Postprint of: Journal of Soils and Sediments 17(8): 2104-2115 (2017)

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1	SOILS, SEC # • RESEARCH ARTICLE	
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3	Humic acid composition and humification processes in wetland soils of a Mediterranean	
4	semiarid wetland	
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18 Abstract

Purpose: The present study focuses on a compositional characterization of the <u>humic acid (HA)</u> fraction of several wetland soils using solid-state ¹³C NMR spectroscopy. The data were analy<u>s</u>ed using the molecular mixing model (MMM), based on an empirical approach by Nelson and Baldock. The compositional data from HAs <u>obtained with this model</u> were used to obtain a wider assessment of the process of humification from comparison of total soil wetland <u>organic matterOM</u> composition and HA composition

25 Materials and methods: Twenty samples of humic acids (HAs) isolated from a Mediterranean semiarid wetland ('Tablas de Daimiel', central Spain) have been were studied using elementary 26 elemental analysis and CPMAS (cross polarization magic angle spinning) ¹²C (CPMAS)¹³C 27 28 nuclear magnetic resonance (NMR) spectroscopy. The NMR data have been were analysed 29 with a-the molecular mixing model (MMM) considering up to six generic components 30 (carbohydrate, protein, lignin, lipid, char, and 'carbonyl'). HAs are considered a conceptual 31 mixture of these model componentsunds, and the MMM determines the proportions of the 32 characteristic biomolecules contributing to the-HA composition.

33 Results and discussion: The composition of the HAs in question under study depends on local 34 factors such as site vegetation and occurrence of fire. Correlations between the proportions of 35 the six generic components and further comparison with those determined for the original entire, unfractionated OM (whole sample, WS), gave information on HA origin and 36 37 humification mechanisms. In particular, the contents-proportions of char and carbohydrate (R^2 38 0.637) and contents of lignin and protein (R^2 0.471) in the HAs were negatively correlated (P <39 0.05). Significant correlations (R^2 0.439) also existed for char contents in whole sample (WS) compared to HA, and for carbohydrates in WS compared to HA (R^2 0.558). Char contents 40 proportion grew in HA with respect to the WS, and -carbohydrates dropped to a half on 41 42 average in HA compared to WS.

43 Conclusions: Two different humification mechanisms could be identified for no-fire and fire
44 areas. In the former, HA-char was preserved selectively from char in the sample, whereas in
45 the latter, char was mainly newly formed by fire effect from preexisting carbohydrates.

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Keywords CPMAS ¹³C NMR spectroscopy • Humification mechanisms with and without fire •
 Molecular mixing method • Wetland soil humic acids

49

51 1. Introduction

52 Humification is assumed to consist of widespread biochemical and abiotic diagenetic reactions 53 leading to recalcitrant, very complex macromolecular compounds (Aiken et al. 1985; 54 Almendros 2008). The resulting humic substances (HS) can originate from alteration of pre-55 existing biomacromolecules polymers but also from abiotic or enzymatic condensation of 56 soluble biogenic precursors (Almendros 2008). Both processes result in the accumulation of 57 material which is more complex than the corresponding precursors and comparatively much 58 more resistant to microbial attack (Almendros 2008). However, it has also been pointed out, 59 that the recalcitrance of natural organic matter (OM), including HS, depends on association 60 with soil minerals, and may not be an intrinsic characteristic of the substrate (Kleber 2010). In 61 any case, humification can be seen as a key step within the wetland carbon cycle, strongly 62 contributing to the important role of wetlands as carbon sinks (Bridgham et al. 2006).

63 The HS are generally divided according to their solubility in aqueous solution (Hayes et al. 64 1989). The fraction only soluble under alkaline conditions is called the humic acid (HA) fraction. 65 HAs are polydisperse, heterogeneous substances. There are several conceptions concerning 66 their nature and composition: According to an early view, they can be seen as a system of 67 micelles of a polymeric nature, with high average molecular masses (20,000-50,000 Da; Aiken 68 et al. 1985). Some authors have considered that HAs do not exist in soil as a definite entity, 69 being mere mixtures of plant and microbial remains and their degradation products (Burdon 70 2001). As a matter of fact, specific chemical structures belonging to HS have not been 71 identified (Kelleher and Simpson 2006; Lehmann et al. 2008); an extended discussion on this 72 topic is given by Kleber and Johnson (2010). The polymeric character of HS is also a matter of 73 discussion: Some authors have introduced terminological insights which consider that HS are 74 supramolecular associations of self-assembling heterogeneous and relatively small molecules 75 (Piccolo 2001). This view is shared by others (Sutton and Sposito 2005), who consider that HS 76 would contain tightly bonded biomolecules or recognizable fragments of them.

77 The present study focuses on a compositional characterization of the HA fraction using solid-78 state ¹³C NMR spectroscopy. The data (from twenty samples of HAs) were analysed using the 79 molecular mixing model (MMM), based on an empirical approach by Nelson and Baldock 80 (Nelson and Baldock (2005) in an attempt to describe the molecular composition of natural OM from quantitative data obtained from ¹³C nuclear magnetic resonance (NMR) 81 spectroscopy. These authors showed that detrital OM, derived from different sources, can be 82 83 described in terms of six generic components (carbohydrate, protein, lignin, lipid, 'pure 84 carbonyl' and char). The contribution of each to the total OM can be inferred from the 85 respective ¹³C NMR spectra by way of a linear combination of spectra of the six generic organic 86 components. The approach of Nelson and Baldock (2005) allows a more specific assignment of 87 the C-groups assigned from NMR to various compound classes. In particular, a better 88 discrimination between sources of aromatic C may help differentiate pyrogenic C forms, i.e., 89 char from other-plant-derived aromatic components. This improves the analysis of the origin of 90 HAs from different precursor macromolecular entities, which is especially appropriate for 91 wetland soils (in most cases peat or histosols) where selective preservation of weakly 92 unaltered domains of biomass-macromolecular biomass constituents is a classically recognized 93 factor in the C sequestration in these ecosystems.

The compositional data from HAs were used to obtain a wider assessment of the process of humification <u>taking advantage of from</u> comparison<u>s between</u>—of total soil wetland OM composition and HA composition. To our knowledge, <u>although</u> MMM has been applied to HAs only_in the study by once (Buurman et al. 2009), there is no previous research on MMM applied to HAs from wetland soils.

99 and never to HAs from wetland soils.

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101 **2.** Materials and methods

102 2.1 Study site

The Tablas de Daimiel is a freshwater wetland in Central Spain on the River Guadiana drainage
basin (39° 08΄ N, 3° 43΄ W; Fig. 1). Its detailed description and environmental history were
recently reported by Rodríguez Murillo et al., (2011) and by Sánchez-Carrillo and Angeler
(2010) and -Rodríguez-Murillo et al., (2011).

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108 2.2 Sample preparation and HA extraction

109 Samples (20) from the upper 20 to 30 cm of the wetland soils from Las Tablas de Daimiel, 110 corresponding to different local environments in the wetland, were collected in March 2007 111 (Fig. 1). In the case of organic soils or sediments, sampling was carried out collecting 30 cm of 112 the uppermost horizon. Only in sandy sediments with low content of organic C the material 113 was collected in the organc horizon, i.e., 4-5 cm thickness topsoil layer. The samples y-were 114 air-dried, ground and sieved to pass a 2 mm sieve. The HAs were extracted following a 115 standard procedure described by (Duchaufour and Jacquin, 1975). Briefly, the soil sample was 116 shaken with 0.1 M Na₄ P₂ O₇ for 5–6 h. After centrifugation at 4,000 rpm –for 5 min₋, the 117 supernatant was separated from the sedimentsolid pellet. The operation was repeated 3+X, 118 and the supernatant solutions were combined. The solid residue was further extracted with 119 0.1 M NaOH by shaking the mixture for 3-4 h and centrifuging at 4,000 rpm for 5 min to 120 separate the supernatant solution from a pellet with the solid phase. The procedure was 121 repeated until the final extract showed a pale colour that did not change in successive extractions (5-10X + depending on the concentration and extractability of organic C). 122 After acidification with 6M HCl of the extract with 6M HCl to pH = 1.5, the HAs were isolated 123 124 as a precipitate from the supernatant which formed the fulvic fraction, after allowing to settle

125 <u>overnight</u>. The HA fraction was re-dissolved in 0.5*M* NaOH, and centrifuged at 20,000 rpm for 5
 126 min to remove clays and particulate OM. The HAs were precipitated again with HCl,

127 centrifuged, and dialyzed for purification with distilled water under continuous agitation until

131 2.3 Sample analysis

Total C, H and N content (%) were determined with a Perkin-Elmer 2400 Series 2 CNHS/O elemental analyszer. <u>The</u>O content was obtained as the difference between 100% and the sum of measured C, N, and H proportions. <u>Based on previous analysis in Tablas de Daimiel</u> wetland, as well as in the examination of the FTIR spectra of the HAs (lack of intense signal in the 600-400 cm⁻¹ range), we assume that ash content is ca. 1% or less, so it should not affect significantly to our results.

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Solid state ¹³C NMR spectra were acquired with a Bruker DSX 200 (4.7 Tesla) operating at 50.29 139 MHz for ¹³C, with HA samples contained in zirconium dioxide rotors of 7 mm diameter in a 140 141 commercial double bearing probe. The magic angle spinning speed was 6.8 kHz. The recycle 142 delay was 300 ms. Cross polarization contact time was 1 ms, and a ramped ¹H-pulse was 143 applied to account for Hartmann-Hahn mismatches. Depending on sample, the number of 144 free-induction decays was between 3000 and 20,000 and the line broadening ranged between 145 25 and 75 Hz. The chemical shifts were calibrated to tetramethylsilane (0 ppm) using glycine as 146 external reference. For spectral interpretation the following ranges and assignations were 147 taken into account-considered (Wilson 1987; Wilson et al. 1988; Preston 1992; Preston et al. 148 1997; Huang et al. 1998): 0–46 ppm, alkyl (13, methyl; 21, acetate; 30, polymethylene), 46–110 149 ppm, O/N-alkyl (56, methoxyl/ α -amino; 73, glucopyranosyde-derived; 103, anomeric C in 150 carbohydrate; 105, quaternary aromatic C in tannins); 110-160 ppm, aromatic/unsaturated 151 [ca. 135: unsubstituted, ca. 145, heterosubstituted: guaiacyl (G) lignins/dihydroxyls of tannins;

 ^{128 &}lt;u>free of chlorides (AgNO₃ test), which took about 8 days</u>. The 20 soil samples resulted in only 18
 129 HAs extracts, since samples 2 and 4 yielded insignificant quantities of HAs.

ca. 153: ether-linked (syringyl (S) lignins)/tannins]; 160–200 ppm, carbonyl (172,
carboxyl/amide; 198, ketone/aldehyde).

Areas in nine spectral regions were measured with ImageJ software (http://rsb.info.nih.gov/ij/)
and normalized to 100.

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157 2.4 Data treatment

158 The molecular mixing model (MMM) of Nelson and Baldock (2005) was applied to the NMR data as described by Rodríguez-Murillo et al., (2011). The used ¹³C_spectral-intensity 159 160 distribution among the seven chemical shift regions for the six components was obtained from 161 Baldock et al. (2004), who also supplied the appropriate elemental atomic H/C, N/C and O/C ratios for the six model components. Based on these data and the ¹³C spectral intensity 162 distribution determined from the ¹³C NMR spectra of our samples, HAs composition was 163 164 calculated in terms of the six generic biomolecules plus 'pure carbonyl' by solving a system of 165 equations for each sample using the Generalized Reduced Gradient nonlinear optimization 166 code in the Solver add-in of Microsoft Excel with two restrictions: (i) the sum of component fractions was forced to be 100 and, (ii) the component fractions were forced to be equal to or 167 168 greater than zero.

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170 3. Results

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171 3.1 <u>Yield of HAs yield</u> and elemental composition

<u>The</u> HA content and composition varied <u>considerably amongwithin</u> the different sampling
 points (Table 1). From the Van Krevelen diagram (Van Krevelen 1950) shown <u>on-in</u> Fig. 2, we
 could distinguish up to three different <u>types-classes</u> of samples: predominantly aliphatic (high

H/C atomic ratio), predominantly aromatic, mature (low H/C), and highly oxidized (high O/Catomic ratio).

The ¹³C NMR CPMAS spectra of the HAs are depicted in Fig. 3<u>a</u>. The respective <u>spectral</u>
intensity distribution, as well as total aromaticity and aliphaticity, are given in Table 2. <u>Two</u>
pairs of spectra (WS and HA), from two differentiated sampling points, are given in Fig. 3b.

Clear <u>geographic</u> differences were evident between HAs, with the highly aliphatic HA corresponding to the upper Tablas sector, with non-permanent flooding and charophyte vegetation in open water areas whereas comparatively more aromatic HAs dominate in lower Tablas, in which the permanent vegetation is dominated by vascular plants. Our data show an increase in C/N of the HAs compared to the <u>whole sample-WS</u> (Rodríguez-Murillo et al. 2011) in 16 of the 18 samples. Only for samples 16 and 19 was the C/N of the <u>whole sample-WS</u> greater than that of the HA.

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188 3.2 HA-composition of the HAs inferred from the MMM

189 The HAs composition obtained from applying the MMM is shown in Fig. 4. We did not 190 introduce the experimental C/N values as a condition in our system of equations, as could be 191 done (Nelson and Baldock 2005), because in the MMM scheme, only peptides are significant N-bearing compounds. This has indeed been confirmed in ¹⁵N NMR spectroscopic studies 192 193 (Knicker 2000), despite the fact that in fire-affected soils, as many of our samples are, a sizeable fraction of organic N could be present in heterocyclic compounds (Almendros et al. 194 195 2003; Knicker 2011), and we would overestimate the proteinaceous contribution assigning all 196 N to this fraction.

As it-is the case in <u>WSwhole-soil</u> samples (Rodríguez- Murillo et al. 2011), the six organic components of the HAs were widespread. Structural components as carbohydrate, protein,

and lipid dominate the HA composition across the wetland, and variable quantities of ligninand char were also present in all the HAs.

201 The reliability of the MMM applied to HAs was assessed in two independent ways:

202 (i) Calculating errors in total spectrum area as the sum of absolute values of differences
 203 between experimental (spectrum) areas and calculated (MMM) areas for each
 204 chemical shift region (Nelson and Baldock 2005).

205 (ii) Calculating H/C and N/C ratios from the composition obtained with the MMM and
 206 comparing the values with the corresponding H/C and N/C values from elementary
 207 elemental analysis.

208 Mean error in total area across HA samples was 2.22 ± 0.75 , greater than that (0.93) found 209 in whole samples WS (Rodríguez- Murillo et al. 2011; Fig. 5a). The spectral regions 210 reproduced most poorly were 95–110 ppm (C1 region of carbohydrate-like structures) and 145-165 ppm (N/O aromatic region). In the former, the mean error for calculated vs. 211 212 measured area was 32% and in the second 4.5%. In the remaining chemical shift regions, 213 differences between calculated and measured spectral intensities were lower. It was 214 estimated that a mean error "x" in determining intensity gives a double error ("2x") in the 215 composition of the HA.

A strong correlation between MMM-calculated and experimental H/C ratio was obtained-, but with some bias;-: calculated H/C values were bigger than experimental H/C ones, i.e. real samples were more aromatic than MMM estimated composition (Fig. 5b). The opposite happened with the N/C ratio, the calculated values being generally lower than the experimental-ones

, but with a less systematic trend in comparison with the H/C <u>ratio</u> case (Fig. 5c). Possible
 causes of the discrepancy between calculated and experimental ratios <u>areis</u>-discussed
 below in Section 4.1.

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3.3. Relationship between among relative proportions of HA components and comparison
 between HAs composition and the whole OM composition of wetland <u>WSsoil samples</u>

The most significant correlations between-among HA components (Table 3) was-were between char and carbohydrate (R^2 -0.637<u>r</u>-0.798), carbonyl and protein (R^2 -0.610<u>r</u>-0.781), lipid and lignin (R^2 -0.502<u>r</u>-0.709), lignin and protein (R^2 -0.471<u>r</u>_0.686), and carbohydrate and protein (R^2 0.463<u>r</u>_0.680). Every significant relationship was negative, except in the case of carbohydrateprotein, carbohydrate-lipid, and carbonyl-char. In the whole-wetland-soil samplesWS, the strongest correlation was lignin-protein, but the char-carbohydrate correlation was also noticeable (Rodríguez-Murillo et al. 2011).

The C/N ratio was positively related to lignin ($R^2 - 0.622 r 0.789$) and carbonyl ($R^2 - 0.273 r 0.522$), and negatively related to the other constituents (mainly to protein, $-R^2 - 0.487 r - 0.698$). No significant relationship was found with char.

237 In Table 4, the HA composition is compared with the composition of the OM of the corresponding whole WS sample. The HAs differ from WSs mainly because of the higher 238 239 aromaticity (relative area of 110–165 ppm region of ¹³C NMR spectra; Fig. 6a). This increase in 240 aromaticity in HA compared to WS as a consequence of progressive humification was fairly 241 regular linear (HA aromaticity = 0.912_x (WS aromaticity) + 10.39; $R^2 - 0.681_r - 0.825_r P < 0.825_r$ 242 0.0001), indicating that HA aromaticity is controlled to a great extent by WS aromaticity 243 indicating that WT and HA aromaticities are significantly related. In the data set of Mahieu et 244 al. (1999) the mean increase in aromatic C content of HAs vs. WSs was 10 %, which is in line Con formato: Fuente: Sin Cursiva

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with our results. The total aromaticity depends strongly upon the sum of lignin content and char content <u>:</u>(94% is explained by lignin + char, which were by far the most significant contributors).

248 The sum of lignin and char in HAs was strongly related to that of WSs (HA = $0.879 \times WS + 15.39$, R^2 -0.570. <u>r</u> 0.755. Significant correlation also existed (Fig 6b) between WS and HA for char 249 250 alone (R^2 -0.439 r 0.663, P = 0.0027) which increased from WS to HA, and for carbohydrates, 251 which decreased sharply in the HA compared to the WS for HA vs. WS-(Fig 6c). In the latter, the 252 253 the HAs, the content of this compound class depended was strongly significantly related to on that of WS from the different sites. Lipid constituents in WS and HA were barely related (R^2) 254 255 0.278 r 0.527), and protein, lignin, and carbonyl showed non-significant relationships (at P < 0.05) between their contribution to HA and to WS (protein, $R^2 - 0.161 r 0.401$; lignin, $R^2 - 0.140 r$ 256 257 <u>0.374;</u> carbonyl, <u>r 0.346</u>).

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259 3.4 Influence of environmental factors in HA-WS component relationships

260 Humification is to a large extent influenced by environmental factors such as aerobic or semi-261 anaerobic conditions for plant detritus diagenesis. Wetlands are subjected to very different 262 diverse local physicochemical processes of variable intensity. In the former case of covidizing 263 environment) the OM tends towards a progressive depletion in carbohydrate, whereas in the 264 latter-case of anaerobic conditions the reducing environment leads to a selective increase of 265 alkyl and lignin constituents (Filip and Alberts 1994). On the other hand, vegetation fires could 266 produce severe thermal modification of existing OM, as well as the differentiation formation of 267 new carbon forms. During a fire, considerable rearrangement of C forms occurs and slightly 268 more refractory and oxidation-resistant organic C forms with increased mean residence time, 269 including black carbon, are formed (Schulze et al. 2000; Almendros et al. 2003).

We analyszed the effect of environmental conditions on humification (i.e. as reflected in the composition of the HA fraction), by studying the relationship between HA and WS separately in the different groups of sampling sites, which could be <u>distinguished based on defined by</u> attending to two factors, viz. flooding regime and fire occurrence, by taking advantage of information recorded from Tablas de Daimiel (Cirujano and Álvarez-Cobelas, personal communication) and our own observations (Table 1).

276 Dividing our sample set into those groups (permanent/non-permanent flooded; fire-277 affected/not fire-affected) and recalculating the correlations between the aromaticity, char 278 and carbohydrate contents for WS and HA (as the most significantly related variables in the 279 whole set of samples) clearly demonstrated the impact of environmental conditions. Further 280 differentiation between sampling sites with permanent and non-permanent flooding (irregular 281 or seasonal in Table 1), showed strong correlation between the aromaticity of HS and of WS 282 for the latter-non-permanent flooding group (r 0.964, P = 0.0001), but only a weak relationship 283 for the former-permanent flooding group (r 0.591, P = 0.072). If fire-affected and unaffected 284 areas were are differentiated, the aromaticity of HA and of WS wss-are not related in fire areas 285 $(r \ 0.533, P = 0.14)$, but shows a tight relationship for no fire areas $(r \ 0.941, P = 0.0002)$. HA 286 aromaticity is higher and more constrained (30 - ca. 45%) in fire areas in comparison with no 287 fire areas (15 - ca. 40%).

288 No significant relationship (Fig. 7b) between char content of HA and of WS char was observed for permanently flooded sites, as well as fire affected areas (r 0.055), but turned into being 289 fairly significant (Fig. 7a) for non-permanently flooded samples (r 0.964, P = 0.0008) and fire-290 291 unaffected areas (r 0.940, P = 0.0002). Char content of HA and WS are significantly correlated 292 in non-permanently flooded samples (r 0.964, P = 0.0008) and fire-unaffected areas (r 0.940, P 293 = 0.0002) (Fig. 7a), but no significant relationship between char content of HA and of WS char 294 was observed for permanently flooded sites, as well as fire-affected areas (r 0.055) (Fig. 7b). 295 The relationship between carbohydrates in HA and WS was relatively strong and positive for 296 <u>non-permanently flooded areas (r 0.821, P = 0.013) (not shown) and fire-unaffected areas (r</u> 297 <u>0.733, P = 0.025) (Fig. 8a), but not significant (Fig. 8b) for in the case of permanently flooded</u> 298 areas and fire-affected areas (r 0.644, P = 0.061) (not shown). -but relatively strong and 299 positive (Fig. 8a) for non-permanently flooded areas (r 0.821, P = 0.013) and fire-unaffected 300 areas (r 0.733, P = 0.025). Insignificant correlation was apparent for the other components 301 (lipid, protein and lignin) of WS and of HA.

302 At the same time, data corresponding to subtraction between the amounts of the different 303 constituents (Fig. 9, a and b) in WS and in HA (WS content minus HA content) in the case of 304 aromaticity (HA aromaticity minus WS aromaticity), showed that there were significant 305 relationships between the increase in aromaticity and the decrease in carbohydrate (r 0.680, P = 0.044), and the increase in char (r 0.764, P = 0.017) for fire-affected areas, and no significant 306 307 relationship for the fire unaffected ones. This confirms that aromaticity increase in fireaffected areas implies a parallel decrease in carbohydrate content (increased difference in 308 carbohydrate content of HA vs. WS). 309

310

311 4 Discussion and conclusions

Previously (Rodríguez-Murillo et al. 2011), we studied the composition of OM in wetland soils 312 of Tablas de Daimiel. Based on a combination of Combining solid-state CPMAS ¹³C NMR 313 spectroscopy with MMM (Nelson and Baldock 2005; Baldock et al. 2004), significant 314 315 differences were found in the composition of wetland soil OM derived from different areas. 316 The differences were ascribed mainly to the nature of the original detrital OM (non-vascular, 317 vascular, mixed and bacterialmicrobial). Additional perturbations such as fires, allochtonous 318 input and movement of sediment (resuspension) could be invoked to justify the presence of 319 lignin and perhaps of char in areas with non-vascular vegetation. Thus, Tablas de Daimiel 320 wetland can be considered as an interesting, mosaic-like ecosystem, with large variation in soil 321 OM characteristics caused by varying vegetation, environmental conditions and disturbances322 in the different areas.

323 Humification processes are also likely to differ in local environments depending on the spatial 324 variability of the microtopography and the periodic or permanent duration of flooding. 325 Carbohydrates are the most labile macromolecular components, and tend to be depleted 326 during the microbial and abiotic reworking of vegetal detritus. Proteins are more resistant, but 327 are also depleted in part, with an effect of increasing the C/N ratio in the HAs vs.versus that of 328 the whole sample. Progressive accumulation of aromatic structures in the course of HA 329 formation is apparent in the sharp increase in lignin-type structures, as well as carbonyl and aromatic char. A Van Krevelen diagram offers some information on HA characteristics. 330 331 Decreasing H/C ratio is frequently interpreted as progressive transformation of the OM 332 towards a greater proportion of aromatic carbon, whereas higher H/C implies a molecular 333 composition more similar to that of plant biomass. Besides, aliphatic samples have low C/N, 334 and aromatic oneshigh C/N, and this can be related to the C/N of original OM (predominantly 335 non-vascular in aliphatic samples and more vascular in mature samples) and/or different 336 degree of transformation.

337 Fig. 2 illustrates the fact that in the Las Tablas ecosystem there are HAs of very different 338 nature: predominantly aliphatic HAs would correspond with WSs containing OM of aliphatic 339 and alkyl character (non-vascular vegetation and microbial origin) (Rodríguez-Murillo et al. 340 2011), indicating that the initial signature of detrital OM is kept through HA extraction. In 341 particular, HA extraction preserves (although more weakly) the negative relationships between 342 lignin and protein found in WSs, perhaps reflecting the original balance between vascular/non-343 vascular vegetation. The main average components of HAs, as calculated with MMM, were 344 lipid > char > protein, whereas in whole sample the main components were protein > lipid > 345 carbohydrate (Table 5). The relative content of carbohydrate is progressively depleted in the course of humification, whereas, as expected, char and—to a lesser extent—lignin, increase in
concentration. Increase in lignin—comparing HA and WS—was not uniform in the samples,
and lignin, often considered the basis for HA structure in the case of peatland soils, was not
the main component identified in our HAs_rapplying the MMM.

350

351 4.1 Error assessment in the determination of composition from MMM

352 An obvious source of error in the determination of HA composition using the MMM could be 353 the occurrence of compounds other than the six generic constituents considered, for instance, 354 tannins. These are These polyphenols, present in higher plants (Kögel-Knabner 2002),- Tannins 355 display an intense NMR signal as two peaks in the 95-110 ppm region-which was the worst 356 reproduced signal with MMM-, due to their aromatic quaternary carbon (Preston et al. 357 1997). The 95–110 ppm region would then show considerable overlapping between anomeric 358 O-alkyl carbons in carbohydrates and aromatic quaternary carbon atoms in tannins and 359 dilignols. Although tannins would not be are usually far lessas abundant as than the other 360 MMM constituents, only 16.7% of carbohydrates, 8.6% of lignin and 6.3% of char peak in the 95-110 ppm region, making it possible that which is compatible with a small proportion of 361 362 tannins carbons could makeing a significant contribution to the signal in the 95-110 ppm 363 range. Incomplete agreement between experimental and simulated spectra (i.e. errors in total spectrum area) might also be attributed to experimental errors in ¹³C-NMR spectra of HAs but, 364 365 considering the better agreement obtained in many cases using natural organic materials (Nelson and Baldock 2005; Rodríguez-Murillo et al. 2011) instead of HAs, the occurrence in HAs 366 of specific chemical structures with special ¹³C-NMR signatures not considered in the MMM 367 368 treatment could be an important factor explaining the observed discrepancies. Unfortunately, 369 our data and their analysis can neither prove nor disprove the presence of such structures, 370 which should be quantitatively minor components.

371	Systematic discrepancies in experimental vs. calculated H/C (Fig. 5b) and N/C (Fig. 5c) ratios
372	were also probably due to the above considerations. Problems of NMR in accurately detecting
373	components with low proton content are also known (Knicker et al. 2005). If this were the case
374	here, an underestimation of aromaticity would result. However, we observed neither an
375	unusual lowering of the sensitivity for the sample nor problems during tuning the sample prior
376	to measurement, which indicates that graphenic structures were present. There was no
377	apparent correlationship of differences in experimental and calculated H/C (Fig. 5b) and with
378	sample aromaticity or <u>sample char content</u> , although as H/C (realexperimental) increased (less
379	low proton components), differences in H/C were smaller. The existence of a significant
380	proportion of N in heterocyclic structures could explain discrepancies in N/C ratio.

382 4.2 Humification mechanisms

383 In the areas permanently flooded in the past, fire has been a recent perturbation of Las Tablas, 384 coinciding with the first desiccation events (around 1960). Such fires, especially in peat zones, 385 have affected humus formation, as could be deduced from the different relationships of char 386 and carbohydrate content for fire and no-fire areas. At first sight, differences in humification 387 could be empirically ascribed to either fire action or flooding regime; the existence-occurrence 388 of fire seems to produce in our system seems to parallel the similar effects as of the permanent 389 flooding in HA - WS component relationships but, as discussed below, fire effects could 390 explain these relationships, which is not the case for flooding. This coincidence between HA 391 and WS relationships in areas subjected to such different environmental influences (flooding 392 and fire) is accidental and probably arises from the fact that fires occur mainly in former 393 permanently flooded areas, after dessication. Fire consumes OM, but, at the same time, 394 increases aromaticity and the content of refractory substances, such as char, which is are a 395 potential contributor to the formation of refractory long-term carbon sinks (Forbes et al.,

2006). This is due to selective enrichment of residual heat-resistant, strongly aromatic
components and also to endothermic neoformation reactions involving carbohydrates, but
also amino acids and unsaturated alkyl chains (González-Pérez et al. 2004).

The concentration of char in HA was unrelated to the accumulation of WS char in fire areas, but depended linearly on WS char in no fire areas (Fig. 7a, b). If we plot the differences in carbohydrates and char between WS and HAs vs. the differences in aromaticity between HA and WS, we observe a linear relationship in fire affected areas and no significant relationship in the unaffected ones (Fig. 9a, b).

404 We can hypothesize that, in no-fire areas, HA 'char' comes mainly from char formerly 405 introduced into the sediment, as suggested by Haumaier and Zech (1995), and is selectively 406 preserved, whereas in fire-affected areas, char is mainly newly formed in situ as a 407 consequence of fire action. New formation of char in HAs in fire affected areas could explain 408 the loss of relationship between WS char and HA char in these areas. from preexisting carbohydrates (González-Pérez 2004; Almendros et al. 2003) . The increase in aromaticity with 409 410 humification in fire areas implies a parallel decrease in carbohydrate content (increased 411 difference in carbohydrate of HA vs. WS; Fig. 9a), pointing to an accumulation of newlyformed—pyrogenic—aromatic structures in HA from carbohydrate in the original sample via 412 413 fire. In contrast, change in carbohydrate content in HA in fire unaffected areas with respect to 414 WS does not show a relationship with change in char or aromatic content, as HA 'char' derives 415 mainly from WS char (Fig. 9b). Then, analysis of HA composition using MMM allows some 416 insight into humification mechanisms, comparing HA and whole sample composition: MMM 417 char and carbohydrate determination and constituent comparison between WS and HA allows 418 the detection of two contrasting HA-formation mechanisms, depending of on the occurrence 419 (or not) of fire.

421 Acknowledgements This study was supported by project 81/2005 of the Spanish CICYT and 422 Organismo Autónomo Parques Nacionales, and by project CGL2008-04296. We wish to thank 423 M. Álvarez-Cobelas and S. Cirujano for environmental information about Tablas, and the staff 424 of the National Park "Las Tablas de Daimiel" for their help, as well as the Analytical Unit staff of 425 MNCN and our laboratory assistants. The NMR measurements were performed at the 426 Lehrstuhl für Bodenkunde of the TU-München-Weihenstephan, Germany, in the frame of the 427 Acciones Integradas Program, within which the exchange of involved scientists was financed by 428 the DAAD (Germany)-CICyT(Spain).

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510	FIGURE CAPTIONS	
511	Fig. 1 Location of Tablas de Daimiel wetland and sampling points.	
512	Fig. 2 Van Krevelen's diagram of humic acids (HAs) from the sampling points of Tablas de	
513	Daimiel.	
514	Fig. 3 NMR spectra of the HAs.a) NMR spectra of the HAs. F: At least a fire has been recorded	Con formato: Inglés (Reino Unido)
515	in the area of the sampling point. NF: No fire recorded.	
516	b) Comparison between ¹³ C NMR spectra of whole sample and its humic acids: the spectral	
517	ranges for the main C-types are indicated.	
518	Fig. 4 HA composition according to the molecular mixing model (MMM). The HAs-are ordered	
519	from lower aromaticity (bottom) to higher aromaticity (top). I, irregular; S, seasonal; P,	
520	permanent. Flooding regime refers to that before 1973, year of the onset of large perturbation	
521	in the hydrological regime of the Tablas. Fires are those recorded before sampling year (2007);	
522	at sites 15, 17, and 17', the existence of fires in the past has been inferred from the occurrence	
523	of charred material in the samples. "Y" <u>means that</u> , at least one fire has been recorded <u>and</u> ;	
524	"N", <u>means</u> no fire recorded in the site.	
525	Fig. 5 a) Performance of MMM. Area error is the sum of absolute values of differences (%)	Con formato: Espacio Después: 0
526	between experimental and calculated areas for each chemical shift region in each sample;	latino y asiático, No ajustar espacio entre texto asiático y números
527	b; Performance of MMM. Calculated vs experimental H/C ratios of the samples. The lower line	Con formato: Inglés (Reino Unido)
528	represents the perfect concordance calculated-experimental; c) Performance of MMM.	Con formato: Inglés (Reino Unido)
529	Calculated vs experimental N/C ratios of the samples.	
530	(a) Performance of MMM. Calculated vs. experimental H/C ratio for the samples; point-	
531	and dash line represents perfect agreement between calculated and experimental.	
532	(b) Performance of MMM. Calculated vs. experimental N/C ratios for the samples; point-	
533	and dash line as in (b).	

534	Fig. 6 Comparison of components of HAs with components of OM-organic matter in whole			
535	samples (WS) (aromaticity, char and carbohydrate), according to MMM.			
536	a) Comparison of aromaticities.		Con formato: Inglés (Peino Unido)	1
550		\leq	Con formato: Interlineado: Doble	Ì
537	b) Comparison of char contents.			ĺ
538	c) Comparison of carbohydrate contents.			
539	Fig. 7 Comparison of components of HAs with components of OM-organic matter in whole			
540	samples (WS), according to MMM. Effect of fire on char content change.			
541	(a) Comparison of char content in no fire areas.			
542	(b) Comparison of char content in fire areas.			
543	Fig. 8 Comparison of components of HAs with components of organic matter in whole samples		Con formato: Fuente: 11 pto, Negrita)
		$\langle \rangle$	Con formato: Interlineado: Doble	j
544	(WS) according to MMM. Effects of fire on carbohydrate content change,	\sim	Con formato: Fuente: 11 pto	J
545	Fig. 8 Comparison of components of HAs with components of OM in whole samples (WS),		Con formato: Fuente: 11 pto, Inglés (Reino Unido)	J
546	according to MMM. Effect of fire on carbohydrate content change. (a) Comparison of			
547	carbohydrate content in fire areas. (b) Comparison of carbohydrate content in no fire areas.			
548	Fig. 9 Differences in char content (in %) versus differences in carbohydrate (CH) content between WS			
549	and HA . a) Fire areas. b) No fire areas Differences in carbohydrate and char content (%) between			
550	WS and HA vs. aromaticity differences (%). (a) Fire areas. b) No fire areas			
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<u>7 53</u>	<u>43.5</u>	<u>52.8</u>	<u>4.07</u>	<u>2.96</u>	<u>40.1</u>	<u>17.8</u>	<u>0.57</u>	<u>0.92</u>	1	Y
<u>8 86</u>	<u>13.2</u>	<u>51.7</u>	<u>4.12</u>	<u>3.78</u>	<u>40.4</u>	<u>13.7</u>	<u>0.59</u>	<u>0.96</u>	<u>s</u>	Y
<u>9 365</u>	8.85	<u>52.9</u>	<u>4.06</u>	<u>2.49</u>	<u>40.5</u>	<u>21.3</u>	<u>0.57</u>	<u>0.92</u>	<u>P</u>	<u>Y</u>
<u>0 83</u>	<u>14.0</u>	<u>53.9</u>	<u>4.34</u>	<u>3.03</u>	<u>38.7</u>	<u>17.8</u>	<u>0.54</u>	<u>0.97</u>	P	Y
<u>1 148</u>	<u>8.81</u>	<u>53.3</u>	<u>4.44</u>	<u>2.82</u>	<u>39.4</u>	<u>18.9</u>	<u>0.55</u>	<u>1.00</u>	P	<u>Y</u>
<u>2 92</u>	<u>11.3</u>	<u>50.1</u>	<u>4.11</u>	<u>3.52</u>	<u>42.3</u>	<u>14.2</u>	<u>0.63</u>	<u>0.98</u>	P	Y
<u>3 88</u>	<u>8.13</u>	<u>51.5</u>	<u>4.90</u>	<u>4.49</u>	<u>39.1</u>	<u>11.5</u>	<u>0.57</u>	<u>1.14</u>	P	<u>N</u>
<u>4 133</u>	<u>14.2</u>	<u>51.2</u>	<u>3.84</u>	<u>3.10</u>	<u>41.9</u>	<u>16.5</u>	<u>0.61</u>	<u>0.90</u>	P	<u>N</u>
<u>5 200</u>	<u>5.73</u>	<u>51.4</u>	<u>4.34</u>	<u>3.90</u>	<u>40.3</u>	<u>13.2</u>	<u>0.59</u>	<u>1.01</u>	1	<u>Y</u>
<u>6 19</u>	<u>2.69</u>	<u>49.9</u>	<u>4.64</u>	<u>4.03</u>	<u>41.4</u>	<u>12.4</u>	<u>0.62</u>	<u>1.12</u>	P	<u>N</u>
<u>7 132</u>	24.7	<u>52.5</u>	<u>4.20</u>	<u>3.58</u>	<u>39.8</u>	<u>14.7</u>	<u>0.57</u>	<u>0.96</u>	P	<u>Y</u>
<u>7' 14</u>	<u>28.7</u>	<u>51.9</u>	<u>4.37</u>	<u>3.30</u>	<u>40.5</u>	<u>15.7</u>	<u>0.59</u>	<u>1.01</u>	P	<u>Y</u>
<u>8 37</u>	<u>8.40</u>	<u>50.1</u>	<u>3.93</u>	<u>3.93</u>	<u>42.0</u>	<u>12.8</u>	<u>0.63</u>	<u>0.94</u>	1	<u>N</u>
<u>9 63</u>	<u>11.9</u>	<u>50.8</u>	<u>4.29</u>	<u>4.18</u>	<u>40.7</u>	<u>12.3</u>	<u>0.60</u>	<u>1.01</u>	P	<u>N</u>
	Z 53 B $B6$ 9 365 0 83 1 148 2 92 3 88 4 133 5 200 6 19 7 132 $7'$ 14 8 37 9 63	Z 53 43.5 8 86 13.2 9 365 8.85 0 83 14.0 1 148 8.81 2 92 11.3 3 88 8.13 4 133 14.2 5 200 5.73 6 19 2.69 7 132 24.7 7' 14 28.7 8 37 8.40 9 63 11.9	Z 53 43.5 52.8 8 86 13.2 51.7 9 365 8.85 52.9 0 83 14.0 53.9 1 148 8.81 53.3 2 92 11.3 50.1 3 88 8.13 51.5 4 133 14.2 51.2 5 200 5.73 51.4 6 19 2.69 49.9 7 132 24.7 52.5 7' 14 28.7 51.9 8 37 8.40 50.1 9 63 11.9 50.8	Z 53 43.5 52.8 4.07 8 86 13.2 51.7 4.12 9 365 8.85 52.9 4.06 0 83 14.0 53.9 4.34 1 148 8.81 53.3 4.44 2 92 11.3 50.1 4.11 3 88 8.13 51.5 4.90 4 133 14.2 51.2 3.84 5 200 5.73 51.4 4.34 6 19 2.69 49.9 4.64 7 132 24.7 52.5 4.20 $7'$ 14 28.7 51.9 4.37 8 37 8.40 50.1 3.93 9 63 11.9 50.8 4.29	Z 53 43.5 52.8 4.07 2.96 8 86 13.2 51.7 4.12 3.78 9 365 8.85 52.9 4.06 2.49 0 83 14.0 53.9 4.34 3.03 1 148 8.81 53.3 4.44 2.82 2 92 11.3 50.1 4.11 3.52 3 88 8.13 51.5 4.90 4.49 4 133 14.2 51.2 3.84 3.10 5 200 5.73 51.4 4.34 3.90 6 19 2.69 49.9 4.64 4.03 7 132 24.7 52.5 4.20 3.58 7' 14 28.7 51.9 4.37 3.30 8 37 8.40 50.1 3.93 3.93 9 63 11.9 50.8 4.29 4.18	Z 53 43.5 52.8 4.07 2.96 40.1 B 86 13.2 51.7 4.12 3.78 40.4 9 365 8.85 52.9 4.06 2.49 40.5 0 83 14.0 53.9 4.34 3.03 38.7 1 148 8.81 53.3 4.44 2.82 39.4 2 92 11.3 50.1 4.11 3.52 42.3 3 88 8.13 51.5 4.90 4.49 39.1 4 133 14.2 51.2 3.84 3.10 41.9 5 200 5.73 51.4 4.34 3.90 40.3 6 19 2.69 49.9 4.64 4.03 41.4 7 132 24.7 52.5 4.20 3.58 39.8 7' 14 28.7 51.9 4.37 3.30 40.5 8 37 8.40 50.1 3.93 3.93 42.0 9	Z 53 43.5 52.8 4.07 2.96 40.1 17.8 B 86 13.2 51.7 4.12 3.78 40.4 13.7 9 365 8.85 52.9 4.06 2.49 40.5 21.3 0 83 14.0 53.9 4.34 3.03 38.7 17.8 1 148 8.81 53.3 4.44 2.82 39.4 18.9 2 92 11.3 50.1 4.11 3.52 42.3 14.2 3 88 8.13 51.5 4.90 4.49 39.1 11.5 4 133 14.2 51.2 3.84 3.10 41.9 16.5 5 200 5.73 51.4 4.34 3.90 40.3 13.2 6 19 2.69 49.9 4.64 4.03 41.4 12.4 7 14 28.7 51.9 4.37 3.30 40.5 15.7 8 37 8.40 50.1 3.93 3.	Z 53 43.5 52.8 4.07 2.96 40.1 17.8 0.57 B 86 13.2 51.7 4.12 3.78 40.4 13.7 0.59 9 365 8.85 52.9 4.06 2.49 40.5 21.3 0.57 0 83 14.0 53.9 4.34 3.03 38.7 17.8 0.54 1 148 8.81 53.3 4.44 2.82 39.4 18.9 0.55 2 92 11.3 50.1 4.11 3.52 42.3 14.2 0.63 3 88 8.13 51.5 4.90 4.49 39.1 11.5 0.57 4 133 14.2 51.2 3.84 3.10 41.9 16.5 0.61 5 200 5.73 51.4 4.34 3.90 40.3 13.2 0.57 4 133 14.2 51.5 4.20 3.58 39.8 14.7 0.57 6 19 2.69 49.9	Z 53 43.5 52.8 4.07 2.96 40.1 17.8 0.57 0.92 B 86 13.2 51.7 4.12 3.78 40.4 13.7 0.59 0.92 9 365 8.85 52.9 4.06 2.49 40.5 21.3 0.57 0.92 0 83 14.0 53.9 4.34 3.03 38.7 17.8 0.54 0.97 1 148 8.81 53.3 4.44 2.82 39.4 18.9 0.55 1.00 2 92 11.3 50.1 4.11 3.52 42.3 14.2 0.63 0.98 3 88 8.13 51.5 4.90 4.49 39.1 11.5 0.57 1.14 4 133 14.2 51.2 3.84 3.10 41.9 16.5 0.61 0.99 5 200 5.73 51.4 4.34 3.90 40.3 13.2 0.57 0.91 6 19 2.69 49.9 4.64	Z 53 43.5 52.8 4.07 2.96 40.1 17.8 0.57 0.92 1 B 86 13.2 51.7 4.12 3.78 40.4 13.7 0.59 0.96 S 9 365 8.85 52.9 4.06 2.49 40.5 21.3 0.57 0.92 1 0 83 14.0 53.9 4.34 3.03 38.7 17.8 0.54 0.97 1 1 148 8.81 53.3 4.44 2.82 39.4 18.9 0.55 1.00 1 2 92 11.3 50.1 4.11 3.52 42.3 14.2 0.63 0.98 1 2 92 11.3 50.1 4.11 3.52 42.3 14.2 0.63 0.98 1 3 88 8.13 51.5 4.90 4.49 39.1 11.5 0.57 1.14 1 4 133 14.2 51.2 3.84 3.10 41.9 16.5 0.61

573 ^a g C in the HA/g C in 100 g of total sample from which the HA was extracted;

574 ^b I: irregular; S: seasonal; P: permanent. Flooding regime refers to that before 1973, year of the

575 <u>onset of large perturbation in the hydrological regime of the Tablas. Fires are those recorded</u>

576 before sampling year (2007); at sites 15, 17, and 17', the existence of fires in the past has been

577 <u>inferred from the occurrence of charred material in the samples.</u>

⁵⁷²

579 Table 2

580 ¹³C NMR areas from HAs from wetland soils of Las Tablas de Daimiel^a.

	<u>Chemical</u> <u>shift</u>	<u>Alkyl</u>	<u>Methoxyl</u>	<u>C6</u>	<u>O–</u> <u>alkyl</u>	<u>C1</u>	<u>H-</u> arom	<u>O/N-arom</u>	<u>Carboxil</u>	Carbonyl	Aliphaticity	Aromaticity
ļ	<u>range</u> (ppm)	<u>0–45</u>	<u>46–58</u>	<u>59–65</u>	<u>66–</u> <u>94</u>	<u>95–</u> <u>110</u>	<u>111–</u> <u>146</u>	<u>147–167</u>	<u>168–188</u>	<u>189–200</u>	<u>111–167</u>	<u>0–110</u>
	Sample											
	<u>1</u>	<u>32.02</u>	<u>9.01</u>	<u>4.04</u>	<u>14.47</u>	<u>4.11</u>	<u>16.47</u>	<u>6.03</u>	<u>11.90</u>	<u>1.95</u>	<u>22.50</u>	<u>63.64</u>
	<u>3</u>	<u>29.87</u>	<u>12.12</u>	<u>3.85</u>	<u>13.20</u>	<u>3.18</u>	<u>17.33</u>	<u>5.71</u>	<u>12.46</u>	<u>2.29</u>	<u>23.04</u>	<u>62.21</u>
	<u>5</u>	<u>36.52</u>	<u>8.39</u>	<u>3.58</u>	<u>14.37</u>	<u>3.68</u>	<u>14.01</u>	<u>5.24</u>	<u>12.15</u>	<u>2.06</u>	<u>19.25</u>	<u>66.53</u>
	<u>6</u>	<u>35.85</u>	<u>10.01</u>	<u>4.96</u>	<u>18.31</u>	<u>5.03</u>	<u>11.03</u>	<u>3.28</u>	<u>10.34</u>	<u>1.19</u>	<u>14.31</u>	<u>74.16</u>
	<u>7</u>	<u>17.61</u>	<u>8.57</u>	<u>3.36</u>	<u>8.66</u>	<u>4.99</u>	<u>30.33</u>	<u>12.73</u>	<u>11.64</u>	<u>2.11</u>	<u>43.05</u>	<u>43.20</u>
ļ	<u>8</u>	<u>21.83</u>	<u>8.01</u>	<u>3.74</u>	<u>10.62</u>	<u>4.31</u>	<u>27.55</u>	<u>10.21</u>	<u>11.85</u>	<u>1.88</u>	<u>37.76</u>	<u>48.51</u>
	<u>9</u>	<u>22.31</u>	<u>8.23</u>	<u>3.02</u>	<u>11.42</u>	<u>4.59</u>	<u>23.25</u>	<u>9.94</u>	<u>14.20</u>	<u>3.05</u>	<u>33.19</u>	<u>49.57</u>
ļ	<u>10</u>	<u>24.60</u>	<u>8.33</u>	<u>3.74</u>	<u>10.22</u>	<u>3.53</u>	<u>24.14</u>	<u>9.83</u>	<u>12.89</u>	<u>2.72</u>	<u>33.96</u>	<u>50.42</u>
ļ	<u>11</u>	<u>20.76</u>	<u>9.89</u>	<u>4.05</u>	<u>9.26</u>	<u>4.44</u>	<u>27.81</u>	<u>11.40</u>	<u>10.38</u>	<u>2.01</u>	<u>39.20</u>	<u>48.41</u>
	<u>12</u>	<u>29.12</u>	<u>7.24</u>	<u>2.93</u>	<u>9.50</u>	<u>2.53</u>	<u>24.44</u>	<u>7.03</u>	<u>14.51</u>	<u>2.70</u>	<u>31.47</u>	<u>51.32</u>
	<u>13</u>	<u>30.25</u>	<u>9.02</u>	<u>3.32</u>	<u>11.09</u>	<u>3.43</u>	<u>20.31</u>	<u>7.39</u>	<u>13.14</u>	<u>2.05</u>	<u>27.70</u>	<u>57.11</u>
	<u>14</u>	<u>20.34</u>	<u>7.41</u>	<u>2.98</u>	<u>9.60</u>	<u>4.47</u>	<u>27.62</u>	<u>10.62</u>	<u>14.40</u>	<u>2.56</u>	<u>38.24</u>	<u>44.80</u>
ļ	<u>15</u>	<u>25.68</u>	<u>8.60</u>	<u>2.80</u>	<u>9.57</u>	<u>2.93</u>	<u>24.57</u>	<u>9.05</u>	<u>14.40</u>	<u>2.41</u>	<u>33.62</u>	<u>49.57</u>
	<u>16</u>	<u>30.14</u>	<u>7.89</u>	<u>2.86</u>	<u>11.40</u>	<u>2.94</u>	<u>21.12</u>	<u>6.79</u>	<u>14.29</u>	<u>2.56</u>	<u>27.91</u>	<u>55.24</u>
	<u>17</u>	<u>25.20</u>	<u>8.25</u>	<u>2.96</u>	<u>10.92</u>	<u>3.66</u>	<u>24.23</u>	<u>8.68</u>	<u>13.88</u>	<u>2.22</u>	<u>32.91</u>	<u>50.99</u>
ļ	<u>17´</u>	<u>27.65</u>	<u>8.13</u>	<u>3.20</u>	<u>9.64</u>	<u>2.96</u>	<u>22.75</u>	<u>8.47</u>	<u>14.50</u>	<u>2.71</u>	<u>31.22</u>	<u>51.58</u>
ļ	<u>18</u>	<u>23.32</u>	<u>7.39</u>	<u>3.10</u>	<u>8.86</u>	<u>2.82</u>	<u>28.94</u>	<u>9.90</u>	<u>13.47</u>	<u>2.20</u>	<u>38.84</u>	<u>45.50</u>
	<u>19</u>	<u>29.32</u>	<u>8.41</u>	<u>3.74</u>	<u>12.10</u>	<u>2.91</u>	<u>19.66</u>	<u>6.71</u>	<u>14.72</u>	<u>2.44</u>	<u>26.37</u>	<u>56.47</u>

582	^a Aromaticity obtained from the NMR spectra strongly correlated with the elementary
583	composition; H/C was strongly (negatively) related to HA aromaticity (R^2 0.794, $p < 0.0001$);
584	the relationship between C/N and aromaticity was weak and positive (R^2 0.310, p = 0.016).
585	

<u>Table 3</u>

587 <u>Pearson's linear correlation coefficients *r* for HA components obtained from MMM model and</u>

588 <u>C/N ratio ^a [n.s. non-significant correlation (P > 0.05)].</u>

Carbohydrate						
<u>0.681</u>	<u>Protein</u>					
<u>0.0019</u>						
<u>-0.563</u>	<u>-0.687</u>	<u>Lignin</u>				
<u>0.015</u>	<u>0.0017</u>					
<u>0.649</u>	<u>n.s.</u>	<u>-0.709</u>	<u>Lipid</u>			
<u>0.036</u>		<u>0.001</u>				
<u>-0.691</u>	<u>-0.781</u>	<u>n.s.</u>	<u>n.s.</u>	<u>Carbonyl</u>		
<u>0.0015</u>	<u>0.0001</u>					
<u>-0.797</u>	<u>-0.502</u>	<u>n.s.</u>	<u>-0.571</u>	<u>0.528</u>	<u>Char</u>	
<u>0.0001</u>	<u>0.034</u>		<u>0.013</u>	<u>0.024</u>		
<u>-0.547</u>	<u>-0.698</u>	<u>0.789</u>	<u>-0.497</u>	<u>0.523</u>	<u>n.s.</u>	<u>C/N</u>
<u>0.019</u>	<u>0.0013</u>	<u><0.0001</u>	<u>0.036</u>	<u>0.026</u>		

590 ^a *P* value of correlation in italics.

593 <u>Table 4</u>

594 <u>Pearson's linear correlation coefficients *r* for HA components and corresponding WS</u>

595

components obtained from the MMM model [n.s., non-significant correlation (P > 0.05)].^a

<u>Component</u>	<u>Carbohydrate</u>	<u>Protein</u>	<u>Lignin</u>	<u>Lipid</u>	<u>Carbonyl</u>	<u>Char</u>	<u>Lignin + char</u>
Ĺ	<u>0.747</u>	<u>0.401</u>	<u>0.374</u>	<u>0.527</u>	<u>0.346</u>	<u>0.663</u>	<u>0.755</u>
p	<u>0.0004</u>	<u>n.s.</u>	<u>n.s.</u>	<u>0.025</u>	<u>n.s.</u>	<u>0.0027</u>	<u>0.0003</u>
^a P value of co	orrelation in italics						

598

599 <u>Table 5</u>

600 Average organic composition of whole samples and HAs (MMM results)

	<u>Whole</u>	
	<u>sample^a</u>	Humic acid
<u>Carbohydrate</u>	<u>19.9</u>	<u>10.7</u>
<u>Protein</u>	<u>24.8</u>	<u>20.7</u>
<u>Lignin</u>	<u>14.8</u>	<u>18.8</u>
<u>Lipid</u>	<u>24.6</u>	<u>22.8</u>
<u>Carbonyl</u>	<u>2.2</u>	<u>5.4</u>
<u>Char</u>	<u>13.7</u>	<u>21.7</u>

601 ^a Data from Rodríguez-Murillo et al .,(2011).





Location of Tablas de Daimiel wetland and sampling points





Van Krevelen's diagram of humic acids (HAs) from the sampling points of Tablas de Daimiel



Fig. 3

a NMR spectra of the HAs. *F*: at least a fire has been recorded in the area of the sampling point. *NF* no fire recorded. **b** Comparison between ¹³C NMR spectra of whole sample and its humic acids: the spectral ranges for the main C-types are indicated



Fig. 4

HA composition according to molecular mixing model (MMM). HAs are ordered from lower aromaticity (*bottom*) to higher aromaticity (*top*). *I* irregular; *S* seasonal; *P* permanent. Flooding regime refers to that before 1973, year of the onset of large perturbation in the hydrological regime of the Tablas. Fires are those recorded before sampling year (2007); at sites 15, 17, and 17', the existence of fires in the past has been inferred from the occurrence of charred material in the samples. *Y* means that at least one fire has been recorded and *N* means no fire recorded in the site

605



Fig. 5

a Performance of MMM. Area error is the sum of absolute values of differences (%) between experimental and calculated areas for each chemical shift region in each sample. b Performance of MMM. Calculated versus experimental H/C ratios of the samples. The *lower line* represents the perfect concordance calculated–experimental. c Performance of MMM. Calculated versus experimental N/C ratios of the samples



Fig. 6 Comparison of components of HAs with components of organic matter in whole samples (WS) according to MMM. a Comparison of aromaticities. b Comparison of char contents. ${\bf c}$ Comparison of carbohydrate contents



Con formato: Español (alfab. internacional)