

1 Addressing the recovery of feeding rates in post-exposure feeding bioassays: *Cyathura*
2 *carinata* as a case study

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24 **Abstract**

25 Post-exposure bioassays are used in environmental assessment as a cost-effective tool,
26 but the effects of organism's recovery after exposure to pollutant has not yet been
27 addressed in detail. The recoveries of post-exposure feeding rates after being exposed to
28 two sublethal concentrations of cadmium during two different exposure periods (48 h
29 and 96 h) were evaluated under laboratory conditions using the estuarine isopod
30 *Cyathura carinata*. Results showed that feeding depression was a stable endpoint up to
31 24 h after cadmium exposure, which is useful for ecotoxicological bioassays.

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33 **Keywords:** aquatic invertebrates; Cd; feeding inhibition; sublethal endpoint.

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35

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61 **1. Introduction**

62 The feeding inhibition of aquatic animals has been shown to be a rapid, sensitive and
63 ecologically relevant nonlethal toxicity test endpoint that can be used to assess toxic
64 stress in ecotoxicology (Allen et al., 1995; Maltby et al., 2002). Since directly
65 measuring feeding rates (FR) during an assay can be extremely difficult, post-exposure
66 feeding inhibition has received a great deal of attention during laboratory and field
67 assays in recent years (e.g., McWilliam and Baird, 2002; Moreira et al., 2005). Post-
68 exposure feeding inhibition is based on the fact that the FR of an organism that is
69 exposed to sublethal concentrations of contaminants may remain inhibited after transfer
70 from a contaminated to an uncontaminated medium. However, few studies address the
71 recovery of feeding inhibition in bioassays based on post-exposure feeding depression
72 (McWilliam and Baird, 2002; Taylor et al., 1998).

73 The objective of this research was therefore to determine the time period during
74 which the FR of a test species remains inhibited after the exposure period, i.e., without
75 contact with the pollutant (post-exposure time), in order to establish a safe window in
76 which the FR endpoint signals the contact with the toxicant. This led us to evaluate the
77 stability of the post-exposure FR response up to 24 hours after two different
78 contaminant exposure periods (48 h and 96 h) for two sublethal concentrations of
79 cadmium (Cd, as a reference toxicant) in laboratory conditions. The experiments were
80 performed using the isopod *Cyathura carinata* for which the inhibition of median
81 feeding rate after 48-h post exposure to Cd concentration has been described to be 7-
82 fold lower than the corresponding 48-h LC50 (Martinez-Haro et al., 2014). *Cyathura*
83 *carinata* is a widely distributed and abundant species. It is an ecologically important
84 element of soft bottom benthic communities, playing a major role in estuarine trophic
85 webs (Marques et al., 1994). As an omnivorous species, it feeds on macroalgae, small

86 invertebrates and detritus (Wägele, 1980). These features make to *C. carinata* a good
87 test species for large-scale monitoring of the ecological status of estuaries.

88

89 **2. Material and Methods**

90 *2.1. Sample Collection*

91 Animal sampling was carried out at low tide in the Mondego estuary (Western Coast of
92 Portugal: 40°08' N, 8°50' W), where this species is abundant and well established
93 (Marques et al., 1994). The animals were collected by sieving the first 3 cm of the
94 sediment with local water through a 1-mm mesh. They were then placed in plastic
95 containers with field water and transported to the laboratory in insulated boxes. See
96 Martinez-Haro et al. (2014) for complete details of the sample collection.

97

98 *2.2. Culture conditions*

99 Once in the laboratory, the animals were sorted by size. Mean total body length (\pm SD),
100 which was estimated using cephalic measurements following the equation: total body
101 length = $0.745 + 9.01 \times$ cephalic length (see Marques et al., 1994), was 5.9 ± 0.6 mm, n
102 = 20. Since sexual maturation has been reported to occur from 8.13 mm (Marques et al.,
103 1994), the organisms used in the present work were undifferentiated animals correspond
104 to juvenile phase. The animals were maintained at 20 °C during a 12 h:12 h light:dark
105 photoperiod, in plastic containers filled with a 2-cm layer of clean fine aquarium sand
106 (<0.5 mm) and a 5- to 8-cm height of field water that was continuously aerated (for
107 details, see Martinez-Haro et al., 2014). They were fed daily *ad libitum* with the
108 defrosted nauplii (less than 24-h old) of brine shrimp *Artemia franciscana* Kellogg. The
109 organisms were progressively acclimatized from field to reconstituted seawater
110 (hereafter RSW; ~ 33 g/l salinity; J. McLachlan's modification of the artificial seawater

111 with L. Provasoli's metal mix PI; described in Guillard, 1983) and maintained in RSW
112 at least 48 h before experiments (ASTM, 2002). Finally, the organisms were kept under
113 fasting conditions (in new cultures with burnt sand and fresh RSW) the last 24 h before
114 the experiments in order to standardize their nutritional storage levels.

115

116 *2.3. Test concentrations*

117 In accordance with our previously published paper, we chose two concentrations of Cd,
118 5 and 0.5 mg/l to test two Cd concentrations in which a significant decrease in the FR
119 could be detected with lower mortality (Martinez-Haro et al., 2014). Stock solutions of
120 500 and 50 mg/l Cd were prepared using CdCl₂ (Acros Organic, Geel, Belgium) and
121 nanopure water (conductivity <5 µS/cm; Seralpur PRO 90 CN, Seral, Ransbach-
122 Baumbach, Germany). The test concentrations were obtained by diluting stock solutions
123 with RSW.

124

125 *2.4. Experimental design*

126 Two different exposure periods, 48 h and 96 h, were tested in order to simulate the most
127 common exposure times considered in short-term assays (see e.g., Moreira et al., 2005;
128 Satapornvanit et al., 2009). For this purpose, five different experimental groups were
129 designed: one control group without exposure to pollutant, plus two more for each of
130 the Cd concentrations, one for each exposure period. A total of 440 animals were used
131 for the experiment, 88 in each experimental group. The groups were placed in
132 polyethylene containers filled with 600 ml of test medium and 300 g of sand, and no
133 food was provided to animals during the experiments.

134 Feeding tests were carried out at five different times in each experimental group:

135 a) in the case of the animals in the control group the feeding tests were performed

136 immediately before starting the experiment (0 h - the result from this test represented
 137 the baseline for all experimental groups), and at 48 h, 72 h, 96 h and 120 h after the start
 138 of the experiment; b) in the case of the animals that had been exposed to Cd, the tests
 139 were carried out immediately after the exposure period (EP), and at EP+3 h, EP+6 h,
 140 EP+12 h and EP+24 h (post-exposure times). The experimental design included to
 141 randomly subtracting 14 animals (each replicate one animal) of each experimental
 142 group at each time to carry out the FRs. In Table 1 the number of replicates analysed
 143 per experimental group and time were summarized. Test media and sand were daily
 144 renewed. Dead animals were counted and removed from the containers on a daily basis
 145 along the experiment. At the end of each exposure periods (48 h or 96 h), and prior to
 146 the feeding assays, the animals were washed with RSW and placed in new containers
 147 filled only with RSW.

148

149 Table 1. Number of replicates analysed per experimental group and time.

Experimental group	Time from the start of the experiment				
	0 h	48 h	72 h	96 h	120 h
Control	n=14	n=14	n=14	n=14	n=14
	Post-exposure time				
	EP	EP+3 h	EP+6 h	EP+12 h	EP+24 h
0.5 mg/l – 48 h EP	n=14	n=14	n=14	n=14	n=14
0.5 mg/l – 96 h EP	n=14	n=13	n=14	n=14	n=14
5.0 mg/l – 48 h EP	n=14	n=14	n=14	n=14	n=14
5.0 mg/l – 96 h EP	n=14	n=14	n=14	n=14	n=13

150

151 Post-exposure FRs were then performed by following the methodology
 152 previously described for this species (Martinez-Haro et al., 2014). Briefly, animals were
 153 individually allowed to feed on 100 defrosted nauplii for 30 min at 20 °C and in
 154 darkness. At the end of the feeding period, the remaining nauplii were counted using a

155 stereomicroscope, and FR were estimated as the mean number of nauplii ingested per
156 individual in 30 min.

157 Cadmium concentrations were analysed by means of atomic absorption
158 spectroscopy (AAnalyst800, PerkinElmer) in the test solutions and in a reserved batch
159 of sand that had been used before. The results were 0.4 ± 0.002 mg/l and 5 ± 0.143
160 mg/l, for Cd concentrations of 0.5 and 5 mg/l, respectively. Since the actual and
161 nominal Cd concentrations differed by more than 10% for 0.5 mg/l, the actual Cd
162 concentration was subsequently used. Blanks, and a certified soil reference material
163 (CRM, GBW07406), were also processed in each batch of digestions (to provide quality
164 control data). The Cd concentrations in the control and in the clean sand were below the
165 limit of detection (LOD 0.003 mg/l and 0.004 μ g/g, respectively). The percentage of Cd
166 recovery for soil CRM was 98%.

167

168 *2.5. Data analysis*

169 The stability of the FR over time in the control group was tested using one-way
170 ANOVA, with the individual FR as the response variable and time (5 categories; 0 h, 48
171 h, 72 h, 96 h, and 120 h) as the factor. For each exposure period (48 h or 96 h), the
172 effect of Cd concentrations (0 mg/l, 0.4 mg/l or 5 mg/l) on FR was tested using one-way
173 ANOVA, and a post-hoc Tukey's test to check for differences among Cd
174 concentrations. Differences in the FR between the control and the groups exposed to Cd
175 for each post-exposure time interval (EP, EP+3 h, EP+6 h, EP+12 h and EP+24 h) were
176 checked using a one-way ANOVA. Differences in the FR between post-exposure times
177 were tested using a one-way ANOVA for each exposure period with a Dunnett's post-
178 hoc test to check differences between EP and each one of the remaining post-exposure

179 time intervals. Residuals of the models were checked for normality and
180 homoscedasticity, through Kolmogorov-Smirnov and Levene tests, respectively.

181 The tests were performed using IBM SPSS Statistics 20.0, with a level of
182 statistical significance of 0.05.

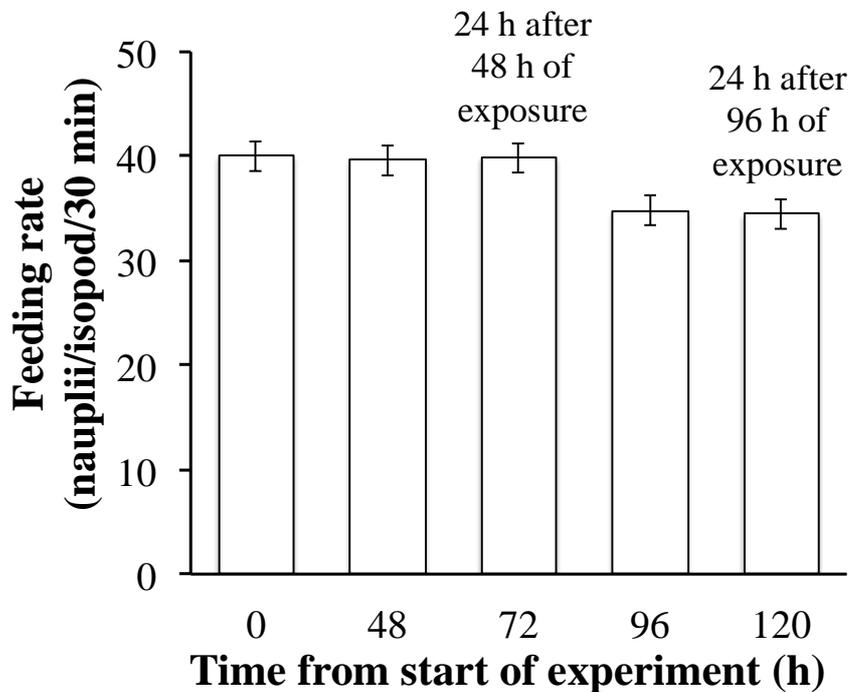
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184 **3. Results**

185 The mortality in the control group was 2.6% of animals at the end of the test (after 120
186 h). For the animals exposed to Cd, a higher metal concentration and longer exposure
187 resulted in higher mortality rates (2.3% and 3.4% for individuals exposed to 0.4 mg/l
188 Cd after 48 h and 96 h of exposure, respectively; and 9.1% and 17.0% for individuals
189 exposed to 5 mg/l Cd after 48 h and 96 h of exposure, respectively).

190 The mean FR in the control group (mean \pm SE: 38 ± 1 nauplii/ind/30 min) did not
191 differ over time ($F_{4,64} = 1.307$, $p = 0.277$; Fig. 1), although a slight decrease was
192 observed at 96 h from the start of the experiment, which may be a sign of the effect of
193 starvation. The FR for this experimental group achieved a mean coefficient of variation
194 of 23% (14% - 34%, min-max). The FR of animals exposed to 5 mg Cd/l for both
195 exposure periods were significantly lower than FR detected in controls ($p < 0.001$; Figs.
196 2a, 2b). The same pattern was not observed in animals exposed to 0.4 mg Cd/l for any
197 of the exposure periods ($p > 0.05$; Figs. 2a, 2b).

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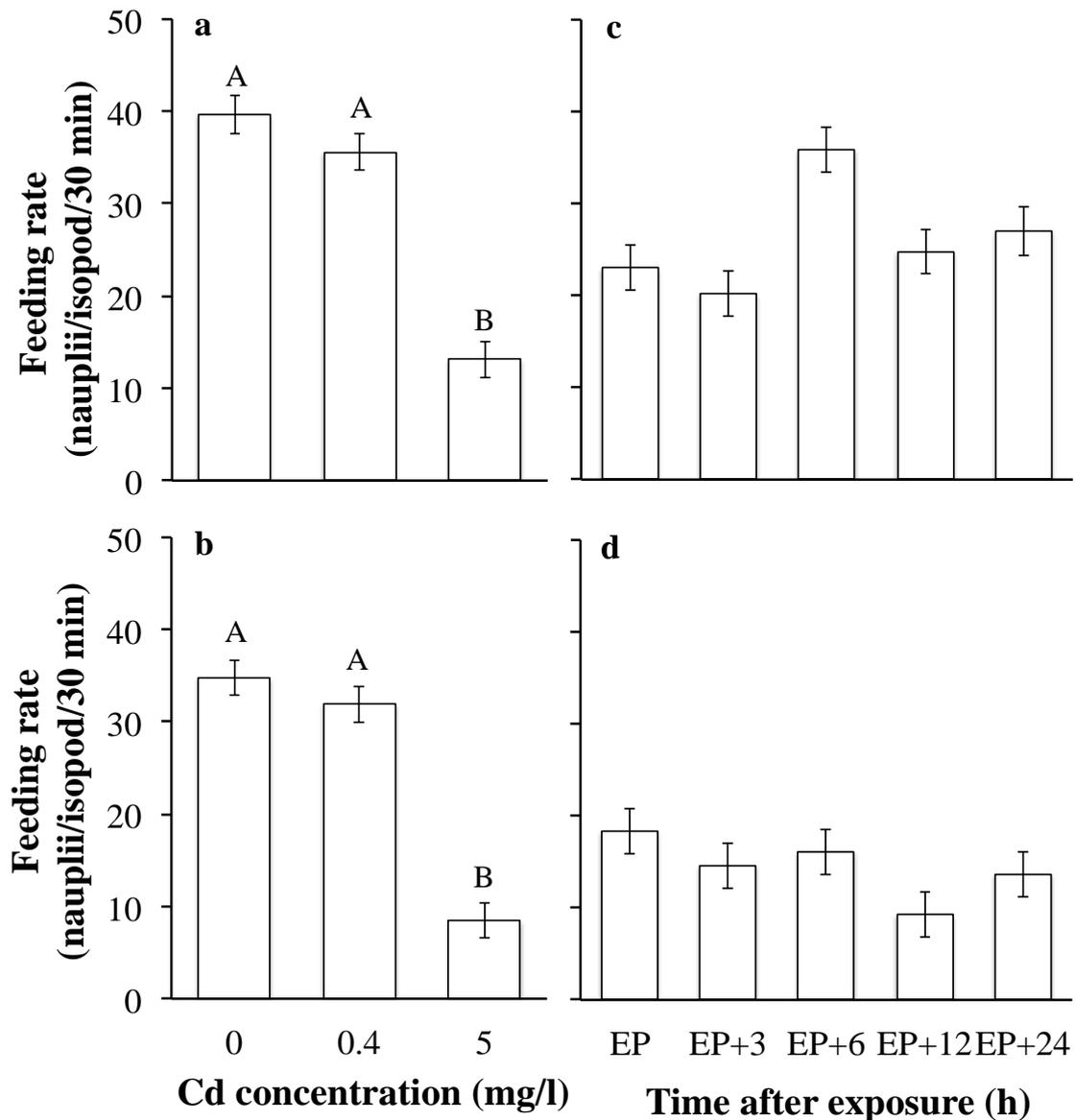


199

200 Fig. 1. Mean (\pm SE) marginal feeding rates (nauplii/individual/30 min) for *Cyathura*
 201 *carinata* in the control group (animals without exposure to cadmium) during the
 202 experiment. Correspondences in time with the exposed animals are depicted.

203

204 With regard to controls, mean FR were statistically lower in the organism
 205 exposed to 5 mg/l for all the post-exposure times tested for both 48 h and 96 h exposure
 206 periods ($p < 0.05$). However, no statistical differences were detected for any of the
 207 organisms exposed to 0.4 mg/l except for the post-exposure times 12 h and 24 h of the
 208 96 h exposure period ($p < 0.05$). For the groups exposed to Cd, no significant
 209 differences between FR at EP and all the remaining post-exposure time intervals were
 210 detected at any of the exposure periods (Dunnett's post-hoc test $p < 0.05$; Figs. 2c, 2d).
 211 Although not significant, a slight increase on the feeding rate was detected at EP+6 h
 212 for 48 h exposure period.



213

214 Fig. 2. Mean (\pm SE) marginal feeding rates (nauplii/individual/30 min) for *Cyathura*
 215 *carinata* after two cadmium (Cd) exposure periods (EP), 48 h and 96 h. Differences
 216 among concentrations are shown in a) and b), for exposure periods of 48 h and 96 h,
 217 respectively (concentrations with the same capital letter did not differ significantly;
 218 Post-hoc Tukey's test, $p > 0.05$). Differences between EP and each post-exposure time
 219 are shown in c) and d), for exposure periods of 48 h and 96 h, respectively (Dunnett's
 220 post-hoc test, $p > 0.05$).

221

222 4. Discussion

223 The recovery of the organism's response after exposure to pollutants is an important
224 issue that must be controlled in assays based on post-exposure endpoints. As expected,
225 higher concentrations and longer exposure period to Cd resulted in higher mortality
226 rates and a higher feeding inhibition of animals, which coincides with previous studies
227 with other invertebrate and vertebrate species that have been exposed to this metal
228 (Pascoe and Shazili, 1986; Pestana et al., 2007). It should be noted that the exposure to
229 5 mg Cd/l for 96 h resulted in a mortality of 17% in *C. carinata*, which may be due to
230 the toxic effect of concentration of that concentration plus the starvation period to
231 which the animals were subjected. The absence of a clear recovery as regards the FR at
232 least 24 h after Cd exposure should be considered as a safe window in which post-
233 exposure feeding assays can be carried out without expectations of punctual feeding
234 reduction as a result of the animal's recovery. Only weak and inconsistent fluctuations
235 in the FR were detected for a punctual post-exposure interval (EP+6 h), which is
236 probably owing to certain uncontrolled factors affecting the feeding assays at this time.

237 The stability in feeding response obtained here is consistent with previous
238 studies focused on metals and some organic pollutants. For instance, feeding depression
239 was detected in *Daphnia* both during the exposure (24 h) and post-exposure (4 h) to
240 sublethal concentrations of contaminants, including Cd and organic pollutants
241 (McWilliam and Baird, 2002; Taylor et al., 1998). Wilding and Maltby (2006) found
242 the FR of *Gammarus pulex* exposed to sublethal zinc concentrations during 6 days did
243 not recover, and even became more depressed, after 6 days of exposure. Similarly,
244 Brent and Herricks (1998) found that *Hyalella azteca* mobility was not recovered for up
245 to 172 h after short (30-240 min) exposure periods to sublethal concentrations of Cd
246 and zinc. Our study corroborates with this previous evidence and supposes the first

247 assessment of the potential recovery effects up to 24 h after exposure, which is quite
248 relevant as regards applying these new generation ecotoxicological tools.

249

250 **5. Concluding remarks**

251 The present study has shown that inhibitions in the feeding rates of *C. carinata* after Cd
252 exposure represented a stable response as a bioassay endpoint since no evidence of
253 recovery was detected in 24 h after the exposure period.

254

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259

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