1

2

New developments and biological insights into the farming of Solea senegalensis 3 reinforcing its aquaculture potential 4 5 Sofia Morais¹, Cláudia Aragão², Elsa Cabrita², Luís E.C. Conceição³, Maria Constenla⁴. 6 Benjamín Costas⁵, Jorge Dias³, Neil Duncan¹, Sofia Engrola², Alicia Estevez¹, Enric 7 Gisbert¹, Evaristo Mañanós⁶, Luísa M.P. Valente^{5,7}, Manuel Yúfera⁸, Maria Teresa Dinis² 8 9 ¹IRTA, Ctra. Poble Nou Km 5.5, 43540 Sant Carles de la Rápita, Spain 10 ²Centre of Marine Sciences, CCMAR, University of Algarve, Campus de Gambelas, edf. 7, 8005-11 12 139 Faro, Portugal ³ Sparos Lda, Área Empresarial de Marim, Lote C, 8700-221 Olhão, Portugal 13 ⁴ Departament de Biologia Animal, de Biologia Vegetal i d'Ecologia and Servei de Diagnòstic 14 15 Patològic en Peixos, Universitat Autònoma de Barcelona, 08193 Barcelona, Spain ⁵CIIMAR/CIMAR LA–Interdisciplinary Centre of Marine and Environmental Research, University 16 17 of Porto, Rua dos Bragas 289, 4050-123 Porto, Portugal ⁶ Instituto de Acuicultura de Torre la Sal (IATS-CSIC), Castellón, Spain 18 ⁷ ICBAS – Institute of Biomedical Sciences Abel Salazar, University of Porto, Rua de Jorge Viterbo 19 20 Ferreira 228, 4050-313 Porto, Portugal ⁸ Instituto de Ciencias Marinas de Andalucía (ICMAN-CSIC), Campus Universitario Río San Pedro 21 22 s/n, 11510 Puerto Real, Cádiz, Spain 23 24 Correspondence 25

27 Email: sofia.morais@irta.cat, Tel: +34 977745427 Ext. 1814, Fax: +34 977744138

28 Abstract

Senegalese sole was one of the earliest identified candidate species with high potential for 29 30 aquaculture diversification in the South of Europe. Its culture has been possible, and 31 commercially attempted, for several decades but intensive production has been slow to take off. This has been explained mostly by serious disease problems, high mortality at weaning, 32 variable growth and poor juvenile quality. However, a strong and sustained research 33 investment that started in the eighties has led to a better understanding of the requirements 34 and particularities of this species. More recently, better management and technical 35 improvements have been introduced, which have led to important progress in productivity 36 and given a new impetus to the cultivation of Senegalese sole. As a result, the last 5 years 37 have marked a probable turning point in the culture of sole towards the development of a 38 knowledge-driven, competitive and sustainable industry. This review will focus on the 39 main technical improvements and advances in the state of knowledge that have been made 40 in the last decade in areas as diverse as reproductive biology, behaviour, physiology, 41 nutritional requirements, modulation of the immune system in response to environmental 42 43 parameters and stress, and characterization and mitigation of the main disease threats. It is now clear that Senegalese sole has important particularities that differentiate it from other 44 current and candidate marine aquaculture species, which bring about important challenges, 45 46 some still unsolved, but also notable opportunities (e.g., a nutritional physiology that is better adapted to dietary vegetable ingredients), as will be discussed here. 47

48

49 Key words: aquaculture industry, diversification, flatfish, Soleidae.

51

52 Introduction

Two Soleidae, Senegalese sole (Solea senegalensis Kaup, 1858) and common sole 53 54 (Solea solea, Linnaeus, 1758) have been identified as being among the most interesting and 55 promising species for marine commercial aquaculture diversification in Europe for at least four decades now. As a result, sole has attracted a lot of research interest and considerable 56 investments have been made to develop rearing methodologies during the last 30 years 57 (Howell 1997; Dinis et al. 1999). Nevertheless, production of common sole in Central and 58 59 Northern Europe (mostly in the Netherlands and in the United Kingdom) and of Senegalese sole in Southern Europe has failed to reach successful commercial development until 60 recently (FAO 2011). This can be mostly explained by a high incidence of diseases, 61 62 particularly Pasteurellosis and Flexibacteriosis, which have plagued quite a few farms. In addition to high vulnerability to diseases, other problems that were reported initially were 63 difficulties in weaning onto formulated diets leading to low growth and high juvenile 64 mortalities, a decreased growth rate at high stocking densities, pigmentation abnormalities 65 and malformations related to eye migration (Dinis et al. 1999). Consequently, poor fry 66 quality and high mortality rates during the weaning stage have led to juvenile scarcity for 67 on-growing purposes. On the upside, several advantages have been identified: natural 68 spawning of wild-caught broodstock readily occurs in captivity in close relation with the 69 70 temperature cycle; larval growth is fast and larval rearing is easily accomplished at high 71 survival rates; and the market value and demand for these species are high and not completely met by fishery catches (Howell 1997; Dinis et al. 1999). Most of these aspects 72 73 have been comprehensively reviewed a decade ago by Imsland et al. (2003). However, in

74 recent years there have been important technical developments in rearing and feed 75 technologies and advances in the state of knowledge of the biology and nutritional requirements of the species in captivity, which have given a new impetus to the culture of 76 these flatfish species. This is particularly the case for Senegalese sole which in the last 5 77 years has shown important signs of expansion and has attracted substantial investment in 78 79 Southern Europe to intensify its production (Howell et al. 2011; APROMAR 2013), in spite of the actual climate of financial restraint. Here we will review the current state of 80 biological knowledge and technical improvements achieved through research conducted 81 during the last decade, pinpointing where scientific advances have been made on early 82 83 bottlenecks to help build a stronger and more competitive sole farming industry. The emphasis on S. senegalensis in the Iberian Peninsula is justified by what has been an 84 unbalanced research and commercial investment in Solea spp. across Europe in recent years 85 but, given the close relationship between both species, some of the basic knowledge 86 presented here could also be relevant to common sole cultivation in Central and Northern 87 Europe. 88

89

90 Senegalese sole market and culture

Senegalese sole is a flatfish of high commercial value and demand not only in the Iberian Peninsula but also in the wider European market, indistinguishable by consumers from common sole, considered as the same species in marketing statistics, and often sold together under the name "sole". These two species are commonly fished along the Eastern Atlantic coast, and common sole also along the Mediterranean basin, in gillnets or trawling nets, but most catches are from the North sea (Bjørndal & Guillen 2014). The biological and culture requirements of these two species are somewhat different (Dinis 1986; Imsland *et al.* 2003) and Southern Europe countries have selected Senegalese sole for farming
mainly due to the species higher growth rates (Howell *et al.* 2009), and also higher
temperature requirements (Howell 1997). It is reared commercially in Spain and Portugal,
where existing facilities were adapted for sole culture (Imsland *et al.* 2003).

102

103 Market

Fisheries landings of sole showed a 43% reduction from 1995 to 2012 and the average 104 size of wild sole has also been declining, while consumers prefer larger fish, with market 105 106 prices proportional to size (Bjørndal & Guillen 2014). On the other hand, average prices of different sole products (fresh, frozen and farmed) in commercialization statistics from one 107 of the most important fish markets in Spain, the MercaMadrid, have been quite stable from 108 2002-2013, with a tendency for fresh sole prices to increase since 2010 (Bjørndal & Guillen 109 110 2014). Furthermore, sole consistently attains high prices in European markets. These 111 factors, associated with Southern European markets being saturated for the main 112 aquaculture species, seabream (Sparus aurata) and seabass (Dicentrarchus labrax), have 113 fuelled interest in sole for marine fish farming diversification.

Global flatfish aquaculture production has increased from 26,300 tonnes in 2000 to 114 115 148,800 tonnes in 2008, the world leading producer being China, which mainly produce 116 turbot (Scophthalmus maximus), Japanese flounder (Paralichthys olivaceus) and tongue sole (*Cynoglossus semilaevis*), while Spain is the main producer in Europe, particularly of 117 118 turbot (FAO 2011). Soleidae presently represent only a small fraction of global flatfish production and, according to EUMOFA, among the 7,752 tonnes of flatfish produced in 119 2011 in the EU, 157 tonnes were from sole. Production of Senegalese sole in the South of 120 121 Europe, although still low due to the limited number of farms, has increased exponentially

from only 60 or 11 tonnes produced in 2005 in Spain and Portugal to 194 or 100 tonnes in 122 123 2012, respectively. France has also started producing sole in 2010 (142 tonnes), almost duplicating production (220 tonnes) by 2012 (Fig. 1; APROMAR 2014, FEAP 2014). For 124 2013, the Spanish Marine Aquaculture Producers Association reported another important 125 126 increase in sole production (343 tonnes, 300 of which produced in Galicia, and 3.9 million juveniles) and, with several new farms being currently under construction, further 127 important expansion is anticipated in the coming years (APROMAR 2014). Spain is the 128 largest market for sole in Europe, with a limited domestic supply meaning that most of the 129 consumed sole is imported. Other important markets in terms of total supply (domestic 130 production plus imports) are France, Italy and the United Kingdom (Bjørndal & Guillen 131 132 2014). In addition, the sole production market is being expanded to Asia; in 2013 the only sole hatchery operating in Portugal and producing eggs year-round sold juveniles (7g 133 weight) to Asia, and increased sales are expected in 2014. 134

Current production costs for this species have been estimated at 9.62 €kg, with just over 135 136 39% being accounted for by juvenile costs (3.77 \notin kg) (Bjørndal & Guillen 2014). These costs are however expected to decrease considerably (possibly down to 7 \notin kg) as the 137 industry expands, due to economy of scale, better domestication and lower feed conversion 138 139 ratios, as commercial cultivation experience builds up and it becomes advantageous for feed companies to produce specialised sole diets. Price of farmed sole in wholesale markets 140 has only been available for two years. The farmed product attained a price of 12.25 €kg in 141 142 MercaMadrid in 2013, which is only slightly higher than wild small (under 500g) sole given that farmed sole have been commercialized at a similar small size, while larger fish 143 are preferred by consumers and sold at substantially higher prices (Bjørndal & Guillen 144 145 2014). Therefore, although there are still many challenges to address, a recent thorough economical analysis by Bjørndal & Guillen (2014) concludes that market prospects forexpansion of sole farming are good.

148

149 Production systems

150 Until recent years, Senegalese sole aquaculture was linked to the salt marshes ("esteros") 151 in the South of Spain (Cádiz and Huelva, Andalusia) and South of Portugal (Algarve) as an added value product in polyculture with semi-intensive seabream and seabass cultivation 152 (Ferreira et al. 2010; Yúfera & Arias 2010). Wild sole juveniles were passively trapped in 153 the ponds or these were stocked with cultured juveniles bought from hatcheries. Sole 154 juveniles would rely on the occurrence of natural prey in the bottom of the ponds with little 155 additional investment by the producers or, in some cases, the natural prey was 156 supplemented to an unknown extent by commercial feed left-over by seabream and seabass. 157 158 In this culture system not only sole juveniles were present, but also other naturally occurring species (e.g., eel and mullets) could also be found. Survival of soles at the end of 159 160 the on-growing process was very variable, depending on the pond sediment condition and 161 seabream/seabass density (Quental-Ferreira et al. 2010). In these conditions, Senegalese sole can grow from 24 to 393g in 16 months and with 99% survival (Quental-Ferreira et al. 162 163 2010).

Nowadays, although salt marshes are still used for semi-intensive aquaculture, the trend is for more intensive on-growing systems using fibreglass or concrete tanks, or shallow raceways (for a more detailed description of the different culture systems see Imsland *et al.* 2013), using commercial feeds and maintaining highly controlled environments. More recently, recirculation aquaculture systems (RAS) have been implemented and now most of the new sole farms in Spain and Portugal use this type of system for a better control of environmental conditions, especially water temperature and nitrogen wastes. This has been an important technological development resulting in increases in production as sole are quite sensitive to environmental changes and the stable environment provided by recirculation technologies, particularly the tight temperature control, can give increased growth and survival.

175

176 Culture conditions and growth

177 The current general parameters that are being used or should be considered for sole178 culture and related existing data on growth performance are as follows:

Temperature: Typical rearing temperature of Senegalese sole is either following natural thermoperiod or maintaining constant around 20°C. Although higher growth occurs at temperatures ranging from 20 to 25°C, temperatures above 22°C entail higher risk of pathological outbreaks (Cañavate 2005).

Salinity: Sole juveniles are partially euryhaline and can tolerate salinities from 5 to 55 (Arjona *et al.* 2007). However, growth was shown to be depressed at a salinity of 15 compared to 25 and 39, with some differences being also noted in terms of ingestion, metabolite levels, energy metabolism and cortisol response at salinities between 25 and 39, with slightly better results at 39 (Arjona *et al.* 2009).

Photoperiod: Sole has a nocturnal activity pattern, with locomotor activity peaking in the first part of the dark period (Bayarri *et al.* 2004) and a higher metabolic rate during the dark phase (Castanheira *et al.* 2011). However, broodstock appeared to habituate to husbandry routines and had the peak of activity during the day from 14:00-18:00, which coincided with feeding at 15:00 (Carazo *et al.* 2013). Aquaculture facilities for indoor ongrowing use mostly a 12hL:12hD photoperiod and/or some shading in the tanks to keep light at the surface between 80 and 350 lux (Salas-Leiton *et al.* 2008; Boluda Navarro *et al.*2009).

Feeding: Senegalese sole displays a nocturnal feeding pattern with 77-85% of feed
demand, when using self-feeders, occurring at night (Boluda Navarro *et al.* 2009).
However, in most farms food is offered in 4-8 meals during the day, or more frequently
over the 24h using automatic feeders. Contrary to other flatfish species like turbot,
Senegalese sole has a strict bottom-feeding behaviour.

Stocking density: Densities of up to 30 kg m⁻² have been tested with no effects on growth (Salas-Leiton *et al.* 2008) although high densities (26.6 kg m⁻²) might affect the shape of the fish (Ambrosio *et al.* 2008). A relationship has been found between high stocking densities and stress in the fish (Costas *et al.* 2008; Salas-Leiton *et al.* 2010), but it is unclear if this is due to density *per se*, or rather to deteriorating water quality.

206 There is little information regarding the growth potential of Senegalese sole in 207 commercial culture given that the growth data available is taken from rearing experiments designed to test nutritional requirements or on-growing conditions (a summary is presented 208 209 in Table 1, taken from Sánchez-Fernández 2012). On the other hand, a major issue is that Senegalese sole typically shows large size variation under culture, increasing with time and 210 leading to high differences between fast and slow growers. Although Senegalese sole does 211 212 not show any aggressive behaviour of large fish over smaller fish, a hierarchical structure exists. Salas Leiton et al. (2011), in a study of the effect of grading on growth, found that 213 214 all the populations of fish (graded or not) tended to reach the same size variation after 60 215 days of culture, and suggested that group heterogeneity improves overall growth, associated with efficient social arrangements. Therefore, it was concluded that the hierarchical 216 217 structure of the population is caused by fish size distribution and not the contrary.

218

219 **Broodstock management**

220

221 Sexual differentiation

222 Senegalese sole females grow faster and mature later than males (Dinis et al. 1999; Imsland *et al.* 2003) and thus female-based stocks would be an advantage for the culture of 223 this species (Viñas et al. 2013). A recent study has demonstrated the influence of 224 225 temperature on sex differentiation in sole, suggesting the potential applicability of 226 environmental sex manipulation to sole aquaculture (Blanco-Vives *et al.* 2011). This work showed that sole larvae grown under three different thermoregimes, a thermophase-227 228 cryophase (TC) thermoregime (daily temperature fluctuation from 21°C during the day to 18°C during the night), an inversed thermoregime (CT: 18°C day to 21°C night) and a 229 230 constant 19.5°C thermoregime, exhibited highly different sex ratios, with the proportion of 231 females being 70% for the TC, 20% for the CT and 40% (approximately the normally 232 observed sex ratio in sole culture) in the constant thermoregime group.

233

234 Spawning of wild sole

Industrial Senegalese sole culture has been so far based on the spawning of wild-origin broodstocks (Dinis *et al.* 1999; Anguis & Cañavate 2005; Martin *et al.* 2014). Wild adult sole have been caught in coastal waters, often coastal lagoons on the southern Atlantic coast of Spain and Portugal, and transported to aquaculture or research facilities. After a period of quarantine and acclimation to captivity, groups of wild broodstocks can spawn naturally within the same year (Dinis *et al.* 1999; Imsland *et al.* 2003). Optimal holding conditions for successful spawning of wild breeders have been described (Table 2). The

principal spawning period is during spring (February-March to June) with a less important 242 243 spawning period in autumn (Howell et al. 2003; Anguis & Cañavate 2005). As in other species (Bromage et al. 2001), both photoperiod and temperature have been implicated in 244 the control of the timing of spawning in Senegalese sole. However, to date there has been 245 246 no published information on the use of artificial photoperiod regimes for the control and manipulation of spawning in Senegalese sole. Natural photoperiod has been most 247 commonly recommended and used for successful spawning (Dinis et al. 1999; Howell et al. 248 249 2003; Anguis & Cañavate 2005) given that constant photoperiod was suggested to disrupt 250 gonadal maturation (García-Lopez et al. 2006a; 2009). Nevertheless, the full role of photoperiod in Senegalese sole maturation has not been investigated and successful 251 252 spawning has also been obtained under a constant photoperiod of 16hL:8hD (Martin et al. 2014). Perhaps more importantly, spawning was observed to be closely related to the 253 254 temperature cycle (Dinis et al. 1999; Anguis & Cañavate 2005) and highest fecundities are 255 between 16 and 21°C (Anguis & Cañavate 2005). Through temperature control (oscillating 256 temperatures between 16 and 18°C) it has been possible to extend the natural spawning season with spawning every month from March to November, although with variable egg 257 production (Martin et al. 2014; Table 2). Microsatellite analysis of parentage has 258 demonstrated that only 8.7% to 51.7% of breeders actually participate in spawning (Porta et 259 260 al. 2006; Martin et al. 2014), which leads to a sub-estimation of relative fecundity (eggs per kg of female biomass) of a broodstock. Individual females were observed to spawn with 261 intervals from 1 to 56 days and fecundities were often over 100,000 eggs kg⁻¹.day⁻¹ and on 262 occasions such high fecundities from a single female were observed on consecutive days 263 (Martin et al. 2014). These observations highlight the potential gains in egg production that 264

265 could be made by management improvements leading to more breeders in a Senegalese266 sole broodstock participating in spawning.

267

268 Spawning of G1 sole

269 A major constraint to the culture of Senegalese sole is that successful spontaneous spawning has not been obtained from generations (G1, G2...) of broodstock bred and reared 270 in captivity. Therefore, the industry at present is threatened by its dependence on the 271 272 capture of wild broodstock, which are in decline due to factors such as fishing pressure. 273 Spawning from G1 broodstock has been infrequent and spawns were almost entirely of eggs that did not hatch (Agulleiro et al. 2007; Guzmán et al. 2008; Howell et al. 2009; 274 Rasines et al. 2012; Norambuena et al. 2012a), and Carazo (2013) later observed that the 275 276 eggs were not fertilized. Many industry and research centres have held G1 broodstocks 277 under the same conditions as successfully spawning wild caught broodstock, both before 278 and during the spawning period, with the same negative result. This has lead to the 279 hypothesis that some aspect (developmental, nutritional, genetic or social / behavioural) in 280 the early rearing (larval or juvenile) might be responsible for the failure of cultured broodstock to spawn viable eggs (Howell et al. 2009, 2011). A significant research effort 281 detailed below has been directed to identify the problem and to develop solutions to 282 283 preferably obtain spontaneous spawning in cultured breeders as in the wild broodstock, but also to develop strip spawning protocols that offer an immediate solution to the problem. 284

285

286 Reproductive behaviour

The reproductive behaviour of Senegalese sole has been studied using non-disrupting
lighting and digital video recording (Carazo *et al.* 2013). Wild Senegalese sole in captivity

exhibited a complex reproductive behaviour that was characterised by an increase in 289 290 activity before paired synchronised spawning at the water surface (Carazo et al. 2011; Carazo 2013), which was similar to other species of flatfish, including common sole 291 292 (Baynes et al., 1994). Conversely, Carazo (2013) identified that the eggs released from G1 293 broodstock were not fertilised by G1 males due to the absence of courtship behaviour. The observation of pre-spawning behaviour of wild breeders, that involved chasing (particularly 294 between males) and fish resting on each other (particularly males on a female), combined 295 296 with microsatellite paternity studies, indicated that spawning was dominated by a few 297 individuals that exhibited fidelity to one another (Porta et al. 2006; Carazo 2013; Martin et al. 2014). Senegalese sole G1 breeders did not exhibit any aspect of the reproductive 298 behaviour observed in wild breeders, either pre-spawning chasing or paired synchronised 299 spawning (Carazo et al. 2011; Carazo 2013). Breeders from different origins were 300 301 combined to establish that G1 and G2 females held with wild males produced viable 302 spawns with typical reproductive behaviour while G1 males held with wild females did not 303 produce viable spawns (Mañanós et al. 2007) and no courtship was observed (Carazo et al. 304 2011; Carazo 2013). These studies have unequivocally established that the reproductive problem is centred on G1 males that lack the reproductive behaviour necessary for 305 successful spawning. 306

307

308 Broodstock nutrition

Achieving good spawning and egg / larval quality results with a specifically formulated broodstock diet is an important step for the development of the commercial aquaculture of sole, which requires consistent and sanitarily safe diets. The standard broodstock feeding regime for this species in captivity consists on natural food items, mostly polychaetes and

frozen squid, mussels, and small shrimp, combined with broodstock diets formulated for 313 314 other marine fish species. This leads to reasonably good spawning results, both in terms of quantity and viability of the eggs produced (Dinis et al. 1999; Imsland et al. 2003). 315 However, the use of natural feeds poses important sanitary risks as these can be a source of 316 317 pathogens or parasites, and in fact mortality outbreaks in common sole have been attributed to the live prey that was fed (Baynes & Howell 1993). Moreover, these wet feeds have 318 variable supply and nutritional composition and contribute to the deterioration of water 319 320 quality, and hence the goal is to completely replace them by formulated feeds. Recently, 321 two new semi-moist diets specifically formulated for S. senegalensis broodstock have been tested during a period of two years, with very little polychaete supplementation (0.3% of 322 323 weight once or twice a week). The most basic diet, formulated with high quality marine ingredients and containing 58.6% crude protein and 16.8% crude fat produced good results, 324 325 comparable to the classical diet, in terms of egg quality and early larval performance, while 326 egg fatty acid composition was improved when compared with previous spawnings from 327 broodstock kept in the same facilities but fed the standard fresh feed diet (Morais et al. 328 2014a). Further supplementation of the broodstock diet with lipids (up to 23.5%), vitamins and long chain polyunsaturated fatty acids (LC-PUFA), mainly docosahexaenoic acid 329 (22:6n-3, DHA) and eicosapentaenoic acid (20:5n-3, EPA), did not improve egg quality or 330 331 larval performance up to 17 days post hatching (dph). Other studies have found that levels of essential fatty acids (EFA) affect gamete and larval quality (Fernandez-Palacios et al. 332 2011; Duncan et al. 2013) and a study on Senegalese sole wild males found that a 333 formulated diet higher in DHA plus antioxidants significantly increased sperm quality, 334 increasing both percentage of progressive spermatozoa and sperm velocity (Beirão et al. 335 336 2015).

Nutrition has also been implicated as a factor that contributes to the reproductive 337 338 behavioural dysfunction in G1 broodstock. Norambuena et al. (2012a) found significant differences in proximate, lipid and fatty acid composition between wild and cultured 339 breeders. The body composition of cultured breeders reflected the diet in captivity, with 340 341 significantly lower levels of the EFA arachidonic acid (20:4n-6, ARA) in liver, testes and muscle, of DHA in liver and male muscle, of EPA in male muscle, and of cholesterol in 342 liver. However, cultured breeders that were fed ARA at 2.3-3.2% of total fatty acids (TFA) 343 (Norambuena et al. 2013a) or that demanded ARA at 3% TFA (Norambuena et al. 2012b) 344 had similar ARA levels in the liver and gonads to those found in wild fish. These 345 differences in nutritional state appeared to affect pathways for the production of 346 prostaglandins (PGs) and possibly steroids given that cultured breeders presented 347 significantly lower levels of total 2-series PGs and higher levels of 3-series PGs compared 348 to wild breeders (Norambuena et al. 2012c). On the other hand, cultured breeders fed 349 increasing contents of ARA (0.7-6.0% TFA) exhibited significantly increasing plasma 350 levels of testosterone (T) and 11-ketotestorone (11KT) and significantly decreasing plasma 351 352 levels of 3-series PGs, while levels of 2-series PGs were similar independently of dietary ARA composition (Norambuena *et al.* 2013b). When ARA content was $\geq 3.2\%$ TFA no 353 differences were observed between levels of 2-series and 3-series PGs. The 2-series PGs 354 have been described to be more active or potent than the 3-series PGs (Tocher 2003) in 355 stimulating steroid production (Wade & Kraak 1993) and acting as pheromones in the 356 control of reproductive behaviour in cyprinids (Sorensen & Stacey 2004). Therefore, 357 nutritional manipulation of ARA levels was shown to stimulate the endocrine system in 358 ways that could affect reproductive success and behaviour and hence special attention 359 360 should be given to the levels of this fatty acid in the formulation of broodstock diets.

Although to date no study has been performed to demonstrate whether feeding G1 fish diets with the EFA levels (and ratios) expected in the diet of wild fish can restore the complete reproductive failure in cultured Senegalese sole, it is believed that the root of the problem may not be simply nutritional.

365

366 Reproductive endocrinology

Further basic knowledge on the physiology and endocrinology of sole reproduction has 367 been obtained in the last years, particularly with the characterization of pituitary 368 gonadotropins (GTHs) and the study of their expression in sole males and females (Cerdà et 369 al. 2008a; Guzmán et al., 2009a). In addition, the recent development of recombinant 370 371 gonadotropins will enable further examination of the endocrine system in the near future (Chauvigné et al. 2012). On the other hand, a good deal of information became available on 372 373 the major sex steroids and their relation with gonad development and spawning. The 374 releasing profiles of sex steroid hormones has been described in sole broodstocks of wild (García-López et al. 2006b; Guzmán et al. 2009a) and cultured origin, including G1 375 376 (García-López et al. 2006c, 2007; Guzmán et al. 2008, 2009a,b; Oliveira et al. 2010) and G2 (Bayarri et al. 2011) generations. Plasma levels of the main androgens (11KT and T) 377 and estrogens (estradiol, E2), in males and females, respectively, were highly correlated 378 379 with gonad development and spawning time and are useful non-lethal markers to estimate the stage of gonad maturation of the broodstock. In males, T and 11KT plasma levels were 380 381 low during the resting period (summer), increased during gametogenesis (winter) and 382 peaked just before the spawning time (early spring) (García-López et al. 2006b,c; Bayarri et al. 2011). A very similar annual profile was observed for E2 in females (García-López et 383 384 al. 2006b, 2007; Guzmán et al. 2008; Bayarri et al. 2011) and for the maturation inducing steroid (MIS) 17,20β-dihydroxy-4-pregnen-3-one (17,20β-P) in both sexes (García-López *et al.* 2006b,c; Bayarri *et al.* 2011), which presented a peak that was slightly delayed with
respect to those of T, 11KT and E2. These results were similar to those obtained in other
fish species and are in agreement with the known role of T, 11KT and E2 in the regulation
of gametogenesis and that of MIS in controlling final gonad maturation and probably
reproductive behaviour (Mañanós *et al.* 2008).

Studies have found that the holding environment can disrupt the endocrine reproductive 391 profiles. Constant temperatures and photoperiod negatively affected steroid profiles and in 392 particular constant temperature significantly reduced steroid levels (García-López et al. 393 2006a) and in females inhibited the steroid rhythm observed under a natural fluctuating 394 temperature regime (Oliveira et al. 2009). On the other hand, wild males maintained in a 395 long-term isolation from females presented lower androgen levels at the peak of the 396 reproductive period, suggesting the absence of male-female communication had disrupting 397 effects on the endocrine system (Cabrita et al. 2011). In addition, evidence of relevant 398 female-to-male communication in sole reproduction was obtained after treating the females 399 400 with MIS and registering a significant increase in sperm viability, velocity and motility in surrounding males (Cabrita et al. 2011). 401

Two aspects should be highlighted from these studies. Under the optimal reproductive conditions for Senegalese sole (Table 2), the synthesis and release of all relevant reproductive hormones, including steroids, vitellogenin (Guzmán *et al.* 2008), gonadotropins and neuropeptides (Guzmán *et al.* 2009a), seems to be correct and well correlated with gonad development, with similar profiles in both cultured and wild soles. Therefore, no evident endocrine failure has been detected in cultured breeders with respect to wild soles. Nevertheless, in spite of this similarity, plasma levels of reproductive 409 hormones seem to be consistently lower in cultured G1 (and G2) stocks compared to wild 410 soles. These observations could be related with the lower degree of gonad development and 411 gamete production in cultured broodstocks, particularly in males. Future research will be 412 necessary to definitively link dysfunctions of the endocrine reproductive axes with failed 413 reproductive performance of the cultured broodstock.

414

415 Exogenous hormonal induction of reproduction

Hormonal induction of reproduction has been extensively studied in cultured sole 416 broodstock and shown to be partially successful in both sexes. In females, different 417 administration methods of gonadotropin-releasing analogue (GnRHa) have been tested. 418 Injection of GnRHa, at doses of 5-25 µg kg⁻¹ body weight (BW), induced a transient 419 elevation of plasma E2 and T (Agulleiro et al. 2006; Guzmán et al. 2009b), slightly 420 421 stimulated final oocyte maturation (Guzmán et al. 2009b) and induced egg spawning at 2-3 422 days after treatment (Agulleiro et al. 2006). Comparatively with the injection method, treatment of females with GnRHa slow-release delivery systems, including implants and 423 424 microspheres, highly stimulated oocyte maturation, induced a higher and prolonged secretion of steroids and a higher number of spawns after a single treatment, thus increasing 425 total egg production (Agulleiro et al. 2006; Guzmán et al. 2009b). However, spawning 426 427 obtained after hormonal induction, as observed for non-induced natural spawning of cultured soles, were unfertilized in all experiments, and therefore the procedure is of 428 429 limited interest for industrial application.

In the case of males, several hormones and administration methods have been tested to
stimulate spermiation and sperm production. Treatment of males with GnRHa injection
induced a transient elevation of androgens, but was ineffective to stimulate testis

maturation or increase milt volume (Agulleiro et al. 2006; Guzmán et al. 2011a,b) and 433 434 increases in sperm quality were also transient (Cabrita et al. 2011). The effectiveness of GnRHa was slightly increased by the use of slow-release implants, which induced higher 435 436 release of steroids and a slight stimulation of spermatogenesis and milt volume (Agulleiro 437 et al. 2007; Guzmán et al. 2011b). Combined treatment of GnRHa with 11ketoandrostenedione, an 11KT precursor (Agulleiro et al. 2007) or pimozide, a dopamine 438 antagonist (Guzmán et al. 2011a), slightly increased the potency of the GnRHa treatments. 439 Treatment with a multiple injection protocol with human chorionic gonadotropin (hCG) 440 also induced steroid release and proved to be more potent than GnRHa treatments to induce 441 spermatogenesis and sperm production (Guzmán et al. 2011b). However, even if these 442 443 combined treatments were effective to stimulate spermatogenesis and sperm production in males, with no detrimental effects on sperm quality, the increased volume produced was 444 445 still insufficient and thus could only be considered as a minor success in terms of potential 446 applicability in aquaculture production.

447

448 Gamete quality and quantity

To date few studies have compared gamete quality between cultured and wild females. 449 450 Reared females held with wild males have produced viable spawns (Mañanós et al. 2007; 451 Carazo et al. 2011; Carazo 2013) and Rasines et al. (2012, 2013) demonstrated that cultured breeders could be induced to ovulate and that ova could be stripped and fertilised. 452 453 Although these studies did not explore the influence of female gamete quality in Senegalese sole reproductive dysfunctions, they indicate that cultured female gamete quality has not 454 been a restrictive factor. On the other hand, sperm characteristics have been recently 455 456 studied (Beirão et al. 2009, 2011), after the detection of low sperm volume, low cell

concentration and production in both types of male breeders - wild and G1 (Cabrita et al. 457 458 2006). Senegalese sole males can produce motile sperm all year round with specific peaks of high spermiation and of high percentage of fluent males, which usually coincide with the 459 460 female breeding season. However, considerable variation was observed in terms of sperm 461 profiles in males maintained under the same conditions. Sperm volume collected ranged from 5 to 20 µl in G1 broodstocks and 10 to 80 µl in wild-captured broodstocks. Cell 462 density and sperm production (total spermatozoa per stripping) ranged from 0.7 to 1.2×10^9 463 spz mL⁻¹ and 20×10⁶ in G1 males to values of $1-2\times10^9$ spz mL⁻¹ and $40-60\times10^6$ 464 spermatozoa for the wild-captured males, respectively. These results demonstrate that 465 sperm production in this species is very low and variable, and that wild males produce 466 sperm of higher quality (Cabrita et al. 2006). Therefore, sperm quality and quantity seems 467 to be at least one of the reasons explaining the reproduction constraints of Senegalese sole. 468 469 Recent studies have focused on gathering basic knowledge on the testis and sperm 470 production (García-López et al. 2005; Marín-Juez et al. 2011, 2013; Chauvigné et al. 2014) and these and further studies will help provide solutions to improve sperm production. 471

472

473 Strip spawning and artificial fertilization

Artificial fertilization in sole is now feasible after the standardization of hormone-based protocols to induce ovulation in females, which allowed obtaining eggs through manual stripping (Liu *et al.* 2008; Rasines *et al.* 2012). A single GnRHa injection ($25 \mu g kg^{-1} BW$) induced ovulation in 70-85% of the treated females at 41-44 h post-treatment, at a water temperature of 16°C (Rasines *et al.* 2012, 2013). The timing of ovulation was temperaturedependent, and only females at advanced stages of maturation were responsive to the treatment. Relative fecundity of Senegalese sole females after hormonal induction and egg

stripping was around 100,000-150,000 eggs kg⁻¹ BW (Rasines et al. 2013) and highest 481 482 fertilization and hatching rates were obtained within 3 h after ovulation (Rasines et al. 2012). Therefore, using stripped eggs and sperm from G1 sole, an artificial fertilization 483 protocol, based on that used for other flatfishes, has been successfully used for the 484 485 production of fertilized eggs and viable larvae (Rasines et al. 2012, 2013). Nevertheless, fertilization and hatching rates were low, highly variable, and most importantly, 486 unpredictable, with typical values around 30% and 15%, respectively. However, 487 fertilization and hatching rates as high as 84.6% and 61.6% have been achieved (Rasines 488 2013), which confirms the potential of this technique. Future experiments are necessary to 489 determine what are the critical parameters to optimize in order to render artificial 490 491 fertilization a predictable and reliable tool.

As described above, one of the main problems in artificial fertilization has been the 492 variable quality and particularly the low quantity of sperm collected. The several hormone 493 494 induction methods that have been attempted resulted in small improvements (Agulleiro et al. 2006, 2007; Guzmán et al. 2011a,b; Cabrita et al. 2011), but the volumes obtained were 495 496 generally not sufficient for aquaculture production. In addition, it is conceivable that the small testis size with gonadosomatic index of <0.15% (García-López et al. 2005) cannot be 497 expected to produce large quantities of sperm. Therefore, in combination with artificial 498 fertilization, cryopreservation of quality-controlled sperm could be used as a tool to 499 guarantee that fish farms are able to store this material and have enough sperm of the 500 501 necessary quality to fertilize the egg batches when necessary. Several protocols have been tested, mostly adapted from other species such as turbot. Briefly, sole sperm has been 502 stored using the Mounib extender with 10% DMSO, packaged in 0.5ml straws and 503 504 cryopreserved using a slower cooling rate (straws set at 5cm from the surface of liquid nitrogen, N_2) (Rasines *et al.* 2012) or packaged in 0.25 ml straws and cooled at a faster rate (2cm from the N_2 surface) (Cabrita, unpublished). Both protocols yielded good results in terms of fertility and sperm motility, but more research is required to optimize the procedure and to ensure that it does not affect progeny quality.

509

510 Larval nutritional physiology

511

As a candidate species for aquaculture, the easiness of larval and post-larval rearing of 512 Senegalese sole compared to other marine species was one of the most remarkable and 513 514 attractive aspects of this species from the beginning. For that reason, culture protocols were 515 established early on (Dinis et al. 1999; Imsland et al. 2003; Conceição et al. 2007) and are nowadays fairly standardized, with post-larvae being routinely produced with good growth 516 517 and at high survival rates. In spite of this, considerable research is still centered during 518 these early life stages given that suboptimal rearing or nutritional conditions could have 519 profound effects on later juvenile quality. Therefore, some of the problems observed during 520 the nursery and on-growing stages, such as difficulties in weaning, variable growth rates, malformations and maybe even susceptibility to diseases, can potentially be improved by a 521 522 better knowledge of larval nutritional physiology.

523

524 Digestive physiology and feeding

As in other vertebrates the digestive function in teleosts is highly associated to the particular anatomical characteristics and feeding habits that are very variable among species. As a flatfish, the Senegalese sole undergoes a remarkable transformation during the second week of life that strongly affects its behaviour, feeding and digestive physiology

(Fernández-Díaz et al. 2001). Conversely, metamorphosis may be affected by feeding and 529 530 nutritional conditions (Yúfera et al. 2005; Villalta et al. 2008a; Engrola et al., 2009b; Engrola et al., 2010). A fair amount of studies have been conducted on the digestive 531 532 physiology of this species, which have set the basis for a comprehensive morphological and 533 functional characterization of its gastrointestinal ontogeny (see Conceição et al. 2007 for a thorough revision of earlier literature). The development of the gut and other organs 534 involved in feeding and digestion is well known in Senegalese sole (Sarasquete et al. 1996; 535 536 Ribeiro et al. 1999a; Fehri-Bedoui et al. 2000; Padrós et al. 2011) and follows the general pattern observed in other marine fish with altricial larval development and particularly that 537 538 observed in other pleuronectids. However, this species has some important particularities. 539 For instance, the mouth opens relatively early in Senegalese sole, at two days after hatching at 19-20°C, and the mouth gape is wide enough to allow the ingestion of large 540 zooplanktonic prey from first feeding. On the other hand, this species is characterized by 541 542 having the smallest stomach and the longest intestine among the different families of 543 flatfishes (De Groot 1971) and there is evidence that it does not have a good capacity for 544 acid digestion, even after the gastric glands are formed in the juvenile (Yúfera & Darias 2007). Therefore, gastric digestion seems to be residual and most of the digestion occurs in 545 546 the long intestine under an alkaline environment. This is likely associated to its omnivorous 547 diet, although crustaceans, bivalves and polychaetes are the preferred prey of juvenile and adults (Cabral 2000; Sá et al. 2003). 548

549 Several studies have examined the activity of the main digestive enzymes during the 550 larval development of Senegalese sole and showed that sole larvae are ready to digest from 551 first feeding (Martínez *et al.* 1999; Ribeiro *et al.* 1999b; Moyano *et al.* 2001). Zymogen 552 granules are present at the opening of the mouth and the activity of pancreatic (trypsin,

amylase and lipase) and intestinal enzymes (alkaline phosphatase and leucine-alanine 553 554 peptidase) exhibits the expected pattern found in most altricial marine fish species (reviewed in Conceição et al. 2007; Rønnestad et al. 2013). The alkaline proteases working 555 in the developing gut exhibit a notable change by the time of metamorphosis (Moyano *et al.* 556 557 2001). Furthermore, settled post-larvae undergo a process of enzymatic maturation of the intestine which occurs between 21 and 27 dph at 22-25°C (Ribeiro et al. 1999b) with an 558 increase in the alkaline phosphatase activity and a decline of the leucine-alanine-peptidase 559 560 activity concomitant with the development of the brush border membrane of the 561 enterocytes. A recent study has shown that the maturation of the digestive system (measured as an increase in N-aminopeptidase/leucine-alanine-peptidase and alkaline 562 563 phosphatase/leucine-alanine-peptidase ratios) was highly affected by fatty acid formulation used for the Artemia enrichment (Boglino et al. 2012a). The alkaline phosphatase activity, 564 565 reflecting the degree of maturation of the digestive tract, was also previously seen to be 566 affected by an early co-feeding of inert diet (Engrola et al. 2007; 2009a). Trypsin activity, 567 on the other hand, appeared to be poorly affected by the feeding regime in sole larvae and 568 post-larvae (Engrola et al. 2007; 2009a). However, in sole juveniles the amount of protein in the diet and the origin of the protein source, animal or vegetal, affected respectively the 569 570 amount and composition of the alkaline proteases secreted in the intestine lumen (Rodiles 571 et al. 2012). Therefore, proteolitic activity seems less dependent on diet composition during the pelagic phase and early post-larval stages while juveniles appear to be able to modulate 572 573 the proteolitic activity in response to changes in dietary protein levels and sources (Sáez de 574 Rodrigáñez et al. 2011; Rodiles et al. 2012).

Recently, genes for several digestive enzymes have been sequenced, which will allow
more in depth studies looking into molecular mechanisms. Manchado *et al.* (2008), using

high throughput transcriptomic techniques, identified six different trypsinogen genes 577 578 grouped in three groups. Group I or anionic trypsinogens (ssetrypla, ssetryplb and ssetryp1c), group II or cationic trypsinogen (ssetryp2) and group III or psychrophilic 579 trypsinogens (ssetryp3 and ssetrypY). These showed a different pattern of expression before 580 581 and after the eye migration stage, with *ssetryp1a* variants and *ssetryp2* transcripts exhibiting higher expression during pre-metamorphosis while ssetryp3 and ssetrypY582 transcripts were more abundant in juveniles. This change had been previously noted in the 583 584 zimograms of proteases (Moyano et al. 2001). In another study, as noted previously, 585 Gamboa-Delgado et al. (2011) found that trypsin and chymotrypsin activities showed a similar pattern in Senegalese sole during early development independently of the dietary 586 treatment but, however, increases in *ssetryp1* transcription were observed at different 587 timings when feeding on rotifers from first feeding or when larvae were fed directly on 588 589 Artemia nauplii (4 or 5 dph, respectively) at 16-22°C. Therefore, there is evidence that the 590 type of diet might modulate trypsinogen gene expression during early larval development, even if this is not reflected in trypsin activity. 591

592 Other recent studies have started looking at daily rhythms of feeding and digestion, as this information is important for optimizing species- and developmental stage-specific 593 feeding protocols. Feeding incidence is high from the opening of the mouth, being 80% of 594 595 larvae hatched from an egg batch able to feed actively in the first day of feeding and practically the totality of larvae in the second day (Navarro-Guillén et al. 2015). Although 596 597 nocturnal habits have been described in juveniles and adults (Bayarri et al. 2004), we have 598 only recently gained insight on the daily rhythms of activity during the larval and postlarval stages, which show a clear switch from diurnal to nocturnal when the larvae finishes 599 600 the eye migration phase and starts the benthic life (Blanco-Vives *et al.* 2012). This has been

related to feeding, as results show that during the early pelagic stage sole larvae exhibit a 601 602 clear diurnal feeding pattern with maximum gut content at sunset hours while after the start of benthic life post-larvae may feed during the whole day (Navarro-Guillén et al. 2015). In 603 another experiment with 35 dph post-larvae fed on Artemia with ¹⁴C-labelled protein, a 604 605 daily feeding and digestive rhythm was observed independent of the illumination cycle. In 606 this experiment, post-larvae fed at 10:00, 14:00, 18:00 and 04:00h presented higher Artemia protein digestibility than sole fed at 23:00h, when Artemia intake was lower. In addition, 607 higher protein retention and lower catabolism occurred in the hours with lower ingestion 608 (18:00 and 23:00h) (Navarro-Guillén et al. 2014a). Finally, recent results on daily rhythm 609 of digestion in sole larvae showed that trypsin activity had a daily pattern following that of 610 611 the gut content, while lipase and amylase activities presented completely independent patterns (Navarro-Guillén et al. 2015). 612

613

614 Fatty acid and lipid requirements

The lack of suitable inert microdiets, with good acceptability and the possibility to manipulate nutrient composition with high precision and reproducibility, has not enabled performing detailed studies on lipid and fatty acid requirements in fish larvae. Therefore, the only available studies are with live prey, either non-enriched or after enriching with different commercial products or oil emulsions, which present important constraints in terms of the accuracy and reproducibility of the factors that can be analysed.

Available evidence suggests that, just as in juvenile stages (see below), high total lipid
levels should be avoided in Senegalese sole larvae and post-larvae diets (Morais *et al.*2005a,b; Conceição *et al.* 2007).

624 Senegalese sole larvae have particularly low requirements for LC-PUFA (reviewed in Conceição et al. 2007). In fact, post-metamorphosed larvae could be grown on diets 625 containing negligible DHA and low EPA levels, such as non-enriched live prey, without 626 obvious detrimental effects (Morais et al. 2004a, 2005a,b; Villalta et al. 2005a). From the 627 628 studies that have been performed so far there is the indication that Senegalese sole larvae perform well, with good growth and survival up to 40 dph, with dietary EPA levels as low 629 as 0.5% TFA (when DHA was provided at 4.3% TFA), while growth was decreased at 630 higher (10.7 and 20.3% TFA) levels of EPA (Villalta et al. 2008a). Regarding DHA, 631 632 Villalta *et al.* (2005a) observed a better growth (only significantly for total length, up to 36 dph) with 7.7% TFA of DHA than with 4.4% or 14.7% TFA but, nonetheless, differences 633 were not significant in the treatment fed Artemia containing no DHA (all treatments 634 contained 1.7% TFA of EPA). On the other hand, in a study by Morais & Conceição 635 636 (2009), growth and metabolic results suggested that DHA requirements of Senegalese sole 637 post-larvae, after settlement, are probably not higher than 3% TFA. Fine studies on EFA requirements are still missing but Boglino et al. (2012a), when testing different Artemia 638 commercial enrichment products to determine the most suitable for Senegalese sole larval 639 development until 38 dph, concluded that larval growth was highest when using a product 640 641 which led to Artemia metanauplii containing intermediate values of ARA, EPA, DHA, total 642 PUFA and monounsaturated fatty acids (32.7, 0.7, 3.1, 9.5, 51.1 and 32.7% TFA, respectively). Conversely, the treatment inducing the lowest growth and delayed 643 644 morphogenesis was the one with highest levels of EPA (7.6% TFA).

Nevertheless, there is evidence suggesting that LC-PUFA requirements are higher in the early larval (pelagic) stage, which is probably a critical period associated with extensive organogenesis and tissue remodelling culminating in metamorphosis, which in flatfishes

involves dramatic functional and morphologic changes (Morais & Conceição 2009; 648 649 Dâmaso-Rodrigues et al. 2010). Therefore, EFA requirements need to be examined independently in the pre- and post-metamorphic stages. In addition, it should be kept in 650 651 mind that the estimation of requirements may vary depending not only on the relative 652 amounts of EFA (particularly DHA/EPA ratio) but also on the available dietary energy 653 (Villalta et al. 2005a, 2008a; Boglino et al. 2012a). A recent study (Navarro-Guillén et al. 2014b) has shown that supplementation of DHA (up to 4% of TFA) to emulsions based on 654 vegetable oils (supplying higher levels of better energy-yielding substrates than typical 655 656 enrichment products) improved larval growth and survival, suggesting that a correct balance needs to be found between dietary energy and EFA. 657

Such unusually low requirements for LC-PUFA during the larval stages, compared to 658 most marine fish species, were quite intriguing. However, a recent discovery revealed the 659 660 physiological reason behind these observations, by uncovering a unique gene activity in the 661 LC-PUFA biosynthesis pathway of this species. The classical pathway for the synthesis of DHA from EPA in vertebrates, known as the "Sprecher" pathway, involves two sequential 662 663 elongations of EPA to 24:5n–3 followed by $\Delta 6$ desaturation and one round of peroxisomal β -oxidation (Voss *et al.* 1991). However, the extent to which a vertebrate species can 664 produce LC-PUFA from lower chain precursors (C18 PUFA) varies according to its 665 repertoire of fatty acyl elongase (Elovl) and desaturase (Fad) enzymes. In teleosts, a long 666 standing paradigm was that species diverge according to the environment and/or trophic 667 668 level they occupy and that marine species (contrary to freshwater/diadromous species) 669 cannot biosynthesise LC-PUFA and hence have a strict dietary requirement for these fatty acids (Tocher 2010). For this reason, the discovery of a gene coding for a fatty acyl 670 671 desaturase with $\Delta 4$ activity ($\Delta 4 fad$) and which is able to produce DHA directly from

desaturation of docosapentaenoic acid (DPA; 22:5n-3), which is synthesised from EPA via 672 elongation through *elov15* (also functionally characterized in the same study), was an 673 extraordinary breakthrough (Morais et al. 2012; Fig. 2). Furthermore, the expression of 674 $\Delta 4 fad$ was shown to be transcriptionally regulated by dietary levels of LC-PUFA, being up-675 676 regulated when sole larvae were fed diets containing low levels of DHA (Morais et al. 2012; Navarro-Guillén et al. 2014b). What was even more surprising, considerable elov15 677 expression was already present in eggs, transcription of $\Delta 4 fad$ started just after hatching, 678 and both transcripts peaked at the start of exogenous feeding (Morais et al. 2012). 679 Furthermore, their levels in eggs and newly hatched larvae were significantly affected by 680 681 the content of DHA in the broodstock diet, which in turn was reflected in the egg biochemical composition (Morais et al. 2014a). These results indicate a high degree of 682 regulation suggesting a true biological relevance of this pathway in reducing LC-PUFA 683 dietary requirements in this species. 684

685

686 Amino acids and protein requirements and metabolism

Early work on amino acid (AA) requirements of Senegalese sole (reviewed in Conceição *et al.* 2007) indicated that the ideal dietary AA composition changes along ontogenesis, probably tightly linked with metamorphosis (Aragão *et al.* 2004). These earlier studies also demonstrated that bioavailability of individual AA changes, and that early life stages of sole have a good capacity to regulate AA catabolism (Conceição *et al.* 2007).

As metamorphosis is a critical point in Senegalese sole development and is regulated by thyroid hormones, recent studies have looked at the importance of their precursors - the aromatic AA phenylalanine and tyrosine. Previous work by Aragão *et al.* (2004) showed that the contribution of aromatic AA to the larval profile decreased significantly after

metamorphosis. Therefore, the fate of aromatic AA was followed by tube-feeding 696 697 radiolabelled phenylalanine and tyrosine at different developmental windows, and the results showed a preferential retention of these AA during metamorphosis (Pinto et al. 698 2009), indicating that physiological requirements for these AA likely increase during this 699 700 stage. In order to verify this, dietary aromatic AA supplementation at this stage was tested by means of short-term tube-feeding experiments (Conceição et al. 2010). The results 701 suggested that tyrosine is probably conditionally indispensable for this species at 702 703 metamorphosis, while dietary tyrosine supplementation may be beneficial to help larvae to cope with metamorphosis-related processes (Pinto et al. 2010a). In post-metamorphic fish, 704 although no clear effects of dietary aromatic AA supplementation were observed under 705 706 normal conditions, it was demonstrated that tyrosine supplementation helps the fish to 707 maintain a normal tyrosine metabolism under stressful conditions (Aragão et al. 2010). Tyrosine, as mentioned before, is the precursor of thyroid hormones, but also of 708 709 catecholamines, both mediating the stress response in fish.

710 In studies of live feed replacement by microencapsulated diets it was found that taurine 711 supplementation increased larval growth potential and metamorphosis success in sole larvae (Pinto et al. 2010b). These effects were not observed during the pelagic phase, but a 712 clear effect of earlier dietary taurine supplementation was observed in post-metamorphic 713 714 larvae. Taurine is not used in protein synthesis but can be found freely in the cytosol. Among other factors, the antioxidant properties of taurine may be of special importance 715 716 during sole metamorphosis. Later work revealed that larvae were fully able to uptake 717 taurine in the digestive tract at the onset of metamorphosis and that metamorphosis was an important developmental trigger to promote taurine gut transport (Pinto et al. 2012). 718

719 Collectively, these results reemphasize that metamorphosis is a critical developmental stage720 in Senegalese sole, during which dietary AA profiles assume an extreme importance.

Recent modelling studies suggest that Senegalese sole are highly sensitive to dietary AA 721 722 imbalances, which result in high AA losses and have a major impact in protein retention 723 (Rønnestad & Conceição 2012). As sole larvae are still largely fed on live preys, which have an imbalanced AA profile (Aragão et al. 2004), and considering the high 724 physiological requirements for some specific AA revealed in the latest studies, dietary 725 726 supplementation with these AA seems paramount to maximize growth. The challenge now 727 is to know how much and for how long these supplements should be provided. It has 728 already been demonstrated in Senegalese sole juveniles that dietary AA supplementation 729 with tryptophan during 14 days presents more benefits than during 28 days (Costas et al. 2012). Therefore, it is essential to evaluate different levels of dietary AA supplementation 730 731 at specific developmental windows, so that the industry can accurately assess the 732 cost/benefits of such an investment in growth and larval quality.

733 Estimates of Artemia protein digestibility in Senegalese sole larvae and post-larvae vary 734 between 57% and 98% (Rønnestad et al., 2001; Morais et al., 2004b; Engrola et al., 2009b; Engrola et al., 2010; Campos et al., 2013a; Navarro-Guillén et al., 2014a). This variation 735 736 can be explained by differences in developmental stage, temperature, time of feeding and 737 feeding regime between studies. Some studies have addressed the effect of live prey replacement by inert diet at different developmental stages. In this respect, sole fed live 738 739 prey alone or co-fed with 20% (low replacement) of inert diet showed a similar protein 740 retention, always above 70%, except at 8 dph, when this was slightly reduced to around 65% (Engrola et al., 2009b). However, in sole co-fed with 58% of inert diet from mouth 741 opening (high replacement), protein digestibility and retention efficiency were significantly 742

reduced between 6 and 15 dph (Engrola et al., 2010). During the metamorphosis climax 743 744 (14-18 dph) both live fed and co-fed sole had a lower protein digestibility than at younger 745 or older ages (Engrola et al., 2009b; Engrola et al., 2010). This is probably explained by a 746 reduced digestive capacity during the metamorphosis climax, as the retention efficiency 747 remained almost constant throughout the experimental period at low replacement levels. 748 Sole larvae do not seem to increase feed intake during this period (Parra & Yúfera, 2001; Engrola et al., 2010), and therefore protein deposition must be sustained by using energy 749 reserves accumulated during the earlier stages (Parra & Yúfera, 2001). 750

751 Studies on the capacity of larvae to digest proteins of different complexities have been performed in view of the long-term aim of developing an inert diet that can be digested 752 753 from mouth opening. These have shown that sole presents a poor capacity to digest complex proteins at mouth opening (Gamboa-Delgado et al., 2008; Engrola et al., 2009a). 754 755 However, when testing peptide fractions, sole showed a high capacity to digest 1.0kDa 756 peptide fractions at all developmental stages. In addition, the retention of a 7.0kDa peptide 757 fraction was improved along development (Richard *et al.*, 2013). These studies support the 758 suggestion that the complexity of the dietary nitrogen is a key issue that needs to be considered to improve marine larval growth (Conceição et al., 2011). 759

Finally, studies have shown that Senegalese sole protein metabolism and retention (hence growth) is highly modulated by thermal conditions during early developmental stages (Campos *et al.*, 2013b,c), as would be expected. It is noteworthy that larval rearing temperatures of 18 and 21°C positively affected *Artemia* protein digestibility during metamorphosis, compared to larvae that were reared at 15°C, indicating that the thermal history modulated physiological pathways (Campos *et al.*, 2013a). However, when the same larvae were analysed at an older age, *Artemia* protein digestibility was higher in larvae reared at lower temperature (15°C), indicating that digestive capacity was no longer
impaired and the improved digestibility could be at least partially responsible for the
compensatory growth that was observed in larval weight (Campos *et al.*, 2013a).

770

771 **On-growing diets**

772

773 Nutrient requirements

774 Senegalese sole inhabits sandy or muddy bottoms of coastal or brackish areas, where it has a low-diversity diet consisting predominantly of polychaetes, but also small crustacean 775 (such as tanaids and amphipods) and bivalve molluscs (Garcia-Franquesa et al. 1996; 776 Cabral 2000). This dietary regime is likely characterized by higher protein and 777 carbohydrate levels and lower total lipid contents than in the zooplanktonic food web. For 778 779 instance, the biochemical composition of one of the main prey items in sole's diet, the 780 polychaete Nereis diversicolor, varies from 47%-60% protein, 6.6%-19.3% lipids and 781 around 8%-20% carbohydrates, depending on the season, in the Tagus estuary (Portugal) 782 (Luis & Passos 1995). Nutritional studies in captivity have shown that sole diets should include a high crude protein level (53% dry matter, DM) to maintain good overall growth 783 performance (Rema et al. 2008). The only available direct estimate for indispensable amino 784 785 acids (IAA) requirement in sole refers to lysine (Lys), as Silva (2010) estimated an optimum dietary supply of 4.7 Lys 16 g⁻¹ N for maximum protein accretion. More recently, 786 Costas (2011) further refined the ideal protein profile in diets for juvenile sole by 787 estimating the bioavailability of the IAA. In this study, the utilization of the 10 individual 788 IAA was evaluated by tube-feeding Senegalese sole juveniles with compound feeds 789 containing ¹⁴C-labelled IAA as tracers. Results showed differences in digestibility, 790

retention and catabolism between individual IAA, and bioavailabilities relative to lysine
were calculated. High relative bioavailabilities were found for histidine, leucine, isoleucine,
valine, methionine, threonine, phenylalanine and arginine, meaning that these IAA are
retained more efficiently than lysine, while tryptophan had the lowest relative
bioavailability among IAA (Costas 2011).

In most marine fishes, a significant protein sparing can be achieved by increasing 796 digestible energy levels through an increase in fats and/or carbohydrates (Helland & 797 798 Grisdale-Helland 1998; Kaushik 1998). However, contrary to most marine fish species, the ability of Senegalese sole juveniles to efficiently use high dietary lipid levels seems limited, 799 in both juvenile (Dias et al. 2004; Borges et al. 2009; Guerreiro et al. 2012) and market-800 801 sized fish (Valente et al. 2011). Borges et al. (2009) clearly demonstrated a low lipid tolerance in this species, and recommended a dietary lipid inclusion of up to 8% for optimal 802 803 growth and feed utilisation efficiency at a protein level of 57% (DM basis). Dietary lipids 804 do not seem to be a good energy source for promoting growth in Senegalese sole as there is no clear evidence of a protein-sparing effect by increasing dietary lipid levels, even when 805 806 the dietary protein level is below this species requirement (Mandrioli et al. 2012; Borges et al. 2013a). Irrespective of the rearing temperature (16 vs. 22°C), Guerreiro el al. (2012) 807 also found that feed efficiency, N retention and energy retention were highest in sole 808 juveniles fed a diet containing 55% protein and 8% lipids. The activity of enzymes 809 involved in key metabolic pathways points towards a lack of metabolic adaptation to high 810 811 lipid levels (Dias et al. 2004; Borges et al. 2013a). Recently, Mandrioli et al. (2012) 812 showed that a concomitant increase in dietary lipids and decrease in dietary protein and was associated with a massive storage of unused lipid within sole hepatocytes. Similarly, 813 814 Valente *et al.* (2011) reported moderate steatosis and some cellular necrosis in large-sized sole (>300g) fed high lipid levels. Moreover, Campos *et al.* (2010) observed a decrease in
the expression of myogenic regulatory factors and myosins in the muscle of Senegalese
sole fed increasing dietary lipid levels, supporting the hypothesis that high lipid levels
somehow depress growth by reducing protein accretion.

819 Even though dietary lipids are not efficiently used as non protein energy sources in 820 Senegalese sole, its digestion and absorption seems identical to other marine species (Dias et al. 2010; Borges et al. 2013b). Different dietary levels equally induced high lipid 821 822 digestibility and intestinal lipase activity, while increased plasma concentrations were observed in fish fed high fat diets compared to those fed the low fat diet, demonstrating 823 effective lipid absorption (Borges et al. 2013b). S. senegalensis is a lean fish (1-4g of 824 825 fat/100g of flesh) with a scarce capacity to accumulate fat even when fed high lipid levels (Borges et al. 2009, 2014; Fernandes et al. 2012). Previous studies (Rueda-Jasso et al. 826 827 2004; Borges et al. 2009; Valente et al. 2011; Fernandes et al. 2012) reported that liver is 828 the preferential site for fat deposition (5.5–37%). Furthermore, sole liver seems to have an 829 important role in clearing plasma triglycerides, showing an increased expression of genes 830 involved in lipid transport (microsomal triglyceride transfer protein, MTP), trafficking 831 (fatty acid binding protein 11, FABP11) and fatty acid uptake (very low density lipoprotein 832 receptor, VLDL-r) in juvenile fish fed high fat diets (Borges et al. 2013b).

As the recommended diets for Senegalese sole have a low lipid level (8%), most energy must be derived from protein and starch substrates. Marine carnivorous fish generally show a low ability to use dietary carbohydrates as an energy source. However, data from Borges *et al.* (2013a) and Guerreiro *et al.* (2014) showed that increased levels of dietary carbohydrates had no detrimental effects on the growth performance of Senegalese sole juveniles. Additionally, Borges *et al.* (2013a) reported that phosphofructokinase 1 (PFK-1)

was up-regulated in sole fed a low fat: high starch diet, suggesting also a role for glucose as 839 840 an energy source in Senegalese sole muscle. To investigate this further, these authors looked at the effect of high dietary lipid levels on glucose metabolism in sole (Borges et al. 841 2014a). Results showed that fish fed on high fat:low starch diet exhibited prolonged 842 843 hyperglycemia and greater liver glucose-6-phosphatase activity than fish fed on low 844 fat:high starch diet. Moreover, the level of major proteins involved in the insulin and nutrient signalling pathway (AKT, p70 ribosomal S6-K1 Kinase and ribosomal protein S6) 845 were reduced in muscle of fish fed on high fat diets suggesting a possible insulin resistance 846 state (Borges et al. 2014a). Further studies on the metabolic abilities of Senegalese sole to 847 848 cope with high dietary carbohydrate levels are needed.

849

850 Alternative vegetable ingredients and nutritional value for human consumption

851 High-quality fish meal (FM) is still the major protein source currently used in sole diets. 852 However, supplies of FM and fish oil (FO) are finite, and their replacement in aquafeed formulations with ingredients from more available plant sources is a major necessity 853 854 (Tacon & Metian 2008). In this respect, Senegalese sole appears to have an important advantage with regards to other marine fish species, as it has been demonstrated to cope 855 856 well with diets in which most FM (89%) was replaced by plant protein (PP) sources (Silva et al. 2010). Furthermore, FM could be totally replaced by a mixture of PP sources without 857 any adverse effects on growth, feed or protein utilization provided that the dietary AA are 858 859 balanced by the addition of small amounts of crystalline AA (Silva et al. 2009). After an 860 intense research effort in recent years, it is now clear that replacement of marine-derived protein sources by practical PP ingredients in Senegalese sole feeds with minimal AA 861 862 supplementation is feasible in both juvenile (Cabral et al. 2011) and large-sized fish

(Valente et al. 2011; Cabral et al. 2013). Present data indicates that Senegalese sole can 863 864 efficiently use diets with up to 75% of FM replacement by PP sources, but growth rate and nutrient gain in juveniles mainly depend on the selection of adequate PP blends, rather than 865 866 on the PP incorporation level (Cabral et al. 2011). The source of dietary protein modified 867 both the amount and composition of the pancreatic proteases secreted into the intestinal lumen of juveniles, without reducing growth (Rodiles et al. 2012), which indicates the 868 capability of Senegalese sole to modulate digestive protease secretion when dietary protein 869 sources are modified. This is further supported by data on large-sized sole, revealing 870 similar nutrient intake and utilization (including similar apparent digestibility coefficients 871 (ADCs) of nutrients) in fish fed either PP or FM based diets (Cabral et al. 2013). It is also 872 noteworthy that increasing FM replacement level has a positive environmental impact as it 873 reduces nitrogen losses and fecal phosphorus (P) waste, as well as the FM used per kg of 874 875 sole produced (Fi:Fo ratio) (Cabral et al. 2011; Cabral et al. 2013). Further studies on the 876 digestibility of alternative ingredients are warranted to help select the most adequate PP combination for this species. 877

878 Some studies have reported a decrease in whole body lipid content of sole fed PP diets with the increasing replacement of FM (Silva et al. 2009; Cabral et al. 2013). However, 879 another study has shown that high dietary PP levels can increase lipid storage in liver 880 881 (Valente et al., 2011), while muscle lipid content is generally not affected by the inclusion of PP. On the other hand, the replacement of FM by PP, even at extremely high levels (75 882 and 100%), is still effective in producing a n-3 LC-PUFA rich product. Muscle fatty acid 883 profile of fish fed PP sources showed higher levels of C18:2n-6 while levels of DHA, 884 DHA/EPA and EPA/ARA ratios were not affected (Cabral et al. 2013). 885

Fish is the main dietary source of DHA and EPA for humans and these fatty acids are 886 887 best known for preventing cardiovascular and inflammatory diseases (Simopoulos 1999). There is a general concern about a possible loss of health beneficial effects for human 888 consumers when replacing FO, rich in EPA and DHA, by vegetable oils (VO) which lack 889 890 these EFA. In Senegalese sole it was possible to substitute up to 100% of FO by either linseed oil and soybean oil (Benítez-Dorta et al. 2013) or blends of rapeseed, soybean and 891 linseed oil (Borges et al. 2014b) without significantly compromising juvenile performance; 892 of these, only the 100% soybean oil diet slightly reduced growth and feed utilization. 893 Although such high substitutions altered muscle fatty acid profile, reflecting the dietary 894 composition, a selective deposition and retention of DHA was observed, as this fatty acid 895 896 was only slightly, but not significantly, reduced in comparison to FO fed fish (Benítez-Dorta et al. 2013; Borges et al. 2014b). Recommended daily intake (RDI) of EPA+DHA is 897 estimated to be at least 0.25 g per day for healthy human individuals (EFSA 2010) and even 898 sole fed on VO based diets can provide consumers with almost two times the RDI, 899 confirming its good nutritional value (Borges et al. 2014b). Nevertheless, it should be noted 900 901 that in all tested diets FM has largely contributed to total dietary fat content and therefore further research is necessary to evaluate vegetable diets during all production cycle and 902 903 with simultaneous substitution of FM and FO in order to clarify the impact of these diets in 904 Senegalese sole flesh quality. However, it is likely that the fatty acid profile could be reestablished with a finishing diet, although this still needs to be demonstrated. 905

The above results are surprising for a marine species but, as mentioned previously, a fatty acyl elongase (*elov15*) and a unique desaturase coding for a protein with $\Delta 4$ activity ($\Delta 4fad$) were discovered and functionally characterized in Senegalese sole and *in vitro* results suggested capability to synthesize DHA from EPA (Morais *et al.* 2012) in this

species. In addition, other circumstantial evidence related to the expression of these genes 910 911 and the fatty acid profile of post-larvae fed Artemia enriched with different VO emulsions 912 supplemented with DHA supported this hypothesis and went further in suggesting that DHA might even be produced from C18 PUFA precursors, even though a gene coding for a 913 914 Δ 6Fad has not been found so far in this species (Navarro-Guillén *et al.* 2014b). However, the in vivo activity and biological relevance of this pathway could not be determined at the 915 time. This was very recently addressed in a study where juvenile sole were fed isoproteic 916 917 diets containing 8% or 18% total lipid supplied by either 100% FO or with 75% of the FO 918 replaced by a VO blend. When incubating hepatocytes and enterocytes isolated from these fish with $[1-^{14}C]18:3n-3$ and $[1-^{14}C]20:5n-3$, it was found that there was no $\Delta 6$ desaturation 919 of ¹⁴C-labeled 18:3n-3 in either tissue and only elongation to 20:3n-3, and therefore the 920 hypothesis of a potential capability of LC-PUFA synthesis from C18 PUFA was not 921 substantiated. In contrast, substantial amounts of radioactivity from ¹⁴C-labeled EPA were 922 923 recovered in 22:5n-3, 24:5n-3 and DHA, and the levels of these metabolites were higher at lower dietary lipid level and clearly stimulated by raising dietary VO levels particularly in 924 925 hepatocytes (Morais et al. 2014b). This unequivocally confirmed the existence of a true capacity to biosynthesize DHA from EPA in Senegalese sole which, furthermore, is 926 modulated by dietary composition both in terms of lipid level and fatty acid composition. 927 928 Finally, considering the highly appreciated sensorial characteristics of Senegalese sole

929 flesh, it is important to evaluate how this can be affected by vegetable-based diets. The 930 sensory evaluation of cooked flesh slices showed that the replacement of FM by PP blends 931 did not have a significant impact on the majority of volatile compounds (Silva *et al.* 2012; 932 Moreira *et al.* 2014) or sensory descriptors (Cabral *et al.* 2013). Nevertheless, the long term 933 impact of high PP incorporation levels on gut integrity, liver function and immune status934 still needs to be addressed.

935

936 Phenotypic abnormalities affecting fish quality

937

Skeletal deformities and, to a lesser extent pigmentary disorders, are two important 938 factors that may reduce the productivity and profitability of Senegalese sole intensive 939 940 production (Koumoundouros 2010; Fernández & Gisbert 2011; Boglione et al. 2013a). The 941 percentage of fish with medium to severe anomalies and/or malpigmentation varies greatly, not only among different farms, but also among different batches within the same hatchery 942 or even within the same batch of eggs (Boglione et al. 2013a; Darias et al. 2013a). 943 944 Hatcheries that provide juveniles grade out malformed fish, while those farms that grow-945 out abnormal fish to market size have either to downgrade the product to a lower value or 946 discard them before their commercialization. Moreover, fish with severe pigmentary 947 disorders cannot be commercialized as these fish do not meet the quality standards set up 948 for fishery products (Bolker et al. 2005; Hamre et al. 2007; Darias et al. 2103a). Therefore, losses during the whole production cycle may be substantial. Hatchery and nursery 949 productions are mostly affected, since the majority of skeletal structures (Gavaia et al. 950 951 2002) and skin chromatophores (Darias et al. 2013b) develop and differentiate during the larval and post-larval stages. 952

953

954 Skeletal deformities

955 Regarding the etiology of deformities, skeletal anomalies are generally the result of 956 genetic factors and/or the incapacity of homeorhetic mechanisms to compensate for

stressful environmental conditions (Boglione et al. 2013a). In this context, the development 957 958 of skeletal disorders has been linked to a poorly understood relationship between environmental and biotic factors affecting fish morphogenesis (see reviews in Lall & 959 Lewis-McCrea 2007; Boglione et al. 2013a). Consequently, understanding the 960 961 environmental needs and nutritional requirements of fish larvae is essential for improving husbandry and rearing practices, and reducing the incidence of skeletal disorders and 962 associated economical costs. Although there is considerable information on the impact of 963 environmental and genetic factors on the development of skeletal deformities in several 964 farmed teleost species (Boglione et al. 2013b), most of the information gathered in 965 Senegalese sole deals with the impact of nutrition on larval morphogenesis (Engrola et al. 966 967 2009a; Fernández et al. 2009; Fernández & Gisbert 2010, 2011; Boglino et al. 2012a,b,c), and there is only one report on the effects of environmental factors on skeletogenesis 968 (Blanco-Vives et al. 2010). Larval nutrition during pre- and pro-metamorphosis is one of 969 970 the key parameters affecting skeletogenesis during sole early development and several studies have shown that suboptimal levels and different forms of nutrients supplied in the 971 972 diet are responsible for the appearance of skeletal deformities (Lall & Lewis-McCrea 2007; Fernández & Gisbert 2011; Boglione et al. 2013a). Most of these studies in Senegalese sole 973 974 have focused on lipids, particularly EFA (Boglino et al. 2012a,b,c, 2013), and liposoluble 975 vitamins, especially vitamin A and K (Fernández et al. 2009; Fernández & Gisbert 2010, 2011; Richard et al. 2014). 976

Different studies have shown a high incidence of skeletal deformities in hatchery-reared
Senegalese sole, with values ranging from 44% (Gavaia *et al.* 2002) to 80% (Engrola *et al.*2009a; Fernández *et al.* 2009; Boglino *et al.* 2012a). Most of the skeletal anomalies are
typically found along the vertebral column and caudal fin complex of pro-metamorphic

larvae and early juveniles. The vertebral column in Senegalese sole is generally composed 981 982 of 45 vertebrae, divided in 8 prehaemal and 37 haemal vertebrae (including the urostyle). The haemal region of the vertebral column, especially the pleural vertebrae, is mostly 983 984 affected by the fusion and/or compression of vertebral bodies, and by abnormalities of the 985 vertebral arches (Engrola et al. 2009a; Fernández et al. 2009; Cardeira et al. 2012). Incomplete and complete fusion of vertebral bodies are not necessarily considered as 986 different stages of the same anomaly, as not always incomplete fusions aggregate in older 987 fish (Deschamps et al. 2009). Although these skeletal disorders are common in reared 988 specimens (Gavaia et al. 2009; Engrola et al. 2009a; Fernández et al. 2009; Fernández & 989 Gisbert 2010; Boglino et al. 2012a,c), they have also been identified in wild animals, but at 990 991 a lower frequency (Gavaia et al. 2009).

992 Among the different body regions, the head is the area less affected by skeletal disorders in Senegalese sole. This differs from round finfish species, where jaw abnormalities are 993 994 very common (Boglione et al. 2013a), and might be explained by differences in the timing of ossification. In Senegalese sole both jaws ossify soon after the onset at exogenous 995 996 feeding (ca. 2 dph at 17-18°C; Wagemans & Vandewalle 2001; Fernández et al. 2009), whereas in other species such as gilthead sea bream or European sea bass this process takes 997 998 place at later stages of development (Koumoundouros 2010; Gisbert et al. 2014). In 999 addition, lower incidences of jaw anomalies were observed when sole were reared at a thermal cycle of 22.1°C day/19.0°C night, instead of 19.2°C day/22.0°C night (Blanco-1000 1001 Vives et al. 2010). Studies in mammals have shown that light and temperature 1002 manipulation changes the pattern of osteoblast and bone mass proliferation in response to changes in circadian pattern, which might have affected the normal development of jaws in 1003 1004 sole (Boglione et al. 2013a).

When different commercial enrichment products differing in biochemical composition 1005 1006 and fatty acid profile were tested, the incidence of skeletal deformities was similar (71.9 -79.3%). However, data from larval performance and degree of ossification suggested that 1007 levels of DHA, EPA and ARA of 9.5, 3.1 and 0.7% TFA, respectively, and $(\pi)/(n-6)$ 1008 1009 PUFA, DHA/EPA, ARA/DHA ratios of 5.2, 3.0, 0.1, respectively, were the most adequate, 1010 among the tested products, for proper larval development (Boglino et al. 2012a). So far most of the nutritional studies dealing with EFA have focused on the effect of ARA on 1011 skeletogenesis. Senegalese sole post-larvae fed high dietary ARA levels (7% TFA) from 8 1012 to 50 dph tended to have a less mineralized skeleton in comparison with those animals fed 1013 1014 lower doses of ARA (4.5% TFA) (Boglino et al. 2012b). Generally, lower mineralization 1015 has been shown to contribute to the development of skeletal disorders in fish, as less mineralized bones tend to be more fragile and prone to abnormal development or get more 1016 1017 easily deformed (Boglione et al. 2013b). Although Boglino et al. (2012b) recommended 1018 4.5% TFA as the optimal level of dietary ARA for proper growth and mineralization in Senegalese sole larvae during the Artemia feeding period, none of the tested dietary ARA 1019 1020 levels (1.0, 4.5 and 7.0% TFA) affected the incidence of skeletal deformities. However, feeding Senegalese sole larvae with high levels of ARA (10.2 and 7.1% TFA in enriched 1021 1022 rotifer and Artemia metanauplii, respectively) at different developmental stages resulted in 1023 differences in the incidence of vertebral column deformities (Boglino et al. 2012b). In brief, skeletal deformities affecting the vertebral column were 25-32% higher when larvae were 1024 1025 fed high dietary ARA levels during both the rotifer and Artemia feeding periods (2-50 dph) 1026 than when these were fed only during the Artemia period (8-50 dph). These results suggest an important effect of ARA intake during the pre-metamorphic stage (3-10 dph) on the 1027 1028 proper development of the vertebral axis. In addition, ARA levels also appeared to affect

the processes of eye migration and cranial bone remodelling as sole juveniles fed high 1029 1030 dietary ARA levels from mouth opening presented a higher incidence of cranial deformities $(95.1 \pm 1.5\%)$. These were mainly associated to the impairment of eye migration, which 1031 1032 resulted in various alterations including shape of the head, disposition of the eyes with 1033 regards to the vertebral column and mouth axes, and a shorter interocular distance. Furthermore, changes in the positioning of both eyes resulted in osteological differences for 1034 some of the skeletal elements from the splanchnocranium and neurocranium (Boglino et al. 1035 2013). A possible explanation for these disorders could be an overproduction of ARA-1036 eicosanoid derivates, like prostaglandin E2 (PGE₂) (Boglino et al. 2013) or leukotriene B4 1037 (LTB₄) (Alves-Martins et al. 2012), which can disrupt the rate of bone mineralization, 1038 formation and resorption. 1039

With respect to vitamins, there is extensive information on the effects of dietary vitamin 1040 1041 A (VA) on skeleton morphogenesis of Senegalese sole as well as of other flatfish species 1042 (see review in Fernández & Gisbert 2011). In general, the incidence of vertebral deformities in Senegalese sole increased with dietary VA content, independently of the 1043 1044 developmental stage (pre-, pro-, and post-metamorphic larvae) at which the VA doses were administered. However, the severity of these deformities was higher when high VA doses 1045 were given at early developmental stages (Férnandez & Gisbert 2011). In particular, high 1046 levels of VA (203,000 IU kg⁻¹) offered to larvae from 6 to 37 dph induced a lower length 1047 and an increase in deformed fish, as well as in the percentage of fish showing more than 1048 1049 one deformity (Fernández et al. 2009). However, no jaw deformities were observed in any larval group, suggesting that the dietary VA imbalance occurred at a non-critical 1050 developmental stage for jaw skeletogenesis. Still, larvae fed the 203,000 IU VA kg-1 1051 1052 showed cranial deformities related to the opercular complex that was significantly

correlated to deformed prehaemal vertebrae. In addition, when such high VA levels were 1053 1054 administered at the pre-metamorphic stage (3-10 dph) the notochord segmentation process was disrupted and the percentage of post-metamorphic specimens with a supranumerary 1055 vertebra increased. In a subsequent study, Fernández and Gisbert (2010) observed that 1056 1057 skeletal structures presented differential sensitivity to dietary VA imbalance in terms of their ossification. Senegalese sole larvae fed Artemia containing more than 42.666 IU kg⁻¹ 1058 VA showed a significant increase in the incidence of deformities in most chondral 1059 structures (neural and haemal spines, epural and parahypural), whereas intramembraneous 1060 1061 bone structures (haemal and caudal vertebrae centra) were only affected when larvae were fed the highest VA dose (203,000 IU total VA kg⁻¹). Comparing these studies in Senegalese 1062 sole with those in other flatfish species like Japanese flounder, summer flounder 1063 (Paralichthys dentatus) and Atlantic halibut, it becomes clear that flatfish larvae present 1064 1065 different developmental timings, and it is likely that the same dietary VA imbalance offered 1066 at the same time after hatching may induce different skeletal deformities or, at least, different deformity incidences and severities for each species (Fernández & Gisbert 2011). 1067 1068 Furthermore, retinoid requirements seem to be quite different between flatfish species; e.g., a safe level for Japanese flounder should be < 50,000 IU kg⁻¹ VA (Dedi et al. 1995), 1069 whereas $< 42,666 \text{ IU kg}^{-1} \text{ VA}$ is advisable for Senegalese sole (Fernández *et al.* 2009). 1070

In a recent study of vitamin K, Richard *et al.* (2014) showed that live prey grown in a commercial enrichment emulsion supplemented with 250 mg kg⁻¹ of vitamin K (phylloquinone) significantly improved larval growth performance, as well as the skeletal quality of post-metamorphic specimens. In particular, authors found that these fish had a lower incidence of deformities and a reduced percentage of fish with malformed/fused haemal and neural arches or spines, and fusions in caudal vertebrae. Dietary supplementation of vitamin K modulated the expression of proteins (MALDI-TOF-TOF
analysis) involved in several biological processes including muscle contraction and
development, cytoskeletal network, skin development, energy metabolism, protein
chaperoning and folding, and bone development (Richard *et al.* 2014).

1081 Flatfishes possess large dorsal and anal fins, supported by a large number of soft rays 1082 that form a semi continuous structure together with the caudal and anal fins, both in adult and larval fish. A significant number of anomalies affecting the fins of Senegalese sole 1083 have been observed, appearing mainly as fusions or anomalies of hypurals in the caudal fin 1084 1085 and as anomalous pterygiophores in the dorsal and anal fins (Gavaia et al. 2002; Engrola et al. 2009a; Fernández et al. 2009; Fernández & Gisbert 2010). The cartilaginous elements, 1086 1087 like those composing the internal skeleton of the dorsal, anal and caudal fins, appear to be more sensitive to high levels of VA or oxidized lipids than dermal bones (Fernández & 1088 Gisbert 2010; Boglino et al. 2014). 1089

1090 What is noteworthy in Senegalese sole is the higher incidence of osteological abnormalities in this species when reared under standard feeding protocols in comparison to 1091 1092 other commonly produced species in the Mediterranean and Atlantic area, like gilthead sea bream or European sea bass (see review in Boglione et al. 2013a). Fernández et al. (2009) 1093 suggested that two different hypotheses might explain this. The first one considered that 1094 1095 this flatfish species is more prone to develop skeletal disorders than other fish species under any rearing conditions, whereas the second hypothesis postulated that since the skeletal 1096 1097 deformities observed in Senegalese sole were not lethal, higher final numbers of Senegalese 1098 sole specimens with deformities would be observed at the juvenile stage than in other species where deformities were lethal at early stages (see review in Koumoundouros 2010). 1099 Since both hypotheses are not mutually exclusive, determining which better explains the 1100

1104

1105 Pigmentation abnormalities

In flatfish species, pigmentation abnormalities are characterized by either a deficiency of 1106 pigment cells on portions of the ocular side (albinism, pseudoalbinism or hypomelanism), 1107 or excess pigmentation on the blind side (staining, spotting, or ambicolouration) (Bolker & 1108 Hill 2000). Pigmentary disorders, especially albinism and ambicoloration, can affect up to 1109 61% of the reared fish of different flatfish species (Estévez & Kanazawa 1995; Estévez et 1110 1111 al. 1999; Bolker & Hill 2000; Villalta et al. 2005b; Hamre et al. 2007; Guillot et al., 2012; Darias et al. 2013a; Boglino et al. 2014). However, pigmentation abnormalities are not as 1112 1113 critical in Senegalese sole as it has been reported in other flatfish species (Seikai & Sinoda 1114 1981; Bolker et al. 2005; Hamre et al. 2007). In fact, under current practical farming conditions malpigmentation in sole seldom occurs. 1115

1116 Most data regarding pigmentation disorders in Senegalese sole are from experimental studies in which pseudoalbinism was induced through a dietary excess of ARA (Villalta et 1117 1118 al. 2005b, 2008b; Boglino et al. 2013; Darias et al. 2013a), although environmental rearing conditions and other nutrients like DHA, EPA and VA have been investigated as factors 1119 possibly related to the occurrence of albinism in hatchery production of other flatfish 1120 species (Bolker & Hill 2000; Hamre & Harboe 2008). Several studies have shown that 1121 when Senegalese sole larvae are fed high dietary ARA levels (7.1 - 10.2% TFA) post-1122 larvae have a high incidence of pigmentary disorders (81.4 – 84.4% of pseudoalbinism), 1123 1124 whereas only 0.3 to 0.9% of malpigmented fish were found when larvae were fed 0.1%

ARA (Villalta et al. 2005b, 2007; Boglino et al. 2013). A likely explanation is that high 1125 1126 ARA levels, by altering the dietary and body ARA/EPA ratio, can modify the relative concentrations of ARA-derived eicosanoids in the developing organism, which might 1127 1128 disrupt the morphogenesis of the skin resulting in pigmentary disorders. In addition, studies 1129 on Soleids have demonstrated that not only dietary levels of ARA, but also the timing of its administration in relation to the larval developmental stage (Lund et al. 2007, 2008; 1130 Boglino et al. 2014), as well as abiotic factors such as tank colour (Lund et al. 2010), can 1131 1132 affect the incidence of pigmentary anomalies. In common sole, a strong light intensity (3600–4500 lx at the water surface) in combination with a transparent tank colour induced 1133 pseudoalbinism, even if on a much lower scale than the observed effects of ARA (Lund et 1134 1135 al. 2010). Boglino et al. (2014) evidenced the existence of a "pigmentation window", with greater larval sensitivity to high dietary ARA levels during pre- and pro-metamorphosis (2-1136 1137 15 dph) than post-metamorphosis (15–50 dph). This was correlated with the aspect and 1138 density of melanophores in the skin of the ocular side, which decreased in the order: normally pigmented individuals > pseudo-albino fish fed high ARA levels during 1139 1140 postmetamorphosis > pseudo-albino fish fed high ARA levels during pre- and prometamorphic stages. 1141

Morphological studies by Darias *et al.* (2013a) in Senegalese sole revealed that ARA did not affect larval pigmentation at the pre-metamorphic stage, but prevented chromatophore terminal differentiation at metamorphosis, leading to the appearance of pseudo-albinism. The authors concluded that the relative proportions between xanthophores and melanophores, cell proximity, size and shape were critical for the correct ontogeny of pigmentation. In this regard, those larvae later becoming pseudo-albino and pigmented individuals developed pigmentation in the same way but, once metamorphosed, the future

pseudo-albinos began to show different relative proportions, allocation patterns, shapes and 1149 1150 sizes of skin chromatophores. In sum, the amount of melanophores and iridophores in pseudo-albinos remained invariable during larval development, whereas normally 1151 1152 pigmented specimens showed an increase in the population of melanophores and 1153 iridophores at post-metamorphosis (47 and 35 dph, respectively), which indicated that the 1154 new population of chromatophores that should appear after metamorphosis was not formed (or cells were not pigmented) in pseudo-albinos. Furthermore, a decrease in xanthophores 1155 1156 was observed from 33 to 35 dph in pseudo-albinos, likely as the result of the degradation of already existent cells. Pseudo-albinos had also lost most of their leucophores, with those 1157 1158 few remaining being located almost exclusively in the distal part of the fins (Darias et al. 2013a,b). In one of these studies, Darias et al. (2013a) has shed some light on the molecular 1159 regulation of some of the events leading to the establishment of the ARA-induced pseudo-1160 1161 albino phenotype, including the expression of several key pigmentation-related genes.

1162 Regarding the establishment of dorsal-ventral pigmentation, pigment cell latent precursors are symmetrically located mainly along the dorsal and ventral margins of the 1163 1164 flank during larval stages, and then migrate from these regions to the lateral sides. After late metamorphic stages, these precursors differentiate into adult-type chromatophores on 1165 1166 the lateral asymmetrical sides. Since the asymmetric body plan, including eye migration, precedes adult pigment pattern formation (Watanabe et al., 2008), pigment asymmetry in 1167 flatfish seems to depend on an asymmetric organizational environment that may regulate 1168 1169 survival, proliferation, distribution and differentiation of latent precursors into adult-type 1170 pigment cells. In this context, the agouti-signalling protein 1 (Asip1) has a key role in proper dorsal-ventral pigment patterning in juvenile and adult fish, since it induces the 1171 1172 regulatory asymmetry involved in precursor differentiation into mature chromatophores.

1176 like differentiation environment (iridophore proliferation) in dorsal regions (Guillot *et al.*,

position, resulting in unbalanced *asip* production levels. These, in turn, generate a ventral-

- 1177 2012; Darias et al., 2013a).
- 1178

1173

1174

1175

- 1179
- 1180 Health issues
- 1181
- 1182 Stress response

1183 Senegalese sole response to acute stress is in line with that previously reported by Barton and Iwama (1991), being the highest increase in plasma cortisol within 0.5-1h after 1184 1185 a stressful disturbance. However, Costas et al. (2011a) observed much higher cortisol 1186 concentrations than those reported in other studies with the same species (López-Olmeda et al. 2013) and those reported in several teleosts following air exposure (Waring et al. 1996; 1187 1188 Arends et al. 1999; Jentoft et al. 2005; Cnaani & McLean 2009). Different cortisol values reported by Costas et al. (2011a) and López-Olmeda et al. (2013) are probably related to 1189 the duration of air exposure (3 min vs. 30 sec, respectively), suggesting that aquaculture-1190 1191 related handling procedures such as grading must be carefully supervised to avoid longterm air exposure and thus higher cortisol release. 1192

1193 Intensively farmed fish may encounter situations or events that they perceive as 1194 potentially threatening such as poor water quality, physical disturbances (i.e. transport, 1195 grading), or sub-optimal stocking densities and social environments (Pottinger 2008). 1196 Cortisol concentrations reported in chronically stressed Senegalese sole reared under

comparable conditions are quantitatively similar (Aragão et al. 2008; Costas et al. 2008, 1197 1198 2012, 2013a; Salas-Leiton et al. 2010; Arjona et al. 2009), and appear to decrease in time, 1199 probably due to a negative feedback of cortisol at the level of the hypothalamus and 1200 pituitary axis, thus modulating adrenocorticotrophic hormone (ACTH) secretion and 1201 consequently cortisol production (Mommsen et al. 1999). Although it is widely accepted that chronic stressful conditions decrease growth in fish and may reduce 1202 immunocompetence leading to enhanced susceptibility to pathogens and parasites 1203 1204 (Wendelaar Bonga 1997), most studies reported that growth in sole is not affected by high 1205 stocking densities and repeated handling stress (Aragão et al. 2008; Costas et al. 2008, 2012, 2013a; Salas-Leiton et al. 2010). Intensively reared Senegalese sole subjected to 1206 different high stocking densities (24 and 30 Kg m⁻² initial stocking densities) initially 1207 showed decreased growth performance (after 30 and 40 days, respectively), developing 1208 1209 thereafter a compensatory growth strategy resulting in no differences in growth after 60 1210 days, compared to specimens reared at low density (Salas-Leiton et al. 2010; Andrade 1211 2012). While most studies presented increased cortisol levels and/or energy metabolism in 1212 parallel with decreased immune parameters and/or eventual mortalities, Andrade (2012) observed no changes in Senegalese sole after 30 and 60 days suggesting that under farming 1213 conditions other chronic stressors related to water quality or rearing system may modulate 1214 1215 the stress response. For instance, Pinto et al. (2007) reported that growth and nutritional state of Senegalese sole juveniles were adversely affected following chronic exposure to 1216 high environmental ammonia (497 mg L^{-1} NH₃) for 52 days. Similarly, Arjona *et al.* (2009) 1217 1218 observed a decrease in growth and feed intake in Senegalese sole reared at salinities lower than 39‰, with the most profound effects observed at 15‰. Therefore, sudden salinity 1219 1220 changes and increases in environmental ammonia represent a potential risk for Senegalese

sole farms and must be avoided to increase productivity, while other intrinsic factors in
intensive sole production such as repeated handling and high stocking density do not seem
to be an issue provided that optimum rearing conditions are guaranteed.

1224 Dietary effects, in particular the effect of ARA/EPA ratio, in the stress response of this 1225 species has also been investigated by Alves-Martins et al. (2011). These authors observed a 1226 quicker recovery from an air exposure acute stress, but not from a repeated stress (determined as the return of cortisol to basal levels) when post-larvae were fed a low 1227 1228 ARA/EPA ratio (0.7). The same authors (Alves-Martins et al. 2013) later found that increases in dietary levels of ARA (up to 1.7% TFA) were positively associated with post-1229 1230 stress cortisol levels, and the highest tested ARA level (2.3% TFA) seemed to slightly 1231 enhance basal cortisol level and alter the response to stress in post-larvae at 35 dph.

On the other hand, differences in Senegalese sole plasma cortisol levels may also be related to genetic factors. Within a single strain or population, variation in stress responses also has a genetic component and some fish may be predisposed to consistently exhibit high or low cortisol responses to stressors (Tort *et al.* 2001). Therefore, selection of phenotypic characters (e.g. high- or low-cortisol responders) could be considered in order to improve production.

1238

1239 Immune response

The vertebrate innate immune system recognises pathogenic and non-pathogenic microorganisms via germline encoded pathogen pattern recognition receptors (PRRs) that sense particular structures of the microorganisms (pathogen-associated molecular patterns, PAMPs) and initiate a well orchestrated immune response (Boltaña *et al.* 2011). The few existing studies on the Senegalese sole immune responses have mainly focused on innate

immunity, particularly on the interactions between bacterial pathogens and parameters 1245 1246 involved in responses to pathogens and stressful rearing conditions. Costas et al. (2013b) reported leucocyte responses to inflammation in both peripheral blood and peritoneal cavity 1247 1248 following challenge with two *Photobacterium damselae* subsp. *piscicida* (*Phdp*) strains 1249 from different geographical origins, thus corroborating the hypothesis of cell migration to the inflammatory focus in fish. In vitro assays showed diverse innate immune responses of 1250 Senegalese sole phagocytes challenged with different strains of either Phdp (PC566.1 and 1251 1252 PP3) or Tenecibaculum maritimum (ACC6.1 and ACC13.1), suggesting that PRRs from 1253 sole macrophages may have detected PAMPs associated with DNA structures unique for 1254 each bacterium isolate (Costas et al. 2013b,c). Lipopolysaccharide, an endotoxin found on 1255 the bacterial cell membrane, is considered to be the prototypical PAMP and was found to increase the level of expression of hepcidin antimicrobial peptide in both intraperitoneally 1256 1257 injected Senegalese sole (Osuna-Jiménez et al. 2009) and primary head-kidney cells culture 1258 (Costas et al. 2013d). Mx protein, an interferon-induced protein that protects against viral infections, has also been studied in Senegalese sole (Fernández-Trujillo et al. 2008a). 1259 1260 Fernández-Trujillo et al. (2008b) reported different Mx expression profiles following both poly I:C injection and solevirus inoculation. Additionally, the Senegalese sole innate 1261 immune machinery also increased following a mixed leucocyte reaction. The level of 1262 1263 expression of interleukin (IL)-1β, IL-8, hepcidin antimicrobial peptide and g-type lysozyme increased after the incubation of blood leucocytes from three different individuals at 24 and 1264 1265 48 h (Costas et al. 2013d).

Several humoral substances and cell secretions also contribute to the natural resistance
of fish to pathogenic and infectious agents. These include complement, transferrins, antiproteases, various lytic enzymes (e.g. lysozyme), lectins, C-reactive protein, interferon and

enzyme inhibitors (Ellis 1999). Moreover, some of these factors, such as lysozyme and 1269 1270 complement, appear to be more potent in fish than in mammals (Ellis 2001). Senegalese sole increased plasma lysozyme and peroxidase activities following challenge with *Phdp*, 1271 1272 consistent with blood neutrophilia and monocytosis and the increase in neutrophils and 1273 macrophage numbers in the peritoneal cavity at 24 h post-challenge (Costas et al. 2013b). This study also showed different responses of the alternative complement pathway 1274 (expressed as ACH50) against two Phdp strains, being higher at 24 h than those reported 1275 1276 for European seabass and gilthead seabream challenged with the same pathogen (Mauri et 1277 al. 2011). Therefore, ACH50 appears to be a mechanism of greater importance, among 1278 other possible responses, against this particular pathogen in Senegalese sole.

1279

1280 The neuroendocrine-immune network

1281 The suppressive effect of stress on the immune system is highly disputable and does not 1282 necessarily translate into decreased resistance to infection in both mammals and fish (Dhabhar 2009; Verburg-van Kemenade et al. 2009). Depending on the duration and 1283 1284 severity of the stressor, increased glucocorticoid levels may enhance innate and adaptive immune responses while similar hormone levels may suppress immune function. Acute and 1285 1286 chronic stressful husbandry conditions induced different responses in several cell-mediated 1287 and humoral innate immune parameters of Senegalese sole. While plasma lysozyme activity decreased at 4 h after air exposure (Costas et al. 2011a) and after 18 days under 1288 1289 high stocking density (Costas et al. 2013a), those levels increased in sole submitted to weekly handling for 14 and 28 days (Costas et al. 2012) or daily handling for 14 days 1290 (Costas et al. 2011b). Similarly, ACH50 was negatively correlated to plasma cortisol levels 1291 1292 in specimens exposed to acute stress or reared at high density (Costas et al. 2011a, 2013a),

whereas daily stressed sole for 14 days presented higher ACH50 values than control fish 1293 1294 (Costas et al. 2011b). Senegalese sole respiratory burst responses to handling stress also 1295 showed different patterns. For instance, the release of reactive oxygen species decreased in 1296 stimulated macrophages from sole submitted to daily handling for 14 days, while nitric 1297 oxide production followed the opposite pattern (Costas et al. 2011b). Although chronically stressed specimens from the latter study showed a higher resistance to *Phdp* than 1298 undisturbed fish, Senegalese sole treated with dexamethasone (a potent glucocorticoid) for 1299 14 days appeared to be more susceptible to the same pathogen (Salas-Leiton et al. 2012). 1300 1301 The modulation of the innate immune system after a stress challenge was similarly observed in several teleosts. For instance, relevant genes associated with acute 1302 1303 inflammation followed similar kinetics and an up-regulation was observed after acute stress or daily handling for 7, 14 and 28 days (Huising et al. 2003; Fast et al. 2008). In contrast, 1304 1305 leucocytes respiratory burst activity decreased in specimens submitted to handling 1306 (Pulsford et al. 1994). Moreover, several in vitro studies demonstrated that cortisol alone inhibits lipopolysaccharides-induced expression of several immune-related genes (Saeij et 1307 1308 al. 2003; Fast et al. 2008; Stolte et al. 2008; Castillo et al. 2009).

1309 In vivo neuroendocrine-immune interactions are thus dependent on the actions of various 1310 hormones (e.g., catecholamines, cortisol, ACTH, β-endorphin) and cytokines (e.g., IL-1β, 1311 IL-6 and TNF- α), as well as on their interactions (Verburg-van Kemenade *et al.* 2009). This could explain the enhanced immune function frequently observed in Senegalese sole 1312 submitted to repeated handling (Costas et al. 2011b, 2012). Other hormones released during 1313 repeated stress responses may influence innate immune mechanisms at a higher degree, 1314 1315 decreasing the suppressive effects of cortisol. In contrast, a different situation probably occurs during constant chronic situations such as high stocking density, where cortisol 1316

plays an immunosuppressive role. This hypothesis is supported by the increased 1317 1318 susceptibility to opportunistic pathogens observed in Senegalese sole reared at high stocking density (Costas et al. 2008), in line with a general decrease in immune function 1319 1320 (Salas-Leiton et al. 2010; Costas et al. 2013a). Although Andrade (2012) did not observe 1321 changes in the immune competence of intensively reared Senegalese sole during 60 days, 1322 especial attention must be given to rearing conditions and handling protocols in order to improve productivity, given that the immune system may be modulated. Further studies 1323 1324 would be instrumental to unravel the mechanisms that Senegalese sole subjected to repeated handling may have adopted during resistance to bacterial challenge. These studies 1325 1326 should consider leucocyte trafficking and redistribution, cytokine kinetics and the release of 1327 hormones other than cortisol.

1328

1329 Current and emergent diseases

1330 One of the main factors that has historically hampered Senegalese sole farming has been the high incidence and intensity of diseases (Padrós et al. 2003; Toranzo et al. 2003). 1331 1332 Although most of the diseases have been described in other species, sole seems quite prone to become infected by cohabitation with other fish species. In the early 90's it was usual to 1333 1334 find gilthead seabream, European seabass or turbot in the same facilities as Senegalese sole. This was a determining factor for the development of diseases in sole and for the economic 1335 viability of the farms, and was one of the reasons explaining the producer's hesitation in 1336 1337 taking on the mass production of this alternative species. Nowadays sole are reared in 1338 specific facilities with controlled environmental factors, highly reducing the risk of transmission. Nonetheless, some diseases are still common in Senegalese sole production 1339 1340 systems, possibly associated to the lack of appropriate standardised rearing techniques and sometimes due to poor husbandry or hygienic conditions of the tanks, or when the
temperature exceeds 22°C (Cañavate 2005). Currently the main pathological problems are
bacterial diseases, mainly Tenacibaculosis (formerly Flexibacteriosis, fin rot or black patch
necrosis), Photobacteriosis (formerly Pasteurellosis) and Vibriosis.

1345 Tenacibaculosis, which is mainly caused by *Tenacibaculum maritimum* (formerly 1346 *Flexibacter maritimum*) can cause significant morbility and mortality in fish farms in many countries, limiting the culture of economically important marine fish species (Santos et al. 1347 1999). The presence of this pathogen in sole in Europe was first described in Scotland from 1348 common sole suffering from the so-called "black patch necrosis" (BPN) (Bernardet et al. 1349 1990), probably the most important problem in the early culture attempts of this species 1350 1351 because of its high incidence (McVicar & White 1979, 1982). Some years later, Cepeda and Santos (2002) isolated for the first time T. maritimum from Senegalese sole in 1352 1353 southwest Spain, where it caused almost 100% mortality of the affected stocks. Affected 1354 sole usually display several external signs including eroded mouth, rotten fins and skin lesions. Although this disease was reported to be highly infectious, it seemed to be both 1355 1356 prevented and controlled by providing a sand substrate in the rearing tanks (McVicar & White 1982). This is paradoxical given that the use of sand in sole farming has been 1357 1358 considered adverse since deficient management and hygiene may favour the emergence of bacterial infections (Howell 1997). However, culturing these fish in smooth hard-bottomed 1359 tanks without sand has been associated to a series of harmful effects (Ottesen & Strand 1360 1361 1996; Ottesen et al. 2007). Recently, Vilar et al. (2012) described particularly severe 1362 ulcerative disease outbreaks in cultured Senegalese sole associated with T. maritimum. Grossly, the affected fish showed total loss of epidermis and dermis and extensive necrosis 1363 1364 of the muscle layers. Many other environmental conditions such as higher temperatures,

salinity, low water quality, excess of UV light and also management factors (i.e. high 1365 1366 density and poor feeding) and host-related factors (stress, skin surface condition) have also been described associated to this disease (Avendaño-Herrera, 2005). Being a disease that 1367 1368 may cause both skin lesions and systemic problems, combined treatments with external 1369 disinfectants (such as formalin or hydrogen peroxide) and oral treatments with antibiotics are usually used. Florfenicol, oxitetracycline, flumequine, potentiated sulfonamides and 1370 especially enrofloxacin were used for controlling T. maritimum outbreaks, although the 1371 rapid appearance of resistant strains has been described (Avendaño-Herrera, 2005). Other 1372 1373 Tenacibaculum species such as T. discolor and T. soleae, have also been isolated from diseased Senegalese sole (Piñeiro-Vidal et al. 2008a,b), showing the typical signs observed 1374 1375 in fish affected by T. maritimum. Regarding vaccination, there is nowadays a commercially available bacterin to prevent the disease caused by T. maritimum in turbot (Icthiovac 1376 1377 TM[®]), which is applied by bath in fish from 1 to 2 g followed by a booster injection in fish 1378 from 20 to 30 g (Avendaño- Herrera et al. 2006). Although no licensed commercial vaccines are yet available for sole, autovaccines made using the strains isolated from the 1379 1380 farms can also be used in this species.

Photobacteriosis, caused by Photobacterium damsela ssp. piscicida, is responsible for 1381 high losses in the aquaculture industry as it provokes massive mortalities in several marine 1382 1383 fish species such as gilthead seabream (Toranzo et al. 1991), seabass (Balebona et al. 1992), and in the flatfish Japanese flounder (Fukuda et al. 1996), among others. Since it 1384 1385 was first recorded in farmed Senegalese sole in southwest Spain (Zorrilla et al. 1999) 1386 several sole farms, mainly in the south of Spain, have suffered mortalities caused by this disease (Magariños et al. 2003). In most cases, peracute mortalities without apparent 1387 1388 lesions are the most typical manifestation found mainly in juveniles. However, in subacute

and chronic cases, external lesions of infected fish included only unspecific symptoms such
as dark skin coloration and swelling of the abdominal cavity. This disease particularly
affects Senegalese sole at temperatures above 18°C and usually triggers severe acute cases
in which mortality can be extremely high (Padrós *et al.* 2003).

1393 Vibrioses affecting Senegalese sole are usually detected as secondary infections 1394 associated with an initial Tenacibaculosis, but often they can also be primary infections and pathogenesis is still unclear (Padrós et al. 2003). Vibrio harveyi and V. 1395 its parahaemolyticus are pathogenic bacteria which were described in an outbreak of farmed 1396 sole by Zorrilla et al. (2003), causing moderate mortalities in the south of Spain. Main 1397 external signs of the disease were skin ulcers and haemorrhagic areas near the fins and 1398 mouth (Zorrilla et al. 2003). Rico et al. (2008) also characterised strains of V. harveyi from 1399 diseased-farmed Senegalese sole in Spain from 2000 to 2004 and, recently, Gomez-Gil et 1400 1401 al. (2012) isolated Vibrio alfacsensis from cultured sole in two regions of Spain.

There are several studies on vaccination against these diseases (e.g. Romalde *et al.* 2005) and a divalent vaccine against *P. damselae* subsp. *piscicida* and *V. harveyi* that provides short-time protection is being studied (Arijo *et al.* 2005). Although no vaccines for sole have been registered at present, autovaccines against *P. damsela* and *Vibrio* species have been used in some farms. In addition, recent studies on probiotics to control *Photobacteriosis* and different *Vibrio* species have given encouraging results (Garcia de la Banda *et al.* 2012; Tapia-Paniagua *et al.* 2012; Batista *et al.* 2013).

Recently, Magariños *et al.* (2011) reported *Aeromona salmonicida* subspecies *salmonicida* as the causative agent of a 'typical' furunculosis outbreak in cultured sole in a marine farm operating in a recirculation system in Galicia. Affected fish showed haemorrhagic areas at the base of the dorsal and ventral fins and, in some cases, ulcerative lesions on the ventral surface. Internally, peritoneal cavities were completely filled with
ascitic fluid and livers were extremely pale and showed petechiae. In this particular case,
soles were grown in a farm which also produced turbot, which pointed towards a potential
crossed infection of this bacterium from one fish species to another.

Similarly, Castro *et al.* (2012) isolated *Edwarsiella tarda* in Senegalese sole growing in a farm which also produced turbot, which again pointed towards a potential crossed infection. Affected fish showed cutaneous lesions in the dorsal surface, tumefactions around the eyes and haemorrhages in their ventral surface, as well as internal lesions abundant ascitic fluid, anaemic liver and kidney with petechial haemorrhages.

Regarding lesions in internal organs, granulomas in the kidney and spleen were observed in this fish species related to acid-resistant bacteria, Ziehl-Neelsen-positive stain (Francesc Padrós, pers. comm., 2013). These bacteria, resembling *Mycobacterium* spp., could represent a potential new hazard for cultured sole. Transmission of mycobacteria in fishes is poorly understood, but water and associated biofilms are natural habitats for *Mycobacterium* spp. (Pedley *et al.* 2004), and therefore recirculation systems may play an important role in its transmission in aquaculture.

Although progress has been made on nutrition and feeding, the lack of specific and standardised diets for some life stages of this species may also facilitate bacterial infections. With respect to this, bacterial enteropathy possibly associated with the administration of contaminated live food has been observed in larvae and juveniles (Padrós *et al.* 2003).

As viral diseases, betanodaviruses, the aetiological agents of the viral nervous necrosis or viral encephalopathy and retinopathy (VER), have also been detected in Senegalese sole (Starkey *et al.* 2001; Thiéry *et al.* 2004; Cutrín *et al.* 2007; Olveira *et al.* 2009; Hodneland *et al.* 2011). Although the typical nervous clinical signs and high mortalities are usually

less common in sole than in other species, fish can show abnormal swimming behaviour 1437 1438 and moderate to high mortalities (Hodneland et al. 2011). However, a serious episode of mortality associated with the presence of VER was detected in juvenile common sole 1439 reared in a farm where it had been previously detected in a batch of croaker, Umbrina 1440 1441 *cirrosa* fry (Borghesan *et al.* 2003). Although this virus can be transmitted horizontally by contact between diseased and healthy fish, the main transmission route is vertical (Baria 1442 2004), which highlights the importance of detecting broodstock carriers that could transmit 1443 the virus to the larvae through fertilised eggs. Currently there are no efficient treatments or 1444 commercial vaccines for nodavirus. The recommendation is that affected stocks are 1445 immediately removed and sacrificed when the virus is detected. However, there are some 1446 1447 promising results from oral, bath and injection vaccination in other fish species such as the sevenband grouper, Epinephelus septemfasciatus (Lin et al. 2007; Kai & Chi 2008; 1448 1449 Nishizawa et al. 2009; Yamashita et al. 2009).

1450 Birnavirus and lymphocystis virus were also detected in cultured sole (Rodríguez et al. 1997; Toranzo et al. 2003; Alonso et al. 2005; Cano et al. 2010). The birnavirus was 1451 1452 described as the agent causing 100% mortality in wild Senegalese sole broodstock introduced into a culture facility in southwest Spain, and its external signs were dark 1453 coloration, hyperactivity and uncoordinated swimming behaviour (Rodríguez et al. 1997). 1454 The characterisation of this virus indicated its similarity to infectious pancreatic necrosis 1455 virus (IPNV). However, both birnavirus and IPNV are widely distributed and affect many 1456 1457 different aquatic organisms, and therefore its role as a primary pathogen in sole species is not so clear. Lymphocystis disease is caused by an iridovirus with a worldwide 1458 geographical distribution that involves a chronic disease characterised by papilloma-like 1459 1460 lesions typically on the skin, fins and tail (Walker & Hill 1980). Although no viral

haemorrhagic septicaemia virus (VHSV) outbreaks have been reported for the time being, 1461 1462 López-Vazquez et al. (2011) demonstrated the susceptibility of Senegalese sole to a VHSV strain isolated from wild Greenland halibut, Reinhardtius hippoglossoides, and farmed 1463 turbot. On the other hand, in wild fish stocks which were held as future breeders, the 1464 1465 presence of intracytoplasmic inclusion bodies in erythrocytes was detected causing low but continuous mortality (Padrós et al. 2003). Until now the presence of viral particles has not 1466 been demonstrated, although the existence of a process similar to VEN (viral erythrocytic 1467 necrosis), or similar processes described in other species, was not ruled out. 1468

In recent years, systemic amoebic disease has become the main parasitic problem in 1469 cultured Senegalese sole. Although the condition was not associated to high mortalities, 1470 1471 reduced growth and high morbidity was noted and fishes show protuberances on the skin surface in addition to unspecific signs of diseases (lethargy with sporadic and erratic 1472 1473 swimming) (Constenla & Padrós 2010). Furthermore, the muscular lesions developed can 1474 later prevent the commercialization of the fish. Endolimax piscium (Archamoeba) is the causative agent of this amoebiasis (Constenla et al. 2014) causing a granulomatous 1475 1476 inflammatory reaction mainly in muscle but also in different internal organs of the host. This parasite was also detected within the intestinal epithelium and submucosa in both 1477 lesioned fishes and apparently healthy fishes (Constenla & Padrós 2010). This location 1478 1479 seems to be an initial stage in the development of the disease and, consequently, early detection of the parasite in the farm should be considered a priority for the management of 1480 1481 this disease in sole culture, since there is no known effective treatment against these 1482 parasites. Amoeboid organisms similar to *Neoparamoeba* sp. have also been observed in cultured Senegalese sole causing a chronic proliferative mucoid inflammation in gills 1483 1484 similar to amoebic gill disease in turbot, with epithelial hyperplasia and fusion of lamellae

1485

1486

in the apical region of some filaments (Francesc Padrós, pers. comm., 2012). At present, the most effective treatment against these gill amoebae seems to be freshwater baths.

Some sporadic infections by protist parasites like flagellates or ciliates (Amyloodinium, 1487 Cryptobia and Cryptocaryon) have also been described (Padrós et al. 2003), usually in 1488 1489 cases where sole were reared in ponds. Albeit infrequently, cases of massive parasitosis can eventually lead to high mortalities, especially in sole grown at high temperatures and in 1490 recirculation systems. Palenzuela et al. (2007) described an infection by Enteromyxum 1491 1492 scophthalmi in sole cohabiting with infected turbot, and other Myxozoa such as E. leei and 1493 Ceratomyxa sp. have also been observed in sole. However, since sole are currently reared in specific facilities, it seems that infections by *Enteromyxum* spp. isolated from highly 1494 1495 susceptible fish species such as gilthead seabream and turbot are not a real threat to sole farming, only a potential risk. Other internal parasites have occasionally been observed in 1496 1497 cultured sole, such as mixosporidia found in renal tubules, xenomas of the microsporidian 1498 Tetramicra sp. in muscle and digenean metacercariae also encysted in the musculature (Padrós et al. 2003). Moreover, ectoparasites such as the leech Hemibdella solea 1499 1500 (Hirudinae) have been identified on broodstock of Senegalese sole in the Virginia Institute of Marine Science (Gloucester Point, USA), but it does not seem to adversely affect the fish 1501 1502 and can be controlled with low salinity (Dinis et al. 1999).

Finally, pathologies of apparently non-infectious origin have also been described by Padrós *et al.* (2003): cell necrosis of the subdermal adipose tissue, related to lipid peroxidation and/or excessive exposure to sunlight, characterized by yellowish areas at the base of dorsal and anal fins which can be associated to secondary bacterial infections; and kidney damage due to the deposition of minerals in the tubular lumen, similar to the processes of nephrocalcinosis. In addition, chronic exophthalmia due to the presence of retrobulbar and periocular bubbles were found in Senegalese soles from ponds under hyperoxic conditions (Salas-Leiton et al. 2009). Affected fish also showed bubbles in gills, causing lamellar obstruction, and under the skin all over the body, and deaths were attributed specially to asphyxia and tissue destruction (Salas-Leiton et al. 2009). Lastly, Senegalese sole seems to be very sensitive to changes in environmental and rearing conditions compared with other farmed species, and therefore even small changes could be more challenging for this species (Francesc Padrós, pers. comm., 2014).

1516

1517 Conclusion and future perspectives

1518 This review clearly demonstrated that important progress has been made in the last 1519 decade towards developing a stronger and sustainable aquaculture industry for Senegalese sole, as a result of a strong and consistent research effort in several biological disciplines. 1520 1521 The industry has now advanced from simply adapting culture protocols and methods that 1522 are routinely used in other marine aquaculture species as it is becoming increasingly clear that Solea spp. have important reproductive, behavioral, nutritional and physiological 1523 1524 particularities that need to be taken into account in the rearing of these species. Some of these particularities result in challenges that are especially difficult to tackle, such as the 1525 reproductive difficulties of G1 stocks and high growth dispersion which, in spite of 1526 considerable advances in knowledge gained on the reproductive biology and behavior of 1527 Senegalese sole in captivity and on its digestive physiology and nutritional requirements, 1528 1529 are still important bottlenecks for its cultivation. On the other hand, a major progress has 1530 been the change in production systems from predominantly earth ponds or salt marshes to dedicated recirculation systems, which has enabled dramatically improving disease issues 1531 1532 in sole farms by eliminating contact with other fish species and enabling a much higher

control of environmental parameters. It has also become apparent that many disease 1533 1534 outbreaks occurred and were intensified when temperatures rise above 20-22°C, which has 1535 led the industry to either procure areas in which the temperature can be easily maintained 1536 below these values or invest in RAS technologies. In addition, recent studies have started 1537 gathering information on the immune system of sole and how the immune response can be modulated or affected by a range of parameters, including the environment and stress. It is 1538 hoped that this knowledge will serve to improve aquaculture practices, leading to improved 1539 1540 well being and reduction of susceptibility to diseases, as well as help develop prophylactic 1541 measures and products, such as vaccines. Moreover, recent research has uncovered an extraordinary capability of Senegalese sole to biosynthesise DHA from EPA which, 1542 1543 possibly associated with lower requirements for LC-PUFA, explains at least partly the 1544 easiness of larval culture and high survival during the hatchery stage, as well as the good 1545 performance results and high flesh quality obtained when on-growing sole with diets 1546 containing high levels of PP and VO. At a time when the global aquaculture industry is struggling with a major sustainability issue related to the need to replace FM and FO in 1547 1548 aquaculture feeds, this unique characteristic represents a very important selling point that differentiates Senegalese sole from other marine aquaculture species. On the other hand, a 1549 lot more is now known regarding the specific nutritional requirements of Senegalese sole 1550 1551 and hopefully this will lead to larval dietary regimes reducing phenotypic abnormalities, as well as to the formulation and commercialization of improved weaning and on-growing 1552 1553 species-specific diets. Advances in knowledge on the digestive physiology and feeding 1554 rhythms of this species are also noteworthy and should help improve weaning protocols and reduce growth dispersion. 1555

Therefore, it is believed that there is now a good knowledge base that should have a 1556 1557 strong impact in reducing many of the classical and specific bottlenecks in Senegalese sole culture. Future research is still required to achieve sustainable industrial production from 1558 G1 stocks of Senegalese sole. This will need a multidisciplinary approach to focus on two 1559 1560 principal aspects: 1) the control and development of male reproductive behaviour from the larvae through to successful courtship and spawning, 2) the control of sperm production 1561 and sperm management. Much work has been done recently that has greatly increased the 1562 understanding of Senegalese sole reproduction and, furthermore, enabled substantial 1563 improvements to protocols that were, however, still insufficient to solve the reproductive 1564 dysfunction of cultured broodstocks. Behavioural research needs to examine social 1565 1566 interactions, the bases of reproductive dominance and development of courtship behaviour. Endocrine research needs to aim at both the control of reproductive behaviour and sperm 1567 1568 production. Chemical communication may also be involved and the olfactory system 1569 should be investigated. Research on larval development will be necessary to determine the influence of external factors (temperature, illumination, nutrition) on sexual differentiation 1570 1571 and sex ratio of the population and on later potential effects on puberty and reproductive performance at adulthood. Research on optimized broodstock, weaning and on-growing 1572 diets will improve performance of farmed sole, make production more cost-effective, and 1573 further reduce quality and welfare problems. 1574

Finally, a noteworthy aspect which will likely shape the next decade of research to come is the arrival of the "genomics era" to aquaculture in general and to flatfishes in particular. In recent years, a strong investment has been made to develop genomic resources for *Solea* spp. which will open up ample possibilities for molecular studies devoted to gain a deeper understanding of biological processes and with potential to also be used in future breeding

programs, to identify molecular markers for traits of economical interest (Cerdà et al. 1580 1581 2008b; Cerdà & Manchado 2013; García-Cegarra *et al.* 2013). An important tool currently 1582 available to the academic community for gene sequence mining is the Solea transcriptome 1583 database (generated from different tissues, developmental stages or stimuli treatments) with 1584 a global assembly containing >1,560 million reads, >694K UniGenes and >337K SNPs (SoleaDB, http://www.juntadeandalucia.es/agriculturaypesca/ifapa/soleadb ifapa/). This 1585 emergent area is expected to boost new research to uncover the molecular basis of many 1586 1587 different physiological processes which might be at the root of some of the main biological bottlenecks that are currently holding up the industry. 1588

1589

1590 Acknowledgements

This review pays tribute to the work carried out by many other researchers that, although 1591 1592 not included as authors in this review, have made a significant contribution to advances in 1593 the research of S. senegalensis, as reflected in the reference list, supported by numerous National and EU-funded projects (too large of a list to be included here). A special word 1594 1595 also to Bari Howell for is life-time contribution to sole farming, and for chairing a series of Sole Workshops, which pushed sole into commercial farming and also inspired this review. 1596 SM holds a Ramón y Cajal post-doctoral contract from the Spanish Ministry of Economy 1597 and Competitiveness (MINECO) and is supported by the European Commission Marie 1598 Curie Actions (FP7-PEOPLE-2010-RG, Project No. 274184). BC and SE are supported by 1599 Fundação para a Ciência e a Tecnologia, Portugal (SFRH/BPD/77210/2011 and 1600 1601 SFRH/BPD/49051/2008, respectively). Author contribution is as follows: SM coordinated the review and all other authors contributed equally. 1602

1603

1604 **References**

1610

Agulleiro MJ, Anguis V, Cañavate JP, Martínez-Rodríguez G, Mylonas CC, Cerdà J (2006)
Induction of spawning of captive reared Senegalese sole (*Solea Senegalensis*) using
different delivery systems for gonadotropin-releasing hormone agonist. *Aquaculture*257: 511–524.

1609 Agulleiro MJ, Scott AP, Duncan N, Mylonas CC, Cerdà J (2007) Treatment of GnRHa

implanted Senegalese sole (Solea senegalensis) with 11-ketoandrostenedione stimulates

- spermatogenesis and increases sperm motility. *Comparative Biochemistry and Physiology A* 147: 885–892.
- 1613 Alonso MC, Cano I, García-Rosado E, Castro D, Lamas J, Barja JL et al. (2005) Isolation
- 1614 of lymphocystis disease virus from sole, *Solea senegalensis* Kaup, and blackspot sea 1615 bream, *Pagellus bogaraveo* (Brünnich). *Journal of Fish Diseases* **28**: 221-228.
- 1616 Alves-Martins D, Engrola S, Morais S, Bandarra N, Coutinho J, Yúfera M et al. (2011)
- 1617 Cortisol response to air exposure in *Solea senegalensis* post-larvae is affected by dietary
- arachidonic acid-to-eicosapentaeonic acid ratio. *Fish Physiology and Biochemistry* 37:
 733-743.
- 1620 Alves-Martins D, Rocha F, Martínez-Rodríguez G, Bell G, Morais S, Castanheira F *et al.*1621 (2012) Teleost fish larvae adapt to dietary arachidonic acid supply through modulation
- of the expression of lipid metabolism and stress response genes. *British Journal of Nutrition* 108: 864-874.
- 1624 Alves-Martins D, Rocha F, Castanheira F, Mendes A, Pousão-Ferreira P, Bandarra N et al.
- 1625 (2013) Effects of dietary arachidonic acid on cortisol production and gene expression in
- stress response of Senegaleses ole (*Solea senegalensis*) post-larvae. *Fish Physiology and*
- 1627 *Biochemistry*, **39**: 1223-1238.

- Ambrosio PP, Costa C, Sánchez P, Flos R (2008) Stocking density and its influence on
 shape of Senegalese sole adults. *Aquaculture International* 16: 333–343.
- 1630 Andrade T (2012) Effects of different stocking densities on growth, feed-intake, oxidative
- stress status and humoral immune parameters of Senegalese sole (*Solea senegalensis*)
- 1632 juveniles. Master Thesis, University of Porto, Portugal.
- Anguis V, Cañavate JP (2005) Spawning of captive Senegal sole (*Solea senegalensis*)
 under a naturally fluctuating temperature regime. *Aquaculture* 243: 133-145.
- 1635 APROMAR (2014) La Acuicultura en España 2014. Asociación Empresarial de
- 1636 Productores de Cultivos Marinos de España (APROMAR) y la Asociación Española de
- 1637 Productores de Acuicultura Continental (ESCUA) (www.apromar.es; www.esacua.com).
- 1638 Aragão C, Conceição LEC, Fyhn HJ, Dinis MT (2004) Estimated amino acid requirements
- during early ontogeny in fish with different life styles: gilthead seabream (Sparus
- aurata) and Senegalese sole (*Solea senegalensis*). *Aquaculture* **242**: 589-605.
- 1641 Aragão C, Corte-Real J, Costas B, Dinis MT, Conceição LEC (2008) Stress response and
- 1642 changes in amino acid requirements in Senegalese sole *Solea senegalensis* Kaup 1758.
- 1643 *Amino Acids* **34**: 143–148.
- 1644 Aragão C, Pinto W, Dinis MT (2010) Dietary tyrosine utilisation in flatfish: the choice
- 1645 between metamorphosis and stress response. In: *Aquaculture Europe 10 Seafarming*
- *tomorrow*, pp. 133-134. European Aquaculture Society, Porto, Portugal.
- 1647 Arends RJ, Mancera JM, Muñoz JL, Wendelaar Bonga SE, Flik G (1999) The stress
- 1648 response of the gilthead sea-bream (*Sparus aurata* L.) to air exposure and confinement.
- 1649 *Journal of Endocrinology* **163**: 149–157.
- 1650 Arijo S, Rico R, Chabrillon M, Diaz-Rosales P, Martínez-Manzanares E, Balelona MC et
- 1651 al. (2005) Effectiveness of a divalent vaccine for sole, Solea senegalensis (Kaup),

against Vibrio harveyi and Photobacterium damselae subsp. piscicida. Journal of Fish
Diseases 28: 33-38.

- Arjona FJ, Vargas-Chacoff L, Ruiz-Jarabo I, Martín del Rio MP, Mancera JM (2007)
 Osmoregulatory response of Senegalese sole (*Solea senegalensis*) to changes in
- 1656 environmental salinity. *Comparative Biochemistry and Physiology Part A: Molecular*
- 1657 *and Integrative Physiology* **148**: 413–421.
- 1658 Arjona FJ, Vargas-Chacoff L, Ruiz-Jarabo I, Gonçalves O, Páscoa I, Martín del Rio MP et
- 1659 *al.* (2009) Tertiary stress responses in Senegalese sole (*Solea senegalensis* Kaup, 1858)
- to osmotic challenge: Implications for osmoregulation, energy metabolism and growth.
- 1661 *Aquaculture* **287**: 419–426.
- Avendaño-Herrera R (2005) Avances en el conocimiento del patógeno de peces
 Tenacibaculum maritimum: implicaciones en el diagnóstico y prevención de la
 enfermedad. PhD thesis, Universidad de Santiago de Compostela
- 1665 Avendaño-Herrera R, Toranzo AE, Magariños B (2006) Tenacibaculosis infection in
- 1666 marine fish caused by *Tenacibaculum maritimum*: a review. *Diseases of aquatic*1667 organisms **71**: 255-266.
- 1668 Balebona MC, Moriñigo MA, Sedano J, Martinez-Manzanares E, Vidaurreta A, Borrego JJ
- 1669 et al. (1992) Isolation of Pasteurella piscicida from Seabass in southwestern Spain.
- 1670 Bulletin of European association of fish pathology **12** (1): 168-170.
- 1671 Barja JL (2004) Report about fish viral diseases. In : Alvarez-Pellitero P, Barja JL, Basurco
- 1672 B, Berthe F, Toranzo AE (eds.) *Mediterranean aquaculture diagnostic laboratories*, pp.
- 1673 91-102. CIHEAM, Zaragoza.
- 1674 Batista S, Tapia-Paniagua ST, Morinigo MA, Nunez-Diaz JA, Goncalves JFM, Barros R et
- 1675 al. (2013) Expression of immune response genes in sole (Solea senegalensis, Kaup

- 1676 1858) induced by dietary probiotic supplementation following exposure to
 1677 *Photobacterium damselae* subsp. *piscicida*. *Fish & shellfish immunology* **34** (6): 16381678 1639.
- 1679 Barton BA, Iwama IW (1991) Physiological changes in fish from stress in aquaculture with
- 1680 emphasis on the response and effects of corticosteriods. *Annual Reviews of Fish*1681 *Diseases* 1: 3–26.
- 1682 Bayarri MJ, Muñoz-Cueto JA, López-Olmeda JF, Vera LM, Rol de Lama MA, Madrid JA
- *et al.* (2004) Daily locomotor activity and melatonin rhythms in Senegal sole (*Solea senegalensis*). *Physiology & Behavior* 81: 577–583.
- 1685 Bayarri MJ, Guzmán JM, Ramos J, Piquer V, Mañanós E (2011) Annual variations of
- maturation inducing steroid in two cultured generations of Senegalese sole, Solea
 senegalensis. In: Proceedings 9th International Symposium on Reproductive Physiology
- 1688 of Fish, pp. 120-121, Thrissur (Cochin), India.
- Baynes SM, Howell BR, Beard TW, Hallam JD (1994) A description of the spawning
 behavior of captive Dover sole, *Solea solea* (L.). *Netherlands Journal of Sea Research*32: 271-275.
- Baynes SM, Howell BR (1993) Observations on the growth, survival and disease resistance
 of juvenile common sole, *Solea solea* (L.), fed *Mytilus edulis* L. *Aquaculture and Fisheries Management* 24: 95-100.
- Benítez-Dorta V, Caballero M, Izquierdo M, Manchado M, Infante C, Zamorano M *et al.*(2013) Total substitution of fish oil by vegetable oils in Senegalese sole (*Solea senegalensis*) diets: effects on fish performance, biochemical composition, and
 expression of some glucocorticoid receptor-related genes. *Fish Physiology and Biochemistry* 39: 335-349.

1700	Beirão J.	Soares F.	, Herráez MI	P. Dinis MT	. Cabrita E	(2009)	Sperm qu	uality	evaluation	in

- Solea senegalensis during the reproductive season at cellular level. *Theriogenology* 72:
 1251–1261.
- 1703 Beirão J, Soares F, Herráez MP, Dinis MT, Cabrita E (2011) Changes in Solea senegalensis
- sperm quality throughout the year. *Animal Reproduction Science* **126**: 122–129.
- 1705 Beirão J, Soares F, Pousão-Ferreira P, Diogo P, Dias J, Dinis MT, Herráez MP, Cabrita E
- (2015) The effect of enriched diets on *Solea senegalensis* sperm quality. *Aquaculture*435: 187-194.
- 1708 Bernardet JF, Campbell AC, Buswell JA (1990) Flexibacter maritimus is the agent of
- 1709 "black patch necrosis" in Dover sole in Scotland. *Diseases of Aquatic Organisms* 8: 2331710 237.
- Bjørndal T, Guillen J (2014) The future of sole farming in Europe: Cost of production and
 markets. *Aquaculture Europe* 39(2): 5-12.
- 1713 Blanco-Vives B, Villamizar N, Ramos J, Bayarri MJ, Chereguini O, Sánchez-Vázquez FJ
- 1714 (2010) Effect of daily thermo- and photo-cycles of different light spectrum on the
- development of Senegal sole (*Solea senegalensis*) larvae. *Aquaculture* **306**: 137–145.
- 1716 Blanco-Vives B, Vera LM, Mañanós E, Ramos J, Bayarri MJ, Sánchez-Vázquez FJ (2011)
- Exposure of larvae to daily thermocycles affects gonad development, sex ratio and
 sexual steroids in *Solea senegalensis*, Kaup. *Journal of Experimental Zoology* **315**: 162-
- 1719 169.
- 1720 Blanco-Vives B, Aliaga-Guerrero M, Cañavate JP, García-Mateos G, Martín-Robles AJ,
- 1721 Herrera-Pérez P et al. (2012) Metamorphosis induces a light-dependent switch in
- 1722 Senegalese sole (*Solea senegalensis*, Kaup) from diurnal to nocturnal behavior. *Journal*
- 1723 *of Biological Rhythms* **27**:135–145.

Boglino A. Darias MJ, Ortiz-Delgado JB, Özcan F, Estévez A, Andree KB *et al.* (2012a)
Commercial products for *Artemia* enrichment affect growth performance, digestive
system maturation, ossification and incidence of skeletal deformities in Senegalese sole
(*Solea senegalensis*) larvae. *Aquaculture* 324-325: 290–302.

- Boglino A, Gisbert E, Darias MJ, Estévez A, Andree KB, Sarasquete C *et al.* (2012b)
 Isolipidic diets differing in their essential fatty acid profiles affect the deposition of
 unsaturated neutral lipids in the intestine, liver and vascular system of Senegalese sole
 larvae and early juveniles. *Comparative Biochemistry and Physiology, Part A* 162: 59–
 70.
- Boglino A, Darias MJ, Estévez A, Andree KB, Gisbert E (2012c) The effect of dietary
 arachidonic acid during the Artemia feeding period on larval growth and skeletogenesis
 in Senegalese sole, *Solea senegalensis. Journal of Applied Ichthyology* 28: 411–418.
- 1736 Boglino A, Wishkerman A, Darias MJ, Andree KB, De la Iglesia P, Estévez A et al. (2013)
- 1737 High dietary arachidonic acid levels affect the process of eye migration and head shape
- in pseudo-albino Senegalese sole *Solea senegalensis* early juveniles. *Journal of Fish Biology* 83: 1302–1320.
- 1740 Boglino A, Wishkerman A, Darias MJ, de la Iglesia P, Andree KB, Gisbert E et al. (2014)
- 1741 Senegalese sole (*Solea senegalensis*) metamorphic larvae are more sensitive to pseudo-
- albinism induced by high dietary arachidonic acid levels than post-metamorphic larvae.
- 1743 *Aquaculture* doi: 10.1016/j.aquaculture.2014.06.012
- 1744 Boglione C, Gisbert E, Gavaia P, Witten PE, Moren M, Fontagné S et al. (2013a) A review
- 1745 on skeletal anomalies in reared European larvae and juveniles. Part 2: Main typologies,
- 1746 occurrences and causative factors. *Reviews in Aquaculture* **5**: 1-47.

- 1747 Boglione C, Gavaia P, Koumoundouros G, Gisbert E, Moren M, Fontagné S et al. (2013b)
- 1748 A review on skeletal anomalies in reared European fish larvae and juveniles. 1: normal
- and anomalous skeletogenic processes. *Reviews in Aquaculture* **5**: 99–120.
- 1750 Bolker JA, Hill CR (2000) Pigmentation development in hatchery-reared flatfishes. Journal
- 1751 *of Fish Biology* **56**: 1029–1052.
- Bolker JA, Hakala TF, Quist JE (2005) Pigmentation development, defects, and patterning
 in summer flounder (*Paralichthys dentatus*). *Zoology* 108: 183–193.
- 1754 Boltaña S, Roher N, Goetz FW, MacKenzie SA (2011) PAMPs, PRRs and the genomics of
- gram negative bacterial recognition in fish. *Developmental and Comparative Immunology* 35: 1195–1203.
- 1757 Boluda Navarro D, Rubio VC, Luz RK, Madrid JA, Sánchez-Vázquez FJ (2009) Daily
- feeding rhythms of Senegalese sole under laboratory and farming conditions using selffeeding systems. *Aquaculture* 291: 130–135.
- 1760 Borges P, Oliveira B, Casal S, Dias J, Conceição L, Valente LMP (2009) Dietary lipid level
- affects growth performance and nutrient utilisation of Senegalese sole (*Solea senegalensis*) juveniles. *British Journal of Nutrition* 102: 1007-1014.
- 1763 Borges P, Medale F, Dias J, Valente LMP (2013a) Protein utilisation and intermediary
- 1764 metabolism of Senegalese sole (*Solea senegalensis*) as a function of protein:lipid ratio.
- 1765 *British Journal of Nutrition* **109:** 1373-1381.
- 1766 Borges P, Medale F, Veron V, Pires MDA, Dias J, Valente LMP (2013b) Lipid digestion,
- absorption and uptake in Solea senegalensis. *Comparative Biochemistry and Physiology*
- 1768 *Part A: Molecular & Integrative Physiology* **166:** 26-35.

- 1769 Borges P, Valente LMP, Veron V, Dias J, Panserat S, Medale F (2014a) High dietary lipid
- 1770 level is associated with persistent hyperglycaemia and downregulation of muscle Akt-
- 1771 mTOR pathway in Senegalese sole (*Solea senegalensis*). PLoS ONE, *in press*.
- 1772 Borges P, Reis B, Fernandes TJR, Palmas Â, Castro-Cunha M, Médale F *et al.* (2014b)
- 1773 Senegalese sole juveniles can cope with diets devoid of supplemental fish oil while
- 1774 preserving flesh nutritional value. *Aquaculture* **418–419:** 116-125.
- 1775 Borghesan F, Palazzi R, Zanella L, Maltese C, Vascellari M, Mutinelli F et al. (2003)
- 1776 Encefalo-retinopatia nella sogliola comune (Solea solea). X Convegno Nazionale SIPI,
- 1777 Teramo, Octobre 9 to11, p. 26.
- 1778 Bromage N, Porter M, Randall C (2001) The environmental regulation of maturation in
- 1779 farmed finfish with special reference to the role of photoperiod and melatonin.
 1780 *Aquaculture* 197: 63-98.
- 1781 Cabral EM, Bacelar M, Batista S, Castro-Cunha M, Ozório ROA, Valente LMP (2011)
- 1782 Replacement of fishmeal by increasing levels of plant protein blends in diets for
 1783 Senegalese sole (*Solea senegalensis*) juveniles. *Aquaculture* 322–323: 74-81.
- 1784 Cabral EM, Fernandes TJR, Campos SD, Castro-Cunha M, Oliveira MBPP, Cunha LM et
- 1785 *al.* (2013) Replacement of fish meal by plant protein sources up to 75% induces good
- growth performance without affecting flesh quality in ongrowing Senegalese sole. *Aquaculture* 380–383: 130-138.
- 1788 Cabral HN (2000) Comparative feeding ecology sympatric *Solea solea* and *S. senegalensis*,
- 1789 within the nursery areas of the Tagus estuary, Portugal. *Journal of Fish Biology* 57:
- 1790 1550–1562.

- 1791 Cabrita E, Soares F, Dinis MT (2006) Characterization of Senegalese sole, *Solea*1792 *senegalensis*, male broodstock in terms of sperm production and quality. *Aquaculture*1793 261: 967–975.
- 1794 Cabrita E, Soares F, Beirão J, García-López A, Martínez-Rodríguez G, Dinis MT (2011)
- 1795 Endocrine and milt response of Senegalese sole, Solea senegalensis, males maintained
- in captivity. *Theriogenology* **75**: 1–9.
- 1797 Campos C, Valente LMP, Borges P, Bizuayehu T, Fernandes JMO (2010) Dietary lipid
- 1798 levels have a remarkable impact on the expression of growth-related genes in Senegalese
- sole (*Solea senegalensis* Kaup). *Journal of Experimental Biology* **213:** 200-209.
- 1800 Campos C, Castanheira MF, Engrola S, Valente LMP, Fernandes JMO, Conceição LEC
- (2013a) Rearing temperature affects Senegalese sole (*Solea senegalensis*) larvae protein
 metabolic capacity. *Fish Physiology and Biochemistry* **39**: 1485-1496.
- 1803 Campos C, Valente LMP, Conceicao LEC, Engrola S, Fernandes JMO (2013b)
 1804 Temperature affects methylation of the myogenin putative promoter, its expression and
- 1805 muscle cellularity in Senegalese sole larvae. *Epigenetics* **8**: 389-397.
- 1806 Campos C, Valente LMP, Conceição LEC, Engrola S, Sousa V, Rocha E et al. (2013c)
- 1807 Incubation temperature induces changes in muscle cellularity and gene expression in
 1808 Senegalese sole (*Solea senegalensis*). *Gene* 516: 209-217.
- 1809 Cano I, Valverde EJ, Lopez-Jimena B, Alonso MC, Garcia-Rosado E, Sarasquete C et al.
- 1810 (2010) A new genotype of Lymphocystivirus isolated from cultured gilthead seabream,
- 1811 Sparus aurata L., and Senegalese sole, Solea senegalensis (Kaup). Journal of fish
- 1812 *diseases* **33** (8): 695-700.
- 1813 Cañavate JP (2005) Opciones del lenguado senegalés Solea senegalensis Kaup, 1858 para
- 1814 diversificar la acuicultura marina. Boletín del Instituto Español de Oceanografía 21 (1-

1815 4): 147-154.

1816 Carazo I, Martin I, Hubbard P, Chereguini O, Mañanós E, Canário A *et al.* (2011)
1817 Reproductive behaviour, the absence of reproductive behaviour in cultured (G1
1818 generation) and chemical communication in the Senegalese sole (*Solea senegalensis*).

1819 *Indian Journal of Science Technology* **4**: 96-97.

1820 Carazo I (2013) Reproductive behaviour and physiology of Senegalese sole, (Solea
1821 senegalensis) broodstock in captivity. PhD Thesis, University of Barcelona, Spain.

1822 Carazo I, Norambuena F, Oliveira C, Sánchez-Vázquez FJ, Duncan NJ (2013) The effect of
1823 night illumination, red and infrared light, on locomotor activity, behaviour and
1824 melatonin of Senegalese sole (*Solea senegalensis*) broodstock. *Physiology & Behavior*

1825 118: 201–207.

1826 Cardeira J, Bensimon-Brito A, Pousão-Ferreira P, Cancela ML, Gavaia PJ (2012) Lordotic-

1827 kyphotic vertebrae develop ectopic cartilage-like tissue in Senegalese sole (*Solea*1828 senegalensis). Journal of Applied Ichthyology 28: 460–463.

- 1829 Castanheira MF, Martins CIM, Engrola S (2011) Daily oxygen consumption rhythms of
- 1830 Senegalese sole *Solea senegalensis* (Kaup, 1858) juveniles. *Journal of Experimental*
- 1831 *Marine Biology and Ecology* **407**: 1–5.
- 1832 Castillo J, Teles M, Mackenzie S, Tort L (2009) Stress-related hormones modulate cytokine
 1833 expression in the head kidney of gilthead seabream (*Sparus aurata*). *Fish and Shellfish*
- 1834 *Immunology* **27**: 493–499.
- 1835 Castro N, Toranzo AE, Devesa S, González A, Núñez S, Magariños B (2012) First
 1836 description of *Edwardsiella tarda* in Senegalese sole, *Solea senegalensis* (Kaup).
 1837 *Journal of Fish Diseases* 35: 79-82.
- 1838 Cepeda C, Santos Y (2002) First isolation of Flexibacter maritimus from farmed

- 1839 Senegalese sole (Solea *senegalensis*, Kaup) in Spain. *Bulletin of European Association*1840 *Fish Pathologists* 22: 388-392.
- 1841 Cerdà J, Manchado, M (2013) Advances in genomics for flatfish aquaculture. *Genes & Nutrition* 8: 5-17.
- 1843 Cerdà J, Chauvigné F, Agulleiro MJ, Marin E, Halm S, Martínez-Rodríguez G et al.
- 1844 (2008a) Molecular cloning of Senegalese sole (*Solea senegalensis*) follicle-stimulating
- hormone and luteinizing hormone subunits and expression pattern during
 spermatogenesis. *General and Comparative Endocrinology* 156: 470-481.
- 1847 Cerdà J, Mercadé J, Lozano JJ, Manchado M, Tingaud-Sequeira A, Astola A et al. (2008b)
- 1848 Genomic resources for a commercial flatfish, the Senegalese sole (*Solea senegalensis*):
- 1849 EST sequencing, oligo microarray design, and development of the bioinformatic
 1850 platform Soleamold. *BMC Genomics* 2008: 9,508.
- 1851 Chauvigné F, Verdura S, Mazon MJ, Duncan N, Zanuy S, Gomez A et al. (2012) Follicle-
- 1852 stimulating hormone and luteinizing hormone mediate the androgenic pathway in leydig
- 1853 cells of an evolutionary advanced teleost. *Biology of Reproduction* **87**: 35.
- 1854 Chauvigné F, Zapater C, Gasol JM, Cerdà J (2014) Germ-line activation of the luteinizing
- 1855 hormone receptor directly drives spermiogenesis in a nonmammalian vertebrate.
- 1856 *Proceedings of the National Academy of Sciences of the United States of America* **111**:
- 1857 1427-1432.
- 1858 Cnaani A, McLean E (2009) Time-course response of cobia (*Rachycentron canadum*) to
 1859 acute stress. *Aquaculture* 289: 140–142.
- 1860 Conceição LEC, Ribeiro L, Engrola S, Aragão C, Morais S, Lacuisse M et al. (2007)
- 1861 Nutritional physiology during development of Senegalese sole (*Solea senegalensis*)
- 1862 *Aquaculture* **268**: 64–81.

- 1863 Conceição LEC, Aragão C, Richard N, Engrola S, Gavaia P, Mira S et al. (2010) Novel
- methodologies in marine fish larval nutrition. *Fish Physiology and Biochemistry* 36: 116.
- 1866 Conceição LEC, Aragão C, Rønnestad I (2011) Proteins. In: Holt J (ed.) Larval Fish
 1867 Nutrition, pp. 83-116. John Wiley & Sons, Inc., UK.
- 1868 Constenla M, Padrós F (2010) Histopathological and ultrastructural studies on a novel
 pathological condition in *Solea senegalensis*. *Diseases of Aquatic Organisms* 90: 1911870 196.
- 1871 Constenla M, Padrós F, Palenzuela O (2014) Endolimax piscium sp. nov. (Amoebozoa),
- 1872 causative agent of systemic granulomatous disease of cultured sole, *Solea senegalensis*
- 1873 Kaup. Journal of Fish Diseases **37**: 229-240.
- 1874 Costas B, Aragão C, Mancera JM, Dinis MT, Conceição LEC (2008) High stocking density
- 1875 induces crowding stress and affects amino acid metabolism in Senegalese sole *Solea*
- 1876 *senegalensis* (Kaup 1858) juveniles. *Aquaculture Research* **39**: 1–9.
- 1877 Costas B (2011) Stress mitigation in sole (Solea senegalensis) through improved nitrogen
- 1878 *nutrition: amino acid utilization, disease resistance and immune status.* PhD Thesis,
- 1879 University of Porto, Portugal.
- 1880 Costas B, Conceição LEC, Aragão C, Martos JA, Ruiz-Jarabo I, Mancera JM et al. (2011a)
- 1881 Physiological responses of Senegalese sole (*Solea senegalensis* Kaup, 1858) after stress
- 1882 challenge: effects on non-specific immune parameters, plasma free amino acids and
- 1883 energy metabolism. *Aquaculture* **316**: 68–76.
- 1884 Costas B, Conceição LEC, Dias J, Novoa B, Figueras A, Afonso A (2011b) Dietary
- arginine and repeated handling increase disease resistance and modulate innate immune

- mechanisms of Senegalese sole (*Solea senegalensis* Kaup, 1858). *Fish and Shellfish Immunology* 31: 838–847.
- 1888 Costas B, Aragão C, Soengas JL, Míguez JM, Rema P, Dias J *et al.* (2012) Effects of
 1889 dietary amino acids and repeated handling on stress response and brain monoaminergic
 1890 neurotransmitters in Senegalese sole (*Solea senegalensis* Kaup, 1858) juveniles.
 1891 *Comparative Biochemistry and Physiology Part A* 161: 18–26.
- 1892 Costas B, Aragão C, Dias J, Afonso A, Conceição LEC (2013a) Interactive effects of a high
 1893 quality protein diet and high stocking density on the stress response and some innate
 1894 immune parameters of Senegalese sole *Solea senegalensis*. *Fish Physiology and*1895 *Biochemistry* **39**: 1141–1151.
- 1896 Costas B, Rêgo PCNP, Simões I, Marques JF, Castro-Cunha M, Afonso A (2013b) Cellular
- and humoral immune responses of Senegalese sole (*Solea senegalensis* Kaup, 1858)
- 1898 following challenge with two *Photobacterium damselae* subsp. *piscicida* strains from
- different geographical origins. *Journal of Fish Diseases* **36**: 543–553.
- Costas B, Simões I, Castro-Cunha M, Afonso A (2013c) Non-specific immune responses of
 Senegalese sole, *Solea senegalensis* (Kaup), head-kidney leucocytes against
 Tenacibaculum maritimum. Journal of Fish Diseases, doi: 10.1111/jfd.12171
- Costas B, Simões I, Castro-Cunha M, Afonso A (2013d) Antimicrobial responses of
 Senegalese sole (*Solea senegalensis*) primary head-kidney leucocytes against *Tenacibaculum maritimum. Fish and Shellfish Immunology* 34: 1702–1703.
- 1906 Cutrín JM, Dopazo CP, Thiéry R, Leao P, Olveira JG, Barja JL et al. (2007) Emergence of
- 1907 pathogenic betanodavirus belonging to the SJNNV genogroup in farmed fish species
- 1908 from the Iberian Peninsula. *Journal of Fish Diseases* **30**: 225-232.

	010) Lack of essential fatty acids in live feed during larval and post-larval
1911 rearing: eff	
	fect on the performance of juvenile Solea senegalensis. Aquaculture
1912 Internationa	<i>al</i> 18 : 741–757.

- 1913 Darias MJ, Andree KB, Boglino A, Rotllant J, Cerdá-Reverter JM, Estévez A *et al.* (2013a)
- Morphological and molecular characterization of dietary-induced pseudo-albinism
 during post-embryonic development of *Solea senegalensis* (Kaup, 1858). PLoS ONE 8:
 e68844.
- 1917 Darias MJ, Andree KB, Boglino A, Fernández I, Estévez A, Gisbert E (2013b) Coordinated
- 1918 regulation of chromatophore differentiation and melanogenesis during the ontogeny of
- skin pigmentation of *Solea senegalensis* (Kaup, 1858). PLoS ONE 8: e63005.
- 1920 Dedi J, Takeuchi T, Seikai T, Watanabe T (1995) Hypervitaminosis and safe levels of
- 1921 vitamin A for larval Japanese flounder (*Paralichthys olivaceus*) fed Artemia nauplii.
- **1922** *Aquaculture* **133**: 135–146.
- 1923 De Groot SJ (1971) On the interrelationships between morphology of the alimentary tract,
- 1924 food and feeding behaviour in flatfishes (PISCES: Pleuronectiformes). *Netherlands*
- 1925 *Journal of Sea Research* **5**: 121–196.
- 1926 Deschamps M-H, Labbe L, Baloche S, Fouchereau-Peron M, Dufour S, Sire J-J (2009)
- 1927 Sustained exercise improves vertebral histomorphometry and modulates hormonal levels
- in rainbow trout. *Aquaculture* **296**: 337–346.
- 1929 Dhabhar FS (2009) A hassle a day may keep the pathogens away: The fight-or-flight stress
- 1930 response and the augmentation of immune function. Integrative and Comparative
- 1931 *Biology* **49**: 215–236.

1932	Dias J, Rueda-Jasso R	, Panserat S, Co	nceição LEC, Gome	es EF, Dinis MT	(2004)) Effect of
------	-----------------------	------------------	-------------------	-----------------	--------	-------------

- dietary carbohydrate-to-lipid ratios on growth, lipid deposition and metabolic hepatic
- 1934 enzymes in juvenile Senegalese sole (*Solea senegalensis*, Kaup). *Aquaculture Research*
- **35:** 1122-1130.
- 1936 Dias J, Yúfera M, Valente LMP, Rema P (2010) Feed transit and apparent protein,
- phosphorus and energy digestibility of practical feed ingredients by Senegalese sole
 (*Solea senegalensis*). *Aquaculture* 302: 94-99.
- 1939 Dinis MT (1986) Quatre Soleidae de l'estuaire du Tage. Reproduction et croissance. Essai
- 1940 d'élevage de Solea senegalensis. Thèse D'État ès-Sciences Naturelles, Université de
- 1941 Bretagne Occidentale, Brest, France.
- Dinis MT, Ribeiro L, Soares F, Sarasquete MC (1999) A review on the cultivation potential
 of *Solea senegalensis* in Spain and Portugal. *Aquaculture* 176: 27-38.
- 1944 Duncan NJ, Sonesson AK, Chavanne H (2013) Principles of finfish broodstock
- 1945 management in aquaculture: control of reproduction and genetic improvement. In: Allan
- 1946 G, Burnell G (ed.) Advances in Aquaculture Hatchery Technology, pp. 23-75.
- 1947 Woodhead Publishing Limited, Cambridge, UK.
- 1948 EFSA (2010) Scientific Opinion on Dietary Reference Values for fats, including saturated
- 1949 fatty acids, polyunsaturated fatty acids, monounsaturated fatty acids, trans fatty acids,
- and cholesterol. *EFSA Journal* **8:** 107.
- 1951 Ellis AE (1999) Immunity to bacteria in fish. *Fish and Shellfish Immunology* **9**: 291–308.
- 1952 Ellis AE (2001) Innate host defense mechanisms of fish against viruses and bacteria.
- 1953 *Developmental and Comparative Immunology* **25**: 827–839.

- 1954 Engrola S, Conceição LEC, Dias L, Pereire R, Ribeiro L, Dinis MT (2007) Improving
- weaning strategies for Senegalese sole: effects of body weight and digestive capacity. *Aquaculture Research* 38: 696-707.
- 1957 Engrola S, Figueira L, Conceição LEC, Gavaia PJ, Ribeiro L, Dinis MT (2009a) Co-
- 1958 feeding in Senegalese sole larvae with inert diet from mouth opening promotes growth at
- 1959 weaning. *Aquaculture* **288**: 264-272.
- 1960 Engrola S, Mai M, Dinis MT, Conceição LEC (2009b) Co-feeding of inert diet from mouth
- opening does not impair protein utilization by Senegalese sole larvae. *Aquaculture* 287:
 185-190.
- 1963 Engrola S, Dinis MT, Conceição LEC (2010) Senegalese sole larvae growth and protein
- utilization is depressed when co-fed high levels of inert diet and *Artemia* since first
 feeding. *Aquaculture Nutrition* 16: 457-465.
- 1966 Estévez A, Kanazawa A (1995) Effect of (n-3) PUFA and vitamin A Artemia enrichment
- 1967 on pigmentation success of turbot, *Scophthalmus maximus* (L.). *Aquaculture Nutrition* **1**:
- 1968 159–168.
- 1969 Estévez A, McEvoy LA, Bell JG, Sargent JR (1999) Growth, survival, lipid composition
- 1970 and pigmentation of turbot (Scophthalmus maximus) larvae fed live-prey enriched in
- arachidonic and eicosapentaenoic acids. *Aquaculture* **180**: 321–343.
- 1972 FAO (2006) available on http://www.fao.org/fishery/statistics/en.
- 1973 FAO (2011) World aquaculture 2010. FAO Fisheries and Aquaculture Department,
- 1974 *Technical Paper*. No. 500/1, FAO, Rome.
- 1975 Fast MD, Hosoya S, Johnson SC, Afonso LOB (2008) Cortisol response and immune-
- 1976 related effects of Atlantic salmon (Salmo salar Linnaeus) subjected to short- and long-
- term stress. *Fish and Shellfish Immunology* **24**: 194–204.

- 1978 FEAP (2013) European Aquaculture Production Report 2003-2012. Federation of
 1979 European Aquaculture Producers.
- 1980 Fehri-Bedoui R, Dinis MT, Maamouri F (2000) Étude histologique de devéloppement de
- 1981 l'appareil digestif et de la vesie gazeuse de la sole Solea senegalensis (Soleidae).
- 1982 *Cybium* **24**(4): 343-358.
- 1983 Fernandes TJR, Alves RC, Souza T, Silva JMG, Castro-Cunha M, Valente LMP et al.
- 1984 (2012) Lipid content and fatty acid profile of Senegalese sole (*Solea senegalensis* Kaup,
- 1858) juveniles as affected by feed containing different amounts of plant protein
 sources. *Food Chemistry* 134: 1337-1342.
- Fernández I, Gisbert E (2010) Senegalese sole bone tissue originated from chondral
 ossification is more sensitive than dermal bone to high vitamin A content in enriched *Artemia. Journal of Applied Ichthyology* 26: 344-349.
- 1990 Fernández I, Gisbert E (2011) The effect of vitamin A on flatfish development and
 1991 skeletogenesis: a review. *Aquaculture* 315: 34-48.
- 1992 Fernández I, Pimentel MS, Ortiz-Delgado JB, Hontoria F, Sarasquete C, Estévez A et al.
- 1993 (2009) Effect of dietary vitamin A on Senegalese sole (*Solea senegalensis*)
 1994 skeletogenesis and larval quality. *Aquaculture* 295: 250–265.
- Fernández-Díaz C, Yúfera M, Cañavate JP, Moyano FJ, Alarcón FJ, Díaz M (2001)
 Growth and physiological changes during metamorphosis of Senegal sole reared in
 laboratory. *Journal of Fish Biology*. 58: 1086-1097.
- 1998 Fernández-Palacios H, Norberg B, Izquierdo M, Hamre K (2011) Effects of Broodstock
- 1999 Diet on Eggs and Larvae, in Larval Fish Nutrition (ed G. J. Holt), Wiley-Blackwell,
- 2000 Oxford, UK. 153-181. doi: 10.1002/9780470959862.ch5

- *et al.* (2008a) In vitro inhibition of sole aquabirnavirus by Senegalese sole Mx. *Fish and Shellfish Immunology* 24: 187–193.
- 2004 Fernández-Trujillo A, Ferro P, Garcia-Rosado E, Infante C, Alonso MC, Bejar J et al.
- 2005 (2008b) Poly I:C induces Mx transcription and promotes an antiviral state against sole
- aquabirnavirus in the flatfish Senegalese sole (*Solea senegalensis* Kaup). *Fish and Shellfish Immunology* 24: 279–285.
- 2008 Ferreira HQ, Ramalho A, Dias J, Yúfera M, Arias AM, Falcão M et al. (2010) Sustainable
- semi-intensive polyculture of seabream and sole in earth ponds. *Aquaculture Europe* **35**:
- 2010 17-21.
- Fukuda Y, Matsuoka S, Mizuno Y, Narita K (1996) *Pasteurella piscicida* infection in
 cultured juvenile Japanese flounder. *Fish Pathology* 31: 33-38.
- 2013 Gamboa-Delgado J, Cañavate JP, Zerolo R, Le Vay L (2008) Natural carbon stable isotope
- 2014 ratios as indicators of the relative contribution of live and inert diets to growth in larval
- 2015 Senegalese sole (*Solea senegalensis*). *Aquaculture* **280**: 190-197.
- 2016 Gamboa-Delgado J, Le Vay L, Fernández-Díaz C, Cañavate P, Ponce M, Zerolo R et al.
- 2017 (2011) Effect of different diets on proteolytic enzyme activity, trypsinogen gene
 2018 expression and dietary carbon assimilation in Senegalese sole (*Solea senegalensis*)
- 2019 larvae. *Comparative Biochemistry and Physiology* **158B**: 251-258.
- 2020 García-Cegarra A, Merlo MA, Ponce M, Portela-Bens S, Cross I, Manchado M et al.
- 2021 (2013) A preliminary genetic map in *Solea senegalensis* (Pleuronectiformes, Soleidae)
- 2022 using BAC-FISH and next-generation sequencing. *Cytogenetic and Genome Research*
- 2023 DOI: 10.1159/000355001.

2024 García-Franquesa E, Molinero A, Valero J, Flos R (1996) Influence of sex, age an	e and season
---	--------------

- 2025 on the feeding habits of the flatfish Solea senegalensis. Environmental Biology of Fishes 2026 **47**: 289–298.
- García-Lopez A, Martinez-Rodriguez G, Sarasquete C (2005) Male reproductive system in 2027
- 2028 Senegalese sole *Solea senegalensis* (Kaup): Anatomy, histology and histochemistry.
- *Histology and Histopathology* **20**: 1179-1189. 2029
- García-López A, Pascual E, Sarasquete C, Martínez-Rodríguez G (2006a) Disruption of 2030
- 2031 gonadal maturation in cultured Senegalese sole Solea senegalensis Kaup by continuous

2032 light and/or constant temperature regimes. Aquaculture 261: 789–798.

- García-López A, Anguis V, Couto E, Canario AVM, Cañavate JP, Sarasquete C et al. 2033
- 2034 (2006b) Non-invasive assessment of reproductive status and cycle of sex steroid levels in a captive wild broodstock of Senegalese sole Solea senegalensis (Kaup). Aquaculture 2035 2036 **254**: 583-593.
- 2037 García-López A, Fernández-Pasquier V, Couto E, Canario AVM, Sarasquete C, Martínez-
- Rodríguez G (2006c) Testicular development and plasma sex steroid levels in cultured 2038
- 2039 male Senegalese sole Solea senegalensis Kaup. General and Comparative Endocrinology 147: 343–351. 2040
- García-López A, Couto E, Canario AVM, Sarasquete C, Martínez-Rodríguez G (2007) 2041
- 2042 Ovarian development and plasma sex steroid levels in cultured female Senegalese sole
- (Solea senegalensis). Comparative Biochemistry and Physiology Part A 146: 342-354. 2043
- 2044 García-López A, Sarasquete C, Martínez-Rodríguez G (2009) Temperature manipulation
- stimulates gonadal maturation and sex steroid production in Senegalese sole Solea
- senegalensis Kaup kept under two different light regimes. Aquaculture Research 40: 2046

2047 103-111.

Garcia de la Banda I, Lobo C, Chabrillo M, León-Rubio JM, Arijo S, Pazos G *et al.* (2012)
Influence of dietary administration of a probiotic strain *Shewanella putrefaciens* on
Senegalese sole (*Solea senegalensis*, Kaup 1858) growth, body composition and
resistance to *Photobacterium damselae* subsp piscicida. *Aquaculture Research* 43: 662669.

- Gavaia PJ, Dinis MT, Cancela ML (2002) Osteological development and abnormalities of
 the vertebral column and caudal skeleton in larval and juvenile stages of hatchery-reared
 Senegal sole (*Solea senegalensis*). *Aquaculture* 211: 305–323.
- 2056 Gavaia PJ, Domingues S, Engrola S, Drake P, Sarasquete C, Dinis MT et al. (2009)
- 2057 Comparing skeletal development of wild and hatchery-reared Senegalese sole (*Solea*
- 2058 senegalensis, Kaup 1858): evaluation in larval and postlarval stages. Aquaculture
 2059 Research 40: 1585–1593.
- 2060 Gisbert E, Fernández I, Villamizar N, Darias MJ, Zambonino-Infante JL, Estévez A (2014)
- 2061 European sea bass larval culture. In: Sánchez-Vázquez FJ (eds.) *Biology of European*
- 2062 *sea bass*, pp. 162-202. Wiley, London.
- 2063 Gomez-Gil B, Roque A, Chimetto L, Moreira APB, Lang E, Thompson F (2012) Vibrio
- 2064 *alfacsensis* sp nov., isolated from marine organisms. *International journal of systematic*
- and evolutionary microbiology **62**: 2955-2961 doi: 10.1099/ijs.0.033191-0
- 2066 Guerreiro I, Peres H, Castro-Cunha M, Oliva-Teles A (2012) Effect of temperature and
- 2067 dietary protein/lipid ratio on growth performance and nutrient utilization of juvenile
- 2068 Senegalese sole (*Solea senegalensis*). *Aquaculture Nutrition* **18**: 98-106.
- 2069 Guerreiro I, Peres H, Castro C, Pérez-Jiménez A, Castro-Cunha M, Oliva-Teles A (2014)
- 2070 Water temperature does not affect protein sparing by dietary carbohydrate in Senegalese
- sole (*Solea senegalensis*) juveniles. *Aquaculture Research* **45:** 289-298.

2072	Guillot R, Ceinos RM, Cal R, Rotllant J, Cerdá-Reverter JM (2012) Transient ectopic
2073	overexpression of agouti-signalling protein 1 (asip1) induces pigment anomalies in
2074	flatfish. PLoS ONE 7: e48526.

2075 Guzmán JM, Norberg B, Ramos J, Mylonas CC, Mañanós E (2008) Vitellogenin, steroid

plasma levels and spawning performance of cultured female Senegalese sole (*Solea senegalensis*). *General and Comparative Endocrinology* **156**: 285–297.

- Guzmán JM, Rubio M, Ortiz-Delgado J, Klenke U, Kight K, Cross I *et al.* (2009a)
 Comparative gene expression of gonadotropins (FSH and LH) and peptide levels of
 gonadotropin-releasing hormones (GnRHs) in the pituitary of wild and cultured
 Senegalese sole (*Solea senegalensis*) broodstocks. *Comparative Biochemistry and Physiology Part A* 153: 266-277.
- Guzmán JM, Ramos J, Mylonas CC, Mañanós E (2009b) Spawning performance and
 plasma levels of GnRHa and sex steroids in cultured female Senegalese sole (*Solea senegalensis*) treated with different GnRHa-delivery systems. *Aquaculture* 291: 2002086 209.
- Guzmán JM, Cal R, García-López A, Chereguini O, Kight K, Olmedo M *et al.* (2011a)
 Effects of *in vivo* treatment with the dopamine antagonist pimozide and gonadotropinreleasing hormone agonist (GnRHa) on the reproductive axis of Senegalese sole (*Solea senegalensis*). *Comparative Biochemistry and Physiology Part A* 158: 235-245.
- 2091 Guzmán JM, Ramos J, Mylonas CC, Mañanós E (2011b) Comparative effects of human
- 2092 chorionic gonadotropin (hCG) and gonadotropin-releasing hormone agonist (GnRHa)
- 2093 treatments on the stimulation of male Senegalese sole (*Solea senegalensis*) reproduction.
- 2094 *Aquaculture* **316**: 121-128.

- Hamre K, Holen E, Moren M (2007) Pigmentation and eye migration in Atlantic halibut
- 2096 (*Hippoglossus hippoglossus* L.) larvae: new findings and hypotheses. *Aquaculture*2097 *Nutrition* 13: 65-80.
- 2098 Hamre K, Harboe T (2008) Critical levels of essential fatty acids for normal pigmentation
- in Atlantic halibut (*Hippoglossus hippoglossus* L.) larvae. *Aquaculture* **277**: 101-108.
- 2100 Helland SJ, Grisdale-Helland B (1998) Growth, feed utilization and body composition of
- juvenile Atlantic halibut (*Hippoglossus hippoglossus*) fed diets differing in the ratio
 between the macronutrients. *Aquaculture* 166: 49-56.
- 2103 Hodneland K, García R, Balbuena JA, Zarza C, Fouz B (2011) Real-time RT-PCR
- 2104 detection of betanodavirus in naturally and experimental infected fish from Spain.
- 2105 *Journal of fish disease* **34**: 189-202.
- Howell BR (1997) A re-appraisal of the potential of the sole, *Solea solea* (L.), for
 commercial cultivation. *Aquaculture* 155: 359-369.
- 2108 Howell B, Prickett R, Baynes S, Cañavate P (2003) The cultivation of soles. Report of a
- 2109 2nd Sole Workshop held at CICEM El Toruño, Cadiz, Spain 29 September to 1 October2100 2003.
- 2111 Howell B, Conceição L, Prickett R, Cañavate P, Mañanos E (2009) Sole farming: nearly
- there but not quite? A report of 4th workshop on the cultivation of soles. *Aquaculture Europe* 34: 24-27.
- 2114 Howell B, Pricket R, Cañavate P, Mañanós E, Dinis MT, Conceição L, Valente LMP
- 2115 (2011) Sole farming: There or thereabouts! A report of the 5th workshop on the
- cultivation of soles. *Aquaculture Europe* **36**: 42-45.

- Huang Z, Song X, Zheng Y, Peng L, Wan R, Lane T *et al.* (2013) Design and evaluation of
- a commercial recirculating system for half-smooth tongue sole (*Cynoglossus semilaevis*)
- 2119 production. *Aquacultural Engineering* **54**: 104-109.
- 2120 Huising MO, Guichelaar T, Hoek C, Verburg-van Kemenade BML, Flik G, Savelkoul HFJ
- *et al.* (2003) Increased efficacy of immersion vaccination in fish with hyperosmotic
- 2122 pretreatment. *Vaccine* **21**: 4178–4193.
- Imsland AK, Foss A, Conceição LEC, Dinis MT, Delbare D, Schram E et al. (2003) A
- review of the culture potential of *Solea solea* and *S. senegalensis*. *Reviews in Fish*
- 2125 *Biology and Fisheries* **13**: 379-407.
- 2126 Jentoft S, Aastveit AH, Torjesen PA, Andersen Ø (2005) Effects of stress on growth,
- cortisol and glucose levels in non-domesticated Eurasian perch (*Perca fluviatilis*) and
- domesticated rainbow trout (Oncorhynchus mykiss). Comparative Biochemistry and
- 2129 *Physiology Part A* **141**: 353–358.
- 2130 Kai YH, Chi SC (2008) Efficacies of inactivated vaccines against betanodavirus in grouper
- larvae (*Epinephelus coioides*) by bath immunization. *Vaccine* **26**: 1450-1457.
- 2132 Kaushik SJ (1998) Nutritional bioenergetics and estimation of waste production in non-
- salmonids. *Aquatic Living Resources* **11:** 211-217.
- 2134 Koumoundouros G (2010) Morpho-anatomical abnormalities in Mediterranean marine
- aquaculture. In: Koumoundouros G (ed.) *Recent Advances in Aquaculture Research*, pp.
- 2136 125–148. Transworld Research Network, Kerala, India.
- Lall SP, Lewis-McCrea LM (2007) Role of nutrients in skeletal metabolism and pathology
- in fish an overview. *Aquaculture* **267**: 3–19.
- Lin CC, Lin JHY, Chen MS, Yang HL (2007) An oral nervous necrosis virus vaccine that
- induces protective immunity in larvae of grouper (*Epinephelus coioides*). Aquaculture

- Liu X, Liu X, Lian J, Wang Y, Zhang F, Yu H (2008) Large scale artificial reproduction
 and rearing of senegal sole, *Solea senegalensis* (Kaup). *Marine Fisheries Research* 29:
 10-16.
- 2145 López-Olmeda J F, Blanco-Vives B, Pujante IM, Wunderink YS, Mancera JM, Sánchez-
- Vázquez FJ (2013) Daily rhythms in the hypothalamus-pituitary-interrenal axis and
 acute stress responses in a teleost flatfish, *Solea senegalensis*. *Cronobiology International* **30**: 530–539.
- 2149 López-Vazquez C, Conde M, Dopazo CP, Barja JL, Bandín I (2011) Susceptibility of
- juvenile sole *Solea senegalensis* to marine isolates of viral haemorrhagic septicaemia
 virus from wild and farmed fish. *Diseases of Aquatic Organisms* **93**: 11-116.
- Luis OJ, Passos AM (1995) Seasonal changes in lipid content and composition of the
 polychaete Nereis (Hediste) diversicolor. Comparative Biochemistry and Physiology
- **111B**: 579–586.
- Lund I, Steenfeldt SJ, Hansen BW (2007) Effect of dietary arachidonic acid,
 eicosapentaenoic acid and docosahexaenoic acid on survival, growth and pigmentation
 in larvae of common sole (*Solea solea*). *Aquaculture* 273: 532–544.
- Lund I, Steenfeldt SJ, Banta G, Hansen BW (2008) The influence of dietary concentrations
- of arachidonic acid and eicosapentaenoic acid at various stages of larval ontogeny on
- eye migration, pigmentation and prostaglandin content of common sole larvae (Solea
- 2161 *solea* L.). *Aquaculture* **276**: 143-153.
- Lund I, Steenfeldt SJ, Hansen BW (2010) Influence of dietary arachidonic acid combined
- with light intensity and tank colour on pigmentation of common sole (*Solea solea* L.)
- 2164 larvae. *Aquaculture* **308**: 159-165.

2165	Magariños B, Romalde JL, López-Romalde S, Moriñigo MA, Toranzo AE (2003)
2166	Pathobiological characterisation of Photobacterium damselae subsp. piscicida isolated
2167	from cultured sole (Solea senegalensis). Bulletin of European Association of Fish
2168	Pathologists 23 (4): 183-190.
2169	Magariños B, Devesa S, González A, Castro N, Toranzo AE (2011) Furunculosis in
2170	Senegalese sole (Solea senegalensis) cultured in a recirculation system. Veterinary

2171 *Record* **168**: 431.

2181

- 2172 Mañanós E, Ferreiro I, Bolón D, Guzmán JM, Mylonas CC, Riaza A (2007) Different
- 2173 responses of Senegalese sole (*Solea senegalensis*) broodstock to a hormonal spawning
- 2174 induction therapy, depending on their wild or captive-reared origin. In: *Proceedings of*
- 2175 *Aquaculture Europe 07*, pp. 330-331. European Aquaculture Society, Istanbul, Turkey.
- 2176 Mañanós E, Duncan N, Mylonas CC (2008) Reproduction and control of ovulation,
- spermiation and spawning in cultured fish. In: Cabrita E, Robles V, Herráez MP (ed.)
- 2178 *Methods in Reproductive Aquaculture: Marine and Freshwater Species*, pp. 3–80. CRC
- 2179 Press, Taylor and Francis Group, Boca Raton.
- 2180 Manchado M, Infante C, Asensio E, Crespo A, Zuasti E, Cañavate JP (2008) Molecular

characterization and gene expression of six trypsinogens in the flatfish Senegalese sole

- 2182 (Solea senegalensis Kaup) during larval development and in tissues. Comparative
 2183 Biochemistry and Physiology 149B: 334–344.
- 2184 Mandrioli L, Sirri R, Gatta PP, Morandi F, Sarli G, Parma L et al. (2012) Histomorphologic
- 2185 hepatic features and growth performances of juvenile Senegalese sole (Solea
- 2186 *senegalensis*) fed isogeneric practical diets with variable protein/lipid levels. *Journal of*
- 2187 *Applied Ichthyology* **28**: 628-632.

- Marín-Juez R, Castellana B, Manchado M, Planas JV (2011) Molecular identification of
 genes involved in testicular steroid synthesis and characterization of the response to
 gonadotropic stimulation in the Senegalese sole (*Solea senegalensis*) testis. *General and Comparative Endocrinology* 172: 130–139.
- 2192 Marín-Juez R, Vinas J, Mechaly AS, Planas JV, Piferrer F (2013) Stage-specific gene
- expression during spermatogenesis in the Senegalese sole (*Solea senegalensis*), a fish
- 2194 with semi-cystic type of spermatogenesis, as assessed by laser capture microdissection
- and absolute quantitative PCR. *General and Comparative Endocrinology* **188**: 242-250.
- 2196 Martin I, Rasines I, Gomez M, Rodriguez C, Martinez P, Chereguini O (2014) Evolution of
- egg production and parental contribution in Senegalese sole, *Solea senegalensis*, during
- four consecutive spawning seasons. *Aquaculture*, doi:10.1016/j.aquaculture.2013.12.042
- 2199 Martínez I, Moyano FJ, Fernández-Díaz C, Yúfera M (1999) Digestive enzyme activity
- during larval development of Senegal sole (*Solea senegalensis*). *Fish Physiology and Biochemistry* 21: 317–323.
- 2202 Martins CIM, Eding EH, Verdegem MCJ, Heinsbroek LTN, Schneider O, Blancheton JP et
- 2203 *al.* (2010) New developments in recirculating aquaculture systems in Europe: A
- perspective on environmental sustainability. *Aquacultural Engineering* **43**: 83-93.
- 2205 Mauri I, Romero A, Acerete L, MacKenzie S, Roher N, Callol A *et al.* (2011) Changes in
- complement responses in gilthead seabream (Sparus aurata) and European seabass
- 2207 (Dicentrarchus labrax) under crowding stress, plus viral and bacterial challenges. Fish
- *and Shellfish Immunology* **30**: 182–188.
- McVicar AH, White PG (1979) Fin and skin necrosis of cultivated Dover sole *Solea solea*(L.). *Journal of Fish Diseases* 2: 557-562.
- 2211 McVicar AH, White PG (1982) The prevention and cure of an infectious disease in

- cultivated juvenile dover sole, *Solea solea* (L). *Aquaculture* **26**: 3-4.
- 2213 Mommsen TP, Vijayan MM, Moon TW (1999) Cortisol in teleosts: dynamics, mechanisms
- of action, and metabolic regulation. *Reviews in Fish Biology and Fisheries* **9**: 211–268.
- 2215 Morais S, Narciso L, Dores E, Pousão-Ferreira P (2004a) Lipid enrichment for Senegalese
- sole (*Solea senegalensis*) larvae: effect on larval growth, survival and fatty acid profile.
- 2217 *Aquaculture International* **12**: 281–298.
- 2218 Morais S, Lacuisse M, Conceição LEC, Dinis MT, Rønnestad I (2004b) Ontogeny of the
- digestive capacity of Senegalese sole (Solea senegalensis), with respect to digestion,
- absorption and metabolism of amino acids from *Artemia*. *Marine Biology* **145**: 243-250.
- 2221 Morais S, Koven W, Rønnestad I, Dinis MT, Conceição LEC (2005a) Dietary protein/lipid
- ratio and lipid nature affects fatty acid absorption and metabolism in a teleost larva. *British Journal of Nutrition* **93**: 813–820.
- Morais S, Koven W, Rønnestad I, Dinis MT, Conceição LEC (2005b) Dietary protein/lipid ratio affects growth and amino acid and fatty acid absorption and metabolism in
- Senegalese sole (Solea senegalensis Kaup 1858) larvae. *Aquaculture* **246**: 347–357.
- Morais S, Conceição LEC (2009) A new method for the study of essential fatty acid
 requirements in fish larvae. *British Journal of Nutrition* 101: 1564-1568.
- 2229 Morais S, Castanheira F, Martinez-Rubio L, Conceição LEC, Tocher DR (2012) Long 2230 chain polyunsaturated fatty acid synthesis in a marine vertebrate: ontogenetic and 2231 nutritional regulation of a fatty acyl desaturase with $\Delta 4$ activity. *Biochimica et* 2232 *Biophysica Acta - Molecular and Cell Biology of Lipids* **1821**:660-671.
- Morais S, Candeias Mendes A, Castanheira MF, Coutinho J, Bandarra N, Dias J *et al.*(2014a) New formulated diets for *Solea senegalensis* broodstock: effects of parental
 nutrition on biosynthesis of long-chain polyunsaturated fatty acids and performance of

- Morais S, Mourente G, Tocher DR (2014b) LC-PUFA biosynthesis in Senegalese sole and
 its modulation by dietary lipid level and fatty acid composition. Proceedings of the 16th
 International Symposium on Fish Nutrition and Feeding, Cairns, Australia, p 44.
- 2241 Moreira N, Soares S, Valente LMP, Castro-Cunha M, Cunha LM, Guedes De Pinho P
- (2014) Effect of two experimental diets (protein and lipid vegetable oil blends) on the
- volatile profile of Senegalese sole (*Solea senegalensis* Kaup, 1858) muscle. *Food Chemistry* 153: 327-333.
- 2245 Moyano FJ, Alarcón FJ, Díaz M, Abellán E, Yúfera M, Fernández-Díaz C (2001)
- Capacidad digestiva comparada durante el desarrollo larvario de tres peces marinos:
 Dorada (*Sparus aurata*), denton (*Dentex dentex*) y lenguado senegalés (*Solea senegalensis*). Implicaciones para el desarrollo de alimento artificial. *Serie Monografías del Instituto Canario de Ciencias Marinas* 4: 340-345.
- Navarro-Guillén C, Yúfera M, Engrola S (2014a) Senegalese sole post-larvae exhibit daily
 rhythmicity of feed intake and protein metabolism. Aquaculture Europe 2014, San
- 2252 Sebastian, Spain 14-18 October 2014.
- 2253 Navarro-Guillén C, Engrola S, Castanheira F, Bandarra N, Hachero-Cruzado I, Tocher DR
- *et al.* (2014b) Effect of varying dietary levels of LC-PUFA and vegetable oil sources on
- 2255 performance and fatty acids of Senegalese sole post larvae: Puzzling results suggest
- complete biosynthesis pathway from C18 PUFA to DHA. *Comparative Biochemistry*
- *and Physiology Part B* **167**: 51-58.
- 2258 Navarro-Guillén C, Moyano FJ, Yúfera M (2015) Diel food intake and digestive enzyme
- production patterns in *Solea senegalensis* larvae. *Aquaculture* **435**: 33-42.

2260	Nishizawa T, Takami I, Kokawa Y, Yoshimizu M (2009) Fish immunization using a
2261	synthetic double-stranded RNA Poly (I:C), an interferon inducer, offers protection
2262	against RGNNV, a fish nodavirus. Diseases of Aquatic Organisms 83: 115-122.

- 2263 Norambuena F, Estevez A, Bell G, Carazo I, Duncan N (2012a) Proximate and fatty acid
- 2264 composition in muscle, liver and gonads of wild versus cultured broodstock of
- 2265 Senegalese sole (*Solea senegalensis*). *Aquaculture* **356–357**: 176–185.
- Norambuena F, Estévez A, Sánchez-Vázquez FJ, Carazo I, Duncan N (2012b) Selfselection of diets with different contents of arachidonic acid by Senegalese sole (*Solea senegalensis*) broodstock. *Aquaculture* 364-365: 198-205.
- 2269 Norambuena F, Mackenzie S, Bell JG, Callol A, Estevéz A, Duncan N (2012c)
- 2270 Prostaglandin (F and E, 2-and 3-series) production and cyclooxygenase (COX-2) gene
- 2271 expression of wild and cultured broodstock of Senegalese sole (*Solea senegalensis*).

2272 *General and Comparative Endocrinology* **177**: 256–262.

- 2273 Norambuena F, Morais S, Estevez A, Bell JG, Tocher DR, Navarro JC et al. (2013a)
- Dietary modulation of arachidonic fatty acid metabolism in Senegalese sole (*Solea senegalensis*) broodstock reared in captivity. *Aquaculture* 372–375: 80–88.
- Norambuena F, Estévez A, Mañanós E, Bell JG, Carazo I, Duncan N (2013b) Effects of
 graded levels of arachidonic acid on the reproductive physiology of Senegalese sole
 (*Solea senegalensis*): Fatty acid composition, prostaglandins and steroid levels in the
 blood of broodstock bred in captivity. *General and Comparative Endocrinology* 191:
 92–101.
- 2281 Oliveira C, Vera LM, López-Olmeda JF, Guzmán JM, Mañanós E, Ramos J et al. (2009)
- 2282 Monthly day/night changes and seasonal daily rhythms of sexual steroids in Senegal sole

- (Solea senegalensis) under natural fluctuating or controlled environmental conditions.
 Comparative Biochemistry and Physiology Part A 152: 168–175.
- Oliveira C, Duncan NJ, Pousão-Ferreira P, Mananos E, Sanchez-Vazquez FJ (2010)
 Influence of the lunar cycle on plasma melatonin, vitellogenin and sex steroids rhythms
- in Senegal sole, *Solea senegalensis*. *Aquaculture* **306**: 343-347.
- 2288 Olveira JG, Souto S, Dopazo CP, Thiéry R, Barja JL, Bandín I (2009) Comparative
- analysis of both genomic segments of betanodaviruses isolated from epizootic outbreaks
- in farmed fish species provides evidence for genetic reassortment. Journal of General
- 2291 *Virology* **90**: 2940-2951.
- 2292 Osuna-Jiménez I, Williams TD, Prieto-Álamo M-J, Abril N, Chipman JK, Pueyo C (2009)
- 2293 Immune- and stress-related transcriptomic responses of *Solea senegalensis* stimulated
- with lipopolysaccharide and copper sulphate using heterologous cDNA microarrays. *Fish and Shellfish Immunology* 26: 699–706.
- 2296 Ottesen OH, Strand HK (1996) Growth, development and skin abnormalities of halibut,
- 2297 *Hippoglossus hippoglossus*, L., juveniles kept on different bottom substrates.
 2298 *Aquaculture* 146, 17–25.
- 2299 Ottesen OH, Noga EJ, Sandaa W (2007) Effect of substrate on progression and healing of
- skin erosions and epidermal papillomas of Atlantic halibut, *Hippoglossus hippoglossus*
- 2301 (L.). *Journal of Fish Diseases* **30**: 43-53.
- 2302 Padrós F, Zarza C, Estévez A, Crespo S, Furones MD (2003) La patología como factor
- 2303 limitante para el desarrollo del cultivo del lenguado. IX Congreso nacional de
- 2304 *acuicultura*, Cádiz, May 12 to 16, pp. 343-345.

- 2305 Padrós F, Villalta M, Gisbert E, Estévez A (2011) Morphological and histological study of
- larval development of the Senegal sole *Solea senegalensis*: an integrative study. *Journal*
- 2307 *of Fish Biology* **79**: 3-32.
- 2308 Palenzuela O, Redondo MJ, López E, Álvarez-Pellitero P (2007) Cultured sole, Solea
- *senegalensis* is susceptible to *Enteromyxum scophthalmi*, the myxozoan parasite causing
- turbot emaciative enteritis. *Parassitologia* **49**:73
- 2311 Parra G, Yúfera M (2001) Comparative energetics during early development of two marine
- fish species, Solea senegalensis (Kaup) and Sparus aurata (L.). Journal of Experimental
- 2313 *Biology* **204**: 2175-2183.
- 2314 Pedley S, Bartram J, Rees G, Dufour A, Cotruvo J (2004) Pathogenic Mycobacteria in
- Water: A Guide to Public Health Consequences, Monitoring, and Management. IWA
 Publishing, London, UK.
- 2317 Piñeiro-Vidal M, Riaza A, Santos Y (2008a) Tenacibaculum discolor sp. nov. and
- 2318 *Tenacibaculum gallaicum* sp. nov., isolated from sole (*Solea senegalensis*) and turbot
- 2319 (*Psetta maxima*) culture systems. International Journal of Systematic and Evolutionary
- 2320 *Microbiology* **58**: 21- 25.
- 2321 Piñeiro-Vidal M, Carballas CG, Gómez-Barreiro O, Riaza A, Santos Y (2008b)
- 2322 *Tenacibaculum soleae* sp. nov., isolated from diseased sole (*Solea senegalensis* Kaup).
- 2323 International Journal of Systematic and Evolutionary Microbiology 58: 881-885.
- 2324 Pinto W, Aragão C, Soares F, Dinis MT, Conceição LEC (2007) Growth, stress response
- and free amino acid levels in Senegalese sole (Solea senegalensis Kaup 1858)
- chronically exposed to exogenous ammonia. *Aquaculture Research* **38**: 1198–1204.
- 2327 Pinto W, Figueira L, Dinis MT, Aragão C (2009) How does fish metamorphosis affect
- aromatic amino acid metabolism? *Amino Acids* **36**: 177-183.

- Pinto W, Rodrigues V, Dinis MT, Aragão C (2010a) Can dietary aromatic amino acid
 supplementation be beneficial during fish metamorphosis? *Aquaculture* 310: 200-205.
- 2331 Pinto W, Figueira L, Ribeiro L, Yúfera M, Dinis MT, Aragão C (2010b) Dietary taurine
- supplementation enhances metamorphosis and growth potential of *Solea senegalensis*
- 2333 larvae. *Aquaculture* **309**: 159-164.
- 2334 Pinto W, Rønnestad I, Jordal A-E, Gomes A, Dinis M, Aragão C (2012) Cloning, tissue and
- ontogenetic expression of the taurine transporter in the flatfish Senegalese sole (*Solea senegalensis*). *Amino Acids* 42: 1317-1327.
- 2337 Porta J, Porta JM, Martínez-Rodriguez G, Alvarez MC (2006) Development of a
- 2338 microsatellite multiplex PCR for Senegalese sole (*Solea senegalensis*) and its 2339 application to broodstock management. *Aquaculture* **256**: 159-166.
- Pottinger TG (2008) The stress response in fish-mechanisms, effects and measurement. In:
 Branson EJ (ed.) *Fish welfare*, pp. 32–48. Blackwell Publishing, Oxford.
- 2342 Pulsford AL, Lemaire-Gony S, Tomlinson M, Collingwood N, Glynn PJ (1994) Effects of
- acute stress on the immune system of the dab, Limanda limanda. Comparative
- 2344 Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology
- **109**: 129–139.
- 2346 Quental-Ferreira H, Ramalho-Ribeiro A, Dias J, Yúfera M, Arias A, Falcão M et al. (2010)
- Sustainable semi-intensive polyculture of seabream and sole in earthen ponds. *Aquaculture Europe* 35: 17-21.
- 2349 Rasines I, Gómez M, Martín I, Rodríguez C, Mañanos E, Chereguini O (2012) Artificial
- 2350 fertilization of Senegalese sole (*Solea senegalensis*): Hormone therapy administration
- 2351 methods, timing of ovulation and viability of eggs retained in the ovarian cavity.
- 2352 *Aquaculture* **326-329**: 129-135.

- 2353 Rasines I (2013) Reproducción en cautividad del lenguado senegalés (Solea senegalensis):
- 2354 inducción hormonal a la ovulación y obtención de huevos para la fecundación artificial.
- 2355 PhD Thesis, Universidad de Oviedo, Oviedo, Spain.
- 2356 Rasines I, Gomez M, Martin I, Rodriguez C, Mañanós E, Chereguini O (2013) Artificial
- fertilization of cultured Senegalese sole (*Solea senegalensis*): effects of the time of day
- of hormonal treatment on inducing ovulation. *Aquaculture* **392–395**: 94–97.
- 2359 Rema P, Conceição LEC, Evers F, Castro-Cunha M, Dinis MT, Dias J (2008) Optimal
- 2360 dietary protein levels in juvenile Senegalese sole (*Solea senegalensis*). Aquaculture
 2361 *Nutrition* 14: 263-269.
- Ribeiro L, Sarasquete C, Dinis MT (1999a) Histological and histochemical development of
- the digestive system of *Solea senegalensis* (Kaup, 1858) larvae. *Aquaculture* 171: 293–
 308.
- Ribeiro L, Zambonino-Infante JL, Cahu C, Dinis MT (1999b) Development of digestive
 enzymes in larvae of *Solea senegalensis*, Kaup 1858. *Aquaculture* 179: 465–473.
- 2367 Richard N, Engrola S, Palma P, Pinto W, Simes D, Conceição LEC (2013) Assessment of
- 2368 protein digestive capacity and utilisation during ontogeny of Senegalese sole: a tracer
- study using in vivo produced radiolabelled peptide fractions. In: Larvi'13 Fish &
- 2370 *Shellfish Larviculture Symposium*, Ghent, Belgium.
- 2371 Richard N, Fernández I, Wulff T, Hamre K, Cancela L, Conceição LEC, Gavaia PJ (2014)
- 2372 Dietary supplementation with vitamin K affects transcriptome and proteome of
- 2373 Senegalese sole, improving larval performance and quality. *Marine Biotechnology* DOI:
- 2374 10.1007/s10126-014-9571-2.

- 2376 Characterization of *Vibrio harveyi* strains recovered from diseased farmed Senegalese
- sole (*Solea senegalensis*). *Journal of Applied Microbiology* **105** (3): 752-760.
- 2378 Rodiles A, Santigosa E, Herrera M, Hachero-Cruzado I, Cordero M, Martínez-Llorens S et
- al. (2012) Effect of dietary protein level and source on digestive proteolytic enzyme
- activity in juvenile Senegalese sole, Solea senegalensis Kaup 1850. Aquaculture *International* 20: 1053-1070.
- 2382 Rodríguez S, Vilas MP, Gutierrez MC, Pérez-Prieto I, Sarasquete MC, Rodríguez B (1997)
- 2383 Isolation and preliminary characterization of a Birnavirus from the sole Solea
- senegalensis in southwest Spain. *Journal of Aquatic Animal Health* **9**: 295-300.
- 2385 Romalde JL, Raveo C, Lopez-Romalde S, Avendano-Herrera R, Magariños B, Toranzo AE
- (2005) Vaccination strategies to prevent emerging diseases for Spanish aquaculture.
 Progress in fish vaccinology 125: 85-95.
- Rønnestad I, Conceição LEC (2012) *Artemia* protein is processed very fast in *Solea senegalensis* larvae: A dynamic simulation model. *Aquaculture* 350–353: 154-161.
- 2390 Rønnestad I, Rojas-García CR, Tonheim SK, Conceição LEC (2001) In vivo studies of
- digestion and nutrient assimilation in marine fish larvae. *Aquaculture* **201**: 161-175.
- 2392 Rønnestad I, Yúfera M, Ueberschär B, Ribeiro L, Sæle Ø, Boglione C (2013) Feeding
- behaviour and digestion physiology in larval fish current knowledge, and gaps and
- bottlenecks in research. *Reviews in Aquaculture* 5 (Suppl. 1): 59-98.
- 2395 Rueda-Jasso R, Conceição LEC, Dias J, De Coen W, Gomes E, Rees JF et al. (2004) Effect
- of dietary non-protein energy levels on condition and oxidative status of Senegalese sole
- 2397 (*Solea senegalensis*) juveniles. *Aquaculture* **231:** 417-433.

²³⁷⁵ Rico M, Tapia-Paniagua S, Martínez-Manzanares E, Balebona MC, Moriñigo MA (2008)

2398	Sá R, Bexiga C, Vieira L, Veiga P, Erzini K (2003) Diets of the sole Solea vulgaris
2399	Quensel, 1806 and Solea senegalensis Kaup, 1858 in the lower estuary of the Guadiana
2400	River (Algarve, southern Portugal): Preliminary results. Boletin del Instituto Español de
2401	<i>Oceanografía</i> . 19 : 505-508.
2402	Saeij JPJ, Verburg-van Kemenade LBM, van Muiswinkel WB, Wiegertjes GF (2003) Daily
2403	handling stress reduces resistance of carp to Trypanoplasma borreli: in vitro modulatory
2404	effects of cortisol on leukocyte function and apoptosis. Developmental and Comparative
2405	Immunology 27 : 233–245.
2406	Sáez de Rodrigáñez MA, Medina E, Moyano FJ, Alarcón FJ (2011) Evaluation of protein
2407	hydrolysis in raw sources by digestive proteases of Senegaleses sole (Solea
2408	senegalensis, Kaup 1858) using a combination of an in vitro assay and sodium dodecyl
2409	sulphate polyacrylamide gel electrophoresis análisis of products. Aquaculture Research

42: 1639-1652.

- 2411 Salas-Leiton E, Anguís V, Manchado M, Cañavate JP (2008) Growth, feeding and oxygen
- consumption of Senegalese sole (*Solea senegalensis*) juveniles stocked at different
 densities. *Aquaculture* 285: 84–89.
- Salas-Leiton E, Cánovas-Conesa B, Zerolo R, López-Barea J, Cañavate JP, Alhama J.
 (2009) Proteomics of juvenile Senegal sole (*Solea senegalensis*) affected by gas bubble
 disease in hyperoxygenated ponds. *Marine Biotechnology* 11: 473-487.
- 2417 Salas-Leiton E, Anguis V, Martín-Antonio B, Crespo D, Planas JV, Infante C et al. (2010)
- 2418 Effects of stocking density and feed ration on growth and gene expression in the
- 2419 Senegalese sole (*Solea senegalensis*): Potential effects on the immune response. *Fish*
- 2420 *and Shellfish Immunology* **28**: 296–302.

Salas-Leiton E, Anguís V, Rodríguez-Rúa A, Cañavate JP (2011) High stocking density
and food restriction have minimum impact on size dispersal of cultured Senegalese sole
(*Solea senegalensis*, Kaup 1858) juveniles. Evidence for individual growth being
regulated by population structure. *Aquacultural Engineering* 45: 43–50.

- 2425 Salas-Leiton E, Coste O, Asensio E, Infante C, Cañavate JP, Manchado M (2012)
- 2426 Dexamethasone modulates expression of genes involved in the innate immune system,
- growth and stress and increases susceptibility to bacterial disease in Senegalese sole
- 2428 (Solea senegalensis Kaup, 1858). Fish and Shellfish Immunology **32**: 769–778.
- 2429 Sánchez-Fernández P (2012) Growth and size variation of Senegalese sole (Solea
 2430 senegalensis). PhD Thesis, Universidad Politécnica de Barcelona, Barcelona, Spain.
- 2431 Santos Y, Pazos F, Barja JL (1999) *Flexibacter maritimus*, causal agent of flexibacteriosis
- 2432 in marine fish. In: Olivier G (ed.) ICES Identification Leaflets for Diseases and
- 2433 *Parasites of Fish and Shellfish. No. 55*, pp. 1–6. International Council for the
 2434 Exploration of the Sea, Copenhagen, Denmark.
- 2435 Sarasquete MC, González de Canales ML, Arellano JM, Muñoz-Cueto JA, Ribeiro L, Dinis
- 2436 MT (1996). Histochemical aspects of the yolk-sac and digestive tract of larvae of the
- 2437 Senegal sole, *Solea senegalensis* Kaup, 1858. *Histology and Histophatology* 11: 881–
 2438 888.
- 2439 Seikai T, Sinoda M (1981) Variations of frequency occurrence of color anomalies in
- hatchery-reared flounder, *Paralichthys olivaceus*, due to duration of *Artemia salina* fed.
- 2441 Bulletin of the Kyoto Institute of Ocean and Fishery Science 5: 29–37.
- 2442 Silva JMG, Espe M, Conceição LEC, Dias J, Valente LMP (2009) Senegalese sole
- juveniles (Solea senegalensis Kaup, 1858) grow equally well on diets devoid of fish
- meal provided the dietary amino acids are balanced. *Aquaculture* **296**: 309-317.

- Silva JMG (2010) Use of alternative protein sources in diets for Senegalese sole (*Solea senegalensis* Kaup, 1858) juveniles. PhD thesis, University of Porto. 152 p.
- 2447 Silva JMG, Espe M, Conceição LEC, Dias J, Costas B, Valente LMP (2010) Feed intake
- and growth performance of Senegalese sole (Solea senegalensis Kaup, 1858) fed diets
- 2449 with partial replacement of fish meal with plant proteins. *Aquaculture Research* **41:** e20-
- e30.
- 2451 Silva JMG, Valente LMP, Castro-Cunha M, Bacelar M, Guedes De Pinho P (2012) Impact
- of dietary plant protein levels on the volatile composition of Senegalese sole (Solea
- senegalensis Kaup, 1858) muscle. *Food Chemistry* **131:** 596-602.
- 2454 Simopoulos AP (1999) Essential fatty acids in health and chronic disease. *The American*2455 *Journal of Clinical Nutrition* 70: 560S-569S.
- Sorensen PW, Stacey NE (2004) Brief review of fish pheromones and discussion of their
 possible uses in the control of non-indigenous teleost fishes. *New Zealand Journal of*
- 2458 *Marine and Freshwater Research* **38**: 399–417.
- 2459 Starkey WG, Ireland JH, Muir KF, Jenkins ME, Roy WJ, Richards RH et al. (2001)
- Nodavirus infection in Atlantic cod and Dover sole in the UK. 2001. *Veterinary record*149(6): 179-181.
- 2462 Stolte EH, Nabuurs SB, Bury NR, Sturm A, Flik G, Savelkoul HFJ et al. (2008) Stress and
- innate immunity in carp: Corticosteroid receptors and pro-inflammatory cytokines.
- 2464 *Molecular Immunology* **46**: 70–79.
- Tacon AGJ, Metian M (2008) Global overview on the use of fish meal and fish oil in
 industrially compounded aquafeeds: Trends and future prospects. *Aquaculture* 285: 146-

2467 158.

- Tapia-Paniagua ST, Díaz-Rosales P, León-Rubio JM, García de La Banda I, Lobo C,
 Alarcón JF *et al.* (2012) Use of the probiotic *Shewanella putrefaciens* Pdp11 on the
 culture of Senegalese sole (*Solea senegalensis*, Kaup 1858) and gilthead seabream
- 2471 (*Sparus aurata* L.). *Aquaculture International* **20**: 1025-1039.
- 2472 Thiéry R, Cozien J, de Boisseson C, Kerbart-Boscher S, Nevarez L (2004) Genomic
- 2473 classification of new betanodavirus isolates by phylogenetic analysis of the coat protein
- 2474 gene suggests a low host-fish species specificity. *Journal of general virology* **85**: 3079-
- 2475 3087.
- 2476 Tocher DR (2003) Metabolism and functions of lipids and fatty acids in teleost fish.
- 2477 *Reviews in Fisheries Science* **11**: 107–184.
- Tocher DR (2010) Fatty acid requirements in ontogeny of marine and freshwater fish. *Aquaculture Research* 41: 717–732.
- 2480 Toranzo AE, Barreiro S, Casal JF, Figueras A, Magariños B, Barja JL (1991) Pasteurellosis
- 2481 in cultured gilthead seabream (*Sparus aurata*): first report in Spain. Aquaculture **99**: 1-
- 2482 15
- 2483 Toranzo AE, Avendaño R, López-Vázquez C, Magariños B, Dopazo CP, Romalde JL et al.
- 2484 (2003) Principales patologías bacterianas y víricas en lenguado cultivado:
- 2485 caracterización de los agentes etiológicos. *IX Congreso nacional de acuicultura*, Cádiz,
- 2486 May 12 to 16, pp. 354-356.
- 2487 Tort L, Montero D, Robaina L, Fernández-Palacios H, Izquierdo MS (2001) Consistency of
- stress response to repeated handling in the gilthead sea bream *Sparus aurata* Linnaeus,
- 2489 1758. Aquaculture Research **32**: 593–598.
- 2490 Valente LMP, Linares F, Villanueva JLR, Silva JMG, Espe M, Escórcio C et al. (2011)
- 2491 Dietary protein source or energy levels have no major impact on growth performance,

- nutrient utilisation or flesh fatty acids composition of market-sized Senegalese sole. *Aquaculture* **318**: 128-137.
- 2494 Verburg-van Kemenade BML, Stolte EH, Metz JR, Chadzinska M (2009) Neuroendocrine-
- 2495 immune interactions in teleost fish. In: Bernier NJ, Van Der Kraak G, Farrell AP,
- 2496 Brauner CJ (ed.) Fish Neuroendocrinology. Fish physiology, vol. 28, pp. 313–364.
- 2497 Academic Press, San Diego.
- 2498 Vilar P, Faílde LD, Bermúdez R, Vigliano F, Riaza A, Silva R et al. (2012)
- 2499 Morphopathological features of a severe ulcerative disease outbreak associated with
- 2500 *Tenacibaculum maritimum* in cultivated sole, *Solea senegalensis* (L.). *Journal of Fish*
- 2501 *Diseases* **35**: 437-445.
- 2502 Villalta M, Estévez A, Bransden MP, Bell JG (2005a) The effect of graded concentrations
- of dietary DHA on growth, survival and tissue fatty acid profile of Senegal sole (Solea
- senegalensis) larvae during the Artemia feeding period. Aquaculture **249**: 353–365.
- Villalta M, Estévez A, Bransden MP (2005b) Arachidonic acid enriched live prey induces
 albinism in Senegal sole (*Solea senegalensis*) larvae. *Aquaculture* 245: 193-209.
- alolitistit ili Sellegal sole (*Soled sellegalensis*) fatvae. Aquaculture **245**. 195-209.
- 2507 Villalta M, Estévez A, Bransden MP, Bell JG (2007) Effects of dietary eicosapentaenoic
- acid on growth, survival, pigmentation and fatty acid composition in Senegalese sole
- 2509 (*Solea senegalensis*) larvae during the *Artemia* feeding period. *Aquaculture Nutrition* **13**:
- 2510 1–9.
- Villalta M, Estévez A, Bransden MP, Bell JG (2008a) Effect of dietary eicosapentanoic
 acid on growth, survival, pigmentation and fatty acid composition in Senegal sole (*Solea senegalensis*) larvae during the *Artemia* feeding period. *Aquaculture Nutrition* 14: 232-
- 2514 241.

- Villalta M, Estévez A, Bransden MP, Bell JG (2008b) Arachidonic acid,
 arachidonic/eicosapentaenoic acid ratio, stearidonic acid and eicosanoids are involved in
 dietary-induced albinism in Senegal sole (*Solea senegalensis*). *Aquaculture Nutrition* 14:
 120–128.
- 2519 Viñas J, Asensio E, Cañavate JP, Piferrer F (2013) Gonadal sex differentiation in the
- Senegalese sole (*Solea senegalensis*) and first data on the experimental manipulation of
 its sex ratios. *Aquaculture* 384–387: 74–81.
- 2522 Voss A, Reinhart M, Sankarappa S, Sprecher H (1991) The metabolism of 7,10,13,16,19-
- docosapentaenoic acid to 4,7,10,13,16,19-docosahexaenoic acid in rat liver is
- independent of a 4-desaturase. *The Journal of Biological Chemistry* **266**: 19995–20000.
- Wade MG, Kraak GVD (1993) Arachidonic acid and prostaglandin E2 stimulate
 testosterone production by goldfish testis *in vitro*. *General and Comparative Endocrinology* 90: 109–118.
- 2528 Wagemans F, Vandewalle P (2001) Development of the bony skull in common sole: brief
- survey of morpho-functional aspects of ossification sequence. *Journal of Fish Biology*59: 1350-1369.
- Walker DP, Hill BJ (1980) Studies on the culture assay of infectivity and some in vitro
 properties of Lymphocystis virus. *Journal of General Virology* 51: 385-395.
- 2533 Waring CP, Stagg RM, Poxton MG (1996) Physiological responses to handling in the 2534 turbot. *Journal of Fish Biology* **48**: 161–173.
- 2535 Watanabe K, Washio Y, Fujinami Y, Aritaki M, Uji S, Suzuki T (2008) Adult-type pigment
- cells, which color the ocular sides of flounders at metamorphosis, localize as precursor
- cells at the proximal parts of the dorsal and anal fins in early larvae. *Development*
- 2538 *Growth and Differentiation* **50**: 731–741.

- Wendelaar Bonga SE (1997) The stress response in fish. *Physiological Reviews* 7: 591–
 625.
- 2541 Yamashita H, Mori K, Nakai T (2009) Protection conferred against viral nervous necrosis
- by simultaneous inoculation of aquabirnavirus and inactivated betanodavirus in the
- seven band grouper, *Epinephelus septemfasciatus* (Thunberg). Journal of Fish Diseases
- **32**: 201-210.
- Yúfera M, Fernandez-Diaz C, Pascual E (2005) Food microparticles for larval fish prepared
 by internal gelation. *Aquaculture*, 248, 253-262.
- 2547 Yúfera M, Darias MJ (2007) Changes in the gastrointestinal pH from larvae to adult in
- 2548 Senegal sole (*Solea senegalensis*). *Aquaculture* **267**: 94–99.
- Yúfera M, Arias AM (2010) Traditional polyculture in "esteros" in the Bay of Cádiz
 (Spain). Hopes and expectancies of a unique activity in Europe. *Aquaculture Europe* 35:
 22-25.
- 2552 Zorrilla I, Balebona MC, Moriñigo MA, Sarasquete C, Borrego JJ (1999) Isolation and
- characterization of the causative agent of pasteurellosis, *Photobacterium damsela* spp.
- *piscicida*, from sole, Solea senegalensis. *Journal of Fish Diseases* **22**: 167-172.
- 2555 Zorrilla I, Arijo S, Chabrillon M, Diaz P, Martinez-Manzanares E, Balebona MC et al.
- 2556 (2003) Vibrio species isolated from diseased farmed sole, Solea senegalensis (Kaup),
- and evaluation of the potential virulence role of their extracellular products. *Journal of*
- 2558 *Fish Diseases* **26**:103-108.

2560 Figure legends

2562	Figure 1 Schematic representation of the long-chain polyunsaturated fatty acid (LC-PUFA)
2563	biosynthesis pathway, including the complement of enzymes intervening in the different
2564	steps (not all are necessarily present in a same species). In bold and underligned are the two
2565	enzymes that have been found and characterized in Senegalese sole (Morais et al. 2012).
2566	Dashed arrows correspond to the Sprecher pathway (Voss et al. 1991).
2567	
2568	Figure 2 Total production of farmed sole in the period 2005-2012 (in tonnes) in Europe
2569	(FEAP 2013), including different Spanish regions (APROMAR 2014).
2570	

Initial	Final	SGR	FCR	Time	Temp	Protein	Lipids	Density $(K \mathrm{cm}^{-2})$	Authors
weight (g)	weight (g)			(days)	(°C)	(%)	(%)	(Kgm^{-2})	
9.5	39.8	1.5	0.9	84	20	57	6	-	Silva <i>et al</i> . 2009
9.9	42.5-43.7	1.2	1.0-1.2	84	20	56	4-8	1	Borges et al. 2009
11.9	31.6-33	1.2	1.0	84	21	53-59	10-13	-	Rema et al. 2008
23.6	37.7-43.8	0.7-0.9	2.5-3.1	67	20	51	11-21	-	Rueda Jasso et al. 2004
39.7	-	0.3	0.8	21	22.5	68	15.7	-	Rubio et al. 2009
70	100	-	1.0-1.5	60	19.4	-	-	2-30	Salas Leiton et al. 2008
78.8	113-121.3	0.6-0.66	-	63	20	-	-	4-14	Costas et al. 2008
148.3	-	0.6-0.65	-	60	18.4	-	-	7-30	Salas Leiton et al. 2010
180	323-354	0.5-0.6	2.0-2.4	240	19	59	8-15	36	Valente et al. 2011
317	495.9	0.35	-	126	20	55	15	8.6	Ambrosio et al. 2008

Table 1 Senegalese sole growth data according to bibliography. From Sánchez-Fernández
(2012)

2576 naturally spawning captive wild Senegalese sole (*Solea senegalensis*) broodstock





