddiqi & Castillo, 1992 and *B. teeni* (Hashim, 1984) Siddiqi, 1986. The new species differs from *B. dubius* and *B. tobari* in the number of lip region annuli (5-7 vs 6-9 and 8-10, respectively), epiptygma (present vs absent), fasciculi (absent vs present), unique shape of tail terminus (suddenly narrowing near terminus giving a bluntly digitate shape to tail terminus vs normal hemispherical tail terminus), and cuticle anterior to the vulva wrinkled (vs usually normal). It can be further distinguished from *B. tobari* by difference in c' ratio (2.8 (2.3-3.3) vs 3.8 (3.1-4.4)) and reproduction behavior (spermatheca filled with sperm and males are common vs spermatheca empty and males absent).

From *B. parvus*, *B. serranus* and *B. teeni*, It differs by having a distinctly offset lip region (vs continuous or slightly offset) and female tail shape (digitate vs not digitate). It can be further distinguished from *B. hispaniensis* and *B. parvus* by presence a distinct epiptygma and absence of intestinal fasciculi (vs presence), from *B. huesingi* by stylet knobs shape (sloping backward vs rounded), from *B. serranus* by slightly shorter stylet (17.7 (17.0-18.5) vs 19.0-22.0 μm), slightly posterior position of vulva ($V = 56.0$ (54.5-60.0) vs 49-54 %) and slightly shorter spicules (23.2 (21.5-25.0) vs 26-31 μm), and from *B. teeni* by absence of intestinal fasciculi (vs presence).

Iranian population of *Bitylenchus parvus* (Allen, 1955) Jairajpuri, 1982

(Fig. 6)

Measurements

The morphometrics of *B. parvus* are presented in Table 2.

This population was found around the rhizosphere of rice (*Oryza sativa* L.) in Dezful, Khuzestan province, southwestern Iran (GPS coordinates: 32°27'87"N, 48°46'11"E). The morphological and morphometric characters of the Iranian population of *B. parvus* well fit with other populations recorded in literature (Allen, 1955; Geraert, 2011; Ghaderi *et al.*, 2014), except that tail terminus is usually smooth.

Iranian population of *Sauertylechus maximus* (Allen, 1955) Siddiqi, 2000

(Figs. 4 & 5)

Measurements

The morphometrics of *Sauertylechus maximus* are presented in Table 3.

Description

Female. Body ventrally curved to C-shaped, about 1 mm in length. Lateral field with four incisures, about one-fourth to one-third or 30.0 (25.5-35.5)% of body diameter, with areolation at entire body. Body annuli deep, 2.2 (2.0-2.5) μm wide at pharynx, 1.7 (1.5-2.0) μm wide at mid-body. Lip region high, non-offset or offset by a slight depression, with 6-8 distinct annuli,

8.4 (8.0-9.0) μm wide and 4.3 (4.0-5.0) high. Stylet slender, conus 11.5 (11.0-12.0) μm , occupying 53 (51-56)% of total stylet length, knobs sloping backward, 2.9 (2.5-3.0) μm wide. DGO 2.2 (1.5-3.0) μm posterior to stylet knobs. Procorpus cylindrical, about as long as isthmus. Median bulb masculine with distinct refractive valve, 14 (13-15) μm wide and 21.0 (19.0-23.5) μm long, occupying 66 (60-70)% of body diameter. Nerve ring at middle of isthmus, 118 (111-125) μm from anterior end. Excretory pore at level with anterior end of basal bulb. Hemizonid 3-8 annuli anterior to excretory pore, or 133 (127-143) μm from anterior end. Basal bulb pyriform and offset, 14.7 (13.5-16.0) μm wide and 30 (28-36) μm long. Cardia small. Post-rectal sac occupies entire tail cavity. Ovaries paired, vulva a transverse slit, without epiptygma, spermatheca indistinct, uterus short, vagina perpendicular to the body axis, 12 (9-16) μm or 47 (35-57)% of vulval body diameter. Phasmids at 43 (37-52) % of tail length. Tail cylindrical, curved ventrally ending to a rounded striated terminus.

Male. Not found.

Locality

This population was recovered from the rhizosphere of milk vetch (*Astragalus* sp.) in Sendan mountain, Zanjan Province, northwestern Iran (GPS coordinates: 36°42'31"N, 49°07'23"E).

Remarks

Sauertylenchus maximus is very similar to *Bitylenchus canalis* (Thorne & Malek, 1968) Jairajpuri, 1982; *B. serranus* and *S. ibericus* (Mahajan & Nombela, 1987) Siddiqi, 2000. It differs from *B. canalis* by lip region morphology (rounded vs truncated), stylet knobs shape (sloping posteriorly vs anteriorly) and the number of tail annuli (25-52 vs 66). It differs from *B. serranus* by annuli characters in tail tip (as wide as other tail annuli vs wider), and post-rectal sac length (occupied almost whole of tail cavity vs short post-rectal sac, 10-19 μm long). The present population fits well with *S. maximus* but differs by annuli on tail terminus as same as other tail annuli (vs finer than other tail annuli). Mahajan & Nombela (1987) differentiated *S. ibericus* from *S. maximus* in having an offset lip region and lacking areolation in the caudal region. However, lip region is illustrated as slightly offset similar to that observed for *S. maximus*. Therefore, areolation of the lateral field on tail region and the width of tail terminus annuli remain as the only diagnostic characters between *S. maximus* and *S. ibericus*. The present population shows areolation of the lateral field on tail region (similar to *S. maximus*)

but tail terminus annuli are generally as wide as other tail annuli (similar to *S. ibericus*). As the present population of *S. maximus* has an offset labial disc distinctly separable from lip region annuli, and the first cephalic annuli divided into six sectors, we prefer to follow Siddiqi (2000) for maintaining this species in *Sauertylenchus*, not in *Bitylenchus*.

Iranian population of *Tylenchorhynchus agri* Ferris, 1963

(Figs. 7 & 8)

Measurements

The morphometrics of *Tylenchorhynchus agri* are presented in Table 4.

Description

Female. Body slightly ventrally curved to C-shaped after heat fixation. Lateral field occupies 30 (28-33)% of the corresponding body diameter. Body annuli delicate, 2.2 (2.0-2.5) μm wide at pharynx, and 1.7 (1.5-2.0) μm at mid-body. Labial region slightly offset, bearing four to five annuli, 7.5 (7.4-7.7) μm wide and 3.4 (3.5-4.0) μm high. Stylet conus 8.8 (8.5-9.0) μm , 53 (50-56)% of total stylet length, knobs rounded, 3.9 (3.5-4.5) μm wide. DGO 2.4 (2.0-2.7) μm posterior to stylet knobs. Procorpus cylindrical, about as long as isthmus. Median bulb ovate with distinct valve, 11 (10-12) μm wide, 16.5 (16-17) μm long, occupying 61 (55-70) % of the corresponding body diameter. Isthmus slender. Basal bulb pyriform to slightly elongate, offset from intestine, 12 (11-14) μm wide and 32 (29-36) μm long. Cardia hemispherical. Nerve ring at anterior half of isthmus, at 85 (83-89) μm from anterior end. Excretory pore at anterior part of basal bulb. Hemizonid one to two annuli anterior to excretory pore, at 99 (96-106) μm from anterior end. A small post-rectal sac occupying 30 (26-36)% of tail cavity. Ovaries paired and straight, spermatheca rounded and non-offset, without sperm. Uterus short, vagina perpendicular to the body axis, 7.5 (7.0-8.0) μm , occupying 38 (34-43)% of the vulval body width, vulva with small epiptygma. Phasmids located at 37 (23-30) % of tail length. Tail sub-cylindrical ending to a smooth terminus.

Male. Not found.

Locality

This population was recovered from the rhizosphere of desert poplar (*Populus euphratica* Oliv.) in Dezful region, Khuzestan province, southwestern Iran (GPS coordinates: 32°11'36"N, 48°42'67"E).

Remarks

Tylenchorhynchus agri is similar to *T. alami* Shaw & Khan, 1996; *T. claytoni* Steiner, 1937; *T. crassicaudatus* Williams, 1960; *T. ewingi* Hopper, 1959; *T. haki* Fotedar & Mahajan, 1971 and *T. mexicanus* Knobloch & Laughlin, 1973. It is different from *T. alami* by lip region morphology (slightly offset vs continuous), tail shape and length (sub-cylindrical, 32-39 µm long vs conoid, 48 µm long), spicules length (21-25 vs 17-18 µm), and post-rectal sac (filling 30% vs less than 20% of tail length). From *T. claytoni*, it can be distinguished by longitudinal striae (absent vs present) and post-rectal sac (present vs absent). It differs from *T. crassicaudatus* by tail length (32-39 vs 38-63 µm) and post-rectal sac (present vs absent), from *T. ewingi* by annuli of lip region (4-5 vs 3 annuli), spicules (21-25 vs 18 µm) and stylet knobs shape (with concave anterior surfaces vs rounded), from *T. haki* by slightly longer stylet (17-23 vs 16-18 µm), tail characters (sub-cylindrical, 32-39 µm long vs conoid, 27 µm long), and from *T. mexicanus* by lip region characters (slightly offset vs distinctly offset), tail shape (sub-cylindrical vs conoid) and post-rectal sac (present vs absent).

Iranian population of *Tylenchorhynchus graciliformis* Siddiqi & Siddiqui, 1983

(Figs. 9 & 10)

Measurements

The morphometrics of *Tylenchorhynchus graciliformis* are presented in Table 4.

Description

Female. Body ventrally curved to C-shape. Lateral field occupied one fourth to one third of body width, outer bands crenate, 32 (23-39)% of body width in diameter. Body annuli delicate, 1.9 (1.0-2.3) µm at pharynx, and 1.7 (1.5-1.9) µm at mid-body. Lip region slightly offset bearing usually five, rarely six annuli, 6.8 (6.5-7.5) µm wide, and 3.2 (3.0-4.0) µm high. Stylet delicate, conus 8.7 (8.0-9.5) µm, 52 (50-54)% of total stylet length, knobs laterally directed,

4.1 (3.5-4.5) μm wide. DGO 2.2 (1.5-3.0) μm posterior to stylet knobs. Procorpus cylindrical, 745
about as long as isthmus. Median bulb ovate with distinct valve, 10.7 (10.0-11.0) μm wide, 746
16.8 (15.5-18.0) long, occupying 64 (59-69)% of body width in diameter. Isthmus slender. 747
Basal bulb elongate-saccate, offset from intestine to slightly overlapping anterior end of 748
intestine, 12.3 (11.0-14.0) μm wide, 31 (27-38) μm long. Cardia hemispherical. Nerve ring at 749
middle of isthmus, 86 (76-93) μm from anterior end. Excretory pore at anterior part of basal 750
bulb. Hemizonid one to three annuli anterior to excretory pore, 104 (97-109) μm from anterior 751
end. Deirid indistinct. Post-rectal sac absent. Reproductive system didelphic-amphidelphic, 752
gonads straight, spermatheca rounded and non-offset, filled with spheroid sperm, 2-3 μm in 753
diameter. Uterus short, vagina perpendicular to body axis, 9.0 (7.5-12.0) μm , occupying 44 754
(35-53)% of vulval body diameter. Vulva prominent equipped with small epiptygma. Phasmids 755
located at 32 (23-40)% of tail. Tail sub-cylindrical bearing 22 (18-26) cuticular annuli on its 756
ventral side, ending to a smooth terminus. 757

Male. General characters similar to those of female. Body ventrally curved to J-shaped. Body 758
annuli 1.6 (1.5-2.0) μm wide at mid-body. Lateral field with four incisures, 29 (23-36)% of 759
body width. Lip region bearing four to five annuli, 6.1 (6.0-6.5) μm wide and 2.9 (2.5-3.0) μm 760
high. Stylet comparable with that of female, conus 51 (50-52)% of total stylet length; knobs 761
sloping backward. DGO 1.9 (1.5-2.0) μm posterior to stylet knobs. Median bulb 10.0 (9.5- 762
11.0) μm wide and 13.5 (12.0-15.0) μm long. Basal bulb elongate-saccate, slightly shorter than 763
female, 11.4 (10.0-13.0) μm wide and 28 (25-30) μm in length. Nerve ring located at 87 (80- 764
93) μm from anterior end. Deirids indistinct. Spicules tylenchoid. Gubernaculum with crest. 765
Bursa extending to end of tail, 61 (53-67) μm in length. 766

Locality

 767

The present population of *T. graciliformis* was recovered from the rhizosphere of tamarisc 769
(*Tamarix* sp.) in Dezful region, Khuzestan province, southwestern Iran (GPS coordinates: 770
32°16'66"N, 48°47'56"E). 771

Remarks

 772

All morphological and morphometric characteristics of the present population fit well with 774
those of the original population of *Tylenchorhynchus graciliformis* differing only in the lip 775
region demarcation; in the present population, lip region is generally slightly offset from the 776
rest body (drawn and illustrated as distinctly offset in the original description). *T. graciliformis* 777

is similar to *T. aduncus* de Guiran, 1967; *T. brassicae* Siddiqi, 1961; *T. clarus* Allen, 1955; *T. contractus* Loof, 1964; *T. aerolatus* Tobar Jiménez, 1970, *T. striatus* Allen, 1955 and *T. ismaili* Azmi & Ahmad, 1991. It differs from *T. aduncus* by tail shape (sub-cylindrical vs conical), shape of stylet knobs (laterally directed vs posteriorly directed), spermatheca shape (spherical vs ovoid), spicules length (19-21 vs 22-25 μm) and shape of gubernaculum (with a simple crest vs characteristic hammer-shaped). It can be distinguished from *T. aerolatus* by absence of areolation in lateral field (vs having regular areolation), the shape of gubernaculum (with simple crest vs hammer-shaped), and body length (678-780 vs 540-700 μm). It differs from *T. brassicae* by annuli of lip region (5-6 vs 4 annuli), stylet length (17-19 vs 16-17 μm) and tail characters (sub-cylindrical, 36-51 μm vs conical, 23-40 μm in length). The present population can be differentiated from *T. clarus* by lip region morphology (offset vs non offset), number of tail annuli (16-20 vs 7-20 annuli), presence of males and spermatheca filled with sperm (vs absence of males and empty spermatheca), from *T. contractus* and *T. striatus* by lip region characters (offset vs non offset), and from *T. ismaili* by body size (678-780 vs 460-660 μm) and tail terminus striation (smooth vs striated).

Molecular characterization of Tylenchorhynchus, Bitylenchus and Sauertylenchus species from Iran

The amplification of D2-D3 expansion segments of 28S, and partial 18S rRNA genes yielded single fragments of 669 and 1695 bp, respectively after discarding primer sequences. D2-D3 for *B. parvulus* n. sp. (MK473884) differed from the closest related species, *B. parvulus* (KJ585431) by 14 different nucleotides and 0 indels (98% similarity), from *B. hispaniensis* (KJ461548; KJ461544; KJ461547 and MG770479) by 13-15 different nucleotides and 1 indels (98% similarity); from *B. huesingi* (KX789756) by 18 different nucleotides and 1 indel (97% similarity); and from *B. dubius* (DQ328707 and EU368590) by 20 different nucleotides and 0 indel (97% similarity). Two sequences for *B. parvulus* n. sp. (MT232322-MT232323) differing in 3 nucleotides (99,8% similar) were obtained for the 18S rRNA marker. These sequences closely matched with several species, such as *B. parvulus* (KX789742) and *B. teeni* (KJ636394) by 19, 21 different nucleotides and 1, 3 indels (99% similarity), respectively. D2-D3 from Iranian population of *B. parvulus* (MT193835) was identical to *B. parvulus* (KJ585431) from Fars, Iran. D2-D3 from the Iranian population of *S. maximus* (MK473883) closely matched with several populations of *S. maximus* (KX789749, KX789755 and KX789748) differing by 1-2

bp and 0 indel (99% similarity). Finally, D2-D3 from Iranian populations of *T. agri* and *T. graciliformis* (MT193837 and MT193836) showed that these isolates had the highest similarity (100% and 97.2%) with *T. agri* and *T. zaeae*, respectively.

Phylogenetic relationships of studied species

Phylogenetic relationships within *Bitylenchus*, *Sauertylenchus*, *Tylenchorhynchus* and representatives of other genera of the subfamily Telotylenchinae from the analysis of the partial 28S rRNA and 18S rRNA gene sequences are given in Figures 11 and 12, respectively. The phylogenetic analysis of the 28S rRNA separated the genus *Bitylenchus* and *Sauertylenchus* in different clades, at exception of *B. brevilineatus* (Williams, 1960) Jairajpuri, 1982 (KJ461533). The D2-D3 sequences of *B. parvulus* n. sp. clustered well (PP = 99) with two sequences of *B. dubius* (DQ328707 and EU368590). *B. parvulus* n. sp. formed a well-supported clade (PP = 100) with *B. parvus* (KJ585431 and MT193835), *B. huesingi* (KX789756) and four sequences of *B. hispaniensis* (KJ461548; KJ461544; KJ461547 and MG770479). Our isolate of *S. maximus* (MK473883) formed a well-supported clade (PP = 99) with populations of *S. maximus* (KX789749, KX789755 and KX789748) deposited in GenBank. Iranian isolate of *B. parvus* (MT193835) clustered well (PP = 100) with *B. parvus* (KJ585431). The Iranian isolate of *T. agri* (MT193837) formed a clade (PP = 99) with three isolates of *T. agri* (KX622690, MG491667 and KJ475549). The Iranian isolate of *T. graciliformis* (MT193837) formed a clade (PP = 100) with four isolates of *T. zaeae* Sethi & Swarup, 1968 (MN757911, KJ461563, KJ461565 and KM068058).

The 18S rRNA phylogeny showed that *B. parvulus* n. sp. (MT226922 and MT226923) formed a moderately supported clade (PP = 84) with *B. parvus* (KX789742) and a high supported clade (PP = 100) with other species of the genus *Bitylenchus*, as *B. dubius* (EU306352, AY284601 and EU306352), *B. teeni* (KJ636395 and KJ636394), *B. huesingi* (KX789739) and *B. serranus* (KX789743), .

Discussion

In the present study, we provided a key for identification of the all known species treated in the three closely related genera *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus*. A new species, *B. parvulus* n. sp., was described and illustrated during this study, morphological and

additional molecular data were obtained for some previously described species (*T. agri*, *B. parvus* and *S. maximus*), and new molecular and morphological data were presented for *T. graciliformis*.

Some morphological (Gomez-Barcina *et al.*, 1992; Siddiqi, 2000) and molecular data (Handoo *et al.*, 2014; Ghaderi *et al.*, 2014; present study) support that *Bitylenchus* and *Sauertylenchus* are separable genera from *Tylenchorhynchus*. However, the validity of the genus *Sauertylenchus* still needs to be tested with more studies, particularly by inclusion of the type species of this genus (Handoo *et al.*, 2014).

Although *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus* are treated as the three separated valid genera in literature, there is no adequate information on the important diagnostic characters such as lateral field areolation, lip region structure, post-rectal sac presence, or the shape of gubernaculum on many of described species in literature. Moreover, there is no agreement on the taxonomic position of certain species; for instance, *S. maximus* in Siddiqi (2000) has been considered as *B. maximus* in Andr assy (2007) or *T. maximus* in Geraert (2011). Which is why, the identification of species in *Tylenchorhynchus*, *Bitylenchus* and *Sauertylenchus* remains as a challenging task and certain species from one genus may come close to certain species from the other. Considering these problems in identification of species, all known species of these three genera are treated in a single identification key here.

The following studies have tried to provide identification keys or diagnostic compendiums for identification of species in the genus *Tylenchorhynchus*: Ganguly *et al.*, 2013 (compendium to 158 species), Geraert, 2011 (key for 133 species including *Bitylenchus* species), Andr assy, 2007 (key for 16 European species), Handoo, 2000 (key and compendium for 111 species), Brzeski & Dolinski, 1998 (compendium for 177 species of *Tylenchorhynchus sensu lato*), Brzeski, 1998 (key for 9 European species and compendium for 160 species of *Tylenchorhynchus sensu lato*), Hooper, 1978 (key and compendium for 55 species), Tarjan, 1973 (key and compendium for 46 species), Tarjan, 1964 (key and diagnostic compendium for 88 species of *Tylenchorhynchus sensu lato*), Thorne & Malek, 1968 (key for 9 species), Allen, 1955 (key for 34 species of *Tylenchorhynchus sensu lato*). For the genus *Bitylenchus*, only two keys are provided separately: Jairajpuri (1982) and Andr assy (2007).

According to D2-D3 of 28S, and partial 18S rRNA genes phylogeny, we accept the hypothesis of *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus* as valid genera and reject the 'large-genus idea' advocated by Fortuner & Luc (1987). Similar results were obtained in some recently published molecular phylogenies (Handoo *et al.*, 2014, Ghaderi *et al.*, 2014; Azimi *et*

al., 2016). Diagnosis and identification of *Bitylenchus*, *Sauertylechus* and *Tylenchorhynchus* species based solely on morphometric features is quite problematic because there is a continuous range in values of morphological-morphometric data within populations of the same species, as well as amongst species (Handoo *et al.*, 2014). As a result, it is important to include molecular markers in the description of topotypes from already described and new species, as it has been suggested by other authors (Gutiérrez-Gutiérrez *et al.*, 2010, Zamora Araya *et al.*, 2016) and the probability of cryptic speciation in plant-parasitic nematodes (Palomares-Rius *et al.*, 2014; Gharakhani *et al.*, 2019).

Bitylenchus parvulus n. sp. is molecularly closely related to *B. dubius* and *B. parvus*, and in both phylogenetic trees (D2-D3 expansion segments of 28S rRNA and partial 18S rRNA) formed a monophyletic position as valid species inside the genus *Bitylenchus*. This species moreover could be separated from these closely related species in regard to morphological and morphometric characteristics. The validity of *Bitylenchus* as a separated genus from *Tylenchorhynchus* has been already discussed in Handoo *et al.* (2014) and the addition of more representatives of this genus, as *B. parvulus* n. sp., reinforce this idea..

Morphological characters of the Iranian isolate of *T. agri* (MT193837) were similar to type species, and in D2-D3 tree formed a clade with three isolates of *T. agri* (KX622690, MG491667 and KJ475549). *Tylenchorhynchus agri* formed a sister clade with *T. annulatus* in the 28S rRNA tree. Additionally, these species could be separated morphologically because *T. annulatus* (MH142612) differs by the number of lip region annuli and absence of post-rectal sac from *T. agri*. Our isolate of *T. graciliformis* formed a clade with *T. zaeae*, which could be also separated morphologically by slightly shorter stylet length and the number of tail annuli. .

Tylenchorhynchus iranensis (KU248449 and KU248450) was transferred to the genus *Bitylenchus* by Ghaderi & Karegar (2018). The authors noticed that this species is not different from *B. parvus*, *B. teeni* and *B. huesingi*, but according to the shape of gubernaculum, having a distinct crest in the relevant description (Azimi *et al.*, 2016) and its phylogenetic position within *Tylenchorhynchus* species in the 28S rRNA tree, it could be considered as a valid species in the genus *Tylenchorhynchus* and not in *Bitylenchus*.

Sauertylechus ibericus is very close to *S. maximus* and differs only by tail terminus annuli (as wide as vs smaller than other tail annuli). *Sauertylechus maximus* (MK473883) isolate from Iran has the annuli of tail terminus as wide as the other tail annuli (seldom smaller than tail annuli). This sequence is identical to and formed a clade with *S. maximus*. As no stable diagnostic character can be observed and sequences of *S. maximus* and *S. ibericus* are identical,

synonymy of these two species is proposed here, being *S. maximus* the valid species and *S. ibericus* a junior synonym. However, the study of topotypes of both species may clarify the exact taxonomic status of them.

Conclusions

1. In this paper, we provided an identification key to all known species treated in closely related genera, *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus*.
2. Confirmed the Siddiqi's classification for transferring *S. maximus* to the genus *Sauertylenchus* as representatives of this species formed a separate clade from *Bitylenchus* species.
3. Described a new species namely *B. parvulus* n. sp. and characterized four known species (*B. parvus*, *S. maximus*, *T. agri* and *T. graciliformis*) by morphological and molecular characters, two of them (*T. agri* and *T. graciliformis*) were new records from Iran.
4. Proposed the synonymization of *S. ibericus* with *S. maximus*.

Acknowledgements. The authors thank anonymous reviewers and the editor of this manuscript for their valuable suggestions to improve the paper. J.E.P.R. acknowledges Spanish Ministry of Economy and Competitiveness for the “Ramon y Cajal” Fellowship RYC-2017-22228.

Conflict of interest. The authors declare no conflicts of interest.

Ethical standards. No specific permits were required for the indicated fieldwork studies. The soil samples were obtained in public areas, forests, and other natural areas and do not involve any species endangered or protected in Iran, nor are the sites protected in any way.

ORCID

Manouchehr Hosseinvand: <https://orcid.org/0000-0002-0472-1513>

Ali Eskandari: <https://orcid.org/0000-0003-3902-7422>

Reza Ghaderi: ID: <https://orcid.org/0000-0001-7647-1413>

Pablo Castillo: ID: <https://orcid.org/0000-0003-0256-876X>

Juan E. Palomares-Rius: <https://orcid.org/0000-0003-1776-8131>

References	944
Akhter S and Zarina B (2014) <i>Bitylenchus capsicum</i> sp. n. and one known species of the genus <i>Aphelenchus</i> from chilli plants in Sindh, Pakistan. <i>International Journal of Biological Research</i> 2 , 135-141.	945 946 947
Allen MW (1955) A review of the nematode genus <i>Tylenchorhynchus</i> . <i>University of California Publications in Zoology</i> 61 , 129-166.	948 949
Andrássy I (1954) Drei neue Arten aus der Superfamilie Tylenchoidea. Nematologische Notizen 3. <i>Annales Biologicae Universitatum Hungariae</i> 2 , 9-15.	950 951
Andrássy I (1969) The scientific results of the Hungarian soil zoological expedition to Brazzaville-Congo. 40. Vier neue Bodemnematoden Arten. <i>Opuscula Zoologica Budapestinensis</i> 9 , 15-29.	952 953 954
Andrássy I (2007) <i>Free-living Nematodes of Hungary, II. Pedozoologica Hungarica No.4.</i> Hungarian Natural History Museum, 496 pp.	955 956
Archidona-Yuste A, Navas-Cortés JA, Cantalapiedra-Navarrete C, Palomares-Rius JE and Castillo P (2016) Molecular phylogenetic analysis and comparative morphology resolve two new species of olive-tree soil related dagger nematodes of the genus <i>Xiphinema</i> (Dorylaimida: Longidoridae) from Spain. <i>Invertebrate Systematics</i> 30 , 547-565.	957 958 959 960 961
Azimi S, Mahdikhani-Moghadam E, Rouhani H and Rajabi Memari H (2016) Description of <i>Tylenchorhynchus iranensis</i> sp. n. (nematoda telotylenchidae) from iran. <i>Redia</i> XCIX , 3-8.	962 963 964
Azizi K, Eskandari A, Karegar A, Ghaderi R, van den Elsen S, Holterman M and Helder J (2016) Morphological and molecular data support the monophyletic nature of the genus <i>Pratylenchoides</i> Winslow, 1958 (Nematoda: Merliniidae) and reveal its intrageneric structuring. <i>Nematology</i> 18 , 1165-1183. DOI: 10.1163/15685411-00003023	965 966 967 968
Azmi MI and Ahmad ST (1991) <i>Tylenchorhynchus ismaili</i> n. sp. from India. <i>Indian Journal of Nematology</i> 19 , 279-281.	969 970
Azmi MI (1991) <i>Tylenchorhynchus leucaenus</i> sp. nov. (Nematoda: Tylenchida) from Jhansi, India. <i>Current Nematology</i> 2 , 81-82.	971 972
Bastian HC (1865) Monograph on the Anguillulidae, or free nematoids, marine, land and freshwater; with descriptions of 100 new species. <i>Transactions of the Linnean Society</i> 25 , 73-184.	973 974 975

Bloemers GF and Wanless FR (1998) Description of <i>Tylenchorhynchus paulettae</i> sp. n. (Dolichodoridae) from Cameroon with comments on the status of the genus <i>Triversus</i> Sher, 1974. <i>International Journal of Nematology</i> 8 , 53-56.	976 977 978
Brzeski MW and Dolinski CM (1998) Compendium of the genus <i>Tylenchorhynchus</i> Cobb, 1913 <i>sensu lato</i> (Nematoda: Belonolaimidae). <i>Russian Journal of Nematology</i> 6 , 189-199.	979 980 981
Brzeski MW (1992) <i>Tylenchorhynchus paratriversus</i> sp. n. and comments on three other species of the genus (Nematoda: Belonolaimidae). <i>Nematologia Mediterranea</i> 19 , 213-220.	982 983 984
Brzeski MW (1998) <i>Nematodes of Tylenchina in Poland and Temperate Europe</i> . Muzeum i Instytut Zoologii Polska Akademia Nauk, Warszaw, 397 pp.	985 986
Bütschli O (1873) Beiträge zur Kenntis der freilebenden Nematoden. Beiträge zur Kenntnis der freilebenden Nematoden. <i>Nova acta Academiae Caesareae Leopoldino-Carolinae Germanicae Naturae Curiosorum</i> 36 , 1-124.	987 988 989
Cassidy GH (1930) Nematodes associated with sugarcane in Hawaii. <i>Hawaiian Planters Record</i> 34 , 379-387.	990 991
Castillo P, Siddiqi MR and Gomez-Barcina A (1989) Studies on the genus <i>Paratrophurus</i> Arias (Nematoda: Tylenchina) with descriptions of two new species. <i>Nematologia Mediterranea</i> 17 , 83-95.	992 993 994
Castresana J (2000) Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. <i>Molecular Biology and Evolution</i> 17 , 540-552.	995 996
Chawla MI, Bhamburkar BL, Khan E and Prasad SK (1968) One new genus and seven new species of nematodes from India. <i>Labdev Journal of Science and Technology</i> 6 , 86-100.	997 998 999
Cobb NA (1913) New nematode genera found inhabiting fresh water and non-brackish soils. <i>Journal of the Washington Academy of Sciences</i> 3 , 432-444.	1000 1001
Darriba D, Taboada GL, Doallo R and Posada D (2012) jModelTest 2: more models, new heuristics and parallel computing. <i>Nature Methods</i> 9 , 772.	1002 1003
Das V (1960) Studies on the nematode parasites of plants in Hyderabad (Andhra Pradesh, India). <i>Zeitschriftfür Parasitenkunde</i> 19 , 553-605.	1004 1005
De Grisse A (1969) Redescription ou modification de quelques techniques utilisées dans l'étude des nematodes phytoparasitaires. <i>Mededelingen Rijksfaculteit Landbouwwetenschappen Gent</i> 34 , 351-369.	1006 1007 1008

De Guiran G (1967) Description de deux espèces nouvelles du genre <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Tylenchinae) accompagnée d'une clé des femelles, et précisions sur <i>T. mamillatus</i> Tobar-Jimenez, 1966. <i>Nematologica</i> 13 , 217-230.	1009 1010 1011
De Ley P, Félix MA, Frisse LM, Nadler SA, Sternberg PW and Thomas WK (1999) Molecular and morphological characterisation of two reproductively isolated species with mirror-image anatomy (Nematoda: Cephalobidae). <i>Nematology</i> 1 , 591-612.	1012 1013 1014
De Man JG (1921) Nouvelles recherches sur les nematodes terricoles de la Hollande. <i>Capita Zoologica</i> 1 , 3-62.	1015 1016
Decker H, Yassin AM and El-Amin ETM (1975) Zur Gattung <i>Paratrophurus</i> Arias, 1970 (Nematoda: Dolichodoridae). In: Vortragstagung (1) zu Aktuellen Problemender Phytonematologie am 29.5.1975 in Rostock. Manuskriptdruck der Vortrage. Biologische Gesellschaft der DDR, Sektion Phytopathologie und Universität Rostock. Rostock. DDR. pp. 89-102.	1017 1018 1019 1020 1021
Elbadri GAA, Moon IS, Lee DW and Choo HJ (2010) Two new species of <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Belonolaimidae) from Sudan. <i>Korean Journal of Applied Entomology</i> 49 , 57-60.	1022 1023 1024
Eliashvili TS (1971) Two new soil-inhabiting nematode species (<i>Amphidelus paramonovi</i> and <i>Tylenchorhynchus georgiensis</i> n.sp.) of eastern Georgia. <i>Bulletin of the Academy of Sciences of the Georgian SSR</i> 61 , 213-216.	1025 1026 1027
Ferris VR (1963) <i>Tylenchorhynchus silvaticus</i> sp. nov. and <i>Tylenchorhynchus agri</i> sp. nov. (Nematoda: Tylenchida). <i>Proceedings of the Helminthological Society of Washington</i> 30 , 165-168.	1028 1029 1030
Filipjev IN (1934) <i>Harmful and Useful Nematodes in Rural Economy</i> . Moscow & Leningrad, 440 pp.	1031 1032
Filipjev IN (1936) On the classification of the Tylenchinae. <i>Proceedings of the Helminthological Society of Washington</i> 3 , 80-82.	1033 1034
Fortuner R and Luc M (1987) A reappraisal of Tylenchina (Nemata). 6. The family Belonolaimidae Whitehead, 1960. <i>Revue de Nématologie</i> 10 , 183-202.	1035 1036
Fotedar DN and Mahajan R (1971) On a new species of the genus <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Tylenchorhynchidae) from Kashmir, India. <i>Kashmir Science</i> 8 , 120-122.	1037 1038 1039
Geraert E (2011) <i>The Dolichodoridae of the world, identification of the family Dolichodoridae</i> (Nematoda: Tylenchida). Ghent, Belgium, Academia Press, 520 pp.	1040 1041

- Gharakhani A, Pourjam E and Pedram M** (2019) Occurrence of *Neodolichodorus persiangulfus* n. sp. (Nematoda: Dolichodoridae) in mangrove forest of southern Iran, Forest Pathology DOI: 10.1111/efp.12563. 1042-1044
- Ghaderi R, Karegar A, Niknam G and Subbotin SA** (2014) Phylogenetic relationships of Telotylenchidae Siddiqi, 1960 and Merliniidae Siddiqi, 1971 (Nematoda: Tylenchida) from Iran, as inferred from the analysis of the D2D3 expansion fragments of 28S rRNA gene sequences. *Nematology* **16**, 863-877. 1045-1048
- Ghaderi R and Karegar A** (2018) Nematodes of the families Merliniidae and Telotylenchidae. In: Ghaderi, R., Kashi, L. & Karegar, A. (Eds). *Plant-parasitic nematodes in Iran*. Marjae-e-elm Publication and Society of Iranian Nematology, Tehran, Iran, pp. 193-294. 1049-1052
- Golden AM** (1971) Classification of the genera and higher categories of the order Tylenchida (Nematoda). In B. M. Zuckerman, W. F. Mai & R. A. Rohde (Eds.), *Plant parasitic nematodes. Volume 1. Morphology, Anatomy, Taxonomy and Ecology* (pp. 191-232). New York: Academic Press. 1053-1056
- Golden AM, Baldwin JG and Mundo-Ocampo M** (1995) Description of *Tylenchorhynchus thermophilus* sp. nov. (Nematoda: Tylenchina) from Saltgrass in Death Valley, California. *Journal of Nematology* **27**, 312-319. 1057-1059
- Golden AM, Maqbool MA and Handoo ZA** (1987) Description of two new species of *Tylenchorhynchus* Cobb, 1913 (Nematoda: Tylenchida), with details on morphology and variation of *T. claytoni*. *Journal of Nematology* **19**, 58-68. 1060-1062
- Gomez-Barcina A, Siddiqi MR and Castillo P** (1992) The genus *Bitylenchus* Filipjev, 1934 (Nematoda: Tylenchida) with descriptions of two new species from Spain. *Journal of the Helminthological Society of Washington* **59**, 96-110. 1063-1065
- Gupta NK and Uma** (1980) On a new species of the genus *Tylenchorhynchus* Cobb, 1913 (Family Tylenchorhynchidae (Eliava, 1964) Golden, 1971). *Revista Ibérica de Parasitología* **40**, 423-427. 1066-1068
- Gutiérrez Gutiérrez C, Palomares-Rius JE; Cantalapiedra-Navarrete C, Landa Del Castillo BB, Esmenjaud D and Castillo P** (2010) Molecular analysis and comparative morphology to resolve a complex of cryptic *Xiphinema* species. *Zoologica Scripta* **39**, 483-498. 1069-1072
- Handoo ZA, Palomares-Rius JE, Cantalapiedra-Navarrete C, Liébanas G, Subbotin SA and Castillo P** (2014) Integrative taxonomy of the stunt nematodes of the genera 1073-1074

- Bitylenchus* and *Tylenchorhynchus* (Nematoda, Telotylenchidae) with description of two new species and a molecular phylogeny. *Zoological Journal of the Linnean Society* **172**, 231–264.
- Hashim Z** (1984) Description of *Tylenchorhynchus teeni* sp. nov. and observations on *Rotylenchus cypriensis* Antoniou, 1980 (Nematoda: Tylenchida) from Jordan. *Systematic Parasitology* **6**, 33-38.
- Holterman M, van der Wurff A, van den Elsen S, van Megen H, Bongers T, Holovachov O, Bakker J and Helder J** (2006) Phylum-wide analysis of SSU rRNA reveals deep phylogenetic relationships among nematodes and accelerated evolution toward crown clades. *Molecular Biology and Evolution* **23**, 1792-1800.
- Hopper BE** (1959) Three new species of the genus *Tylenchorhynchus* (Nematoda: Tylenchida. *Nematologica* **4**, 23-30.
- Hosseinvand M, Eskandari A and Ghaderi R** (2019a) Additional data on *Pratylenchooides riparius* (Andrássy, 1985) Luc, 1986 (Nematoda: Merliniidae) from Iran. *Nematology* **21**, 827-836.
- Hosseinvand M, Eskandari A and Ghaderi R** (2019b) Morphological and Molecular Characterization of *Coslenchus paramaritus* sp. nov. and *C. cancellatus* (Cobb, 1925) Siddiqi, 1978 (Nematoda: Tylenchidae) from Iran. *Journal of Nematology* **51**, e2019-59.
- Hosseinvand M, Eskandari A and Ghaderi R** (2020) Morphological and molecular characterisation of *Telotylenchus tamariscus* n. sp. (Nematoda: Telotylenchinae) recovered from Khuzestan, south-western Iran. *Nematology* DOI: 10.1163/15685411-bja10016.
- Huelsenbeck JP and Ronquist F** (2001) MrBayes: Bayesian inference of phylogeny. *Bioinformatics* **17**, 754-755.
- Ivanova TS** (1983) New species of nematodes from the family Tylenchorhynchidae (Eliava, 1964) Golden, 1971 from the southern Pamir-Alai. *Izvestiya Akademii Nauk Tadzhikskoi SSR, Biologicheskie Nauki* **1**, 40-45.
- Ivanova TS** (1989) Phytoparasitic nematodes of the Tylenchina suborder of the Eastern Pamirs. *Doklady Akademii Nauk Tadzhikskoi SSR* **32**, 785-788.
- Jairajpuri MS** (1982) Some studies on Tylenchorhynchinae: The subgenus *Bitylenchus* Filipjev, 1934, with description of *Tylenchorhynchus (Bitylenchus) depressus* sp. nov. and a key to species of *Bitylenchus*. *Mededelingen van de Fakuliteit Landbouwwetenschappen Rijksuniversiteit Gent* **47**, 765-770.

Karapetjan JA (1979) Four new species of nematodes (Nematoda: Tylenchorhynchidae) from greenhouses in Armenia. <i>Doklady Akademii Nauk Armyanskoi SSR</i> 68 , 124-128.	1108 1109
Kaul V and Waliullah MIS (1995) <i>Tylenchorhynchus oryzae</i> sp. n. (Tylenchorhynchidae: Nematoda) from Kashmir, India. <i>Annals of Plant Protection Science</i> 3 , 155-157.	1110 1111
Khakimov NK (1973) Nematodes of the soil in cotton fields of Golodnaya steppe of Uzbekistan. (<i>Tylenchorhynchus bicaudatusi</i>). <i>Voprosy Fitogel'mintologii v Uzbekistane</i> . Book 3, Izdatel'stvo "FAN", Tashkent, USSR, pp. 161-214.	1112 1113 1114
Khan A (2004) <i>Tylenchorhynchus handooi</i> n.sp. (Nematoda: Tylenchida) from Khuzdar, Balochistan. <i>Proceedings of Parasitology</i> 38 , 57-61.	1115 1116
Khan E and Darekar KS (1979) Soil and plant-parasitic nematodes from Maharashtra, India. IV. Two new species of <i>Telotylenchus</i> Siddiqi, 1960 (Tylenchida: Nematoda). <i>Indian Journal of Nematology</i> 8 , 13-18.	1117 1118 1119
Khan E and Nanjappa CK (1974) <i>Tylenchorhynchus nordiensis</i> nomen novum for <i>T. aerolatus</i> Khan & Nanjappa, 1971. <i>Indian Journal of Nematology</i> 2 , 216.	1120 1121
Khan FA (1971) <i>Merlinius bijnorensis</i> sp. nov. and <i>Tylenchorhynchus hordei</i> sp. nov. (Nematoda: Tylenchida) from North India. <i>Proceedings of the 59th Session of the Indian Science Congress Association, Calcutta, 1972, Part III</i> , pp. 594-595.	1122 1123 1124
Khan FA (1990) Two new species of the genus <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Tylenchorhynchinae) from India. <i>Pakistan Journal of Nematology</i> 8 , 73-78.	1125 1126
Khan FA (1991) <i>Tylenchorhynchus paracanalisis</i> sp. nov. (Nematoda: Tylenchida) from Nigerian Savanna. <i>Pakistan Journal of Nematology</i> 9 , 87-90.	1127 1128
Khan HA, Saeed H and Akhter M (2004) A new species of <i>Tylenchorhynchus</i> with comments on <i>Geocenamus rugosus</i> (Thome and Malek, 1968) Brzeski, 1991 from Sindh. <i>Pakistan Journal of Scientific and Industrial Research</i> 47 , 446-450.	1129 1130 1131
Khurma U and Mahajan R (1988) Two new species of <i>Tylenchorhynchus</i> Cobb, 1913 from Punjab, India. <i>Indian Journal of Nematology</i> 17 , 202-207.	1132 1133
Kleynhans KPN (1984) <i>Tylenchorhynchus natalensis</i> sp. nov. from South Africa (Nematoda: Tylenchorhynchinae). <i>Phytophylactica</i> 16 , 71-72.	1134 1135
Knobloch NA and Laughlin CW (1973) A collection of plant parasitic nematodes (Nematoda) from Mexico with descriptions of three new species. <i>Nematologica</i> 19 , 205-217.	1136 1137 1138

Knobloch NA (1975) <i>Tylenchorhynchus aspericutis</i> sp. nov. (Nematoda) from Mexico with an emended description of <i>Psilenchus hilarulus</i> de Man, 1921 and two intersexes. <i>Nematologica</i> 21 , 287-295.	1139 1140 1141
Kumar AC (1981) Studies on nematodes in coffee soils of South India. 5. Descriptions of three new species of <i>Tylenchorhynchus</i> and occurrence of four other Tylenchid species. <i>Journal of Coffee Research</i> 11 , 89-99.	1142 1143 1144
Kumar P (1982) On new species of <i>Tylenchorhynchus</i> and <i>Helicotylenchus</i> from cauliflower (<i>Brassica oleracea</i> var. <i>botrytis</i>) at Lucknow. <i>Kanpur University Research Journal</i> 1 , 185-192.	1145 1146 1147
Kumar S, Stecher G and Tamura K (2016) MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. <i>Molecular Biology and Evolution</i> 33 , 1870-1874.	1148 1149 1150
Lin M (1992) A new species of <i>Tylenchorhynchus</i> (Nematoda: Tylenchorhynchinae). <i>Journal of Nanjing Agricultural University</i> 15 , 41-44.	1151 1152
Litvinova NF (1946) Four new species of <i>Tylenchorhynchus</i> (Nematoda) from Kazakhstan. <i>Proceedings of the Zoological society of London</i> 116 , 120-128.	1153 1154
Loof PAA (1960) Miscellaneous notes on the genus <i>Tylenchorhynchus</i> (Tylenchinae: Nematoda). <i>Nematologica</i> 4 , 294-306.	1155 1156
Loof PAA (1964) Free-living and plant parasitic Nematodes from Venezuela. <i>Nematologica</i> 10 , 201-300.	1157 1158
Luqman M and Khan SH (1986) Three new nematode species attacking fruit trees in India. <i>Indian Journal of Nematology</i> 15 , 202-208.	1159 1160
Mahajan R and Nombela G (1987) <i>Tylenchorhynchus ibericus</i> sp. n. (Nematoda: Tylenchorhynchinae) from Spain. <i>Phytophylactica</i> 19 , 47-48.	1161 1162
Mahajan R (1974) <i>Tylenchorhynchus kashmirensis</i> sp. n. and <i>Quinisulcius himalayae</i> sp. n. (Nematoda: Tylenchorhynchinae) from India. <i>Proceedings of the Helminthological Society of Washington</i> 41 , 13-16.	1163 1164 1165
Mathur VK, Sanwal KC and Lal A (1979) <i>Tylenchorhynchus neoclavicaudatus</i> sp. nov. in soil washings from imported potato tubers. <i>Indian Journal of Nematology</i> 8 , 148-150.	1166 1167
Mavlyanov OM (1978) Two new species of Nematodes of the family Dolichodoridae (Nematoda: Tylenchida) from the root soil of cotton. <i>Parazitologiya</i> 12 , 138-142.	1168 1169
Minagawa N (1995) <i>Bitylenchus iphilus</i> sp. n. and <i>Tylenchorhynchus kagasawai</i> sp. n. (Nematoda: Tylenchida) from Japan. <i>Afro-Asian Journal of Nematology</i> 5 , 151-160.	1170 1171

- Monteiro AR and Lordello LGE** (1976) *Tylenchorhynchus queirozi* sp. nov. (Nematoda, Tylenchorhynchidae) de solo do Brasil. *Revista Brasileira de Biologia* **36**, 697-699. 1172
1173
- Nasira K and Maqbool MA** (1996) Description of *Tylenchorhynchus gossypii* sp. nov. (Nematoda: Dolichodoridae) with observations on *T. elegans* Siddiqi, 1961 from Pakistan. *Pakistan Journal of Nematology* **14**, 33-40. 1174
1175
1176
- Nobbs JM** (1990) Four new species of plant parasitic nematodes (Nematoda: Tylenchida) from the arid region of South Australia. *Nematologica* **35**, 399-412 1177
1178
- Paetzold D** (1958) Beitrage zur Nematodenfauna mitteldeutscher Salzstellen im Raum von Halle. *Wissenschaftliches Zeitschrift Universitiit Hall, Mathematisch-naturwissenschaftliche Reihe* **8**, 17-48. 1179
1180
1181
- Palomares-Rius JE, Cantalapiedra-Navarrete C and Castillo P** (2014) Cryptic species in plant-parasitic nematodes. *Nematology* **16**, 1105-1118. 1182
1183
- Pathak M and Siddiqui AU** (1997) One new and five known species of *Tylenchorhynchus* Cobb, 1913 from ornamental crops in Udaipur region of Rajasthan. *Indian Journal of Nematology* **27**, 99-103. 1184
1185
1186
- Rambaut A** (2014) Figtree, a graphical viewer of phylogenetic trees [Internet]. Available from: <http://tree.bio.ed.ac.uk/software/figtree>. 1187
1188
- Ramzan M, Handoo ZA and Fayyaz S** (2008) Description of *Tylenchorhynchus qasimii* sp. n. with a new report of *T. kegasawai* Minagawa, 1995 from Pakistan. *Journal of Nematology* **40**, 20-25. 1189
1190
1191
- Rashid F and Heyns J** (1990) *Tylenchorhynchus* species from Namibia (Nematoda: Belonolaimidae). *Phytophylactica* **22**, 403-412. 1192
1193
- Ray S and Das SN** (1983) Three new and five nominal species in the family Tylenchorhynchidae (Tylenchoidea: Nematoda) from Orissa, India. *Indian Journal of Nematology* **13**, 16-25. 1194
1195
1196
- Ronquist F, Teslenko M, van der Mark P, Ayres D, Darling A, Höhna S, Larget B, Liu L, Suchard MA and Huelsenbeck JP** (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* **61**, 539–542. 1197
1198
1199
1200
- Saha M, Gaur HS and Lal M** (1998) A new species of the stunt nematode, *Tylenchorhynchus iarius* sp.nov. (Tylenchorhynchidae: Tylenchida) from rice field at New Delhi. *Annals of plant Protection Science* **6**, 63-65. 1201
1202
1203

Saha M, Singh M, Lal M and Kaushal KK (2002) Two new tylenchid nematode species associated with wheat and rice cropping system in North India. (<i>Tylenchorhynchus karnalensis</i>). <i>Annals of plant Protection Science</i> 10 , 355-359.	1204 1205 1206
Sara M and Khan E (1982) <i>Duotylenchus bilineatus</i> gen. n., sp. n. and <i>Tylenchorhynchus badliensis</i> sp. n. (Tylenchida: Nematoda) from Haryana, India. <i>Indian Journal of Nematology</i> 11 , 205-211.	1207 1208 1209
Sauer MR and Annells CM (1981) Three new Tylenchs (Nematoda) from Australia. <i>Nematologica</i> 27 , 422-431.	1210 1211
Seinhorst JW (1963) Five new <i>Tylenchorhynchus</i> species from West Africa. <i>Nematologica</i> 9 , 173-180.	1212 1213
Sethi CL and Swarup G (1968) Plant parasitic nematodes of northwestern India. I. The genus <i>Tylenchorhynchus</i> . <i>Nematologica</i> 14 , 77-88.	1214 1215
Shaw SP and Khan E (1992) Tylenchorhynchidae Golden, 1971 (Nematoda: Tylenchida) in India. 1. <i>Amplimerlinius sikkimensis</i> sp. n. from Sikkim with key to <i>Amplimerlinius</i> species and <i>Tylenchorhynchus shivanandi</i> sp. n. from Nagaland. <i>Bulletin of Entomology</i> 33 , 7-13.	1216 1217 1218 1219
Shaw SP and Khan E (1996) Two new species of Tylenchorhynchidae Golden, 1971 (Nematoda: Tylenchida) from India. <i>Journal of Research Birsa Agricultural University</i> 8 , 1-8.	1220 1221 1222
Sher SA (1974) <i>Sauertylenchus labiodiscus</i> n. gen., n. sp; from Australia (Nematoda: Tylenchorhynchinae). <i>Journal of Nematology</i> 6 , 37-40.	1223 1224
Siddiqi MR (1960) <i>Telotylenchus</i> , a new nematode genus from North India (Tylenchida Telotylenchinae n. subfam.). <i>Nematologica</i> 5 , 73-77.	1225 1226
Siddiqi MR (1961) Studies on <i>Tylenchorhynchus</i> spp. (Nematoda: Tylenchida). <i>Zeitschrift für Parasitenkunde</i> 21 , 46-64.	1227 1228
Siddiqi MR (1985) Nematodes from Colombian rain forest. 2. <i>Tylenchorhynchus (Bitylenchus) botrys</i> sp. nov. and <i>T. (B.) colombianus</i> sp. nov. <i>Fitopatologia Colombiana</i> 11 , 29-31.	1229 1230 1231
Siddiqi MR (1986) <i>Tylenchida: parasites of plants and insects</i> . Commonwealth Agricultural Bureaux, Farnham Royal, Slough SL2 3BN, UK, 645 pp.	1232 1233
Siddiqi MR (2000) <i>Tylenchida: parasites of plants and insects. 2nd edition</i> . CABI Publishing, Wallingford, UK, 833 pp.	1234 1235

Siddiqi MR (2008) Descriptions of five new species of <i>Tylenchorhynchus</i> Cobb (Nematoda: Tylenchida: Telotylenchinae). <i>International Journal of Nematology</i> 18 , 159-168.	1236 1237
Siddiqi MR and Basir MA (1959) On some plant parasitic nematodes occurring in South India with description of two new species of the genus <i>Tylenchorhynchus</i> Cobb, 1913. <i>Proceedings 46th Indian Science Congress Part IV</i> (Abstracts), p. 35.	1238 1239 1240
Siddiqi MR, Mukherjee B and Dasgupta MK (1982) <i>Tylenchorhynchus microconus</i> sp. nov., <i>T. crassicaudatus leviterminalis</i> n. subsp. and <i>T. coffeae</i> Siddiqi & Basir, 1959 (Nematoda: Tylenchida). <i>Systematic Parasitology</i> 4 , 257- 262.	1241 1242 1243
Siddiqi MR and Patel DJ (1990) <i>Tylenchorhynchus microcephalus</i> sp. n. and <i>Paurodontus simms</i> Siddiqi, 1961 (Nematoda: Tylenchida) from Gujarat state, India. <i>Current Nematology</i> 1 , 7-10.	1244 1245 1246
Siddiqi MR and Sharma SB (1994) <i>Scutellonema paralabiatum</i> sp. n., <i>S. propeltatum</i> sp. n. and <i>Bitylenchus singularis</i> sp. n. found associated with pigeonpea in Kenya. <i>Afro-Asian Journal of Nematology</i> 4 , 35-39.	1247 1248 1249
Siddiqi MR and Siddiqui ZA (1983) <i>Paratrophurus acristylus</i> sp. n. and <i>Tylenchorhynchus graciliformis</i> sp. n. (Nematoda: Tylenchida) from wheat fields in Libya. <i>Proceedings of the Helminthological Society of Washington</i> 50 , 301-304.	1250 1251 1252
Singh M, Lal M, Rathour KS and Ganguly S (2010) Two New Tylenchid Nematode Species from India. <i>Indian Journal of Nematology</i> 40 , 216-220.	1253 1254
Singh SP (1976) A new species of the genus <i>Deladenus</i> Thorne, 1941 and <i>Tylenchorhynchus</i> Cobb, 1913 from Lucknow, India. <i>Indian Journal of Zootomy</i> 15 , 187-192.	1255 1256
Singh RV and Khera S (1978) Plant parasitic nematodes from the rhizosphere of vegetable crops around Calcutta. 2. Family Tylenchorhynchidae. <i>Bulletin of Zoological Survey of India</i> 1 , 25-28.	1257 1258 1259
Singh SD (1971) Studies on the morphology and systematic of plant and soil nematodes mainly from Andra Pradesh. I. Tylenchoidea. <i>Journal of Helminthology</i> 45 , 353-369	1260 1261
Singh SP and Jain VK (1983) Two plant parasitic nematodes of the genus <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Tylenchida) from Lucknow. <i>Journal of the Zoological Society of India</i> 34 , 62-68.	1262 1263 1264
Steiner G (1937) Opuscula miscellanea Nematologica V. <i>Tylenchorhynchus claytoni</i> sp. nov. an apparently rare nemtic parasite of the tobacco plant. <i>Proceedings of the Helminthological Society of Washington</i> 4 , 33-34.	1265 1266 1267

Sturhan D (1966) Uber Verbreitung, Pathogenität und Taxonomie der Nematodengattung <i>Tylenchorhynchus</i> . <i>Mitteilungen der Biologischen Anstalt für Land- und Forstwirtschaft</i> 118 , 82-99.	1268 1269 1270
Sturhan D (2014) Plant-parasitic nematodes in Germany - an annotated checklist. <i>Soil Organisms</i> 86 , 177–198.	1271 1272
Subbotin S, Waeyenberge AL and Moens M (2000) Identification of cyst forming nematodes of the genus <i>Heterodera</i> (Nematoda: Heteroderidae) based on the ribosomal DNA-RFLPs. <i>Nematology</i> 2 , 153-164.	1273 1274 1275
Sultan MS, Singh I and Sakhuja PK (1991) Plant parasitic nematodes of Punjab, India III. <i>Tylenchorhynchinae</i> Eliava, 1964 with proposal of <i>Macrorhynchus</i> n. gen. and <i>Tylenchorhynchus persicus</i> sp. nov. <i>Indian Journal of Nematology</i> 19 , 215-222.	1276 1277 1278
Talavera M and Siddiqi MR (1995) <i>Bitylenchus mediocris</i> sp. nov. from Malawi and <i>B. equatorialis</i> sp. nov. from Ecuador (Nematoda: Telotylenchinae). <i>Afro-Asian Journal of Nematology</i> 5 , 181-185	1279 1280 1281
Talavera M, Watanabe T and Mizukubo T (2002) Description of <i>Tylenchorhynchus shimizui</i> sp. nov. from Paraguay and notes on <i>T. leviterminalis</i> Siddiqi, Mukhetjee & Dasgupta from Japan (Nematoda: Tylenchida: Telotylenchidae). <i>Systematic Parasitology</i> 51 , 171-177.	1282 1283 1284 1285
Thorne G and Malek RB (1968) Nematodes of the Northern Great Plains. Part I. Tylenchida (Nematoda: Secernentea). <i>South Dakota State University Agricultural Experiment Station Technical Bulletin</i> 31 , 1-111.	1286 1287 1288
Tobar Jimenéz A (1969) Descripción del <i>Tylenchorhynchus ventrosignatus</i> sp. nov. (Nematoda: Tylenchida). <i>Revista Ibérica de Parasitología</i> 29 , 399-403.	1289 1290
Tobar Jimenéz A (1970) Descripción de dos nuevas especies del género <i>Tylenchorhynchus</i> Cobb, 1913 (Nematoda: Tylenchidae), con algunos datos adicionales sobre el <i>T. sulcatus</i> de Guiran, 1967. <i>Revista Ibérica de Parasitología</i> 30 , 215-228.	1291 1292 1293
Upadhyay KD, Swarup G and Sethi CL (1972) <i>Tylenchorhynchus vulgaris</i> sp. n. associated with maize roots in India, with notes on its embryology and life history. <i>Indian Journal of Nematology</i> 2 , 129-138.	1294 1295 1296
Whitehead AG and Hemming JR (1965) A comparison of some quantitative methods extracting small vermiform nematodes from the soil. <i>Annals of Applied Biology</i> 55 , 25-38.	1297 1298 1299

Williams JR (1960) Studies on the nematode soil fauna of sugar cane fields in Mauritius. 4.	1300
Tylenchoidea (Partim). <i>Occas. Paper. Mauritius Sugar Industry Research Institute</i> 4 , 1-	1301
30.	1302
Wouts WM and Sher SA (1981) Two new <i>Tylenchorhynchus</i> species (Nematoda:	1303
Tylenchorhynchidae) from the sub-antarctic region. <i>Nematologica</i> 27 , 253-257.	1304
Wu LY (1969) Three new species of the genus <i>Tylenchorhynchus</i> Cobb, 1913 (Tylenchida:	1305
Nematoda) from Canada. <i>Canadian Journal of Zoology</i> 47 , 563-567.	1306
Zamora Araya T, Peraza Padilla W, Archidona Yuste A, Cantalapiedra Navarrete C,	1307
Liébanas G, Palomares-Rius JEP and Castillo P (2016) Root-lesion nematodes of the	1308
genus <i>Pratylenchus</i> (Nematoda: Pratylenchidae) from Costa Rica with molecular	1309
identification of <i>P. gutierrezii</i> and <i>P. panamaensis</i> topotypes. <i>European Journal of Plant</i>	1310
<i>Pathology</i> 145 , 973-998.	1311
Zarina B and Maqbool MA (1994) Description of <i>Tylenchorhynchus tuberosus</i> sp. nov. and	1312
observations on <i>Dolichorhynchus phaseoli</i> (Nematoda: Dolichodoridae) from	1313
ornamental plants of Karachi, Pakistan. <i>Pakistan Journal of Nematology</i> 12 , 51-57.	1314
Zarina B, Siddiqi MR and Shahina F (2004) <i>Bitylenchus microcephalus</i> sp. n. and <i>B.</i>	1315
<i>maximus pakistanensis</i> subsp. n. (Nematoda: Telotylenchidae). <i>International Journal of</i>	1316
<i>Nematology</i> 14 , 85-90.	1317
	1318
	1319

Figure legends	1320
Fig. 1. <i>Bitylenchus parvulus</i> n. sp. Female (A, C, E, F, H-K) and male (B, D, G, L). A, B: Entire body; C: Anterior end; D, E: Anterior end and pharyngeal region; F: reproductive system; G: Spicules and Gubernaculum; H-L: Posterior end.	1321 1322 1323
Fig. 2. <i>Bitylenchus parvulus</i> n. sp. Female (A, B, D, E, F, I-K) and male (C, G-H). A: Anterior body and pharyngeal region; B-C: Entire body; D: Anterior end; E: Lateral field; F: Pharyngeal basal bulb; G-H, K: Posterior end; I-J: Vulva region. (Scale-bars: B, C = 100 μ m, A, D-K = 10 μ m).	1324 1325 1326 1327
Fig. 3. <i>Bitylenchus parvulus</i> n. sp. Female (A-N). A-E: Anterior end; F-H: Vulva region; I-N: Posterior end. (Scale-bars: A-N = 10 μ m).	1328 1329
Fig. 4. <i>Sauertylenchus maximus</i> . Female (A-I). A: Entire body; B-C: Anterior end; D: Reproductive system; E: Anterior end and pharyngeal region; F-H: Posterior end; I: Lateral field.	1330 1331 1332
Fig. 5. <i>Sauertylenchus maximus</i> . Female (A-L) and male (B-C). A: Anterior body and pharyngeal region; B-C: Anterior end; D: Entire body; E: Vulva region; F: Lateral field; G-L: Posterior end. (Scale-bars: A-G = 10 μ m).	1333 1334 1335
Fig. 6. <i>Bitylenchus parvus</i> . Female (A, D-G) and male (B-C). A-B: Anterior end; C-G: Posterior end. (Scale-bars: A-G = 10 μ m).	1336 1337
Fig. 7. <i>Tylenchorhynchus agri</i> . Female (A-F). A: Entire body; B: Anterior end and pharyngeal region; C: Anterior end; D-E: Posterior end; F: Reproductive system.	1338 1339
Fig. 8. <i>Tylenchorhynchus agri</i> . Female (A-G). A: Entire body; B: Anterior end and pharyngeal region; C: Anterior end; D: Pharyngeal basal bulb; E-G: Posterior end. (Scale-bars: A = 100 μ m, B-G = 10 μ m).	1340 1341 1342
Fig. 9. <i>Tylenchorhynchus graciliformis</i> . Female (A, C-E, F-H) and male (B, D, I-J). A-B: Entire body; C-D: Anterior end and pharyngeal region; E: Reproductive system; F-I: Posterior end; J: Spicules and gubernaculum.	1343 1344 1345
Fig. 10. <i>Tylenchorhynchus graciliformis</i> . Female (A, D, E-J) and male (B-C, H). A-B: Entire body; C-D: Anterior end; E: Vulva region; F-H: Posterior end. (Scale-bars: A-B = 100 μ m, C-H = 10 μ m).	1346 1347 1348
Fig. 11. Phylogenetic relationships among <i>Bitylenchus</i> , <i>Sauertylenchus</i> and <i>Tylenchorhynchus</i> species with representatives of other Telotylenchinae genera. Bayesian 50 % majority rule consensus tree as inferred from D2-D3 expansion segments of 28S rRNA sequence alignment under the general time-reversible model of sequence evolution with correction for invariable	1349 1350 1351 1352

sites and a gamma-shaped distribution (GTR + I + G). Newly obtained sequences in this study are shown in bold. (Scale bar = expected changes per site).

Fig. 12. Phylogenetic relationships among *Bitylenchus*, *Sauertylenchus* and *Tylenchorhynchus* species with representatives of other Telotylenchinae genera. Bayesian 50 % majority rule consensus tree as inferred from the partial 18S rRNA gene sequence alignment under the general time-reversible model of sequence evolution with correction for invariable sites and a gamma-shaped distribution (GTR + I + G). Newly obtained sequences in this study are shown in bold. (Scale bar = expected changes per site).

Table 1. Morphometric characters of *Bitylenchus parvulus* n. sp. recovered from Iran. All measurements are in μm , and in the form: mean \pm s.d. (range) and coefficient of variation.

1362

1363

n	Holotype	Paratypes			
	-	Females 18	CV	Males 7	CV
L	816	697 \pm 77.6 (542-834)	11.1	633 \pm 32.9 (582-672)	5.2
H-V	448	390 \pm 40.0 (323-455)	10.2	-	-
H-a	761	649 \pm 73.9 (498-775)	11.3	593 \pm 29.6 (545-627)	4.9
Stylet	18.5	17.7 \pm 0.4 (17.0-18.5)	2.7	16.7 \pm 0.5 (16-18)	3.3
m	52.0	51.4 \pm 1.7 (49-56)	3.4	53 \pm 1.1 (52-55)	2.1
b	5.3	5.0 \pm 0.5 (4.2-5.8)	9.9	4.7 \pm 0.1 (4.4-4.9)	3.1
a	32.5	30.7 \pm 3.0 (26-38)	9.9	34 \pm 1.8 (32-38)	5.4
c	14.8	14.6 \pm 1.0 (12-17)	6.8	15.7 \pm 1.5 (13.1-17.7)	9.7
c'	2.9	2.8 \pm 0.2 (2.3-3.3)	8.8	2.2 \pm 0.2 (1.9-2.5)	9.0
V	54.9	56.0 \pm 1.3 (54.5-60.0)	2.4	-	-
Median bulb	76.0	66.9 \pm 5.3 (58-77)	7.9	63.2 \pm 2.8 (60-69)	4.4
MB	49.7	48.8 \pm 2.5 (45-57)	5.1	48.3 \pm 2.0 (46-51)	4.2
Excretory pore	121	107 \pm 8.7 (93-126)	8.2	102 \pm 3.0 (97-107)	3.0
Pharynx	153	137 \pm 9.3 (123-155)	6.8	131 \pm 5.9 (123-140)	4.5
Body width	25.0	22.6 \pm 1.5 (20-25)	6.9	18.5 \pm 0.6 (17.7-19.5)	3.5
Lateral field	7.8	6.6 \pm 1.0 (5-8)	16.4	5.4 \pm 0.8 (4.5-7.0)	15.8
Vulva body width	25.0	23.1 \pm 1.8 (19-26)	7.9	-	-
Anal body width	19.0	16.5 \pm 2.1 (13-20)	12.8	17.6 \pm 0.9 (16.5-19.5)	5.5
Tail length	55.0	47.5 \pm 4.5 (42-59)	9.5	40.5 \pm 5.4 (34.8-50.0)	13.4
Phasmid	22.0	19.8 \pm 3.3 (13.5-26)	16.9	17.9 \pm 3.6 (13-23)	20.3
Tail annuli	48.0	46 \pm 2.9 (42-52)	6.3	-	-
Spicules	-	-	-	23.2 \pm 1.4 (21.5-25.0)	6.2
Gubernaculum	-	-	-	10.2 \pm 1.1 (9-12)	10.8
Bursa length	-	-	-	58 \pm 7.1 (52-73)	12.3

1364

1365

Table 2. Morphometric characters of *Bitylenchus parvus* recovered from Iran. All measurements are in μm , and in the form: mean \pm s.d. (range).

	Present study		Geraert, 2011	Ghaderi <i>et al.</i> , 2014
	Females	Males	Female/Male	Female/Male
n	7	5	?	26
L	758 \pm 60.3 (644-834)	622 \pm 32.8 (582-672)	650-910	745 (630-863)
H-V	419 \pm 31.2 (358-455)	-	-	-
H-a	707 \pm 55.7 (600-775)	583 \pm 29.2 (545-626)	-	-
Stylet	18.2 \pm 0.2 (18.0-18.7)	16.6 \pm 0.4 (16-17)	16.5-19.5	17.6 (16.5-19)
b	5.2 \pm 0.3 (4.7-5.8)	4.7 \pm 0.1 (4.4-4.9)		6.0 (5.1-7.3)
a	32.6 \pm 3.7 (26-38)	34 \pm 2.1 (32-38)	25-31	31.3 (28-34.9)
c	14.9 \pm 0.6 (14-16)	16 \pm 1.1 (14.6-17.7)	12-17	14.6 (12-17)
c'	2.9 \pm 0.1 (2.5-3.0)	2.2 \pm 0.2 (1.9-2.5)	2.3-3.3	2.9 (2.4-3.4)
V	55.3 \pm 0.8 (54.5-57.0)	-	50-57	53 (50-56)
Median bulb	72 \pm 2.6 (68-77)	62.1 \pm 1.6 (60-64)	-	-
MB	50.4 \pm 3.2 (46-57)	47.4 \pm 1.6 (46-49)	-	-
Excretory pore	115 \pm 7.8 (101-126)	101 \pm 2.7 (97-103)	-	-
Pharynx	144 \pm 6.4 (135-155)	131 \pm 5.1 (128-140)	125-148	-
Body width	23 \pm 1.8 (20-25)	18.5 \pm 0.5 (18-19)	23-33	-
Lateral field	7.7 \pm 0.2 (7.5-8.0)	5.0 \pm 0.3 (4.5-5.5)	-	-
Anal body width	17.5 \pm 1.9 (15-20)	17 \pm 0.6 (16-18)	-	-
Tail length	51 \pm 5.0 (44-59)	39 \pm 4.2 (35-46)	43-63	33-48
Phasmid	21.8 \pm 2.3 (19-26)	17 \pm 3.4 (13-21)	-	-
Tail annuli	48 \pm 2.5 (45-52)	-	42-58	45-50
Spicules	-	23.5 \pm 0.8 (23-25)	28-31	23.9 (22-26)
Gubernaculum	-	9.5 \pm 0.6 (9.0-10.5)	13-13.5	-
Bursa length	-	56 \pm 3.0 (52-60)	-	-

1366

1367

1368

1369

Table 3. Morphometric characters of *Sauertylenchus ibericus* recovered from Iran and its comparison with original description, as well as with populations of *S. maximus*. All measurements are in μm , and in the form: mean \pm s.d. (range) and coefficient of variation.

	<i>S. ibericus</i>			<i>S. maximus</i>	
	Present study		Mahajan & Nombela, 1987	Geraert, 2011	Panahandeh & Pourjam, 2014
	Females	CV	Females	Females	Females
n	13		10	?	10
L	1108 \pm 55.5 (1012-1191)	5.	1130 (990-1270)	940-1620	1329 (1120-1348)
H-V	582 \pm 34.8 (496-620)	5.9	-	-	-
H-a	1056 \pm 53.2 (966-1137)	5.	-	-	-
Styilet	21.8 \pm 0.8 (20-23)	3.6	24.1 (21-26)	20-24.5	-
a	42 \pm 2.1 (39-46)	5.1	43 (40-48)	28-58	41.6 (36.2-46.5)
b	6.1 \pm 0.3 (5.5-6.6)	5.2	6.4 (5.6-7.1)	-	6.7 (6.0-7.3)
c	21.3 \pm 1.3 (19.0-23.1)	6.2	21.8 (18.4-25.1)	16-26	20.4 (18.7-23.5)
c'	2.7 \pm 0.1 (2.5-3.0)	5.6	2.7 (2.3-3.3)	1.9-4.1	2.9 (2.3-4.0)
V	52.5 \pm 2.3 (46-58)	4.5	53.1 (51-56)	47-58	52 (50-54)
Median bulb	96 \pm 3.3 (90-100)	3.5	-	-	-
MB	53.5 \pm 1.6 (51-57)	2.9	57.6 (53-61)	51-58	-
Excretory pore	137.1 \pm 5.1 (130-148)	3.7	143 (111-176)	-	-
Pharynx	179 \pm 5.1 (170-189)	2.8	-	153-192	-
Body width	26.5 \pm 1.5 (24-29)	5.7	-	23-32	-
Lateral field	8.0 \pm 0.8 (6.8-10.0)	10.7	-	8-11	-
Vulva body width	25.7 \pm 1.5 (24-28)	6.	-	-	-
Anal body width	18.8 \pm 1.3 (17-21)	7.1	-	-	-
Tail length	52 \pm 4.0 (46-59)	7.7	-	42-83	-
Phasmid	22.5 \pm 3.3 (19-28)	14.7	-	19-30	-
Tail annuli	32 \pm 1.8 (28-35)	5.9	32-36	25-52	39-47

1373

1374

Table 4. Morphometric characters of *Tylenchorhynchus graciliformis* and *T. agri* recovered from Iran. 1376All measurements are in μm , and in the form: mean \pm s.d. (range) and coefficient of variation. 1377

n	<i>T. graciliformis</i>				<i>T. agri</i>	
	females	CV	males	CV	females	CV
	16		7		8	
L	722 \pm 33.5 (678-779)	4.6	703 \pm 17.4 (682-729)	2.4	659 \pm 31.3 (615-709)	4.7
H-V	405 \pm 24.2 (374-471)	5.9	-		358 \pm 14.0 (338-381)	3.9
H-a	680 \pm 31.0 (636-728)	4.5	660 \pm 17.9 (636-686)	2.7	608 \pm 29.1 (567-654)	4.7
Stylet	16.8 \pm 0.8 (15.5-18.5)	4.9	16.9 \pm 0.3 (16.5-17.5)	2.3	17.6 \pm 0.4 (17-18)	2.6
m	52 \pm 1.2 (50-54)	2.4	51.6 \pm 0.9 (50-53)	1.7	50.0 \pm 0.2 (50.0-50.5)	0.5
a	37 \pm 2.9 (32-45)	8.0	36 \pm 1.3 (34-39)	3.7	31 \pm 1.6 (29-34)	5.3
b	5.3 \pm 0.4 (4.6-6.1)	7.7	5.2 \pm 0.1 (5.0-5.5)	2.7	4.6 \pm 0.2 (4.2-4.9)	4.8
c	17.3 \pm 1.1 (15.2-19.1)	6.4	16.3 \pm 0.7 (14.8-17.1)	4.8	12.9 \pm 0.2 (12.5-13.3)	2.1
c'	3.1 \pm 0.3 (2.6-3.8)	11.2	2.6 \pm 0.1 (2.5-2.8)	4.6	3.6 \pm 0.1 (3.4-3.8)	3.6
V	56 \pm 2.4 (53-64)	4.2	-		54 \pm 1.0 (53-56)	1.9
Median bulb	60 \pm 2.8 (56-65)	4.7	61 \pm 2.7 (57-64)	4.5	63 \pm 2.1 (61-67)	3.3
MB	44 \pm 1.6 (41-48)	3.6	46 \pm 1.5 (44-48)	3.3	44 \pm 1.2 (42-46)	2.7
Excretory pore	104 \pm 3.9 (97-109)	3.8	106 \pm 6.1 (102-120)	5.8	103 \pm 3.3 (99-109)	3.2
Pharynx	136 \pm 6.1 (125-145)	4.5	133 \pm 3.7 (127-139)	2.8	143 \pm 5.8 (134-150)	4.0
Body width	19.5 \pm 1.4 (16-22)	7.3	19 \pm 0.7 (18-20)	3.8	21 \pm 0.9 (19.5-22.0)	4.7
Lateral field	6.0 \pm 0.9 (5.0-7.5)	14.9	5.6 \pm 0.7 (4.5-7.0)	12.7	6.4 \pm 0.3 (6-7)	5.7
Vulva body width	20.0 \pm 1.4 (17.5-22.5)	7.3	-		19.5 \pm 1.2 (17.5-21.0)	6.2
Anal body width	13.5 \pm 1.4 (11-16)	10.6	16 \pm 0.4 (15-16.5)	2.8	14 \pm 0.8 (13-15)	5.9
Tail length	42 \pm 3.7 (36-51)	8.8	43 \pm 1.5 (41-46)	3.7	51 \pm 2.4 (48-55)	4.8
Phasmid	13.5 \pm 2.5 (9-18)	19.1	15 \pm 1.6 (13.5-18.0)	10.6	19 \pm 1.7 (17-22)	9.1
Tail annuli	22.5 \pm 2.2 (19-26)	10.1	-		26.4 \pm 2.5 (23-30)	9.4
Spicules	-		19.7 \pm 0.6 (19-21)	3.5	-	
Gubernaculum	-		10.3 \pm 0.6 (9.5-11.0)	6.6	-	
Bursa length	-		61.4 \pm 4.7 (53-67)	7.7	-	
Post-rectal sac	-		-		15.3 \pm 2.3 (13-20)	15.5
PAS/Tail	-		-		30.1 \pm 3.3 (26.5-36.0)	11.2