Effects of aripiprazole on pupillometric parameters related to pharmacokinetics and pharmacogenetics after single oral administration to healthy subjects

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Short title

Aripiprazole influence on pupil contraction

Key words - aripiprazole, pupillometry, pharmacogenetics, pharmacokinetics

Abstract

Background: Pupillometry is used for the detection of autonomic dysfunction related to numerous diseases and drug administration. Genetic variants in cytochrome P450 (CYP) (*CYP2D6*, *CYP3A4*), dopamine receptor (*DRD2*, *DRD3*), serotonin receptor (*HTR2A*, *HTR2C*) and ATP-binding cassette subfamily B (*ABCB1*) genes were previously associated with aripiprazole response.

Aims: Our aim was to evaluate if aripiprazole affects pupil contraction and its relationship with pharmacokinetics and pharmacogenetics.

Methods: Thirty-two healthy volunteers receiving a 10 mg single oral dose of aripiprazole were genotyped for 15 polymorphisms in *ABCB1*, *CYP2D6*, *DRD2*, *DRD3*, *HTR2A* and *HTR2C* genes by RT-PCR. Aripiprazole and dehydro-aripiprazole plasma concentrations were measured by HPLC-MS/MS. Pupil examination was performed by automated pupillometry.

Results: Aripiprazole caused pupil constriction and reached the peak value at C_{max} . *HTR2A* rs6313 T allele carriers and *HTR2C* rs3813929 C/T subjects showed higher maximum constriction velocity (MCV) and maximum pupil diameter. Besides, Gly/Gly homozygotes for *DRD3* rs6280 showed significantly lower MCV values. A/G heterozygotes for *DRD2* rs6277 showed higher total time taken by the pupil to recover 75% of the initial resting size values. *CYP2D6* intermediate metabolizers showed higher AUC, C_{max} and $T_{1/2}$ than extensive metabolizers. *ABCB1* G2677T/A A/A homozygotes had greater $T_{1/2}$ in comparison with C/C homozygotes. *ABCB1* C3435T T allele carriers and C1236T C/T subjects showed greater AUC than C/C homozygotes.

Conclusions: Aripiprazole affects pupil contraction, which could be a secondary effect through dopamine and serotonin receptors. Pupillometry could be a useful tool to assess autonomic nervous system activity during antipsychotic treatment.

Introduction

Schizophrenia is a chronic and relapsing disorder with generally incomplete remissions, variable degrees of functional impairment and social disability (Alves et al., 2005). Aripiprazole (ARI) is a quinolinone derivative which belongs to the atypical (second generation) antipsychotics commonly prescribed for patients with acute relapse of schizophrenia or schizoaffective disorders. Doses from 15 to 30 mg once a day are effective and well tolerated by patients (McGavin and Goa, 2002). This drug has partial agonist activity at dopamine D2, D3 and serotonin 5-HT1A receptors, and acts also as 5-HT2A and 5-HT7 receptor antagonist (de Bartolomeis et al., 2015; Lawler et al., 1999; Stip and Tourjman, 2010). The main active metabolite, dehydro-aripiprazole (DARI), amounts to 40% of the parent compound in plasma, has high affinity towards dopamine D2 receptors and is an antagonist at 5-HT2A receptors. Therefore, DARI has some pharmacological activity similar to ARI (Kirschbaum et al., 2008; Taylor, 2003; Wood et al., 2006).

The metabolism and elimination of ARI is mainly mediated through two cytochrome P450 enzymes, CYP2D6 and CYP3A4. CYP3A4 is responsible for N-dealkylation and both CYP2D6 and CYP3A4 mediate dehydrogenation and hydroxylation of the drug leading to the main metabolite DARI (Kirschbaum et al., 2008; Urichuk et al., 2008). The mean elimination half-life is about 75 hours for ARI, however, for individuals who do not have sufficient CYP2D6 activity (poor metabolizers; PMs), this can be about 146 hours (Dean, 2012a). Approximately 8% of Caucasians cannot metabolize CYP2D6 substrates and are classified as PMs (Gaedigk et al., 2017). Patients carrying three or more active CYP2D6 alleles (up to 7% in Caucasians), are called CYP2D6 ultra-rapid metabolizers (UMs) (de Leon et al., 2006; Dean, 2012b).

Genetic variants of different genes related to ARI response were analysed in previous studies. A1/A1 genotype for DRD2 Taq1A genetic variant (rs1800497) was reported as a predictor of positive symptom improvement on ARI treatment in patients with schizophrenia (Kwon et al., 2008). In case of -141C Ins/Del (rs1799732) polymorphism, Del allele carriers were associated with lower antipsychotic drug response, compared to Ins/Ins genotype (Zhang et al., 2010). Another study showed that Gly/Gly genotype was more common among schizophrenia patients than Ser/Ser genotype for DRD3 Ser9Gly (rs6280) polymorphism in comparison with healthy controls, therefore, Gly/Gly can be considered as a risk factor for the disease (Loch et al., 2015). Moreover, between pediatric patients under antipsychotic treatment the ATP Binding Cassette Subfamily B Member 1 (ABCB1, coding for P-glycoprotein) T allele carriers for G2677T/A and C3435T polymorphisms had significantly lower plasma concentration/dose ratio for ARI related to patients with other ABCB1 genotypes (Rafaniello et al., 2017). Furthermore, it was found that subjects carrying G/G genotype for HTR2A A-1438G (rs6311) and C/C genotype for T102C (rs6313) polymorphisms have a poorer ARI response, especially for negative symptoms (Chen et al., 2009). Additionally, carrying the C allele of HTR2C -759C/T (rs6318) polymorphism was confirmed to influence weight gain following antipsychotic treatment (Ma et al., 2014).

The iris size, and therefore the pupillary diameter, is controlled by two muscles, the sphincter pupillae and the dilator pupillae, which are mainly under the regulation of the parasympathetic nervous system and the sympathetic nervous system, respectively. Contraction of the sphincter, accompanied by the relaxation of the dilator, produces pupil constriction (miosis), while contraction of the dilator, accompanied by the relaxation of the sphincter, produces pupil dilation (mydriasis) (McDougal and Gamlin, 2014).

Since pupillography was developed in 1958 (Lowenstein and Loewenfeld, 1958), it is used for the detection of autonomic dysfunction associated with numerous diseases, such as diabetes (Piha and Halonen, 1994), heart failure (Keivanidou et al., 2010), Parkinson's disease (Jain et al., 2011), Alzheimer's disease (Fotiou et al., 2000) and hypertension (Koike et al., 2016). Pupillometry is a non-invasive and cost-effective method to determine autonomic nerve activity (Granholm and Steinhauer, 2004). The majority of studies analysing the effect of drugs on the pupil diameter were performed with opioids finding that they cause pupil constriction (miosis) (Harris et al., 2017; Levy-Cooperman et al., 2016; Mangas-Sanjuan et al., 2016; Pickworth et al., 1990). Furthermore, it was shown that carriers of *CYP2D6* EM and PM phenotypes treated with tramadol experienced a decrease of response amplitude, constriction velocity and reaction duration as well as an increase of latency of the pupil (Fliegert et al., 2005), however, the effect was minor in subjects carrying PM phenotype.

In a previous study, the refraction and eye anterior segment parameters were investigated in schizophrenic patients under antipsychotic treatment including ARI and healthy controls. Overall, no difference was found in their pupil diameter (Cumurcu et al., 2015). However, in another study it was confirmed that olanzapine causes a decrease in pupil/iris ratio compared to placebo (Kleinloog et al., 2012). Therefore, only two studies were performed to analyse antipsychotic effects on the pupil and only one included ARI. This drug could affect the pupil size due to its partial agonist activity at dopamine D2, D3 and serotonin 5-HT1A receptors, and antagonist activity at 5-HT2A and 5-HT7 receptors (de Bartolomeis et al., 2015; Lawler et al., 1999; Stip and Tourjman, 2010), as it was proved that serotonin and dopamine are effectors on various types of muscles including the sphincter pupillae and the dilator pupillae (Costagliola et al., 2008). Consequently, ARI administration and the presence of polymorphisms in the genes coding for the receptors involved in its mechanism of action may affect the

response of the pupil.

The aim of the current study was to evaluate if ARI affects various pupillometric parameters in healthy subjects. Furthermore, their connection to pharmacokinetics and pharmacogenetics was also discussed.

Materials and Methods

Study Population

A single-dose bioequivalence trial was performed at the Clinical Trial Unit of Hospital de la Princesa (Madrid, Spain) including thirty-six healthy volunteers (18 men and 18 women) who received 10 mg ARI orodispersible tablets. The protocol was approved by the Research Ethics Committee duly authorized by the Spanish Drugs Agency and under the guidelines of good clinical practice. All subjects were adequately informed about the study and, if agreeing to participate, signed an informed consent form before inclusion.

Subjects were included if they met all the inclusion criteria: male and female volunteers, aged between 18 and 55 years, free from any known organic or psychiatric conditions, normal vital signs and electrocardiogram (ECG), normal medical records and physical examination which were relevant to this trial, and no clinically significant abnormalities in haematology, biochemistry, serology and urine tests. Tests monitoring pregnancy and illicit drug use were performed from urine samples and repeated each hospitalisation day before drug administration. Subjects were excluded if they met any of the following criteria: received prescribed pharmacological treatment in the last 15 days or any kind of medication in the last 48 hours prior to receiving the study medication, body mass index (BMI) outside the 18.5-30.0 kg/m² range, history of drug allergy, suspected

consumers of controlled substances, smokers, daily alcohol consumers and/or acute alcohol poisoning in the previous week, having donated blood in the last month, pregnant or breastfeeding women as well as investigational drug study participants in the previous 3 months. Subjects were free to withdraw from the study at any time.

Study design and procedures

The clinical trial was a phase I, single oral dose, randomized, crossover, two periods, two sequences, open-label, single-centre study with blind determination of the plasma concentrations of ARI and DARI. The volunteers received 10 mg orodispersible ARI tablets. The drug was administered to each healthy volunteer at the same time in each period. Every day the first volunteer received the drug at 9:00 am and the last one at 9:25 am. In the first period, each volunteer received a single dose of one formulation of ARI (test or reference). In the second period, after a washout time frame of 28 days, each volunteer received the same dose of the other formulation. Volunteers fasted from 10 hours before until 5 hours after drug administration. Before drug administration, volunteers rinsed their mouth with 20 ml of water. The reference formulation was Abilify[®] (Otsuka Pharmaceutical Laboratories Europe, London, United Kingdom).

Twenty blood samples were collected from baseline (before administration) until 72 h after dosing. Samples were centrifuged at 3500 rpm (1900G) for 10 minutes and then the plasma was collected and stored at -20° C until the determination of plasma concentrations by the analytical laboratory.

Plasma concentrations of aripiprazole and dehydro-aripiprazole

Plasma concentrations of ARI and DARI were measured by high-performance liquid chromatography tandem mass spectrometry (HPLC-MS/MS) method validated in our laboratory (Wojnicz et al., 2017), with a lower limit of quantification of 0.18 ng/mL for ARI and 0.35 ng/mL for DARI. Stable isotope labelled ARI (ARI-D8) was used as an internal standard for both analytes. For analyte extraction we used three-step microelution-solid-phase extraction (Oasis PRiME HLB 96-well µElution Plate) from 200 µL of human plasma. An octadecyl carbon chain (C18)-bonded silica, reversed phase HPLC column with pentafluorophenyl ligand (to ensure extra selectivity), was used for chromatographic separation. A combination of ammonium formate in Milli-Q water (5 mM; pH 4.0) and acetonitrile (65:35, v/v) was used as the mobile phase at a flow rate of 0.6 mL/min. The total chromatographic run time was 8 minutes, constituting of a 5-minute run time and a 3-minute re-equilibration time. Overall, a volume of 5 µL eluent was injected into the HPLC/MS/MS system. Linearity was confirmed for concentration ranges from 0.18 to 110 ng/mL for ARI and 0.35 to 100 ng/mL for DARI.

Pharmacokinetic Assessments

Pharmacokinetic parameters were calculated by non-compartmental analysis using WinNonlin Professional (Version 2.0, Pharsight Corporation, Palo Alto, California). The maximum plasma concentration (C_{max}) and the elapsed time to reach the maximum plasma concentration (T_{max}) were obtained directly from raw data. The total area under the curve (AUC) from administration to infinity ($AUC_{0-\infty}$) was calculated as the sum of AUC_{0-t} and the residual area (C_t divided by k_e , with C_t as the last measured concentration and k_e as the apparent terminal elimination rate, which was estimated by log-linear regression from the terminal portion of the log-transformed concentration-

time plots). Half-life was calculated by dividing 0.693 by k_e . The total drug clearance adjusted for bioavailability (Cl/F) was calculated by dividing the dose by the AUC_{0-∞} and adjusting for weight. Volume of distribution adjusted for bioavailability (Vd/F) was calculated as Cl/F divided by ke. The AUC and C_{max} were adjusted for dose and weight (AUC/dW and C_{max}/dW) and logarithmically transformed for statistical analysis.

For DARI, only AUC_{0-t} (truncated to 72 hours), C_{max} and T_{max} were determined, because it was not possible to calculate the remaining parameters.

Pupillary light reflex (PLR) measurements

Pupillometric measurements were performed before drug administration and 2, 4, 6, 8, 10, and 12 hours after drug administration recorded with a PRL 200 automated monocular infrared pupillometer (NeurOptics, Irvine, CA, United States). Each measurement was performed in a hospital room with artificial illumination. Before starting the measurement, the subject was instructed to focus on a small target object - a wall chart - that was at least 3 meters away with the eye that was not being tested open (left eye). Stimuli were single light pulses with a fixed intensity of $180 \mu W$ (microwatts) during 154 milliseconds. Visual light stimuli were generated using white light emitting diodes (LED) monocularly. The LED lightning source was placed approximately 2 cm away from the right eye and its largeness was 5×5 mm. Once the device was focused on the target pupil (right eye), a white light stimulus was flashed. The measurements were sampled at a frequency of 32-frames per second and lasted up to 5 seconds, allowing a full or partial recovery of the pupil size after light constriction. The above conditions were identical for all subjects to keep the measurements comparable. Additionally, it was of importance that the eye cup of the pupillometer was correctly fitted as it helps to reduce the possibility of stray light entering the eye during

measurements. In order to adjust for the potential differences in luminosity, consequently in maximum pupil diameter, a digital light (lux) meter (PCE Iberica, S.L., Albacete, Spain) was used to measure the light intensity in lux immediately before each analysis. Light intensity was 96.08 ± 76.89 lux (lm/m²).

We monitored 8 different pupillometric parameters as described in the user guide (NeurOptics, Inc., 2010). Maximum pupil diameter (MAX) and minimum pupil diameter (MIN) represent the pupil diameter before the constriction and just at the peak of the constriction, respectively. CON is the percent of the constriction calculated by (MAX—MIN)/MAX. LAT is the latency and it represents the time of the onset of the constriction. ACV and MCV are the average and the maximum constriction velocity, respectively. The negative sign is to differentiate constriction from dilation, which is the opposite movement. ADV is the average dilation velocity and represents the average pupillary velocity, when, after reaching its constriction, the pupil tends to recover and dilates back to its initial resting size. T75 is the total time taken by the pupil to recover 75% of the initial resting pupil size after it reached the peak of constriction.

Genotyping

DNA was extracted from 1 ml peripheral blood samples using a MagNA Pure LC DNA Isolation Kit in an automatic DNA extractor (MagNa Pure® System, Roche Applied Science, Indianapolis, Indiana) and quantified spectrophotometrically in a NanoDrop® ND-1000 Spectrophotometer (Nanodrop Technologies, Wilmington, Delaware).

The *CYP2D6* *3, *4 and *5 polymorphisms were analysed by RT-PCR using LightCyler® 2.0 instrument (RocheDiagnostics, Mannheim, Germany). The primers and the probes were designed by TIB MOLBIOL (Berlin, Germany) for this purpose.

ABCB1 C3435T, C1236T, G2677T/A, *DRD2* rs6277, rs1800497, *DRD3* rs6280, *CYP2D6* *6, *7 and *9, *HTR2A* rs6313, rs6314 and *HTR2C* rs3813929 polymorphisms were genotyped using a StepOnePlusTM PCR instrument (Applied Biosystems, Foster City, California). To determine the copy number variations (CNVs) in the *CYP2D6* gene a TaqMan Copy Number Assay (Applied Biosystems, Foster City, California) was used, which detects a specific sequence on exon 9. For statistical reasons, all the samples were run in quadruplicates in the instrument. A positive control was used from Coriell Institute with a known copy number of *CYP2D6* (3 copies, Coriell ID: NA17221). The assay was performed with an endogenous control, TaqManRNase P Copy Number Reference Assay (Applied Biosystems, Foster City, California). To determine the exact number of *CYP2D6* copies we used CopyCaller Software (Applied Biosystems, Foster City, California) that uses the comparative $\Delta\Delta$ CT method (Schmittgen and Livak, 2008).

Statistical analysis

Statistical analysis was performed with the SPSS 19.0 software (SPSS Inc., Chicago, Illinois). P values lower or equal than 0.05 were considered significant. The Hardy-Weinberg equilibrium was estimated for all analysed variants. Deviations from the equilibrium were detected by comparing the observed and expected frequencies using a Fisher exact test based on the De Finetti program (available at <u>http://ihg.gsf.de/cgi-bin/hw/hwa1.pl</u>).

The changes between the recorded values in different time points in pupillometric parameters (MAX, MIN, T75, MCV, CON, ACV, ADV, LAT) after the administration of ARI were analysed by repeated measures ANOVA. The values were adjusted for

differences in light intensity immediately before each analysis. Differences in pharmacokinetic and pupillometric parameters between individuals with distinct genotypes were statistically analysed by a parametric univariate analysis (t -test or ANOVA). A simple linear regression was performed to predict pupillometric parameters based on pharmacokinetic data (AUC, C_{max} , $T_{1/2}$, T_{max}). Multiple linear regression models were used to study factors related to all the pupillometric and pharmacokinetic dependent variables. Models for each dependent with standardized (β) regression coefficients are shown in results. *CYP2D6* genotypes were classified in 4 phenotypes (PM, intermediate metabolizer: IM, extensive (normal) metabolizer: EM, and UM) according to the method used by Gaedigk et al., which is based on the functionality of alleles (Gaedigk et al., 2008), but PM and UM were not found in this study.

Results

Demographic and genotypic characteristics

Thirty-six healthy volunteers were included in the bioequivalence study. All of them completed it, however, only 32 (17 men and 15 women) participated in the pharmacogenetic and the pupillometric analyses. Average age was distinctively different between men and women, but it did not reach the significant level (25.6 ± 7.0 years and 31.0 ± 10.7 years, respectively, p = 0.06). Notably, men had greater weight than women (77.4 ± 8.7 kg and 62.4 ± 8.8 kg, respectively, p < 0.001), but the BMI values did not differ significantly (25.5 ± 2.2 kg/m² and 23.8 ± 3.5 kg/m², respectively, p=0.06). All the subjects were Caucasian.

The genotype frequencies of the analysed genes are shown in *Table* 1. All genetic variants were in Hardy-Weinberg equilibrium ($p \ge 0.05$). Differences in genotype frequencies were found between men and women in *CYP2D6* phenotype and *DRD2* rs6277 and *HTR2C* rs3813929 polymorphisms, as shown in *Table* 1. In case of *HTR2C* rs3813929 polymorphism, it is explained by the location of the gene on chromosome X.

Gene / variant	Genotype	Total	Men	Women
CYP2D6 PHENOTYPE	IM	13 (39.4)	4 (30.8)	9 (69.2)
(N=32)	EM	19 (60.6)	13 (68.4)	6 (32.5)*
ABCB1/C3435T (N=32)	C/C	7 (21.8)	5 (23.8)	2 (28.6)
	C/T	16 (50.0)	10 (47.6)	6 (37.5)
	T/T	9 (28.2)	6 (28.6)	3 (33.3)
<i>АВСВ1</i> / С1236Т	C/C	13 (40.6)	7 (41.2)	6 (40.0)
(N=32)	C/T	15 (46.9)	8 (47.1)	7 (46.7)
	T/T	4 (12.5)	2 (11.8)	2 (13.3)
<i>ABCB1</i> / G2677T/A	C/C	11 (34.4)	6 (35.3)	5 (33.3)
(N=32)	C/A	15 (46.9)	8 (47.1)	7 (46.7)
	C/T	2 (6.25)	1 (5.88)	1 (6.67)
	A/A	4 (12.5)	2 (11.8)	2 (13.3)
<i>DRD2</i> / rs6277 (N=32)	A/A	9 (28.1)	3 (17.6)	6 (40.0)
	A/G	17 (53.1)	8 (47.1)	9 (60.0)
	G/G	6 (18.8)	6 (35.3)	0 (0.0)*
<i>DRD2</i> / rs1800497	A1/A1	1 (3.1)	0 (0.0)	1 (6.67)
(N=32)	A1/A2	8 (25.0)	4 (23.5)	4 (26.7)
	A2/A2	23 (71.9)	13 (76.5)	10 (66.7)
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<i>DRD3</i> / rs6280 (N=32)	Ser/Ser	11 (34.3)	6 (35.3)	5 (33.3)
	Ser/Gly	17 (53.1)	9 (52.9)	8 (53.3)
	Gly/Gly	4 (12.5)	2 (11.8)	2 (13.3)
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<i>HTR2A</i> / rs6313 (N=32)	C/C	11 (34.4)	4 (23.5)	7 (46.7)

Table **1.** Genotype frequencies of the analysed polymorphisms. Values are expressed as number of subjects (percentage from the total).

	C/T	11 (34.4)	5 (29.4)	6 (40.0)
	T/T	10 (31.3)	8 (47.1)	2 (13.3)
<i>HTR2A</i> / rs6314 (N=32)	C/C	25 (78.1)	13 (76.5)	12 (80.0)
	C/T	7 (21.9)	4 (23.5)	3 (20.0)
<i>HTR2C</i> / rs3813929 (N=32)	C/C C/T T/T	20 (62.5) 7 (21.9) 5 (15.6)	13 (23.5) 0 (0.0) 4 (47.1)	7 (46.7) 7 (40.0)* 1 (13.3)

*P<0.05 versus men. Abbreviations: *ABCB1*: ATP binding cassette subfamily B member 1; CYP: cytochrome p450 oxidase; *HTR2A*: serotonin receptor 2A; *HTR2C*: serotonin receptor 2C; *DRD2*: dopamine D2 receptor; *DRD3*: dopamine D3 receptor; IM: intermediate metabolizer; EM: extensive metabolizer.

Pharmacokinetic analysis

Mean and standard deviation (SD) of ARI and DARI pharmacokinetic parameters are shown in *Table* 2. After adjusting for weight, sex differences were found in ARI Vd/F (p = 0.01) and in Cl/F (p = 0.01) as women had higher and lower level than men, respectively. Women showed higher T_{1/2}, C_{max} and AUC_{0-t} (p < 0.05), but these differences disappeared after adjusting for weight. No differences were found between sexes in DARI pharmacokinetics.

Table 2. Aripiprazole and dehydro-aripiprazole pharmacokinetic parameters after a sing	;le
oral dose of 10 mg.	

Aripiprazole	All	Men	Women
AUC ₀₋₇₂ (ng·h/mL)	1570.0±335.6	1441.1±255.9	1698.9±371.5
C _{MAX} (ng/mL)	48.1±9.7	44.7±8.6	51.6±9.4
T _{MAX} (h)	2.9±1.7	2.8±1.7	3.0±1.7
T½ (h)	58.3±30.0	48.7±29.0	67.9±28.6

Vd/F (L/kg)	4.45±0.80	4.07±0.50	4.81±0.90*
Cl/F (mL/h·kg)	60.0±20.0	70.0±20.0	50.0±20.0*
Dehydro-aripiprazole			
AUC ₀₋₇₂ (ng·h/mL)	376.6±136.2	389.7±145.9	363.4±128.7
C _{MAX} (ng/mL)	6.6±2.3	6.7±2.5	6.4±2.1
T _{MAX} (h)	49.6±19.0	46.7±20.5	52.6±17.3

*P \leq 0.05 versus men after adjusting for weight.

Abbreviations: C_{max} : maximum plasma concentration; T_{max} : time to reach the maximum plasma concentration; AUC₀. ₇₂: area under the curve between 0 and 72 h after dosing; $T_{1/2}$: half-life; Cl/F: total drug clearance adjusted for bioavailability; Vd/F: volume of distribution adjusted for bioavailability.

The effect of aripiprazole on pupillometry and its relationship with pharmacokinetics

Following oral administration of ARI, significant decrease was observed at 2 h in pupillary parameters MAX, MIN, T75 and MCV, therefore, the pupil showed constriction. The changes reached their peak value at 2-4 hours after drug administration, concurrently when ARI reached the maximum concentration of 48 ng/mL (C_{max}) at 2.9 h (T_{max}). Thereafter, they started to increase linearly in time until the last measurement point (12 h). (*Figure* 1). No differences were found between men and women in any of these pupillometric parameters.

Mean and SD of baseline, 2 h after drug administration and maximum values of the pupillometric parameters are shown in *Table* 3.

Table 3. Pupillometric parameters before and after administration of aripiprazole 10 mg. Values are expressed as mean \pm SD.

Pupillometric parameters	Baseline value (0h)	2h	Last measured value (12h)	Partial eta squared
MAX (mm)	5.00±0.84	4.30±1.01**	5.80±0.88**	0.59
MIN (mm)	3.41±0.69	3.03±0.75*	4.16±0.86*	0.56
T75 (s)	1.52 ± 0.60	1.25±0.60**	1.64 ± 0.63	0.12
MCV (mm/s)	-4.88±0.74	-4.17±1.02**	$-4.74\pm0.71*$	0.26
ACV (mm/s)	-3.79±0.59	-3.20 ± 0.79	-3.71±0.55	0.14
CON (%)	-29±0.04	-26±0.05	-27±0.06	0.06
ADV (mm/s)	$1.00{\pm}0.28$	$0.90{\pm}0.31$	$1.00{\pm}0.22$	0.08
LAT (s)	$0.20{\pm}0.02$	$0.20{\pm}0.03$	$0.20{\pm}0.02$	0.04

Abbreviations: MAX: maximum pupil diameter; MIN: minimum pupil diameter; CON: percent of the constriction; LAT: latency; ACV: average constriction velocity; MCV: maximum constriction velocity; ADV: dilation velocity; T75: total time taken by the pupil to recover 75% of the initial resting pupil size after it reached the peak of constriction. $*p \le 0.05$ related to the baseline value. $**p \le 0.001$ related to the baseline value.

Moreover, C_{max} and $T_{1/2}$ had influence on MAX and MIN pupillometric parameters. When MAX was estimated, it was found that it has an indirect relationship with $T_{1/2}$ (non-standardized β coefficient = -0.01, p = 0.02, r² = 0.16, Cohen's f² = 0.19). However, no significant effect was found with C_{max} (non-standardized β coefficient = -0.04, p = 0.09, r² = 0.10, Cohen's f² = 0.11). Regarding MIN, we detected the same tendency: an indirect relationship with $T_{1/2}$ (non-standardized β coefficient = -0.01, p = 0.01, r² = 0.18, Cohen's f² = 0.22), however, C_{max} did not reach the statistically significant level (non-standardized β coefficient = -0.01, p = 0.10, r² = 0.02, Cohen's f² = 0.01). No interrelation was found with the remaining parameters. Moreover, after analysing heart rate variability as another marker of autonomic function, no association was found with the pupillometric parameters at different time points.

The influence of polymorphisms on pupillometry

TT subjects for the rs6313 SNP in *HTR2A* gene showed higher MCV (p=0.001, $r^2 = 0.399$, Cohen's $f^2 = 0.664$) and MAX (p = 0.030, $r^2 = 0.226$, Cohen's $f^2 = 0.292$) (*Figure* 2). Furthermore, carriers of T allele for the rs3813929 of *HTR2C* showed higher T75 (p = 0.044, $r^2 = 0.131$, Cohen's $f^2 = 0.151$), but this effect disappeared after Bonferroni correction (p = 0.078). Additionally, the multivariate analysis revealed that TT subjects for this SNP show higher MAX (p=0.008, $r^2 = 0.214$, Cohen's $f^2 = 0.272$) and MCV (p = 0.000, $r^2 = 0.348$, Cohen's $f^2 = 0.534$) values. Besides, carriers of Gly/Gly genotype for *DRD3* rs6280 SNP showed significantly lower MCV values (p = 0.013, $r^2 = 0.474$, Cohen's $f^2 = 0.901$). Additionally, AG heterozygotes for *DRD2* rs6277 SNP showed higher T75 values (p = 0.047, $r^2 = 0.125$, Cohen's $f^2 = 0.143$).

However, polymorphisms in *ABCB1* (C3435T, C1236T, G2677T/A) and *DRD2* (rs1800497) genes and *CYP2D6* phenotypes had no influence on pupillometric parameters.

Pharmacokinetics and polymorphisms

Bivariate and multivariate analyses revealed associations between several pharmacokinetic parameters and polymorphisms in the analysed genes. *CYP2D6* intermediate metabolizers (IM) showed higher AUC (p = 0.010, $r^2 = 0.198$, Cohen's $f^2 = 0.247$), C_{max} (p = 0.012, $r^2 = 0.187$, Cohen's $f^2 = 0.230$) and $T_{1/2}$ (p = 0.041, $r^2 = 0.128$, Cohen's $f^2 = 0.147$) than extensive metabolizers (EM) (*Figure 3*). Moreover, G2677T/A polymorphism in *ABCB1* gene had an influence on the $T_{1/2}$ of ARI, but it was only significant in the multivariate analysis (bivariate analysis: p = 0.055, $r^2 = 0.181$, Cohen's $f^2 = 0.221$; multivariate analysis: non-standardized β coefficient = 0.35; p = 0.040, $r^2 = 0.243$, Cohen's $f^2 = 0.321$) as A/A homozygotes had greater value (74.9 ± 39.8) in

comparison with C/C homozygotes (40.4 \pm 10.0). The minor allele carriers (C/T and T/T) of C3435T polymorphism showed greater AUC (p = 0.074, r² = 0.164, Cohen's f² = 0.196) in the univariate analysis, although they did not reach statistical significance. However, in the multivariate analysis, this reached a statistically significant level (non-standardized β coefficient = 0.33; p = 0.045, r² = 0.337, Cohen's f² = 0.508). Moreover, it revealed that heterozygote CT subjects for C1236T SNP had also higher level of AUC (non-standardized β coefficient = 0.39; p = 0.019, r² = 0.234, Cohen's f² = 0.305) (*Figure* 3). No association was found between DARI and the polymorphisms of *CYP2D6* or *ABCB1*.

Discussion

Sex Influence on Pharmacokinetics

ARI pharmacokinetics is affected by sex based on a previous study performed by our group as AUC, C_{max} , $T_{1/2}$, and Vd/F were higher in women than in men (Belmonte et al., 2016). In the current analysis, these results were verified and additionally Cl/F was lower in women than in men. These findings are consistent with previous studies (Islam et al., 2017; Schwartz, 2003). However, they are mostly explained by differences in body weight as it is shown in the drug label. It states that C_{max} and AUC of aripiprazole are 30-40% higher in women than in men (Abilify, Otsuka Pharmaceutical, Co., Ltd., 2014). Sex-related variability in pharmacokinetics of psychotropic drugs can be due to the differences in gastric acidity, intestinal motility, body weight and composition, blood volume, liver enzymes (mainly the cytochrome P450), renal excretion or the sensitivity to side effects and toxicity of drugs (Marazziti et al., 2013).

Pharmacokinetics and pupillometry

Pupillary dilation as a marker of drug occupancy and action has been measured several times in previous studies (Graur and Siegle, 2013; Siepmann et al., 2007). However, mostly the effects of opioid drugs were analysed. Miosis is thought to be caused by a direct opioid effect during the use of these drugs on μ receptors in the autonomic nucleus of the oculomotor nerve (Larson et al., 1997). In addition to agonist activity at the μ -opioid receptor, tramadol, an opioid drug used during acute or chronic pain, also inhibits the re-uptake of the neurotransmitters norepinephrine and serotonin (5-HT) in the descending inhibitory pain pathways, thus, facilitating 5-HT release (Raffa et al., 1992). Based on this information, as ARI has antagonist activity at 5-HT2A and 5-HT7 receptors, we suspected that the drug has a similar effect on the pupil as tramadol has.

According to our knowledge, antipsychotic effects on pupillary motility was explored only in patients treated with olanzapine, where it caused a decrease in pupil/iris ratio (Kleinloog et al., 2012). In another study, antipsychotic drugs (risperidone, olanzapine, aripiprazole, amilsulpiride, clozapine, quetiapine and ziprasidone) did not cause a change in pupil diameter compared with placebo (Cumurcu et al., 2015).

The pupillary parameters MAX, MIN, MCV and T75 significantly changed after 2-4 hours of drug administration, exactly when ARI reached C_{max} . The background of these changes could be rooted in the fact that serotonin is an effector on various types of smooth muscles, functioning also at the level of ciliary muscle and sphincter of the pupil (Costagliola et al., 2008). Serotonin and its receptors 5-HT1A and 5-HT2A are proved to be abundantly expressed in mammalian iris-ciliary body complex (ICB) (Barnett and Osborne, 1993a; Tobin et al., 1988; Tobin and Osborne, 1989). Additionally, it is known that higher levels of serotonin causes pupil dilation when it

binds to its receptors (Yu et al., 2004). Moreover, dopamine can cause pupil dilation or constriction through sympathetic and parasympathetic nerves, respectively (Spiers and Calne, 1969). Based on our results, pupil constriction could be due to the partial antagonist activity of ARI at 5-HT2A and 5-HT7 receptors, because ARI is a partial agonist at dopamine receptors, therefore, it would cause dilated pupils. Therefore, clinicians could assess autonomic nervous system activity in schizophrenic patients treated with antipsychotic drugs. This would allow to estimate subclinical signs with a non-invasive method like pupillometry.

Polymorphisms and pupillometry

So far only a few related studies were carried out to analyse the relationship between pupil miosis and mydriasis in response to drugs and genetic polymorphisms. When evaluating the effects of the *CYP2D6* gene duplication on the pharmacokinetics and pharmacodynamics of tramadol, the mean maximal decrease speed in pupil diameter was 1.5-fold longer in UMs compared with EMs (Fliegert et al., 2005). In contrast, in our study, no association was found between *CYP2D6* phenotypes when analysing ARI pharmacokinetics and the pupillometric parameters. Therefore, the effect of *CYP2D6* on the pupil was not measurable. Additionally, probably due to the low sample size, we could not find UM and PM among the studied group.

No *ABCB1* genetic variants were found which could predict pupil miotic and mydriatic effects while administrating the opioid drug loperamide (Skarke et al., 2003). To the extent of our knowledge, no other study was carried out to analyse the genetic differences in *ABCB1* related to pupil contraction. We could not associate C3435T, C1236T and G2677T/A polymorphisms in the gene with any of the pupillometric parameters. Second-generation antipsychotics such as amisulpride, aripiprazole,

risperidone and paliperidone are transported by P-gp (Moons et al., 2011). A P-gp inhibitor, HM30181, increases the oral bioavailability of co-administered drugs (Kim et al., 2014, p. 30; Kwak et al., 2010). Pupil diameter was slightly decreased or remained almost the same over time when loperamide was administered alone or co-administered with HM30181, respectively, whereas it was markedly decreased when quinidine -a CYP2D6 inhibitor- was co-administered, suggesting that the influence of P-gp could be minor compared to CYP2D6 (Kim et al., 2014, p. 30; Skarke et al., 2003).

It is known that dopamine administration produces marked dilatation of the pupil (Ong and Bruning, 1981; Spiers and Calne, 1969). The administration of neurotensin, which is a neuromodulator of dopamine transmission, induced miosis in the rabbit eye (Hernandez and Jennes, 1990). We found that heterozygotes for *DRD2* rs6277 SNP showed higher T75 values and Gly/Gly homozygotes for *DRD3* rs6280 SNP showed lower MCV values. ARI is a partial agonist for dopamine D2 and D3 receptors (Burris et al., 2002). Based on our previous results, we concluded that ARI effects on the pupil are conceivably due to its antagonism at serotonin receptors. However, as we found a relationship between dopamine receptor genes and MCV and T75 values, presumably the mechanism is more complex and further studies are needed to confirm our findings.

We found that TT subjects of *HTR2A* rs6313 polymorphism showed higher MCV and MAX values and carriers of T allele of *HTR2C* rs3813929 showed higher T75 value. Serotonin receptors, in particular the 5-HTlA subtype, may be associated with the iris sphincter muscle and suggest their involvement in the regulation of pupil size (Barnett and Osborne, 1993b). This notion was further strengthened by the finding that pupil size is decreased by the intravenous injection of ketanserin, which is an antagonist primarily of 5-HT2 receptors (Krootila et al., 1987). ARI has agonist activity at 5-HT1A receptors

and antagonist activity at 5-HT2A receptors, therefore, the polymorphisms in these genes may have a role in showing distinct pupil phenotypes in response to the drug.

Pharmacokinetics and polymorphisms

Our study confirms the impact of *CYP2D6* phenotypes on ARI pharmacokinetics (Hendset et al., 2007; Suzuki et al., 2014). In our previous study, the AUC_{0-t} of ARI was increased by 20% in IM and by 50% in PM compared to EM (Belmonte et al., 2018). In the current analysis, we obtained matching results as *CYP2D6* IM showed higher AUC (32%), C_{max} (26%) and $T_{1/2}$ (42%) values than EM. Based on these results, a dose reduction could be recommended for IM, but U.S. Food & Drug Administration (FDA) only recommends it for PM, a phenotype which we did not find among our subjects (Abilify, Otsuka Pharmaceutical, Co., Ltd., 2014).

To date, there are no consistent findings about the role of polymorphisms in *ABCB1*. In our previous study, we showed that C1236T SNP has an influence on the pharmacokinetics of ARI, which could be caused by the lower P-gp activity of T/T subjects (Belmonte et al., 2018). In a study performed in pediatric patients, an effect of both polymorphisms was revealed on ARI pharmacokinetics as subjects with the 2677TT and 3435TT genotypes had lower ARI concentrations (Rafaniello et al., 2017). On the contrary, we found that T allele carriers of C3435T polymorphism showed greater AUC values. To date, there are no consistent results about the role of *ABCB1* polymorphisms; therefore, further studies should be performed before dose adjustments.

Study limitations

The main limitation of our study is the low sample size, as only 32 healthy subjects were included. Therefore, it is important to interpret these results with caution. This could be the reason why we could not find PM and UM phenotypes for *CYP2D6*.

Studies including more subjects are necessary to increase the statistical reliability of the results. Moreover, we administered single-dose of ARI, which prevents us to assess longer-term effects on the pupil. Furthermore, pharmacokinetic and pharmacogenetic effects on pupillometry could be different in psychotic patients receiving long-term treatment compared to healthy volunteers. Therefore, we are not able to apply pupillometry as a tool to assess autonomic dysfunction in a clinical setting yet, further studies are needed.

Conclusions

ARI administration produced pupil constriction: MAX, MIN, MCV and T75 parameters were decreased to the peak value when ARI reached C_{max} . Additionally, effects of ARI on pupillometric parameters are associated with polymorphisms in serotonin (*HTR2A* and *HTR2C*) and dopamine (*DRD2* and *DRD3*) receptor genes. Pharmacokinetics of ARI is affected by sex as Vd/F was higher, and Cl/F was lower in women than in men. Moreover, *CYP2D6* phenotypes had an impact on AUC, C_{max} and $T_{1/2}$ of aripiprazole. Additionally, *ABCB1* polymorphisms were associated with $T_{1/2}$ and AUC values. In conclusion, pupillometry could be an important and useful tool to assess the autonomic nervous system activity during antipsychotic drug treatment. It could estimate the subclinical signs of adverse effects, therefore predicting them beforehand. Furthermore, the main advantage of these measurements is their non-invasive character.

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References

- Abilify, Otsuka Pharmaceutical, Co., Ltd., 2014. Food and Drug Administration. Aripiprazole label. Available at: http://www.accessdata.fda.gov/drugsatfda_docs/label/2014/021436s038, 021713s030,021729s022,021866s023lbl.pdf.
- Alves, T.M., Pereira, J.C.R., Elkis, H., 2005. The psychopathological factors of refractory schizophrenia. Rev. Bras. Psiquiatr. Sao Paulo Braz. 1999 27, 108–112. https://doi.org//S1516-44462005000200007
- Barnett, N.L., Osborne, N.N., 1993a. The presence of serotonin (5-HT1) receptors negatively coupled to adenylate cyclase in rabbit and human iris-ciliary processes. Exp. Eye Res. 57, 209–216. https://doi.org/10.1006/exer.1993.1116
- Barnett, N.L., Osborne, N.N., 1993b. The effect of serotonin on the rabbit isolated iris sphincter muscle. Curr. Eye Res. 12, 665–673.
- Belmonte, C., Ochoa, D., Román, M., Cabaleiro, T., Talegón, M., Sánchez-Rojas, S.D., Abad-Santos, F., 2016. Evaluation of the Relationship Between Pharmacokinetics and the Safety of Aripiprazole and Its Cardiovascular Effects in Healthy Volunteers. J. Clin. Psychopharmacol. 36, 608–614. https://doi.org/10.1097/JCP.00000000000577
- Belmonte, C., Ochoa, D., Román, M., Saiz-Rodríguez, M., Wojnicz, A., Gómez-Sánchez, C.I., Martín-Vilchez, S., Abad-Santos, F., 2018. Influence of CYP2D6, CYP3A4, CYP3A5 and ABCB1 polymorphisms on pharmacokinetics and safety of aripiprazole in healthy volunteers. Basic Clin. Pharmacol. Toxicol. https://doi.org/10.1111/bcpt.12960
- Burris, K.D., Molski, T.F., Xu, C., Ryan, E., Tottori, K., Kikuchi, T., Yocca, F.D., Molinoff, P.B., 2002. Aripiprazole, a novel antipsychotic, is a high-affinity partial agonist at human dopamine D2 receptors. J. Pharmacol. Exp. Ther. 302, 381–389.

- Chen, S.-F., Shen, Y.-C., Chen, C.-H., 2009. HTR2A A-1438G/T102C polymorphisms predict negative symptoms performance upon aripiprazole treatment in schizophrenic patients. Psychopharmacology (Berl.) 205, 285–292. https://doi.org/10.1007/s00213-009-1538-z
- Costagliola, C., Parmeggiani, F., Semeraro, F., Sebastiani, A., 2008. Selective Serotonin Reuptake Inhibitors: A Review of its Effects on Intraocular Pressure. Curr. Neuropharmacol. 6, 293–310. https://doi.org/10.2174/157015908787386104
- Cumurcu, T., Keser, S., Cumurcu, B.E., Gunduz, A., Kartalci, S., 2015. Refraction and eye anterior segment parameters in schizophrenic patients. Arq. Bras. Oftalmol. 78, 180– 184. https://doi.org/10.5935/0004-2749.20150046
- de Bartolomeis, A., Tomasetti, C., Iasevoli, F., 2015. Update on the Mechanism of Action of Aripiprazole: Translational Insights into Antipsychotic Strategies Beyond Dopamine Receptor Antagonism. CNS Drugs 29, 773–799. https://doi.org/10.1007/s40263-015-0278-3
- de Leon, J., Armstrong, S.C., Cozza, K.L., 2006. Clinical guidelines for psychiatrists for the use of pharmacogenetic testing for CYP450 2D6 and CYP450 2C19. Psychosomatics 47, 75–85. https://doi.org/10.1176/appi.psy.47.1.75
- Dean, L., 2012a. Aripiprazole Therapy and CYP2D6 Genotype, in: Pratt, V., McLeod, H., Dean, L., Malheiro, A., Rubinstein, W. (Eds.), Medical Genetics Summaries. National Center for Biotechnology Information (US), Bethesda (MD).
- Dean, L., 2012b. Codeine Therapy and CYP2D6 Genotype, in: Pratt, V., McLeod, H., Dean, L., Malheiro, A., Rubinstein, W. (Eds.), Medical Genetics Summaries. National Center for Biotechnology Information (US), Bethesda (MD).
- Fliegert, F., Kurth, B., Göhler, K., 2005. The effects of tramadol on static and dynamic pupillometry in healthy subjects--the relationship between pharmacodynamics, pharmacokinetics and CYP2D6 metaboliser status. Eur. J. Clin. Pharmacol. 61, 257–266. https://doi.org/10.1007/s00228-005-0920-y
- Fotiou, F., Fountoulakis, K.N., Tsolaki, M., Goulas, A., Palikaras, A., 2000. Changes in pupil reaction to light in Alzheimer's disease patients: a preliminary report. Int. J. Psychophysiol. Off. J. Int. Organ. Psychophysiol. 37, 111–120.
- Gaedigk, A., Sangkuhl, K., Whirl-Carrillo, M., Klein, T., Leeder, J.S., 2017. Prediction of CYP2D6 phenotype from genotype across world populations. Genet. Med. Off. J. Am. Coll. Med. Genet. 19, 69–76. https://doi.org/10.1038/gim.2016.80
- Gaedigk, A., Simon, S.D., Pearce, R.E., Bradford, L.D., Kennedy, M.J., Leeder, J.S., 2008. The CYP2D6 activity score: translating genotype information into a qualitative measure of phenotype. Clin. Pharmacol. Ther. 83, 234–242. https://doi.org/10.1038/sj.clpt.6100406
- Granholm, E., Steinhauer, S.R., 2004. Pupillometric measures of cognitive and emotional processes. Int. J. Psychophysiol. Off. J. Int. Organ. Psychophysiol. 52, 1–6. https://doi.org/10.1016/j.ijpsycho.2003.12.001

- Graur, S., Siegle, G., 2013. Pupillary Motility: Bringing Neuroscience to the Psychiatry Clinic of the Future. Curr. Neurol. Neurosci. Rep. 13. https://doi.org/10.1007/s11910-013-0365-0
- Harris, S.C., Cipriano, A., Colucci, S.V., Kapil, R.P., Geoffroy, P., Hopyan, T., Levy-Cooperman, N., 2017. Oral Abuse Potential, Pharmacokinetics, and Safety of Once-Daily, Single-Entity, Extended-Release Hydrocodone (HYD) in Recreational Opioid Users. Pain Med. Malden Mass 18, 1278–1291. https://doi.org/10.1093/pm/pnw208
- Hendset, M., Hermann, M., Lunde, H., Refsum, H., Molden, E., 2007. Impact of the CYP2D6 genotype on steady-state serum concentrations of aripiprazole and dehydroaripiprazole. Eur. J. Clin. Pharmacol. 63, 1147–1151. https://doi.org/10.1007/s00228-007-0373-6
- Hernandez, D.E., Jennes, L., 1990. Inhibition of neurotensin-induced miosis by blockade of ocular dopamine pathways. J. Ocul. Pharmacol. 6, 31–36.
- Islam, M.M., Iqbal, U., Walther, B.A., Nguyen, P.-A., Li, Y.-C.J., Dubey, N.K., Poly, T.N., Masud, J.H.B., Atique, S., Syed-Abdul, S., 2017. Gender-based personalized pharmacotherapy: a systematic review. Arch. Gynecol. Obstet. 295, 1305–1317. https://doi.org/10.1007/s00404-017-4363-3
- Jain, S., Siegle, G.J., Gu, C., Moore, C.G., Ivanco, L.S., Jennings, J.R., Steinhauer, S.R., Studenski, S., Greenamyre, J.T., 2011. Autonomic insufficiency in pupillary and cardiovascular systems in Parkinson's disease. Parkinsonism Relat. Disord. 17, 119–122. https://doi.org/10.1016/j.parkreldis.2010.11.005
- Keivanidou, A., Fotiou, D., Arnaoutoglou, C., Arnaoutoglou, M., Fotiou, F., Karlovasitou, A.,
 2010. Evaluation of autonomic imbalance in patients with heart failure: a preliminary study of pupillomotor function. Cardiol. J. 17, 65–72.
- Kim, T.-E., Lee, H., Lim, K.S., Lee, S., Yoon, S.-H., Park, K.-M., Han, H., Shin, S.-G., Jang, I.-J., Yu, K.-S., Cho, J.-Y., 2014. Effects of HM30181, a P-glycoprotein inhibitor, on the pharmacokinetics and pharmacodynamics of loperamide in healthy volunteers. Br. J. Clin. Pharmacol. 78, 556–564. https://doi.org/10.1111/bcp.12368
- Kirschbaum, K.M., Müller, M.J., Malevani, J., Mobascher, A., Burchardt, C., Piel, M., Hiemke, C., 2008. Serum levels of aripiprazole and dehydroaripiprazole, clinical response and side effects. World J. Biol. Psychiatry Off. J. World Fed. Soc. Biol. Psychiatry 9, 212–218. https://doi.org/10.1080/15622970701361255
- Kleinloog, D., Liem-Moolenaar, M., Jacobs, G., Klaassen, E., de Kam, M., Hijman, R., van Gerven, J., 2012. Does olanzapine inhibit the psychomimetic effects of Δ⁹tetrahydrocannabinol? J. Psychopharmacol. Oxf. Engl. 26, 1307–1316. https://doi.org/10.1177/0269881112446534
- Koike, Y., Kawabe, T., Nishihara, K., Iwane, N., Hano, T., 2016. Effects of azelnidipine and amlodipine on exercise-induced sympathoexcitation assessed by pupillometry in hypertensive patients. Hypertens. Res. Off. J. Jpn. Soc. Hypertens. 39, 863–867. https://doi.org/10.1038/hr.2016.94

- Krootila, K., Uusitalo, H., Palkama, A., 1987. Effect of alpha-adrenergic and serotonergic blockers on the acute irritative response in the rabbit eye. Exp. Eye Res. 45, 721–729.
- Kwak, J.-O., Lee, S.H., Lee, G.S., Kim, M.S., Ahn, Y.-G., Lee, J.H., Kim, S.W., Kim, K.H., Lee, M.G., 2010. Selective inhibition of MDR1 (ABCB1) by HM30181 increases oral bioavailability and therapeutic efficacy of paclitaxel. Eur. J. Pharmacol. 627, 92–98. https://doi.org/10.1016/j.ejphar.2009.11.008
- Kwon, J.S., Kim, E., Kang, D.-H., Choi, J.S., Yu, K.-S., Jang, I.-J., Shin, S.-G., APLUS study group, 2008. Taq1A polymorphism in the dopamine D2 receptor gene as a predictor of clinical response to aripiprazole. Eur. Neuropsychopharmacol. J. Eur. Coll. Neuropsychopharmacol. 18, 897–907. https://doi.org/10.1016/j.euroneuro.2008.07.010
- Larson, M.D., Kurz, A., Sessler, D.I., Dechert, M., Bjorksten, A.R., Tayefeh, F., 1997. Alfentanil blocks reflex pupillary dilation in response to noxious stimulation but does not diminish the light reflex. Anesthesiology 87, 849–855.
- Lawler, C.P., Prioleau, C., Lewis, M.M., Mak, C., Jiang, D., Schetz, J.A., Gonzalez, A.M., Sibley, D.R., Mailman, R.B., 1999. Interactions of the novel antipsychotic aripiprazole (OPC-14597) with dopamine and serotonin receptor subtypes. Neuropsychopharmacol. Off. Publ. Am. Coll. Neuropsychopharmacol. 20, 612–627. https://doi.org/10.1016/S0893-133X(98)00099-2
- Levy-Cooperman, N., McIntyre, G., Bonifacio, L., McDonnell, M., Davenport, J.M., Covington, P.S., Dove, L.S., Sellers, E.M., 2016. Abuse Potential and Pharmacodynamic Characteristics of Oral and Intranasal Eluxadoline, a Mixed μ- and κ-Opioid Receptor Agonist and δ-Opioid Receptor Antagonist. J. Pharmacol. Exp. Ther. 359, 471–481. https://doi.org/10.1124/jpet.116.236547
- Loch, A.A., van de Bilt, M.T., Bio, D.S., Prado, C.M. do, de Sousa, R.T., Valiengo, L.L., Moreno, R.A., Zanetti, M.V., Gattaz, W.F., 2015. Epistasis between COMT Val158Met and DRD3 Ser9Gly polymorphisms and cognitive function in schizophrenia: genetic influence on dopamine transmission. Rev. Bras. Psiquiatr. Sao Paulo Braz. 1999 37, 235–241. https://doi.org/10.1590/1516-4446-2014-1553
- Lowenstein, O., Loewenfeld, I.E., 1958. Electronic pupillography; a new instrument and some clinical applications. AMA Arch. Ophthalmol. 59, 352–363.
- Ma, X., Maimaitirexiati, T., Zhang, R., Gui, X., Zhang, W., Xu, G., Hu, G., 2014. HTR2C polymorphisms, olanzapine-induced weight gain and antipsychotic-induced metabolic syndrome in schizophrenia patients: a meta-analysis. Int. J. Psychiatry Clin. Pract. 18, 229–242. https://doi.org/10.3109/13651501.2014.957705
- Mangas-Sanjuan, V., Pastor, J.M., Rengelshausen, J., Bursi, R., Troconiz, I.F., 2016. Population pharmacokinetic/pharmacodynamic modelling of the effects of axomadol and its Odemethyl metabolite on pupil diameter and nociception in healthy subjects. Br. J. Clin. Pharmacol. 82, 92–107. https://doi.org/10.1111/bcp.12921
- Marazziti, D., Baroni, S., Picchetti, M., Piccinni, A., Carlini, M., Vatteroni, E., Falaschi, V., Lombardi, A., Dell'Osso, L., 2013. Pharmacokinetics and pharmacodynamics of

psychotropic drugs: effect of sex. CNS Spectr. 18, 118–127. https://doi.org/10.1017/S1092852912001010

- McDougal, D.H., Gamlin, P.D., 2014. Autonomic Control of the Eye, in: Terjung, R. (Ed.), Comprehensive Physiology. John Wiley & Sons, Inc., Hoboken, NJ, USA, pp. 439–473. https://doi.org/10.1002/cphy.c140014
- McGavin, J.K., Goa, K.L., 2002. Aripiprazole. CNS Drugs 16, 779–786; discussion 787-788.
- Moons, T., de Roo, M., Claes, S., Dom, G., 2011. Relationship between P-glycoprotein and second-generation antipsychotics. Pharmacogenomics 12, 1193–1211. https://doi.org/10.2217/pgs.11.55

NeurOptics, Inc., 2010. Neuroptics PLR-200[™] Pupillometer Instruction Manual.

- Ong, G.L., Bruning, H.A., 1981. Dilated fixed pupils due to administration of high doses of dopamine hydrochloride. Crit. Care Med. 9, 658–659.
- Pickworth, W.B., Lee, H., Fudala, P.J., 1990. Buprenorphine-induced pupillary effects in human volunteers. Life Sci. 47, 1269–1277.
- Piha, S.J., Halonen, J.P., 1994. Infrared pupillometry in the assessment of autonomic function. Diabetes Res. Clin. Pract. 26, 61–66.
- Rafaniello, C., Sessa, M., Bernardi, F.F., Pozzi, M., Cheli, S., Cattaneo, D., Baldelli, S., Molteni, M., Bernardini, R., Rossi, F., Clementi, E., Bravaccio, C., Radice, S., Capuano, A., 2017.
 The predictive value of ABCB1, ABCG2, CYP3A4/5 and CYP2D6 polymorphisms for risperidone and aripiprazole plasma concentrations and the occurrence of adverse drug reactions. Pharmacogenomics J. https://doi.org/10.1038/tpj.2017.38
- Raffa, R.B., Friderichs, E., Reimann, W., Shank, R.P., Codd, E.E., Vaught, J.L., 1992. Opioid and nonopioid components independently contribute to the mechanism of action of tramadol, an "atypical" opioid analgesic. J. Pharmacol. Exp. Ther. 260, 275–285.
- Schmittgen, T.D., Livak, K.J., 2008. Analyzing real-time PCR data by the comparative C(T) method. Nat. Protoc. 3, 1101–1108.
- Schwartz, J.B., 2003. The influence of sex on pharmacokinetics. Clin. Pharmacokinet. 42, 107– 121. https://doi.org/10.2165/00003088-200342020-00001
- Siepmann, T., Ziemssen, T., Mueck-Weymann, M., Kirch, W., Siepmann, M., 2007. The effects of venlafaxine on autonomic functions in healthy volunteers. J. Clin. Psychopharmacol. 27, 687–691. https://doi.org/10.1097/jcp.0b013e31815a255b
- Skarke, C., Jarrar, M., Schmidt, H., Kauert, G., Langer, M., Geisslinger, G., Lötsch, J., 2003. Effects of ABCB1 (multidrug resistance transporter) gene mutations on disposition and central nervous effects of loperamide in healthy volunteers. Pharmacogenetics 13, 651–660. https://doi.org/10.1097/01.fpc.0000054133.14659.e2
- Spiers, A.S., Calne, D.B., 1969. Action of dopamine on the human iris. Br. Med. J. 4, 333–335.
- Stip, E., Tourjman, V., 2010. Aripiprazole in schizophrenia and schizoaffective disorder: A review. Clin. Ther. 32 Suppl 1, S3-20. https://doi.org/10.1016/j.clinthera.2010.01.021
- Suzuki, T., Mihara, K., Nakamura, A., Kagawa, S., Nagai, G., Nemoto, K., Kondo, T., 2014. Effects of genetic polymorphisms of CYP2D6, CYP3A5, and ABCB1 on the steady-state plasma concentrations of aripiprazole and its active metabolite, dehydroaripiprazole, in

Japanese patients with schizophrenia. Ther. Drug Monit. 36, 651–655. https://doi.org/10.1097/FTD.000000000000070

- Taylor, D.M., 2003. Aripiprazole: a review of its pharmacology and clinical use. Int. J. Clin. Pract. 57, 49–54.
- Tobin, A.B., Osborne, N.N., 1989. Evidence for the presence of serotonin receptors negatively coupled to adenylate cyclase in the rabbit iris-ciliary body. J. Neurochem. 53, 686–691.
- Tobin, A.B., Unger, W., Osborne, N.N., 1988. Evidence for the presence of serotonergic nerves and receptors in the iris-ciliary body complex of the rabbit. J. Neurosci. Off. J. Soc. Neurosci. 8, 3713–3721.
- Urichuk, L., Prior, T.I., Dursun, S., Baker, G., 2008. Metabolism of atypical antipsychotics: involvement of cytochrome p450 enzymes and relevance for drug-drug interactions. Curr. Drug Metab. 9, 410–418.
- Wojnicz, A., Belmonte, C., Koller, D., Ruiz-Nuño, A., Román, M., Ochoa, D., Abad Santos, F., 2017. Effective phospholipids removing microelution-solid phase extraction LC-MS/MS method for simultaneous plasma quantification of aripiprazole and dehydro-aripiprazole: Application to human pharmacokinetic studies. J. Pharm. Biomed. Anal. 151, 116–125. https://doi.org/10.1016/j.jpba.2017.12.049
- Wood, M.D., Scott, C., Clarke, K., Cato, K.J., Patel, N., Heath, J., Worby, A., Gordon, L., Campbell, L., Riley, G., Davies, C.H., Gribble, A., Jones, D.N.C., 2006. Pharmacological profile of antipsychotics at monoamine receptors: atypicality beyond 5-HT2A receptor blockade. CNS Neurol. Disord. Drug Targets 5, 445–452.
- Yu, Y., Ramage, A.G., Koss, M.C., 2004. Pharmacological studies of 8-OH-DPAT-induced pupillary dilation in anesthetized rats. Eur. J. Pharmacol. 489, 207–213. https://doi.org/10.1016/j.ejphar.2004.03.007
- Zhang, J.-P., Lencz, T., Malhotra, A.K., 2010. D2 receptor genetic variation and clinical response to antipsychotic drug treatment: a meta-analysis. Am. J. Psychiatry 167, 763–772. https://doi.org/10.1176/appi.ajp.2009.09040598



Figure 1. Changes in the pupillometric parameters after administration of aripiprazole 10

mg. Abbreviations: MAX: maximum pupil diameter; MIN: minimum pupil diameter; MCV: maximum constriction velocity; T75: total time taken by the pupil to recover 75% of the initial resting pupil size. $*p \le 0.05$ related to the baseline value. $**p \le 0.001$ related to the baseline value.



Figure 2. Association of *HTR2A* rs6313 genotypes with MCV and MAX pupillometric parameters, *HTR2C* rs3813929 genotypes with MCV and MAX, DRD2 rs6277 genotypes with T75 and *DRD3* rs6280 genotypes with MCV. Abbreviations: MCV: maximum constriction velocity; MAX: maximum pupil diameter; T75: total time taken by the pupil to recover 75% of the initial resting pupil size after it reached the peak of constriction; HTR2A: 5-hydroxytryptamine receptor 2A; HTR2C: 5-hydroxytryptamine receptor 2A; DRD2: dopamine D2 receptor; DRD3: dopamine D3 receptor

* $p \le 0.05$; ** $p \le 0.001$, \cdot shows the outliers.



Figure 3. Relationship of *CYP2D6* phenotypes (EM, IM) and *ABCB1* genotypes (G2677T/A, C3435T, C1236T) with aripiprazole pharmacokinetic parameters AUC₀₋₇₂, C_{max} and T_{1/2}. Abbreviations: *ABCB1*: ATP binding cassette subfamily B member 1; CYP: cytochrome p450 oxidase; IM: intermediate metabolizer; EM: extensive metabolizer; C_{max}: maximum plasma concentration; AUC: the total area under the curve; T_{1/2}: half-life. * $p \le 0.05$, \cdot shows the outliers.

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