

<b>Zebrafish</b>					
<b>Viewpoints</b>	<b>Left</b>	<b>Right</b>	<b>Anterior</b>	<b>Cluster</b>	<b>Posterior</b>
<i>nfe2l2a</i>	8.8%	91.2%	71.6%	27.1%	1.4%
<i>hnrpa3</i>	45.9%	54.1%	63.9%	33.4%	2.7%
<i>hoxd4a</i>	83.3%	16.7%	/		
<i>hoxd10a</i>	70.7%	29.3%			
<i>hoxd11a</i>	67.4%	32.6%			
<i>hoxd13a</i>	23.4%	76.6%			
<i>evx2</i>	26.8%	73.2%			
<i>lnpa</i>	20.7%	79.3%			
<i>atp5g3a</i>	62.4%	37.6%			
<b>Amphioxus</b>					
<b>Viewpoints</b>	<b>Left</b>	<b>Right</b>	<b>Anterior</b>	<b>Cluster</b>	<b>Posterior</b>
<i>Meox</i>	44.2%	55.8%	100.0%	0.0%	0.0%
<i>Hnrpa</i>	38.5%	61.5%	71.7%	25.6%	2.7%
<i>Mtx2</i>	30.9%	69.1%	41.2%	42.7%	16.1%
<i>Hox2</i>	24.8%	75.2%	24.8%	75.2%	0.0%
<i>Hox5</i>	38.5%	61.5%	18.8%	80.2%	1.0%
<i>Hox6</i>	44.1%	55.9%	24.7%	71.3%	4.0%
<i>Hox7</i>	51.6%	48.4%	21.6%	74.1%	4.3%
<i>Hox9</i>	46.9%	53.1%	14.1%	83.2%	2.6%
<i>Hox11</i>	62.1%	37.9%	0.0%	99.5%	0.5%
<i>Hox13</i>	65.7%	34.3%	1.7%	88.7%	9.5%
<i>Hox15</i>	74.2%	25.8%	5.3%	68.9%	25.8%
<i>Evxa</i>	66.1%	33.9%	11.6%	45.6%	42.8%
<i>Lnp</i>	73.0%	27.0%	6.4%	35.5%	58.1%
<i>Gpatch8</i>	50.6%	49.4%	12.0%	3.5%	84.5%

**Supplementary Table 1**

**Contact distribution for each of the amphioxus viewpoints.** The slashed cell correspond to those viewpoints where the exclusion of reads around the viewpoint overlap with more than 30% of the total length of the zebrafish HoxDa cluster.

Amphioxus		Zebrafish	
Eliminated Viewpoints	Correlation	Eliminated Viewpoints	Correlation
Gpatch88	0,93	atp5g3a	0,84
Lnp	0,96	lnpa	0,90
Evxa	0,95	evx2	0,81
Hox15	0,92	hoxd13a	0,89
Hox13	0,93	hoxd11a	0,90
Hox11	0,95	hoxd10a	0,92
Hox9	0,94	hoxd4a	0,93
Hox7	0,94	hoxd3	0,91
Hox6	0,95	nfe2l2a	0,89
Hox5	0,96	hnrmpa3	0,91
Hox2	0,95	evx2, hoxd10a	0,86
Mtx2	0,93	lnpa, hoxd3	0,92
Hnrmpa	0,94	atp5g3a, nfe2l2a	0,86
Meox	0,94	hoxd13a, hnrmpa3	0,88
Evxa, Hox13	0,94	hoxd11a, hoxd4a	0,91
Hox5, Meox	0,94	evx2, hoxd11a, nfe2l2a, hnrmpa3	0,85
Gpatch8, Hox9	0,88	atp5g3a, hoxd13a, hoxd11a, hoxd10a	0,86
Lnp, Hox2	0,95	lnpa, hoxd13a, hoxd10a, hoxd3	0,91
Hox11, Mtx2	0,93	evx2, hoxd13a, hoxd3, nfe2l2a	0,82
Hox15, Hox13, Hox7, Meox	0,82	atp5g3a, hoxd11a, hoxd4a, hoxd3	0,87
Gpatch8, Hox,11, Hox2, Mtx2	0,87	atp5g3a, evx2, hoxd11a, hoxd10a, hoxd4a, nfe2l2a	0,84
Lnp, Evxa, Hox9, Hox6	0,93	lnpa, hoxd13a, hoxd4a, hoxd3, nfe2l2a, hnrmpa3	0,77
Evxa, Hox11, Hox6, Hnrmpa	0,91	lnpa, hoxd13a, hoxd10a, hoxd4a, hoxd3, hnrmpa3	0,83
Hox15, Hox11, Hox7, Mtx2	0,89	atp5g3a, evx2, hoxd13a, hoxd11a, hoxd3, nfe2l2a	0,80
Evxa, Hox13, Hox6, Hox5, Mtx2, Meox	0,92	evx2, hoxd13a, hoxd11a, hoxd4a, hoxd3, nfe2l2a	0,82
Gpatch8, Hox11, Hox9, Hox6, Hox5, Hox2	0,90		
Lnp, Hox15, Hox11, Hox7, Hox2, Mtx2	0,86	KEY	
Evxa, Hox15, Hox9, Hox7, Mtx2, Hnrmpa	0,81	Number of viewpoints eliminated	
Lnp, Hox13, Hox9, Hox6, Hox2, Hnrmpa	0,88		1
Gpatch8, Evxa, Hox13, Hox7, Hox6, Hox2, Mtx2, Hnrmpa	0,82		2
Evxa, Hox15, Hox11, Hox9, Hox5, Hox2, Hnrmpa, Meox	0,89		4
Lnp, Evxa, Hox11, Hox7, Hox6, Hox5, Mtx2, Hnrmpa	0,85		6
Lnp, Hox15, Hox13, Hox11, Hox9, Hox7, Hox2, Mtx2	0,85		8
Evxa, Hox15, Hox13, Hox9, Hox6, Hox5, Hox, 2Hnrmpa	0,82		10
Lnp, Evxa, Hox13, Hox11, Hox9, Hox6, Hox5, Hox2, Mtx2, Meox	0,89		
Gpatch8, Evxa, Hox15, Hox13, Hox9, Hox7, Hox6, Hox5, Mtx2, Hnrmpa	0,83		
Lnp, Hox15, Hox13, Hox9, Hox7, Hox6, Hox5, Hox2, Mtx2, Hnrmpa	0,68		
Gpatch8, Lnp, Evxa, Hox13, Hox11, Hox9, Hox6, Hox5, Mtx2, Meox	0,88		
Gpatch8, Evxa, Hox15, Hox13, Hox11, Hox7, Hox6, Hox2, Mtx2, Hnrmpa	0,54		

**Supplementary Table 2**  
**Spearman's correlations of the virtual Hi-C jackknife resampling experiments**

<b>Dataset 1</b>	<b>Dataset 2</b>	<b>Pearson's correlation</b>	<b>Spearman's correlation</b>
ESCells-HindIII-rep1	ESCells-HindIII-rep2	0,98	0,97
ESCells-HindIII-rep1	ESCells-Ncol	0,94	0,95
ESCells-HindIII-rep1	Cortex-HindIII-rep1	0,8	0,76
ESCells-HindIII-rep1	Cortex-HindIII-rep2	0,79	0,8
ESCells-HindIII-rep2	ESCells-Ncol	0,93	0,94
ESCells-HindIII-rep2	Cortex-HindIII-rep1	0,79	0,79
ESCells-HindIII-rep2	Cortex-HindIII-rep2	0,79	0,8
ESCells-Ncol	Cortex-HindIII-rep1	0,7	0,76
ESCells-Ncol	Cortex-HindIII-rep2	0,71	0,78
Cortex-HindIII-rep1	Cortex-HindIII-rep2	0,86	0,83

**Supplementary Table 3**

**Correlation coefficients for the mouse HoxD locus between Hi-C datasets corresponding to different experimental replicates and tissues.**

	<b>Viewpoint</b>	<b>Read 5'-&gt;3'</b>	<b>Non-read 5'-&gt;3'</b>
	European amphioxus 4C	Hox2	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGGCCCCCTGTCCCCAGATC
Hox5		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTCGCCAACACTCTGCTATACATGATC	CAAGCAGAAGACGGCATACGAGCGCACGAGATAGTGGGTAC
Hox6		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTATCTCACGTACAGAAAGCTGATC	CAAGCAGAAGACGGCATACGACATCTCGTTGAAATTGCGGAC
Hox7		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGTCCGGATGATACATGGTGACGATC	CAAGCAGAAGACGGCATACGAAGGAGACCCCTTGCCACCACC
Hox9		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTTCATCTCAGGAAAACACGTGATC	CAAGCAGAAGACGGCATACGAATTACCTACATGCCACGGG
Hox11		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTAGGCCTTTTTTTTCAACACAAGGATC	CAAGCAGAAGACGGCATACGAGCGAAAGAAAATCTACGCG
Hox13		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTCAAAAAGCGAACGGCCAGATC	CAAGCAGAAGACGGCATACGAGGATATCACTCCCAGTTCTTGG
Hox15		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTCCGTCTCGGCCCGGGCGATC	CAAGCAGAAGACGGCATACGAATGGCGCAGGAGTGCCTCCG
Evx2		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGCGGTGCACAAGCAAAGATC	CAAGCAGAAGACGGCATACGATTACTGCCTTACCACAGCAC
Mtx2		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTACTTGCCGACGCAGCGATC	CAAGCAGAAGACGGCATACGAGGTCAGGGTTACTGGACGAG
Lnpa		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGTGTTGAACTGTATATCCATGATC	CAAGCAGAAGACGGCATACGATGGACTTACTTTGAAAGCGTG
Gpatch8		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTCTGTGTATGAAAGTCTCTGATC	CAAGCAGAAGACGGCATACGACTTTAACAGAACCTCAGCAC
Meox		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTAACGCCCTTGTGCAAGTGATC	CAAGCAGAAGACGGCATACGAATGCTACGAGCTTACTAACAGGAC
Hnrnpa		AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTTCCCATCCTTTGTAGGACGATC	CAAGCAGAAGACGGCATACGAGGATACTCCGAGTAGTTCTGC
Nfe2l	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTCCATCCCTGTGGGGATC	CAAGCAGAAGACGGCATACGATTGTGCAGTTCCTGAGAAATCTC	
Zebrafish 4C	<b>Viewpoint</b>	<b>Read 5'-&gt;3'</b>	<b>Non-read 5'-&gt;3'</b>
	Hnrnpa3	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTATTAGGCATGGTTGTATACGATC	CAAGCAGAAGACGGCATACGAGAAAAGTACATCACATCTGCT
	Nfe2l2a	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTAAGCAAATGGATGACTGAGGATC	CAAGCAGAAGACGGCATACGAACCTGACTTTTTGCAACCCAC
	Atp5g3a	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGCACAATAACAGTGTATTTGATC	CAAGCAGAAGACGGCATACGATTTTCATTACAAGAGGCCGC
	Evx2	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTGGTTCCCAAATAAGCTGTCGATC	CAAGCAGAAGACGGCATACGACCTAATGTCTGAGTGCTAGCTG
	Lnpa	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTCCTTCACTGCTGTGTCACGTGATC	CAAGCAGAAGACGGCATACGACCCTGTATCAGCTGACAAGTTGC
HoxD4a	AATGATACGGCGACCACCGAACACTCTTCCCTACACGACGCTCTTCCGATCTTTGTTTCATCATCATATGGGAGGATC	CAAGCAGAAGACGGCATACGACAGTTGACAGCAAGTAGGAGG	
Enhancer cloning	<b>Enhancer</b>	<b>Fwd 5'-&gt;3'</b>	<b>Rev 5'-&gt;3'</b>
	1655	CACTTCCAGCCCACTTCCTC	CGGGTGCCAGGTTTATTCTGA
	1739	GCTGAGATTTCCAACAACCACA	GGGACACGGAGGTTGATAAGT
	1784	CCACCCGGAAATCTTTGTCC	TATGCGCTCTGAGATGACGG
	1801	TTCCGCATGCCTTACACACA	CCCCTGATATAAAGCCCACT
2473	AGTACCCGTTAGATTCCCT	ACCATACGCTGCTTATCCATGA	

**Supplementary Table 4**  
**Primers used in the 4C-seq and enhancer cloning experiments.**

## Supplementary Note

Vertebrate Hox gene deserts were ancestrally populated by a conserved array of neighboring genes.

In most jawed vertebrate species, the genomic arrangements of HoxA and HoxD neighborhoods show little similarity: many HoxA long-range cis-regulatory elements (CREs) are embedded in introns of neighboring genes, while HoxD long-range CREs are located in gene deserts<sup>2,8</sup>. Therefore, the long-range regulatory interactions of HoxA and HoxD could have evolved independently<sup>9,10</sup>. Here, we show that these differences are a derived situation and that the genomic organisations of the four Hox clusters neighboring regions were ancestrally very similar. First, out of the 15 gene families consistently found in the vicinity of the four clusters, 6 of them (*Nfe2*, *Hnrnpa*, *Cbx1-3-5*, *Calcoco*, *Atp5g* and *Creb*) have 3 to 4 members in the Hox regions of at least one gnathostome species, confirming that their close Hox linkages predate the WGDs (Fig. 1b). Second, ‘desertification’ process (erosion of flanking genes’ exons<sup>13</sup> while non-coding introns containing CREs are maintained) has probably been rampant thanks to the functionally redundant genes copies generated by the two vertebrate WGD duplications. The genomes of slow evolving species such as elephant shark, still retain additional paralogs of Hox-neighboring genes that have been secondarily lost in mammals and teleosts (such as *Eve1* linked to HoxB, *Lnp2* linked to HoxC, and *Jazf2* linked to HoxD); in turn, the HoxD-linked *Atp5g3* is present in mammals but has been lost in elephant shark (Fig. 1b,<sup>54</sup>). The presence of Hox distal CREs within the introns of some of these lost paralogs further demonstrate that the coding regions of Hox neighboring genes were selectively lost, generating gene deserts in the process. In elephant shark, two of the four conserved distal regulatory islands (I and II) of mouse HoxD posterior desert are located inside the introns of the additional *Jazf2* copy, in an arrangement almost identical to that typically found in HoxA clusters (Supplementary Figure 2). In fact, a pseudogenic relict of a *Jazf2* exon is still detectable in mouse (Supplementary Figure 2). Furthermore, part of the HoxD global control region, the CsB element, has a paralogous copy in HoxA posterior

side inside the intron 4 of the gene *Hibadh*, implying that a paralog of this gene was also present in the HoxD posterior neighborhood (Supplementary Figure 1, <sup>12</sup>). Thus, HoxD 'gene desert' was originally occupied by a *Hibadh2* and a *Jazf2* genes, showing that HoxA and D genomic organisations are in fact almost identical. In sum, these results show that the syntenic organisation of the four clusters was originally very similar and the close linkage of most Hox neighboring genes predate the WGDs.

#### Reconstruction of the syntenic arrangement around the Hox cluster of the pre-WGD vertebrate ancestor.

Comparisons between the four paralogous clusters in different vertebrates species allow us to infer that several Hox-neighboring genes were already present around the single cluster of the pre-WGD vertebrate ancestor. We recovered 6 genes that have at least 3 paralogous copies (*Nfe2*, *Hnrnpa*, *Cbx1-3-5*, *Calcoco*, *Atp5g* and *Creb*). Five other genes only have one or two copies in vertebrates (*Copz*, *Skap*, *Smug1*, *Mtx2* and *Lnp*) but their locations and orientations in amphioxus are almost identical to those of vertebrates, and therefore their syntenic arrangement was already present in the chordate ancestor (Fig. 1b). In the case of three other jawed vertebrate genes with less than 3 copies, *Hibadh*, *Jafz* and *Ube2z*, a conserved tight linkage with their closest vertebrate neighbor is also present in at least one additional species outside vertebrates, indicating that they belong to ancestral microsyntenic pairs that predate the origin of vertebrates (Supplementary Figure 3). Finally, for the two remaining jawed vertebrate neighboring genes, *Snx10-11* and *Ttll6*, we have not found any data that could help to infer their status in the pre-WGD vertebrate ancestor.

#### Most chordate Hox neighboring genes belong to the same ancestral Hox macrosyntenic linkage group.

The majority of chordate Hox neighboring genes, including the conserved anterior genes and the vertebrate and amphioxus specific posterior ones (*Agps*,

*Copz, Nfe2, Skap, Smug1, Mtx2, Evx, Lnp, Hibadh, Calcoco, Jafz, Atp5g, Ube2z, Creb, Nifk, Mdh1b, Dlgap1-4, Ubp1-Tfcp2, Asb1* and *Gpatch8*) are tightly linked to at least another chordate neighboring gene in at least one non-chordate species (Supplementary Figure 3). This indicates that they all belong to the same ancestral large macrosyntenic linkage group that also contained the Hox cluster (or the putative Hox 'ghost' locus, in the case of placozoans <sup>55</sup>) as well as additional related ANTP homeobox genes (such as *Evx, Mnx, Dlx* and *Meox*), and some other genes showing conserved Hox macrosyntenic associations in different non vertebrate species (*Elac1, Mcm6, Tmem169, Nhej1, Smarcc, Myd98*, Supplementary Figure 3). In general, these inferred ancestral linkages between different neighboring genes and the Hox cluster and between neighboring genes with other neighboring genes show an extremely patchy phylogenetic distribution: most of these syntenic associations have been heavily reshuffled in different animal lineages and gene order and orientations are highly variable. Thus, although all these genes were probably located in the same Hox chromosome in the bilaterian ancestor, in most phyla their linkage has only been maintained at the macrosyntenic level.

### **Supplementary References**

54. Ravi, V. *et al.* Elephant shark (*Callorhynchus milii*) provides insights into the evolution of Hox gene clusters in gnathostomes. *Proc Natl Acad Sci U S A* **106**, 16327-32 (2009).
55. Mendivil Ramos, O., Barker, D. & Ferrier, D.E. Ghost loci imply Hox and ParaHox existence in the last common ancestor of animals. *Curr Biol* **22**, 1951-6 (2012).