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1 **Value-added of heat moisture treated mixed flours in wheat-based matrices: a functional and**  
2 **nutritional approach.**

3

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**Abstract** The significance of heat moisture treatment (HMT) of non-wheat –teff (T), chestnut (CN) and chickpea (CP) flours on dough viscoelastic and thermal parameters and on the structural and nutritional pattern of breads was investigated in untreated (-) and HMT (+) associated wheat-based (WT) matrices (WT:T:CN:CP, 66.20:7:7, wt. basis). Suitable trends for the enhancement of the physical characteristics of breads in terms of larger specific volume, higher viscoelastic and textural profiles, with lower and slower staling kinetics on ageing were achieved by the pairs T-CN+, T-CP+, CN-CP+ and CN+CP+. In addition, a fine and uniformly- sized cell structure with similar cell walls thickness were achieved in crumb samples. The pair T-CN+ enhanced extracted bioaccessible polyphenols, and the pair CN+CP+ synergistically promoted the anti-radical activity in breads. Blended breads can be labelled as high-fibre breads ( $\geq 6$  g DF/100 g food), and a recommended daily consumption of 250 g of bread fulfilled from 44% (men) to 67% (women) of dietary fibre requirements.

**Keywords** Heat Moisture Treatment, flour, dough, bread, functionality, nutritional value

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4 49 **1. Introduction**

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6 50 The increasing demand for novel, tasty and healthy foods has boosted a new market in which  
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9 51 products traditionally made from wheat are partially replaced or supplemented with alternative nutrient-  
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11 52 dense and health promoting non-wheat ingredients. In this context, ancient crops (Angioloni & Collar,  
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13 53 2011), minor cereals (Collar and Angioloni, 2013), pseudocereals (Collar and Angioloni, 2014a),  
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15 54 legumes (Angioloni & Collar, 2012), and non-traditional fruit and seed flours (Paciulli et al., 2016) have  
16  
17 55 received much attention over the last years as functional ingredients for potential breadmaking  
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19 56 applications.  
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22  
23 57 Chestnut flour contains high quality proteins with essential amino acids, dietary fibre, low amount  
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25 58 of fat and also vitamin E, vitamin B group, potassium, phosphorous, and magnesium of nutritional  
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27 59 interest on both wheat (Dall'Asta et al., 2013) and gluten-free (Paciulli et al., 2016) breadmaking. It has  
28  
29 60 been reported that wheat breads enriched with the addition of chestnut flour presented an increased  
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31 61 quality from both organoleptic and nutritional points of view (Dall'Asta et al., 2013).  
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35 62 Legumes constitute traditional, ubiquitous and wholesome imaged foods providing nutritional (high  
36  
37 63 protein, mineral and fibre contents, low digestible starch), health (protective and therapeutic effects to  
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39 64 chronic health conditions) and functional promoting effects (body, texture and taste enhancement) to  
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41 65 foods (Angioloni & Collar, 2012). Associated mixtures of grain (chickpea, greenpea) and oilseed  
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43 66 (soybean) legumes replacing wheat flour at 42 % provided, highly nutritious breads—meeting  
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45 67 viscoelastic restrictions and sensory standards (Collar and Angioloni, 2017).  
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49 68 Teff (*Eragrostis tef*) is a nutritious cereal wheat-type gluten-free grain indigenous to Ethiopia, rich  
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51 69 in carbohydrates and fiber, microelements and phytochemicals with superior amounts of iron, calcium  
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53 70 and zinc than wheat, barley and sorghum (Abebe et al., 2007). Teff was successfully applied in  
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55 71 breadmaking matrices up to 40% of wheat replacement (Ronda, Abebe, Pérez-Quirce, & Collar, 2015).  
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4 72 Despite partial wheat flour replacement by nutrient-dense flours constitutes a plausible simple  
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6 73 strategy to create value-added baked goods, making highly replaced wheat flour breads to assure  
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9 74 nutritional and health-related benefits, often encompasses a fall in techno-functional bread quality,  
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11 75 particularly loaf volume and texture. Technological treatments have been applied with variable success  
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14 76 to palliate the adverse effects associated to gluten and starch dilution in wheat matrices. High  
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16 77 Hydrostatic Pressure treatment has been recently used as an alternative to hydrocolloids/gluten  
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19 78 addition for the structure rearrangement of legume batters and consequently for their incorporation, in  
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21 79 high amount, in breadmaking systems (Collar & Angioloni, 2017). Heat moisture treatment (HMT)  
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23 80 constitutes a clean label alternative to chemical modification for altering the gelatinization and  
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25 81 retrogradation properties of starches (Gunaratne and Hoover, 2002), flours and doughs (Collar &  
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27 82 Armero, 2018), and the aggregation/disaggregation equilibrium of proteins (Mann et al., 2013).  
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30 83 Microscopic observations by confocal laser scanning microscopy and light microscopy revealed that  
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33 84 HMT caused the clumping of starch granules and the aggregation of denatured protein (Chen et al.,  
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35 85 2015). HMT of flours has successfully been applied to wheat (Cetiner et al., 2017), sorghum (Marston,  
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38 86 Khouryieh, & Aramouni, 2016), composite oat-wheat (Verdú, Vásquez, Ivorra, Sánchez et al., 2017)  
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40 87 and barley-wheat (Collar & Armero, 2018) flours to improve dough functionality, and both volume and  
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42 88 textural profile of the resulting breads. Improvement was ascribed mainly to the increase of disulphide  
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44 89 cross-linkages of amino acids, and to changes on starch granules conformation, principally in physical  
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46 90 reorganization in their structure resulting in higher dough viscosities (Ovando-Martínez et al., 2013).

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48 91 Significance of HMT on viscoelasticity and functional performance of blended doughs –hydrated  
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51 92 wheat/non-wheat flours- has been recently addressed in diluted wheat: barley binary systems (Collar  
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53 93 & Armero, 2018). Results pointed out the importance of both the water availability and the heat  
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55 94 treatment of compositional flours to obtain a reinforced dough structure with partially restored  
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58 95 viscoelasticity.  
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96           Although the use of clean label treatments is regaining interest among consumers due to an  
97 increasing commitment with the environment, the evaluation of the application of HMT in breadmaking  
98 in associated wheat/non-wheat flours is little explored in the literature. This paper is aimed at  
99 investigating the potencial of HMT to restore and/or improve bread viscoelasticity and functional  
100 performance keeping nutritional value of diluted wheat matrices with incorporation of nutrient-dense  
101 value-added non-gluten forming flours (teff, chestnut, chickpea), by studying the changes induced in  
102 treated blended bread matrices at both functional and nutritional levels.

103

104   **2. Materials and methods**

105

106   **2.1. Materials**

107           Commercial flours from refined common wheat *Triticum aestivum* (WT), teff *Eragrostis tef* (T),  
108 chestnut *Castanea sativa* (CN), and whole decorticated chickpea *Cicer arietinum* (CP) were obtained  
109 from the Spanish market (Navarro, Valencia, Spain). Refined WT (70% extraction rate) of 195 x 10<sup>-4</sup> J  
110 energy of deformation W, 0.57 curve configuration ratio P/L, and 58.8% water absorption in Brabender  
111 Farinograph, was used. Carboxymethylcellulose Aquasorb® A-500 (CMC) was acquired from  
112 Copenhagen Pectin (Denmark), and commercial wheat sour dough *Pie* was kindly supplied by Ireks  
113 (Spain).

114

115   **2.2. Methods**

116    2.2.1. *Chemical and nutritional composition of flours*

117           Chemical and nutritional composition of native WT, T, CN and CP flours (Table 1) was  
118 determined following the ICC methods (ICC, 2014) regarding moisture, protein, and fat contents. Total,  
119 soluble and insoluble dietary fibre contents were determined according to the AOAC method 991.43

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120 (AOAC, 1991). Three replicates were made for each analysis. Digestible carbohydrates were  
121 calculated by indirect determination as  $100 - [\text{Moisture} + \text{Protein} + \text{Fat} + \text{Dietary Fibre}]$  (FAO, 2003).

122

### 123 2.2.2. Heat-moisture treatment (HMT)

124 HMT conditions (15% moisture content, 1 h and 120°C) were selected based on previous  
125 experiments (Collar and Armero, 2018), in which maximization of viscometric profile and minimization  
126 of loss of hydration properties of flour samples were applied as criteria. Single T, CN and CP flour  
127 samples were placed into screw-capped cylindrical glass containers (150 mm  $\square$ , 250 mm height).  
128 Small amount of distilled water was added slowly with frequent stirring until moisture levels (w/w) of the  
129 total mixture reached 15%, and equilibrated for 24 h at room temperature. Hydrated samples that  
130 occupied 13 mm height in containers were kept for 1h at 120 °C in a convection oven (P-Selecta,  
131 Barcelona, Spain). Samples took 20 min to reach the pre-set temperature, and 30 min to reach the  
132 room temperature after heating. Untreated native flours were used as controls. Untreated (-) and HMT  
133 (+) single flours were used in quaternary blends (T:CN:CP:WT) in presence of WT- for doughmaking.

134

### 135 2.2.3. Bread making of wheat -based blended flours

136 Specific flour composition was set after a prospective study on the compositional and functional  
137 characteristics of non-wheat flours (native and HMT) was performed. Results pointed out that besides  
138 the superior nutritional value as compared to wheat, teff, chestnut and chickpea individual flours were  
139 sensitive to HMT in terms of increased water absorption, viscosity after heating-cooling cycles,  
140 increased consistency (forward-extrusion test), and acceptable dough handling ability during  
141 processing. This behaviour made flours interesting candidates to be integrated in wheat diluted  
142 systems with good prediction as dough strengtheners. Percentages of replacement resulted from

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143 experimental studies aimed at knowing maximum amount of each flour without significant deleterious  
144 effect on dough machinability.

145 Doughs and breads were prepared from wheat-based blended flours (T, CP, CN) by WT replacement  
146 at 34%, and incorporation of ternary blends of T (20%, flour basis), CN (7%, flour basis), and CP (7%,  
147 flour basis) flours according to a Multilevel Factorial Design with the following attributes: 3 experimental  
148 factors (T, CP and CN flours) at 2 levels, coded 0 (untreated) and 1 (HMT), and 5 error degrees of  
149 freedom. The model resulted in 8 randomized runs in 1 block. A 3 digit bread sample code was set  
150 referring to no HMT (0) and HMT (1) T (1st digit), CN (2nd digit), and CP (3rd digit) flours in sample  
151 formulation, as it follows: 110, 101, 100, 000, 001, 111, 010, 011. Blended flours (100 g), water (100%,  
152 flour basis), commercial compressed yeast (3%, flour basis), salt (2%, flour basis), commercial sour  
153 dough *Pie* (5%, flour basis), CMC (3%, flour basis), and calcium propionate (0.5%, flour basis) were  
154 mixed in a 10 kg mixer at 60 revolutions min<sup>-1</sup> for 10-13 min up to optimum dough development.  
155 Preliminary tests were performed to know the amount of water necessary to avoid stickiness and  
156 deleterious effects on dough machinability, and 100% of water absorption was enough for all the  
157 formulations to assure dough handling ability during processing. Fermented doughs were obtained  
158 after bulk fermentation (10 min at 28°C), dividing (300 g), rounding, molding, panning and proofing up  
159 to maximum volume increment (50 min at 28°C), and were baked at 225 °C for 25 min to make blended  
160 breads. Two baking trials were conducted per formulation. Bread samples were packaged in co-  
161 extruded polypropylene bags, and stored for 1, 3, 6, and 8 days at room temperature and 56% relative  
162 humidity to describe firming kinetics.

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164 2.2.4. Dough measurements

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166 2.2.4.1. Texture Profile Analysis (TPA)

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167 TPA of quaternary doughs were performed in absence of yeast by applying a double  
168 compression cycle in a TA-XT2 Texture Analyser (Stable Micro Systems, Surrey, UK) equipped with a  
169 30 kg-load cell and operating at 10 mm/s head speed, by using a 1 cm diameter probe, 75 s waiting  
170 period, and 60% compression. The primary textural properties (hardness, cohesiveness, springiness  
171 and resilience) were measured in absence of dough adhesiveness by using a plastic film on the dough  
172 surface to avoid the distortion induced by the negative peak of adhesiveness, while dough  
173 adhesiveness was measured separately by running a second TPA without the plastic film and  
174 disregarding the other parameters. Runs were performed in triplicate per sample.

175

#### 176 2.2.4.2. Stress relaxation test

177 Doughs without yeast were submitted to uniaxial compression in the Texture Analyser using  
178 an acrylic probe (37-mm diameter) to a 10% strain and the change in force with time was measured  
179 for 300 s. A pretest speed of 5 mm/s and test speed of 0.5 mm/s were used. The obtained stress  
180 relaxation curves were normalized and linearized according to the Peleg model:  $F_0t/(F_0-F(t))=k_1 + k_2t$ ,  
181 where  $F_0$  is the initial force,  $F(t)$  is the momentary force at time (t) and  $k_1(s)$ ,  $k_2$  are constants related to  
182 stress decay rate and to residual stress at the end of the experiment, respectively. In addition, percent  
183 stress relaxation (% SR=  $(F_0-F_{300}).100/F_0$ ),  $1/k_1$  (initial rate of relaxation),  $1/k_2$  (extent of relaxation) and  
184 relaxation time (RT) as the time required for  $F_0$  to drop to 36.8% of its values, respectively, were  
185 compared for the different samples. Tests were performed in duplicate per sample.

186

#### 187 2.2.4.3. Thermal analysis

188 The thermal properties of native and HMT flours were determined by using a differential  
189 scanning calorimeter (DSC-7, Perkin-Elmer, Norwalk, CT). Dough samples were prepared by mixing  
190 flour blends with excess water (1:3) to avoid partial gelatinization of samples due to water restrictions.



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191 For DSC analysis, 50–70 mg of dough samples were weighed in large volume pre-weighed, sealed  
192 stainless-steel pans. An empty pan was used as a reference. Simulation of the temperature profile in  
193 the center of the bread crumb during baking was performed in the calorimeter under the following  
194 scanning conditions: samples were kept at 30°C for 2 min, then heated from 30 to 110°C at a rate of  
195 11.7°C/min, kept at 110°C for 5 min, and finally cooled from 110 to 30°C at a rate of 50°C/min. Thermal  
196 transitions of samples for gelatinization were characterised by  $T_o$  (onset temperature),  $T_p$  (peak  
197 temperature),  $T_e$  (end temperature), and  $\Delta H_g$  (enthalpy of gelatinization). The enthalpy calculations  
198 were based on a dry-flour weight. The samples were analyzed three times, and the data were  
199 calculated with a Pyris software (Perkin-Elmer, Norwalk, CT).

201 *2.2.5. Bread measurements*

202 *2.2.5.1. Physico-chemical determinations*

203 Loaf volume was determined using the rapeseed displacement method as in AACC (2005).  
204 Specific loaf volume was calculated dividing the loaf volume by the corresponding loaf weight.

205 Colour determinations were carried out on bread crumbs using a Photoshop system according  
206 to the method previously described by Angioloni & Collar (2009), and results were expressed in  
207 accordance with the Hunter Lab colour space. Parameters determined were L (L = 0 [black] and L =  
208 100 [white]), a (-a = greenness and +a = redness), b (-b = blueness and +b = yellowness),  $\Delta E$  -total  
209 colour difference-, and WI -whiteness index-. All measurements were made in triplicate.

210 Crumb grain characteristics were assessed in bread slices using a digital image analysis  
211 system. Images were previously acquired with a ScanJet II cx flatbed scanner (Hewlett-Packard, Palo  
212 Alto, CA, USA) supported by a Deskscan II software. The analysis was performed on 40 mm × 40 mm  
213 squares taken from the centre of the images. Data were processed using SigmaScan Pro 5 (Jandel  
214 Corporation, San Rafael, CA, USA). The crumb grain features evaluated were mean cell area,

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215 cells/cm<sup>2</sup>, cell/total area ratio, wall/total area ratio and crumb area/total cell ratio. In addition, area  
216 distribution and cell number distribution were counted, and percentage of cell area and cell number  
217 were calculated according to pre-set cell size ranges: <1mm<sup>2</sup>, 1-5mm<sup>2</sup>, 5-50mm<sup>2</sup>, >50mm<sup>2</sup>.

218 Bread mechanical characteristics (TPA in a double compression cycle) of fresh and stored  
219 breads were recorded in a TA-XT2 Texture Analyser using a 10 mm diameter probe, a 30 kg load cell,  
220 50% penetration depth and a 30 s gap between compressions on slices of 15 mm width (Armero &  
221 Collar, 1998). For textural measurements, three slices of two breads were used for each sample. The  
222 obtained firming curves during bread storage were modelled using the Avrami equation, and model  
223 factors were estimated by fitting experimental data of hardness to the nonlinear regression equation  
224  $\theta = \frac{T_{\infty} - T_t}{T_{\infty} - T_0} = e^{-kt^n}$  where  $\theta$  is the fraction of the recrystallisation still to occur;  $T_0$ ,  $T_{\infty}$  and  $T_t$  are  
225 crumb firmness at time zero,  $\infty$  and time  $t$ , respectively,  $k$  is a rate constant, and  $n$  is the Avrami  
226 exponent.

227 The stress relaxation data were collected by applying an instantaneous strain to the sample  
228 and the force required to maintain the formed deformation was observed as a function of time. Samples  
229 from the centre of the crumb slices were cut into cylinders (27 mm diameter x 15 mm thick) and were  
230 compressed using a TA-XT2 Texture Analyser with a load cell of 30 kg. For compression, a cylindrical  
231 upper die of 25 mm diameter was used at a cross speed of 0.5 mm/s and a pretest speed of 5 mm/s.  
232 **Samples were placed in a semi-close cabinet to prevent moisture loss.** The strain used was 20% and  
233 the whole relaxation experiment lasted for 10 min. The obtained stress relaxation curves were  
234 normalized and linearized according to Peleg model as described for doughs, and previously applied  
235 by Angioloni and Collar (2009) for breads. In addition, percent of stress relaxation (% SR=  $(F_0 -$   
236  $F_{600}) \cdot 100 / F_0$ ),  $1/k_1$  (initial rate of relaxation),  $1/k_2$  (extent of relaxation) and relaxation time ( $RT$  as the

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237 time required for  $F_0$  to drop to 60% of its values), were compared for the different bread samples. Stress  
238 relaxation tests were replicated two times.

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#### 240 2.2.5.2. *Nutritional parameters*

241 Bioaccessible phenol determinations were carried out by conducting an “in vitro” digestive  
242 enzymatic mild extraction that mimics the conditions in the gastrointestinal tract according to the  
243 procedure of Glahn et al. (1998) and adapted by Angioloni and Collar (2011) for breads. Calibration  
244 curve was performed using gallic acid and therefore obtained amounts of phenolics were expressed  
245 as gallic acid equivalents.

246 The stable 2,2-diphenyl-1-picrylhydrazyl (DPPH•) radical was used to measure the radical  
247 scavenging capacity of the bioaccessible polyphenol enzymatic extracts of bread samples according  
248 to the DPPH• method modified by Sánchez-Moreno et al. (1998) and adapted by Collar et al. (2014).  
249 **Readings** were taken from two replicates per sample. Plots of  $\mu\text{mol DPPH}$  vs. time (min) were drawn,  
250 and calculations were made to know the antiradical activity (AR).  $AR = \frac{([DPPH]_{INITIAL} - [DPPH]_{PLATEAU})}{[DPPH]_{INITIAL}} \times 100$ .

252

#### 253 2.2.6. *Statistical analysis*

254 Multivariate analysis of variance of data and non-linear regression analysis were performed by  
255 using Statgraphics V.7.1 program (Bitstream, Cambridge, MN). Multiple range test (Fisher’s least  
256 significant differences, LSD) for analytical variables was applied to know the difference between each  
257 pair of means.

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### 259 **3. Results and discussion**

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261           **3.1. Significance of HMT of non-wheat flours on the functional performance of blended**  
262           **doughs**

263           Functional performance of untreated and HMT blended doughs were characterized at macroscopic  
264           and structural levels in terms of their viscoelastic, mechanical and thermal profiles (Table 2), and the  
265           significance of HMT of non-wheat flours on the functional parameters of blended matrices was  
266           determined (Table 3).

267           Significant differences ( $p < 0.05$ ) were found in the physico-chemical patterns of doughs from  
268           untreated and HMT blended flours (Table 2). Dough viscoelasticity parameters ranged from 0.89 N  
269           (000) to 1.73N (001, 100) for the initial force  $F_0$ , from 11.39s (110) to 19.50s (000) for  $k_1$ , and from 1.36  
270           (110) to 1.52 (000, 111.011) for  $k_2$ , providing relaxation times  $RT$  that varied from 53 (110) to 367s  
271           (000), and extent of relaxation  $SR$  ranging from 63% (000) to 71% (110). HMT of single flours in  
272           blended doughs, particularly significant for T and CN, provided a fall in the elastic-like nature of doughs.  
273           This led to a concomitant promotion of the viscous nature of doughs evidenced by the decreased values  
274           of both  $k_1$  and  $k_2$ . As a consequence, increased values for the reciprocal  $1/k_1$  and  $1/k_2$  related to the  
275           initial stress decay rate and the asymptotic level of stress not relaxed at long times, respectively were  
276           obtained. In accordance, effects of HMT led to shorter  $RT$  and higher extent of  $SR$ , particularly for T,  
277           which values changed from 181s to 101s ( $RT$ ) and from 66% to 68% ( $SR$ ), respectively (Table 3). In  
278           the current work, simultaneous presence of thermally treated T+ and CP+ magnified the above  
279           mentioned changes, while treating the pair CN+ and CP+ flours reduced the extent of dough weakening  
280           leading to values for dough viscoelasticity near those of untreated flours (Table 3). Observations are in  
281           agreement with previous changes of the rheological properties at small and large deformations  
282           described for doughs made of heat-treated wheat flour (Mann et al., 2013). Changes comprise dough  
283           reinforcement by increase of the dynamic moduli, an easier destruction of the dough network by  
284           increase of protein solubility, and an unequal change in loss and storage moduli, leading to irreversible

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285 changes in flour. These changes can be ascribed assuming protein aggregation (leading to a weakened  
286 protein network and possibly acting as additional filler particles) and starch surface modifications  
287 (leading to changed starch-protein and starch-starch) interactions (Mann et al., 2013). Later, it was  
288 observed that HMT favoured the strengthening of arrowroot starch gels determined by oscillatory  
289 rheological tests, and gave them greater resistance, particularly to acidification stress (Pepe, Moraes,  
290 Albano, Telis & Franco, 2015).

291 Dough mechanical profile of blended flours underwent little changes with HMT in terms of  
292 cohesiveness (0.788-0.858), springiness (0.822-0.884) and resilience (0.095-0.108) values (Table 2).  
293 On the contrary, hardness and adhesiveness depended on the thermal treatment of the pairs T/CN  
294 and T/CP (Table 3). Increase of dough hardness and adhesiveness by thermal treatment of individual  
295 flours was observed earlier for treated sweet potato starch with HMT (Collado & Corke, 1999). The  
296 observed increase was counteracted after HMT of both T+ and CP+ (4.75N vs 6.22N) and after HMT  
297 of either T/CN (35.75N.s vs 42.37N.s) or T/CP (31.06N.s vs 43.27N.s), respectively (Table 3). It was  
298 found that modification by HMT resulted in a reduction of gel hardness of both starch and flour rice  
299 samples (Puncha-arnon & Uttapap, 2013). In the case of HMT flours, a sharp decrease in gel hardness  
300 was affected by components other than starch. Protein layers formed on the starch surface, as well as  
301 lipid complexes formed during HMT, could both inhibit the swelling of starch granules. Hamaker &  
302 Griffin (1993) earlier explained that proteins with intact disulfide bonds made the swollen granules less  
303 susceptible to breakdown, either by imparting strength to the swollen granules or by reducing the  
304 degree of swelling. The flours used in this study are rich in both protein and lipids, particularly CP (Table  
305 1), and these components may favour the formation of protein layers and starch-lipid complexes on  
306 HMT, boosting the inhibition of starch granules swelling and the dough softening .

307 During gelatinization, at high water concentration (>66 wt% or water/starch ratio >1.5), a single  
308 symmetrical endothermic transition appears in a temperature range of 60–80°C in the DSC profiles

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309 (called endotherm G), as reported by Donovan (1979). In the present research, blended dough samples  
310 contain high water concentration (75 wt%), and only one peak was defined during the DSC scan of all  
311 samples. The corresponding thermal transitions for the peak occurred at close temperature intervals  
312 (°C) of 61.23-62.24 ( $T_0$ ), 69.30-70.50 ( $T_p$ ), and 78.47-80.39 ( $T_e$ ), and similar temperature and enthalpy  
313 (J/g, dry flour) ranges of 17.18-18.09 and 5.548-6.170, respectively (Table 2). According to the different  
314 models, the endotherm G (peak 1) was suggested to result from a) plasticization in amorphous regions  
315 b) swelling-driven crystalline disruption and/or c) melting of the less stable crystallites in sufficient water,  
316 and d) associated with the smectic–nematic/isotropic transition (Wang and Copeland, 2013). The  
317 differences in gelatinization temperatures among doughs can be attributed mainly to differences in size,  
318 form and distribution of starch granules in the blended flours, and to the internal arrangement of starch  
319 fractions within the granule, as stated earlier for legume flours (Kaur & Singh, 2005). In fact, starches  
320 in the flour blends are composed of granules differing in size, from small to large: pea (wrinkled) 5–34  
321  $\mu\text{m}$ , small wheat granules 2–3  $\mu\text{m}$  (Zhou et al. 2004), large wheat granules 22–36  $\mu\text{m}$ , teff 2-6  $\mu\text{m}$   
322 (Bultosa and Taylor, 2004), and chestnut 2.9-21.4  $\mu\text{m}$  (Demiate, Oetterer, & Wosiacki, 2001) that  
323 exhibit irregular oval shaped granules of broad size range. In addition, despite HMT did not alter the  
324 size or shape of the starch granules of some cereals, roots and legumes (Hoover, 2010), the thermal  
325 treatment may affect the aggregation of starch granules as observed for oat starch. Starch granules  
326 that were aggregated in the native state were less compactly packed after heat treatment (Hoover &  
327 Vasanthan, 1994). Significant single effect of HMT of T flour revealed a discreet broadening of the  
328 gelatinization-temperature range (R) and a shifting of the endothermal transition towards higher  
329 temperatures (Table 3), being values 17.34°C vs 17.87°C (R), 61.54 vs 62.28°C ( $T_0$ ), 69.60 vs 70.50°C  
330 ( $T_p$ ), 78.87 vs 80.15°C ( $T_e$ ). Previous studies on tuber, legume, and cereal (normal, waxy, high  
331 amylose) starches (Hoover, 2010) and flours (Satmalee & Matsuki, 2011) ascribed the observations to  
332 amylose-amylose, amylose-amylopectin and amylose-lipid interactions, and as well as other chemical

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333 bonding/interactions that occur during HMT (Watcharatewinkul et al., 2009). Authors stated that  
334 interactions suppress the mobility of starch chains in the amorphous regions. Consequently, the  
335 amorphous regions would require a higher temperature to incur swelling that could contribute to the  
336 disruption of the crystalline regions. Lin et al. (2001) proposed that the increase in gelatinization  
337 temperature was caused by the transformation of the inter-crystalline amorphous regions to amorphous  
338 phases, which may provide the short chains in the crystalline structure more freedom. Thus, the  
339 crystalline micelles undergo a structural transformation towards an increased thermodynamic stability.  
340 Besides, it was probably due to the restriction of water penetration into the granules by the new  
341 superficial protein layer formed by HMT, thus retarding the granule swelling (Chen, He, Fu, & Huang,  
342 2015).

343 In addition, T significantly interacted with CP, modifying in small extent the values of the  
344 gelatinization enthalpies. HMT of either T or CP resulted in a small decrease of  $\Delta H_g$  compared to values  
345 observed for the untreated pair of flours (5.64 vs 6.12 J/g), while HMT of both flours led to intermediate  
346 enthalpy values (5.88 J/g). Enthalpy change reflects the melting of imperfect amylopectin-based  
347 crystals, with potential contributions from crystal-packing and helix melting enthalpies (Lopez-Rubio et  
348 al., 2008). The decrease in  $\Delta H_g$  on HMT reflects disruption of double helices present in crystalline and  
349 non-crystalline regions of the granule (Gunaratne & Hoover, 2002). In addition, most of the semi-  
350 crystalline structure and a considerable extent of the concentrated crystalline region were destroyed  
351 during HMT.

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353 **3.2. Significance of HMT of non-wheat flours on the physical and nutritional profiles of**  
354 **blended breads**

355 In general, mixed HMT breads were visibly similar in colour (Figure 1), and exhibited low-  
356 medium lightness  $L$  (54.8-60.4),  $>0$   $a$  (2.3-3.4) and  $b$  (10.3-12.3) values, resulting in low WI crumbs

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4 357 (53.6-58.6) (Table 4). Colour coordinates of breads were not dependent on the thermal treatment of  
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6 358 any compositional flour, so that all sample crumbs were characterized by a dark orange-brown colour  
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9 359 (Figure 1).

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11 360 Common crumb grain features of breads (Table 4, Figure 1) evidenced main small cells <1  
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13 361 mm<sup>2</sup> accounting for 81% - 92% of total cells, intermediate medium size (1-50 mm<sup>2</sup>) cells (7-19%), and  
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15 362 marginal big cell (>50 mm<sup>2</sup>) proportion (0-0.2%). Mean cell area (mm<sup>2</sup>) ranged from 0.43 to 0.82, and  
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17 363 cell area distribution covered by cells sized 1-50 mm<sup>2</sup> varied from 63% (001) to 87% (100). Cell density  
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19 364 (cells/cm<sup>2</sup>) ranged from 48 (111) to 107 (100), being cell/wall ratio from 38/62 (000, 111, 011) to 50/50  
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21 365 (110). Except for the samples 110, 101, and 100 that exhibited heterogeneous and/or highly packed  
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23 366 crumb structure, a fine and uniformly-sized cell structure with similar cell walls thickness were achieved  
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26 367 in crumb samples (Figure 1). Improvement of the internal grain structure with heat treatment was  
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29 368 reported previously for substandard wheat flour breads (Gélinas, McKinnon, Rodriguez, & Montpetit,  
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31 369 2001) and for sorghum breads (Marston et al., 2016). The high fibre content of chestnut flour was  
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33 370 reported to enhance the viscoelastic properties and to reduce dough expansibility due to the  
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35 371 entanglement of fibres, entrap more air bubbles, and produce pores with a small cell area when added  
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37 372 with a relatively high amount of this flour (Demirkesen, Mert, Sumnu, & Sahin, 2010). Positive effects  
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39 373 of HMT of CN and/or CP on bread crumb grain have not been described so far.

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41 374 Macrostructural properties of HMT breads were characterized in terms of specific volume,  
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43 375 mechanical/ textural behaviour, stress relaxation pattern and firming kinetics on ageing (Table 4), and  
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45 376 the significant (p<0.05) single and interactive effects of thermal treatment of compositional flours were  
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47 377 determined (Table 5). Compared to untreated blended breads (000), HMT breads exhibited in general  
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49 378 from smaller to larger specific volumes (2.59-3.28 mL/g vs 3.07 mL/g), similar percent of stress  
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51 379 relaxation (43-51%), slightly variable stress decay rate  $k_1$  (45.67-66.58s vs 57.24s) and residual stress  
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53 380  $k_2$  (1.86-2.19 vs 1.98), similar or higher cohesiveness (0.460-0.568 vs 0.485) and resilience (0.186-



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381 0.245 vs 0.200), variable fresh crumb firmness (7.89-10.58N vs 9.44N) and rate of staling kinetics  $n_f$   
382 (0.390-1.817 vs 1.097) (Table 4). HMT of single flours T, CN or CP significantly ( $p < 0.05$ ) affected bread  
383 physical characteristics, being the trend and extent of the changes dependent on the treated flour. CN+  
384 and CP+ increased bread specific volume by 4% and 9%, respectively, while T+ led to loaves with 8%  
385 lower specific volume, compared to untreated flours. Effects of CN+ and CP+ on bread viscoelasticity  
386 were of similar extent but followed an opposite trend. CP+ decreased the rate of initial stress decay  
387 rate  $1/k_f$  by 11% with a concomitant increase in the relaxation time of 33%, while CN+ promoted the  
388 decay rate by 12% (Table 5). HMT of T provided a fall in the elastic-like nature of doughs, leading to  
389 increased initial stress decay rate and level of stress not relaxed at long times. This encompasses  
390 higher extent of stress relaxation, and dough weakening. Resulting doughs encompassed lower  
391 viscoelasticity and gas retention ability, and reduced specific volume after fermentation and baking.  
392 Although legume and chestnut proteins, are generally low in methionine, cysteine and tryptophan,  
393 aggregation of proteins can take place on HMT, leading to a reinforced dough structure in some extent,  
394 able to retain carbon dioxide and leading to increased specific volume, as previously observed for  
395 sorghum breads (Marston et al., 2016). Entanglement/disaggregation of fibres, particularly present in  
396 high amount in chickpea flour (Table 1), in the denatured protein network can also play a role. Impact  
397 of HMT flours on firming kinetics parameters were relevant (Figure 2). Both initial  $T_0$  and final  $T_\infty$  bread  
398 firmness were lowered (-7.5%) or promoted (+17-23%) by the respective addition of CP+ or T+ to the  
399 formulation. In addition, the rate of ageing  $n_f$  was drastically reduced (-26 %) or increased (+78%) by  
400 either CP+ or CN+, and  $k_f$  decreased by 72% with T+ (Table 5). Simultaneous presence of native and  
401 treated flours provided variable impact on the physical profile of blended breads, particularly for the  
402 pairs T/CN and T/CP. Suitable trends for the enhancement of the physical characteristics of breads in  
403 terms of larger specific volume, higher viscoelastic and textural profiles, with lower and slower staling  
404 kinetics on ageing (Table 5) were achieved by the pairs T-/CN+, T-/CP+, CN-/CP+ and CN+/CP+, as

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405 in 010, 011, 001 samples. These samples also showed homogeneous and fine crumb grain and  
406 appealing colour features (Figure 1). Caution should be applied to the pair T+/CP+ because of the  
407 adverse effect on the firming of the initial texture of the crumb  $T_0$ , and on the rate of staling denoted by  
408 increased  $n_f$ . It is the case of samples 101 and 111. In fact, comparing staling kinetics parameters  
409 (averaged values) in samples T-CP+ vs T+CP+ (Table 5), values were 7.97N vs 10.84N ( $T_0$ ), and 0.405  
410 vs 1.269 ( $n_f$ ). Experimental values ( $T_0$ ,  $n_f$ ) for bread samples 001 (7.89N, 0.420) and 011 (8.00N, 0.390)  
411 vs 101 (10.54N, 0.758) and 111 (11.25N, 1.775) are in good accordance with the statistical trends  
412 (Figure 2, Table 5).

413 Blended bread samples (100 g) contain 6,71g of dietary fibre DF, while regular white breads  
414 account for 1,67g DF, so that blended breads can be labelled as high-fibre breads (6 g DF/100 g food),  
415 according to Nutritional Claims for DF foods (Regulation EC, 2006). Formulations based on  
416 WT:T:CN:CP flours, 66.20:7:7 fulfilled from 44% (men) to 67% (women) of dietary fibre daily  
417 requirements (Otten, Hellwig, & Meyers, 2006), when a daily consumption of 250 g of bread is  
418 accomplished, following the WHO bread intake recommendation.

419 Chestnut flour is an important source of antioxidant compounds, mainly containing ellagic acid  
420 (Dall'Asta et al., 2013). Teff is generally assumed to contain substantial amounts of phenolics (Dykes  
421 and Rooney 2007), particularly that ferulic acid, and chickpeas exhibited total polyphenol content  
422 ranging from 0.78 to 2.3 mg/ g-1 (Bravo, 1998). Bioaccessible polyphenol content (mg gallic acid/100  
423 g flour, as is) of blended breads in the current work varied from 417 mg to 494 mg (Figure 3). Amount  
424 of bioaccessible polyphenols is dependent on several factors. Mechanical input during mixing and  
425 thermal treatment of flours (HMT) and doughs (baking) may induce depolymerization of constituents,  
426 mainly fibre, and hence may favour bread accessibility to solvents, acids and enzymes and the  
427 subsequent release and extraction of fibre-associated polyphenols. Concomitantly, dietary fibre and  
428 other compounds of proven resistance to the action of digestive enzymes, such as resistant starch,

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429 resistant protein, Maillard compounds and other associated compounds, may reduce the bread phenol  
430 bioaccessibility (Saura-Calixto et al., 2000). Extensive depolymerization of fibre constituents can be  
431 applied to the increase of bioaccessible polyphenols determined in HMT blended breads compared to  
432 the untreated sample 000, since compositional non-wheat flours flours accounted for >10% dietary  
433 fibre content (Table 1). Despite HMT may favour the formation of polymeric aggregates that hinder  
434 enzyme accessibility and further attack (Mann et al., 2013; Watcharatewinkul et al., 2009), it is apparent  
435 that HMT makes bioaccessible polyphenol extraction available, in some cases. In fact, T+ decreased  
436 the amount of bioaccessible polyphenols by 4%, but the pair T/CN modulated polyphenol extractability  
437 (Table 5). T-/CN- and T+/CN+ gave similar amount of extracted polyphenols at about 425 mg, while  
438 T+/CN- led to intermediate amount of 456 mg, and the pair T-CN+ maximized extracted bioaccessible  
439 polyphenols (491 mg). In accordance, samples 010 and 011 exhibited the highest amount of  
440 bioaccessible polyphenols ( $\approx$ 490 mg) while samples 110, 111, 000, and 001 explicated the lowest  
441 values (420-450 mg), and samples 101 and 100 showed intermediate mean values (450-460 mg)  
442 (Figure 3).

443         Anti-radical activity was determined by the extent of the reduction of the stable 2,2-diphenyl-1-  
444 picrylhydrazyl (DPPH•) radical. Results expressed correspond to the remaining unreacted DPPH•  
445 amount when 0.494  $\mu$ mol of the free radical are initially available to react with pepsin/ pancreatin  
446 extracts from 2.4-2.6mg freeze-dried breads. Despite different kinetics of the reduction of DPPH• were  
447 shown among untreated and treated samples (Figure 4), discreet differences in anti-radical activity of  
448 breads (38-45%) were observed. T+ and CN+ respectively induced a depletion (-7%) and an increase  
449 (+3%) of the anti-radical activity, while the simultaneous presence of the pair CN+/CP+ synergistically  
450 promoted the reduction of DPPH• (Table 5). In good accordance samples 011, 010 and 111 gave the  
451 higher values and the more rapid kinetics in terms of anti-radical activity (Figure 4).

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453           **3.3. Correlations between dough physical parameters and bread physical and nutritional**  
454           **parameters of blended matrices**

455           Using Pearson correlation analysis, a range of correlation coefficients ( $r$ ) (from -0.4760 to 0.7352)  
456 were obtained for the relationships between viscoelastic, mechanical and starch gelatinization  
457 transition parameters of doughs, and physical and nutritional parameters of mixed breads from  
458 untreated and HMT matrices (Table 6). Despite  $r$  values were discreet in most correlations, significant  
459 ( $0.01 < p < 0.05$ ) interdependences between the extent of stress relaxation ( $1/k_2$ ), starch gelatinization  
460 transition ( $T_p$ ,  $T_e$ ,  $\Delta H_g$ ) of doughs, and physical (specific volume,  $n_f$ ) and nutritional (%ARA) parameters  
461 of breads were found (Table 6). In accordance, larger specific volume in breads exhibiting higher anti-  
462 radical activity corresponded to higher elastic-like doughs with smaller initial decay rate  $1/k_1$  and lower  
463 extent of stress relaxation  $1/k_2$ , %SR. Clumping of starch granules and aggregation of denatured protein  
464 on HMT were reported (Chen et al., 2015). Crosslinking by oxidizing the free sulfhydryl groups on HMT  
465 result in stronger doughs with a greater resistance to mechanical shock, improved oven spring, and  
466 larger loaf volume, as described for sorghum breads (Marston et al., 2016). In addition, lower  
467 temperatures for starch gelatinization  $T_0$ ,  $T_p$ ,  $T_e$  in hydrated flours led to breads with higher specific  
468 volume ( $r$  -0.6132, -0.6293, -0.6299), while smaller energy for the thermal transition  $\Delta H_g$  was connected  
469 with softer fresh breads ( $r$  0.5696) and slower crumb firming rate on ageing ( $r$  0.718). Modifications of  
470 starch surface interactions take place on HMT (Mann et al., 2013). Most of the semi-crystalline structure  
471 and a considerable extent of the concentrated crystalline region were destroyed during the thermal  
472 treatment (Gunaratne & Hoover, 2002) leading to less compact and rigid structures (Hoover &  
473 Vasanthan, 1994). Resulting doughs are able to expand during fermentation, and provide breads with  
474 more aerated structure and softer crumb, undergoing slower staling kinetics.

475  
476           **Conclusions**

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477 HMT of flours modified viscoelasticity and thermal transitions of doughs and techno-functional and  
478 nutritional profiles of breads from diluted breadmaking wheat matrices made at 34% of wheat flour  
479 replacement by teff (20%), chestnut (7%) and chickpea (7%) flours. The trend and extent of the  
480 changes are mainly dependent on the simultaneous presence of specific untreated and thermally  
481 treated flours, particularly for the pairs T/CN and T/CP. Suitable trends for the enhancement of the  
482 physical characteristics of breads in terms of larger specific volume, higher viscoelastic and textural  
483 profiles, with lower and slower staling kinetics on ageing were achieved by the pairs T-CN+, T-CP+,  
484 CN-CP+ and CN+CP+, Resulting breads showed all homogeneous and fine crumb grain and appealing  
485 colour features. The pair T-CN+ maximized extracted bioaccessible polyphenols, and the pair CN+CP+  
486 synergistically promoted the anti-radical activity in breads. Caution should be applied to the pair T+CP+  
487 because of the adverse effect on the firming of the fresh crumb and on the rate of staling.  
488 HMT of associated non-wheat flours appears as a clean label simple strategy to create added value to  
489 breads from highly diluted wheat flour matrices, provided single and interactive effects of the thermal  
490 treatment of blended flours on the structural features of breads are known.

491  
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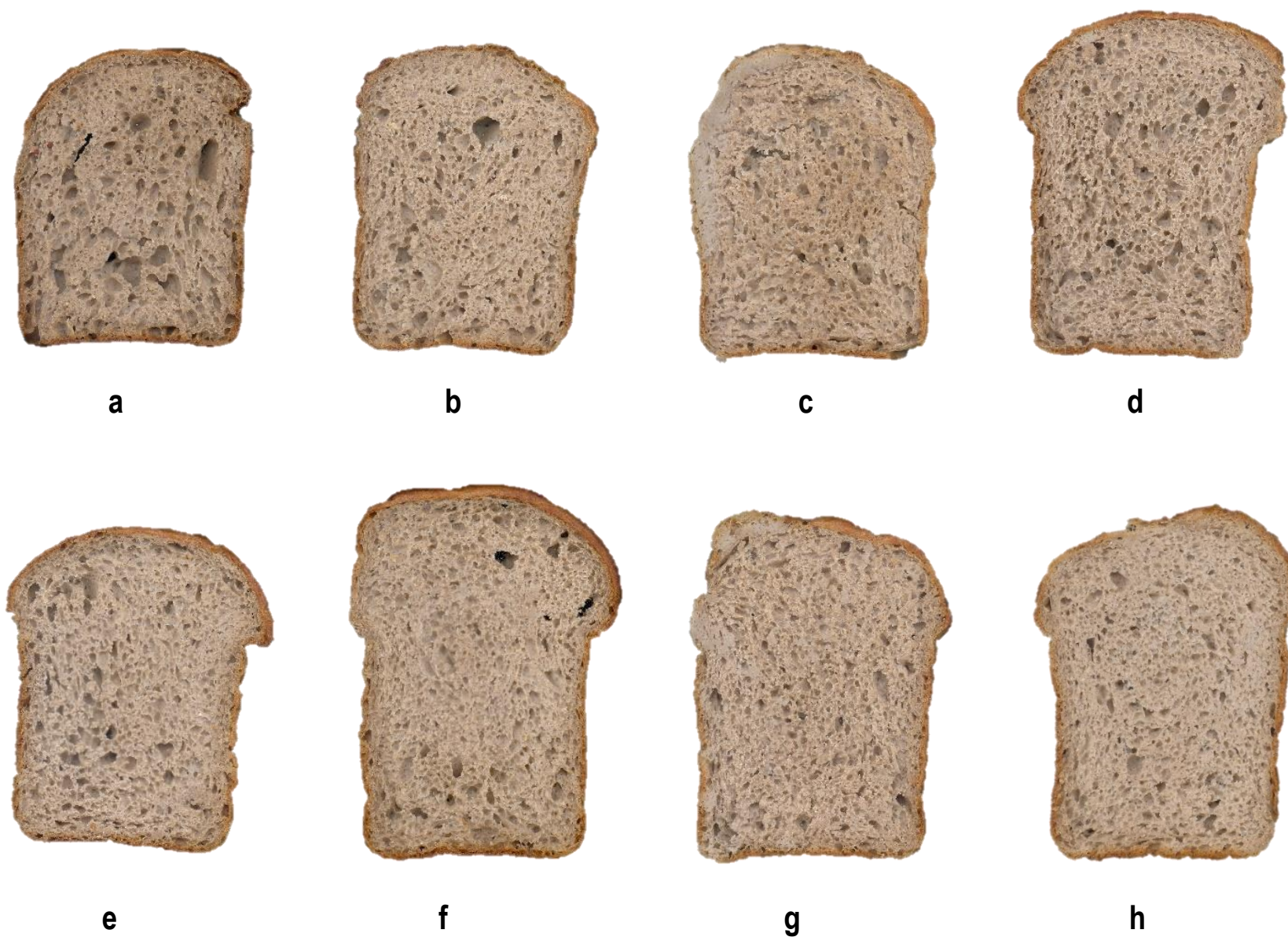
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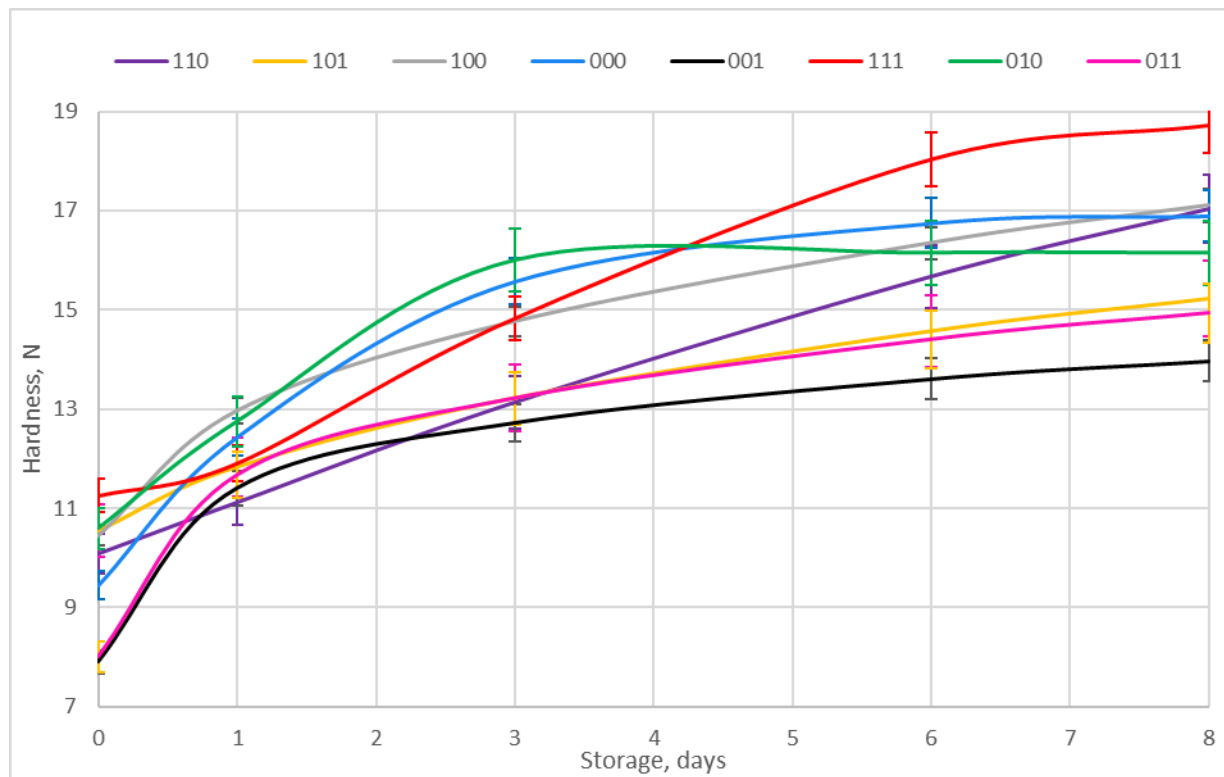
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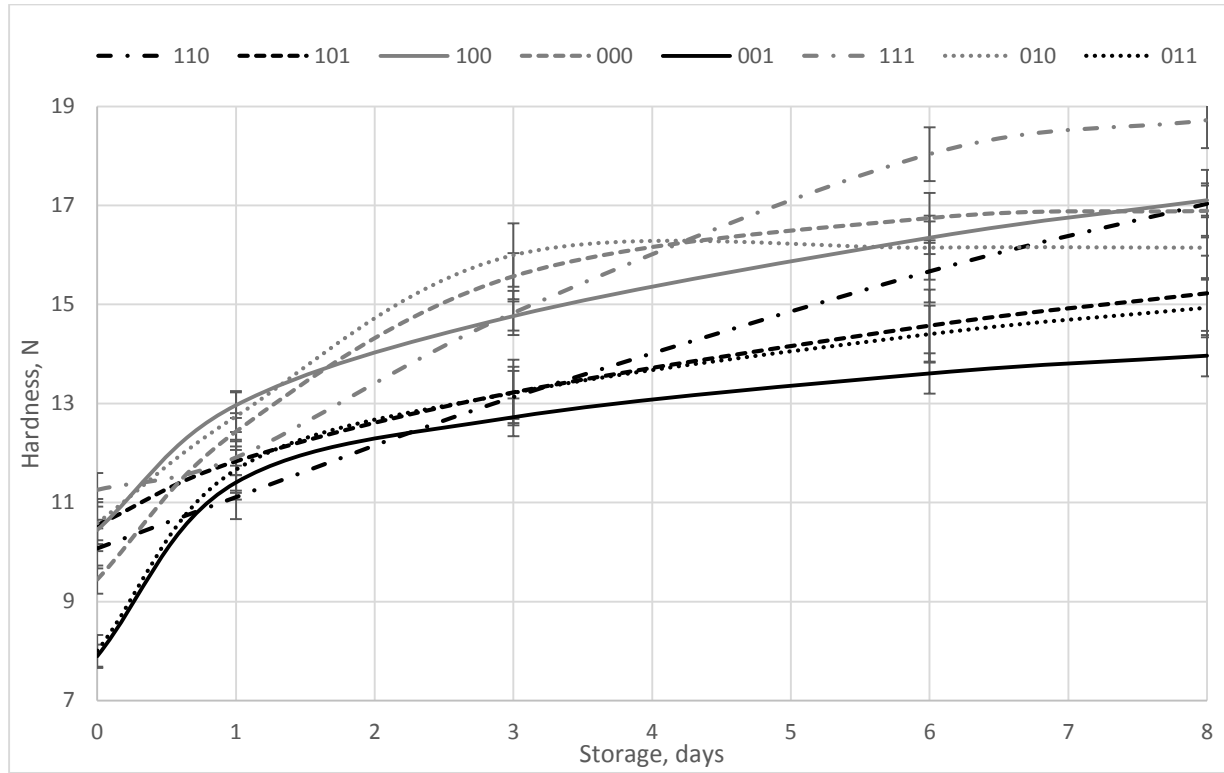
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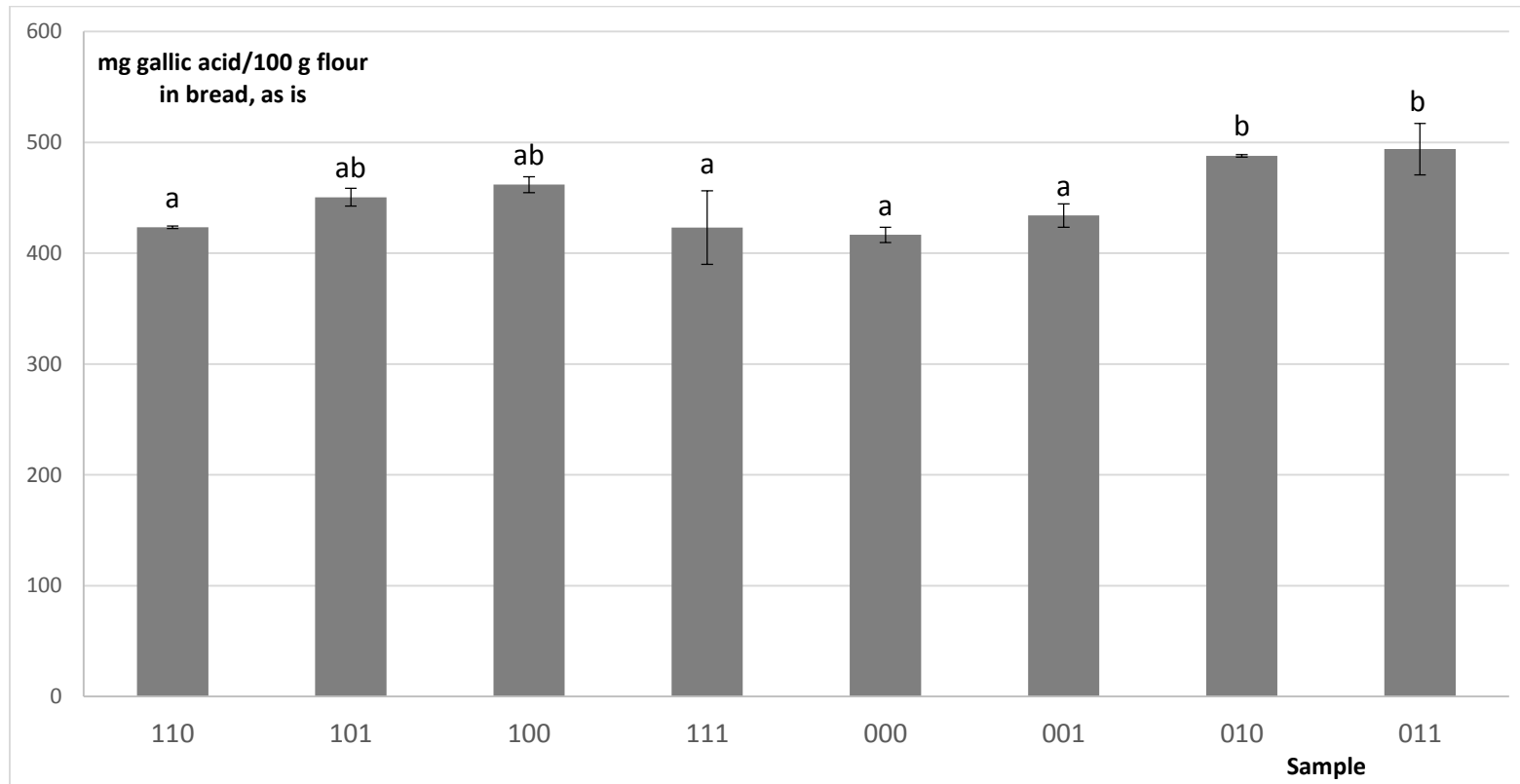
**Figure 1.-** Digitalized images of central slices of wheat-based blended breads formulated with teff (T), chestnut (CN), and chickpea (CP) flours. Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP mixed flours replacing wheat flour in sample formulation. a: 110, b: 101, c: 100, d: 000, e: 001, f: 111, g: 010, h: 011.



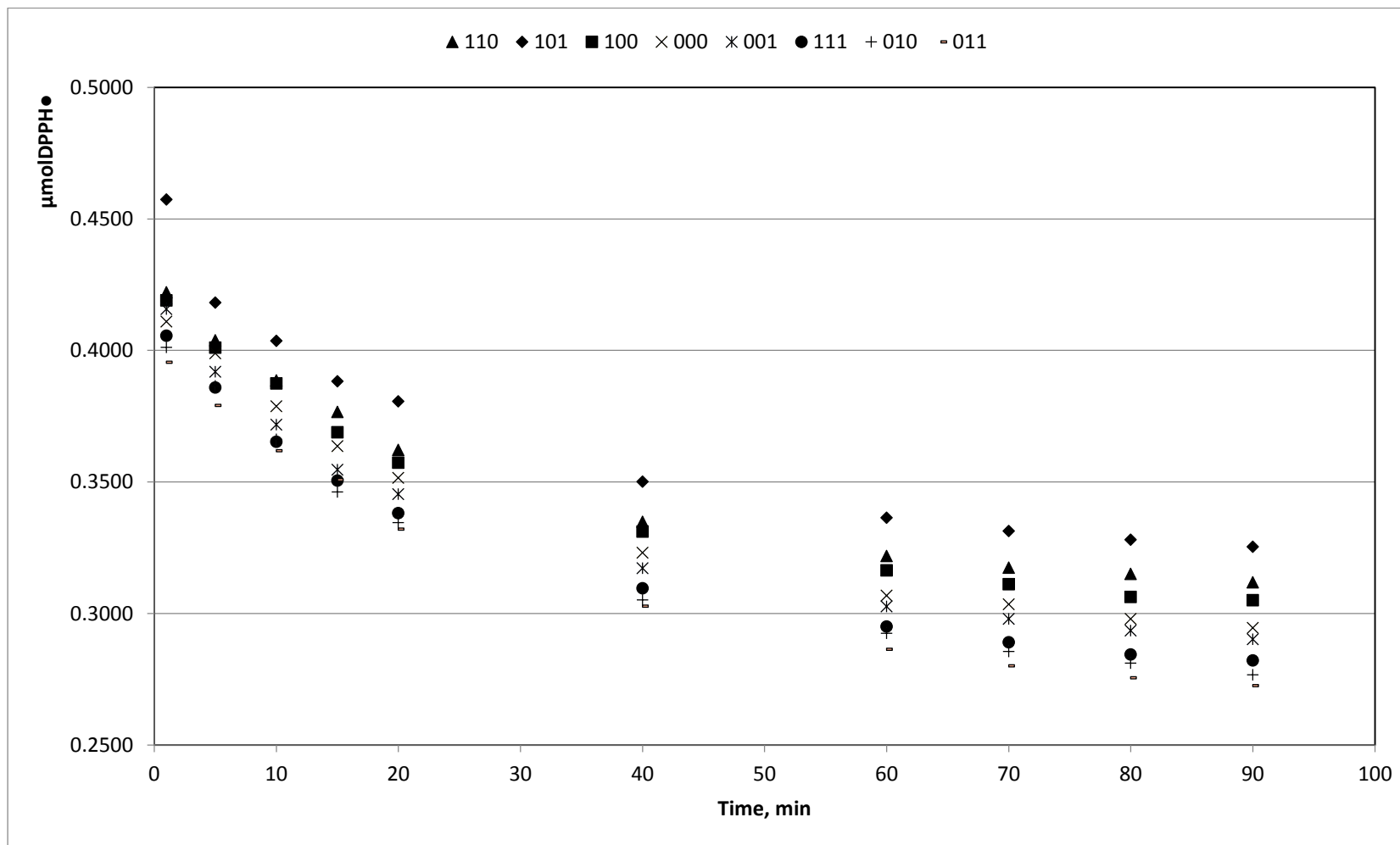
**Figure 2.-** Evolution of crumb firming during ageing (adjusted mean data and error bars) of wheat-based blended breads formulated with teff (T), chestnut (CN), and chickpea (CP) flours. Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP flours replacing wheat flour in sample formulation.



**Figure 2.-** Evolution of crumb firming during ageing (adjusted mean data and error bars) of wheat-based blended breads formulated with teff (T), chestnut (CN), and chickpea (CP) flours. Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP flours replacing wheat flour in sample formulation.



**Figure 3.-** Bioaccessible polyphenol content of wheat-based blended breads formulated with teff (T), chestnut (CN), and chickpea (CP) flours. Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP flours replacing wheat flour in sample formulation.



**Figure 4.-** Kinetics of consumption of 0.494  $\mu\text{mol}$ s DPPH by 2.8 mg freeze-dried wheat-based blended breads formulated with teff (T), chestnut (CN) and chickpea (CP) flours. Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP flours replacing wheat flour in sample formulation.

**Table 1.-** Chemical and nutritional composition<sup>a</sup> of flours

Parameter (g/ 100 g flour, d. b.)	Teff	Chestnut	Chickpea	Wheat
Moisture	12.62±0.13 <sup>c</sup>	6.90±0.05 <sup>a</sup>	11.88±0.09 <sup>b</sup>	14.30±0.12 <sup>d</sup>
Protein	14.08±0.46 <sup>b</sup>	6.44±0.20 <sup>a</sup>	18.82±0.60 <sup>c</sup>	14.12±0.28 <sup>b</sup>
Fat	4.69±0.29 <sup>c</sup>	4.08±0.18 <sup>b</sup>	6.96±0.12 <sup>d</sup>	1.56±0.11 <sup>a</sup>
Digestible carbohydrates	68.7±4.5 <sup>b</sup>	79.5±3.2 <sup>c</sup>	48.8±1.1 <sup>a</sup>	81.7±2.3 <sup>c</sup>
Total Dietary Fibre	12.31±1.42 <sup>c</sup>	9.67±1.03 <sup>b</sup>	25.13±2.33 <sup>d</sup>	2.56±0.23 <sup>a</sup>
Soluble Dietary Fibre	4.84±0.55 <sup>b</sup>	-	6.55±0.42 <sup>c</sup>	1.06±0.11 <sup>a</sup>
Insoluble Dietary Fibre	7.46±1.10 <sup>b</sup>	-	18.62±1.40 <sup>c</sup>	1.49±0.28 <sup>a</sup>

<sup>a</sup>Mean values ± standard deviation. Within rows, values (mean of three replicates) with the same following letter do not differ significantly from each other (p> 0.05).



**Table 2.-** Physicochemical parameters of blended doughs.

Parameter	Dough sample <sup>a, b</sup>							
	110	101	100	000	001	111	010	011
<b>Viscoelasticity</b>								
$F_0, N$	1.16±0.02b	1.03±0.02a	1.73±0.04d	0.89±0.04a	1.68±0.07d	1.25±0.08b	1.49±0.07c	1.14±0.05b
$k_1, s$	11.39±0.43a	13.81±0.13b	16.58±0.30e	19.50±0.35f	14.56±0.30c	15.64±0.25d	15.42±0.30cd	15.05±0.04c
$k_2$	1.36±0.01a	1.41±0.01b	1.44±0.02c	1.52±0.02d	1.42±0.01b	1.47±0.02d	1.44±0.02c	1.48±0.01d
$1/k_1, s^{-1}$	0.087±0.006f	0.072±0.002e	0.060±0.001b	0.051±0.001a	0.068±0.001e	0.063±0.003bc	0.064±0.004cd	0.066±0.003cd
$1/k_2$	0.731±0.001e	0.706±0.002d	0.690±0.001c	0.653±0.011a	0.702±0.005d	0.678±0.006ab	0.691±0.008bc	0.674±0.003ab
$F_{RT}(36,8\%F_0), N$	0.43±0.01b	0.38±0.01a	0.64±0.02d	0.33±0.02a	0.62±0.03d	0.46±0.02b	0.55±0.03c	0.42±0.02b
$RT, s$	53±2a	83±2b	123±4cd	367±23f	91±3b	144±5de	114±7c	151±8e
%SR	71±2d	68±1c	67±0abc	63±1a	68±2cd	66±2bc	67±2bc	65±0ab
<b>Texture</b>								
Hardness, N	5.88±0.41ab	5.64±0.55ab	6.45±1.54b	4.38±0.58ab	6.30±1.22b	3.85±0.21a	5.91±1.11ab	6.14±1.05b
Cohesiveness	0.788±0.045a	0.79±0.051a	0.791±0.083a	0.854±0.002a	0.811±0.061a	0.858±0.001a	0.797±0.068a	0.800±0.053a
Springiness	0.884±0.098h	0.868±0.076f	0.847±0.096e	0.843±0.063c	0.844±0.094d	0.822±0.076a	0.831±0.085b	0.879±0.096g
Resilience	0.108±0.008ab	0.099±0.013a	0.095±0.005a	0.118±0.011b	0.101±0.002ab	0.101±0.005ab	0.103±0.006ab	0.103±0.005ab
Adhesiveness, N.s	39.19±1.00bc	37.87±2.99bc	46.88±9.46c	26.25±1.51ab	45.25±7.68c	24.26±0.99a	43.24±7.50c	41.30±6.98c
<b>Gelatinization</b>								
$T_o, ^\circ C$	62.24±0.67ab	62.30±0.47b	61.93±0.16ab	61.64±0.02ab	61.77±0.21ab	61.42±0.13ab	61.50±0.16ab	61.23±0.88a
$T_p, ^\circ C$	70.50±0.71b	70.50±0.71b	70.10±0.14ab	69.80±0.00ab	69.70±0.14ab	69.50±0.14ab	69.60±0.28ab	69.30±0.99a
$T_e, ^\circ C$	80.31±1.11b	80.39±1.06ab	79.60±0.49ab	79.15±0.10ab	78.95±0.14ab	78.70±0.04ab	78.93±0.40ab	78.47±1.61a
$\Delta H_g, J/g$	5.548±0.064a	5.643±0.004a	5.709±0.117ab	6.071±0.181c	5.694±0.193a	5.996±0.063bc	6.17±0.139c	5.587±0.132a
$R, ^\circ C$	18.07±0.44ab	18.09±0.59b	17.68±0.32ab	17.51±0.12ab	17.18±0.07a	17.29±0.09ab	17.43±0.23ab	17.24±0.73ab

<sup>a</sup> Mean values ± standard deviation. Within rows, values (mean of three replicates) with the same following letter do not differ significantly from each other ( $p > 0.05$ ). (<sup>b</sup>) Three digit bread sample code refers to untreated (0) and HMT (1) teff: chestnut: chickpea flours in sample formulation.

$F_0$  is the initial force,  $k_1(s)$ ,  $k_2$  are constants related to stress decay rate and to residual stress at the end of the experiment, respectively,  $RT$  as the time required for  $F_0$  to drop to 36.8% to its values,  $F_{RT}$ , as the force corresponding to  $RT$ , % SR=  $((F_0 - F_{300}) / F_0) 100$ .  $T_o$ : onset temperature,  $T_p$  peak temperature,  $T_e$  end temperature,  $\Delta H_g$  gelatinization entalpy,  $R$  gelatinization temperature range

**Table 3.-** Significant ( $p < 0.05$ ) single effects and 2<sup>nd</sup> order interactions of HMT compositional flours on dough physical properties.

Parameter	Single effects								Second order interactions							
	Level	Overall mean	Teff	*	Chestnut	*	Chickpea	*	Level	Teff x Chestnut	*	Teff x Chickpea	*	Chestnut x Chickpea	*	
<b>Stress Relaxation</b>																
$k_1$	s	0	15.25	16.14	b	16.12	b	15.72	b	00	ns	17.46	c	18,04	d	
		1		14.36	a	14.38	a	14.77	a	01		14.81	b	14,19	b	
										10		13.98	a	13,41	a	
										11		14.73	b	15,35	c	
$k_2$		0	1.45	1.47	b	1.45	b	ns	00	ns	1.49	d	1,49	d		
		1		1.43	a	1.44	a		01		1.45	c	1,42	b		
									10		1.41	a	1,41	a		
									11		1.44	b	1,48	c		
$1/k_1$	s-1	0	0.067	0.063	a	0.063	a	ns	00	ns	0.058	a	0,056	a		
		1		0.071	b	0.071	b		01		0.068	b	0,071	c		
									10		0.074	d	0,076	d		
									11		0.068	c	0,065	b		
$1/k_2$		0	0.692	0.681	a	0.689	a	0.692	b	00	0.678	a	0.673	a	0,672	a
		1		0.702	b	0.694	b	0.691	a	01	0.683	b	0.689	b	0,705	b
									10	0.699	c	0.711	d	0,711	c	
									11	0.705	d	0.693	c	0,677	a	
RT	s	0	141	181	b	166	b	164	b	00	229	b	241	c	245	c
		1		101	a	115	a	117	a	01	133	a	121	b	87	a
									10	103	a	88	a	83	a	
									11	98	a	113	a	147	b	
%SR		0	67	66	a	66	a	ns	00	ns	65	a	65	a		
		1		68	b	67	b		01		67	b	68	b		
									10		69	c	69	b		
									11		67	b	65	a		
<b>Texture Profile Analysis</b>																
Hardness	N	0	5.57	ns		ns		ns	00	ns	5.14	ab	ns			
		1							01		6.22	b				
									10		6.17	ab				
									11		4.75	a				
Adhesiveness	N.s	0	38.03	ns		ns		ns	00	35.75	ab	34.74	ab	ns		
		1							01	42.27	b	43.27	b			
									10	42.37	b	43.03	b			
									11	31.72	a	31.06	a			
<b>Gelatinization</b>																
$T_o$	°C	0		61.54	a	ns		ns		ns	ns		ns			
		1	61.91	62.28	b											
$T_p$	°C	0		69.6	a	ns		ns		ns	ns		ns			
		1	70.05	70.5	b											
$T_e$	°C	0		78.87	a	ns		ns		ns	ns		ns			
		1	79.51	80.15	b											
$\Delta H_g$	J/g	0		ns		ns		ns	00	ns	6.12	b	ns			
		1	5.82						01		5.64	a				
									10		5.63	a				
									11		5.88	ab				
R	°C	0		17.34	a	ns		ns		ns	ns		ns			
		1	17.6	17.87	b											

$k_1(s)$ ,  $k_2$  are constants related to stress decay rate and to residual stress at the end of the experiment, respectively, RT as the time required for  $F_0$  to drop to 36.8% to its values, % SR=  $((F_0 - F_{300}) / F_0) 100$ .  $T_o$ : onset temperature,  $T_p$  peak temperature,  $T_e$  end temperature,  $\Delta H_g$  gelatinization entalpy, R gelatinization temperature range. Level 0: untreated, level 1: HMT.

**Table 4.-** Physical profile of wheat-based blended breads formulated with untreated and HMT teff (T), chestnut (CN) and chickpea (CP) flours.

Parameter	Bread sample <sup>a</sup>							
	110	101	100	000	001	111	010	011
<b>Volume</b>								
Specific volume, mL/g	2.59±0.09a	2.72±0.09a	2.67±0.06a	3.07±0.01c	3.01±0.08bc	3.28±0.01d	2.89±0.05b	3.18±0.11cd
<b>Stress relaxation</b>								
$F_0$ , N	8.01±0.10b	8.62±0.22c	8.46±0.31c	7.92±0.24b	6.99±0.18a	10.72±0.29d	11.31±0.34e	7.07±0.23a
$k_1$ , s	56.70±1.24c	63.04±1.89d	51.94±2.03b	57.24±1.32c	66.58±2.56e	50.97±1.95b	45.67±0.97a	55.64±0.99c
$k_2$	2.06±0.03c	1.98±0.09abc	2.01±0.06ab	1.98±0.11bc	2.19±0.08d	1.96±0.06abc	1.86±0.05a	2.04±0.07bc
$F_{RT}$ (60% $F_0$ ), N	4.81±0.09	5.17±0.16	5.08±0.21	4.75±0.42	4.20±0.32	6.43±0.53	6.79±0.78	4.24±0.21
RT, s	132±5f	121±3e	107±4c	111±9d	220±12g	96±8b	72±7a	123±14e
%SR	46±5a	48±3a	48±3a	48±5a	43±2a	49±6a	51±4a	47±2a
<b>Mechanical properties</b>								
Cohesiveness	0.568±0.058c	0.460±0.021a	0.484±0.012a	0.485±0.027a	0.532±0.037bc	0.481±0.012a	0.495±0.027ab	0.501±0.019ab
Springiness	0.853±0.080a	0.875±0.050a	0.898±0.052a	0.879±0.052a	0.899±0.075a	0.895±0.032a	0.906±0.026a	0.872±0.055a
Chewiness, N	4.81±0.95ab	4.21±0.80a	4.55±0.62a	4.03±0.48a	5.83±1.03bc	4.86±0.45ab	6.53±0.22c	4.58±0.86a
Resilience	0.245±0.004b	0.186±0.019a	0.202±0.007a	0.200±0.022a	0.231±0.027b	0.196±0.004a	0.197±0.013a	0.207±0.015a
<b>Firming kinetics</b>								
$T_\infty$ , N	23.30e	19.16c	25.30f	16.94b	16.17a	18.93c	16.15a	22.11d
$k_f$	0.0814a	0.161c	0.185d	0.508g	0.551h	0.089b	0.490f	0.300e
$n_f$	1.065e	0.758d	0.559c	1.097f	0.420b	1.775g	1.817h	0.390a
$T_0$ , N	10.07c	10.54d	10.44cd	9.44b	7.89a	11.25e	10.58de	8.00a
<b>Crumb grain</b>								
Mean cell area, mm <sup>2</sup>	0.59b	0.45a	0.46a	0.43a	0.74c	0.82d	0.77c	0.47a
Cell area distribution, %								
<1mm <sup>2</sup>	13a	17b	13a	15ab	12a	16b	13a	18b
1-5mm <sup>2</sup>	20a	36c	29b	30b	28b	41c	34c	37c
5-50mm <sup>2</sup>	47b	47b	58c	45b	35a	43b	46b	45b
>50mm <sup>2</sup>	20c	0a	0a	10b	25c	0a	7b	0a
Cell number distribution, %								
<1mm <sup>2</sup>	92b	91b	92b	92b	88ab	81a	85a	89ab
1-5mm <sup>2</sup>	5a	7a	6a	6a	9a	16b	12b	9a
5-50mm <sup>2</sup>	2a	2a	2a	2a	3a	3a	3a	2a
>50mm <sup>2</sup>	0.1a	0a	0a	0.1a	0.2a	0a	0.1a	0a
Cell density, cells/cm <sup>2</sup>	86c	91c	107d	89c	56b	48a	60b	84c
Cell to total area ratio	50b	41a	49b	38a	41a	39a	46b	39a
Wall to total area ratio	50a	59b	51a	62b	59b	61b	54a	61b
<b>Colour</b>								
L	54.8±a	59.2±e	56.1±b	57.4±c	58.9±de	60.4±f	58.2±d	60.1±f
a	2.4±0.1a	2.8±0.1b	3.4±0.2d	2.5±0.1a	2.5±0.1a	2.5±0.1a	3.0±0.1c	2.3±a
b	10.3±0.3a	11.5±0.1d	12.3±0.1e	10.6±0.1ab	10.4±0.2a	10.7±0.1ab	11.0±0.1c	10.7±ab
Whiteness Index	53.6	57.5	54.3	56.0	57.5	58.9	56.7	58.6

<sup>a</sup>Three digit code refers to untreated (0) and heat-moisture treated (1) T:CN:CP blended breads.

$F_0$  is the initial force,  $k_1$ (s),  $k_2$  are constants related to stress decay rate and to residual stress at the end of the experiment, respectively, RT as the time required for  $F_0$  to drop to 60% to its values,  $F_{RT}$ , as the force corresponding to RT, % SR=  $((F_0-F_{600})/F_0)100$ .  $k_f$  constant of proportion of firming kinetics,  $n_f$  Avrami exponent of firming kinetics,  $T_\infty$ ,  $T_0$  crumb firmness at  $\infty$  and 0 time.

**Table 5.-** Significant ( $p < 0.05$ ) single effects and 2<sup>nd</sup> order interactions of HMT compositional flours on the physical and nutritional profiles of wheat-based blended breads formulated with untreated (0) and HMT (1) teff (T), chestnut (CN) and chickpea (CP) flours.

Parameter	Unit	Single effects								2nd order interactions							
		Overall mean	Level	Teff	*	Chestnut	*	Chickpea	*	Level	Teff x Chestnut	*	Teff x Chickpea	*	Chestnut x Chickpea	*	
<b>Volume</b>																	
SV	mL/g	2.92	0	3.04	b	2.87	a	2.8	a	00	3.04	b	2.98	b	2.87	b	
			1	2.81	a	2.98	b	3.05	b	01	3.03	b	3.10	b	2.87	b	
										10	2.69	a	2.63	a	2.74	a	
										11	2.94	b	3.00	b	3.23	c	
<b>Viscoelastic</b>																	
$F_0, N$		8.59	0	ns		7.97	a	ns		00	ns	9.51	b	ns			
			1			9.21	b			01		7.00	a				
										10		8.24	ab				
										11		9.61	b				
$k_1$	s-1	55.91	0	ns		59.70	b	52.73	a	00	62.45	b	51.15	a	54.25	a	
			1			52.13	a	59.10	b	01	50.65	a	61.95	b	65.15	b	
										10	56.95	ab	54.30	a	51.20	a	
										11	53.60	a	56.25	ab	53.05	a	
RT		122	0	131	b	140	b	105	a	00	165	c	91	a	109	b	
			1	114	a	105	a	140	b	01	97	a	171	d	171	c	
										10	114	b	119	c	101	a	
										11	113	b	108	b	109	b	
%SR		47	0	ns		47	a	48	b	00	45	a	50	d	ns		
			1			48	b	46	a	01	49	c	44	a			
										10	48	b	47	b			
										11	47	b	48	c			
<b>Textural</b>																	
Cohesiveness		0,503	0	ns		ns		ns		00	0.515	b	0.475	a	ns		
			1							01	0.500	ab	0.540	b			
										10	0.459	a	0.534	b			
										11	0.538	c	0.462	a			
Resilience		0.207	0	ns		0.201	a	ns		00	0.216	bc	0.188	a	0.192	a	
			1			0.214	b			01	0.206	b	0.234	b	0.209	b	
										10	0.185	a	0.224	b	0.220	b	
										11	0.222	c	0.183	a	0.208	b	
Chewiness	N	4.59	0	ns		4.11	a	ns		00	ns	ns	ns	ns			
			1			5.06	b			01							
<b>Firming kinetics</b>																	
$T_\infty$	N	19.69	0	17.67	a	ns		20.46	b	00	16.53	a	16.52	a	21.06	c	
			1	21.71	b			18.93	a	01	18.82	b	18.82	b	17.59	a	
										10	22.12	c	24.40	c	19.86	b	
										11	21.31	c	19.03	b	20.26	bc	
$k_f$		0.297	0	0.463	b	0.353	b	ns		00	ns	ns	ns	ns			
			1	0.131	a	0.241	a			01							
$n_f$		0.986	0	ns		0.709	a	1.135	b	00	ns	1.458	c	ns			
			1			1.263	b	0.837	a	01		0.405	a				
										10		0.813	b				
										11		1.269	bc				
$T_0$	N	9.77	0	8.99	a	ns		10.14	b	00	ns	10.01	b	ns			
			1	10.55	b			9.41	a	01		7.97	a				
										10		10.26	b				
										11		10.84	b				
<b>Nutritional</b>																	
BPP		449	0	458	b	ns		ns		00	425	a	ns	ns			
mg gallic acid/100 g flour			1	440	a					01	491	c					
										10	456	b					
										11	424	a					
ARA	%	42	0	44	b	42	a	ns		00	ns	ns	ns	43	b		
			1	41	a	43	b			01				40	a		
										10				43	b		
										11				44	c		

$F_0$  : initial force,  $k_1(s)$ ,  $k_2$  are constants related to stress decay rate and to residual stress at the end of the experiment, respectively, RT: time required for  $F_0$  to drop to 36.8% (dough) and 60% (bread) to its values, % SR=  $((F_0 - F_{300/600}) / F_0) 100$ ,  $T_0$ : onset temperature,  $T_p$ : peak temperature,  $T_e$  end temperature,  $\Delta H_g$ : gelatinization enthalpy,  $T_\infty$ : crumb firmness at  $\infty$  time,  $k_f$ : constant of proportion of firming kinetics,  $n_f$ : Avrami exponent of firming kinetics, BPP: bioaccessible polyphenols, ARA: anti-radical activity.

**Table 6.-** Significant Pearson correlations ( $p < 0.05$  \*,  $p < 0.01$  \*\*) between dough viscoelastic, mechanical, and thermal parameters and bread textural, nutritional, and firming kinetic characteristics from untreated and heat moisture treated blended matrices.

Dough	Bread											
	SV, ml/g	Hardness, N	Cohesiveness	Resilience	$F_0$ , N	RT, s	SR, %	$L$	$a$	% ARA	$T_\infty$ , N	$n_f$
$F_0$ , N									0,5382			
									*			
$k_1$ , s			-0.4975	-0.4873						0.5365		
			*	*						*		
$k_2$	0.7352							0.4832		0.7334		
	**							*		**		
$1/k_1$ , s <sup>-1</sup>	-0.476		0.5074	0.5343						-0.5034		
	*		*	*						*		
$1/k_2$	-0.7341		0.4993					-0.5205		-0.7221		
	**		*					*		**		
RT, s										0.5731		
										*		
SR, %	-0.5436		0.5671							-0.6011		
	*		*							*		
Springiness		-0.4896			-0.6481						0.536	-0.5938
		*			**						*	*
$T_o$ , °C	-0.6132									-0.4835		
	*									*		
$T_p$ , °C	-0.6293							-0.487				
	**							*				
$T_e$ , °C	-0.6299											
	**											
$\Delta H_g$ , J/g		0.5696			0.6498	-0.4886	0.5774				-0.5772	0.718
		*			**	*	*				*	**

$F_0$ : initial force,  $k_1$ (s),  $k_2$  are constants related to stress decay rate and to residual stress at the end of the experiment, respectively, RT: time required for  $F_0$  to drop to 36.8% (dough) and 60% (bread) to its values, % SR=  $((F_0 - F_{300/600}) / F_0) 100$ ,  $T_o$ : onset temperature,  $T_p$ : peak temperature,  $T_e$ : end temperature,  $\Delta H_g$ : gelatinization enthalpy,  $T_\infty$ : crumb firmness at  $\infty$  time,  $n_f$ : Avrami exponent of firming kinetics, SV: specific volume, % ARA: percent of anti-radical activity.

**Abstract** The significance of heat moisture treatment (HMT) of non-wheat –teff (T), chestnut (CN) and chickpea (CP) flours on dough viscoelastic and thermal parameters and on the structural and nutritional pattern of breads was investigated in untreated (-) and HMT (+) associated wheat-based (WT) matrices (WT:T:CN:CP, 66.20:7:7, wt. basis). Suitable trends for the enhancement of the physical characteristics of breads in terms of larger specific volume, higher viscoelastic and textural profiles, with lower and slower staling kinetics on ageing were achieved by the pairs T-CN+, T-CP+, CN-CP+ and CN+CP+. In addition, a fine and uniformly- sized cell structure with similar cell walls thickness were achieved in crumb samples. The pair T-CN+ enhanced extracted bioaccessible polyphenols, and the pair CN+CP+ synergistically promoted the anti-radical activity in breads. Blended breads can be labelled as high-fibre breads ( $\geq 6$  g DF/100 g food), and a recommended daily consumption of 250 g of bread fulfilled from 44% (men) to 67% (women) of dietary fibre requirements.