1 Evolutionary and demographic history of the Californian
2 scrub white oak species complex: An integrative approach

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Abstract

Understanding the factors promoting species formation is a major task in evolutionary research. Here, we employ an integrative approach to study the evolutionary history of the Californian scrub white oak species complex (genus *Quercus*). To infer the relative importance of geographical isolation and ecological divergence in driving the speciation process, we (i) analyzed inter- and intra-specific patterns of genetic differentiation and employed an approximate Bayesian computation (ABC) framework to evaluate different plausible scenarios of species divergence. In a second step, we (ii) linked the inferred divergence pathways with current and past species distribution models, and (iii) tested for niche differentiation and phylogenetic niche conservatism across taxa. ABC analyses showed that the most plausible scenario is the one considering the divergence of two main lineages followed by a more recent pulse of speciation. Genotypic data in conjunction with species distribution models and niche differentiation analyses support that different factors (geography *vs.* environment) and modes of speciation (parapatry, allopatry and maybe sympatry) have played a role in the divergence process within this complex. We found no significant relationship between genetic differentiation and niche overlap, which probably reflects niche lability and/or that multiple factors have contributed to speciation. Our study shows that different mechanisms can drive divergence even among closely related taxa representing early stages of species formation and exemplifies the importance of adopting integrative approaches to get a better understanding of the speciation process.

Keywords: Bayesian inference, genetic structure, hybridization, niche divergence, paleo-distribution modeling, *Quercus*, species distribution modeling

Introduction

Understanding the factors promoting species divergence is a major task in ecological and evolutionary research (Dobzhanski 1937; Mayr 1942; Simpson 1953; Papadopoulos *et al.* 2011; Butlin *et al.* 2012). Opportunities for speciation have been linked to geographic isolation (Mayr 1942), environmental heterogeneity (i.e. ecological speciation; Orr & Smith 1998; Nosil 2012; Schluter 2001) and a combination of both, which can result in different mechanisms of speciation with or without complete reproductive isolation among diverging lineages (Orr & Smith 1998; Schluter 2001; Graham *et al.* 2004; Schluter 2009). Accordingly, two major modes of speciation have been proposed: (i) Allopatric/parapatric speciation, which leads to population divergence in total or partial geographical isolation. Under this model, taxa diverged in vicariance can conserve similar environmental niches or experience ecological specialization after divergence due to either genetic drift or environmentally-mediated divergent selection (Graham *et al.* 2004; Warren *et al.* 2008); (ii) Sympatric speciation, which results from ecological and reproductive isolation due to divergent selection in the absence of physical barriers to gene flow (Graham *et al.* 2004; Papadopoulos *et al.* 2011). Disentangling the relative roles of geography and ecological factors in promoting species/lineage diversification requires analyses of both the historical geographical separation of taxa that could have led to speciation as well as the present-day environmental conditions that shape the current distribution and ecological requirements of species (Rundell & Price 2009; Ribera *et al.* 2011). For hybridizing species, identifying the underlying mechanisms behind divergence is even more challenging, because interspecific gene flow can confound the genetic and phenotypic signals left during the diversification process (e.g. Eaton & Ree 2013; Papadopoulos *et al.* 2011,

2013). In presence of interspecific genetic exchange, the maintenance of species boundaries requires selection against hybrids due to their intrinsic low performance, maladaptation to both parental species environments or habitat-induced variation in assortative mating (e.g. Anderson 1953; Papadopoulos *et al.* 2011; Singhal & Moritz 2012), so that hybrid/introgressed individuals constitute an ephemeral state that only persist under some particular circumstances (e.g. in areas with intermediate environments; Papadopoulos *et al.* 2013; Ortego *et al.* 2014a).

California has a complex biogeographic history, with deep topographic and climatic gradients that have contributed to shape one of the richest floras and faunas on the planet (Raven & Axelrod 1978; Calsbeek *et al.* 2003; Davis *et al.* 2008; Lancaster & Kay 2013). The high biodiversity and local endemism of this region have been linked to the important role of topographic and climatic complexity in creating multiple ecological niches, favoring high rates of intra- and interspecific diversification (Calsbeek *et al.* 2003; Davis *et al.* 2008). In parallel, climate buffering associated with complex topography seems to have contributed to a particularly low extinction rate of species/lineages adapted to contrasting environmental conditions since the Tertiary (Lancaster & Kay 2013). Comparative studies have often revealed common phylogeographic breaks and genetic discontinuities across multiple taxa, suggesting a similar history of diversification by vicariance associated with geological events and climatic fluctuations (Calsbeek *et al.* 2003; Lapointe & Rissler 2005; Rissler *et al.* 2006; Vandergast *et al.* 2008; Gugger *et al.* 2013; Sork & Werth 2014). However, other studies have also pointed to ecological adaptation as an important mode of diversification of California biota (e.g. Ortego *et al.* 2012; Langin *et al.* 2015). This environmental complexity and high diversity makes California Floristic Province an ideal scenario to study the proximate factors underlying the speciation process, which

can ultimately contribute to understand the origin of its high rates of endemism and guide conservation policies aimed to preserve this biologically rich but increasingly threatened ecoregion (Myers *et al.* 2000; Vandergast *et al.* 2008).

Here, we employ an integrative approach to analyze the evolutionary and demographic history of the Californian scrub white oak species complex (genus and section *Quercus*), infer the most plausible modes of speciation underlying the diversification process within the complex, and understand the relative importance of geographical isolation *versus* ecological-mediated selection in driving taxonomic divergence and maintaining species genetic identity. There are six recognized taxa within this complex, namely California interior (*Q. berberidifolia*, Liebmman 1854), leather (*Q. durata*, Jepson 1909), coastal sage (*Q. dumosa*, Nuttall 1842), island (*Q. pacifica*, Nixon & Muller 1994), Muller (*Q. cornelius-mulleri*, Nixon & Steele 1981) and Tucker (*Q. john-tuckeri*, Nixon & Muller 1994) scrub oaks (Nixon 2002). Most of these species are found in Southern California, where they live in close geographical proximity but not usually in mixed stands (Roberts 1995). Many of these taxa have been described in the last few decades (Nixon & Steele 1981; Nixon & Muller 1994) and, with the exception of *Q. durata*, all of them were generally treated before as part of a broad complex of *Q. dumosa* (Nixon 2002; e.g. Forde & Faris 1962). These taxa are edaphic specialist (*Q. durata*), island endemics (*Q. pacifica*) or encountered in different habitats such as coastal sage scrub (*Q. dumosa*), desert margins (*Q. cornelius-mulleri*), and chaparral and mixed forest (*Q. berberidifolia* and *Q. john-tuckeri*), suggesting that different modes of speciation (allopatric speciation in island endemics *vs.* sympatric/parapatric speciation among nearby ecological specialist) may have contributed to the diversity of the complex (Nixon 2002). Thus, this system offers an interesting case study to understand how potentially different mechanisms may have

contributed to taxonomic divergence in a complex of dominant keystone species with a high importance in ecosystem functioning (Sawyer & Keeler-Wolf 1995).

To study the evolutionary and demographic history of the Californian scrub white oak species complex we collected over 800 individuals from all putative species covering their entire distribution ranges across California (Fig. 1). Specimens were genotyped at 16 nuclear and 5 chloroplast simple sequence repeat (SSR) markers in order to infer the demographic history of the complex and the patterns of genetic structure and hybridization within and among species, respectively. We also built species distribution models (SDMs) for the different species that were projected into the last glacial maximum (LGM; ca. 21000 years BP) using different climate models to estimate range shifts and characterize the environmental envelope of each taxon. In particular, (i) we first analyzed patterns of inter- and intra-specific genetic differentiation among all putative species and employed an approximate Bayesian computation (ABC) framework (Beaumont *et al.* 2002; Beaumont & Rannala 2004) to compare different plausible scenarios of divergence. This allowed us to infer the evolutionary and demographic history of the different taxa within the complex. In a second step, to elucidate the potential mechanism of speciation (allopatric/parapatric speciation *vs.* ecological speciation in sympatry), we (ii) linked the inferred divergence pathways with current and past species distribution ranges and (iii) tested for environmental niche differentiation among taxa (Graham *et al.* 2004; Mao & Wang 2011; Papadopoulos *et al.* 2013). Finally, we (iv) analyzed whether pair-wise species genetic differentiation is associated with niche overlap. If niches are highly conserved or if allopatric divergence is the dominant process, it would be expected that closely related species share a similar niche space (Warren *et al.* 2008; Nakazato *et al.* 2010). The opposite pattern would arise if phylogenetically distant taxa tend to share more

similar environments than those representing earlier stages of species formation and would point to niche divergence as an important mechanism promoting divergence (Cavender-Bares *et al.* 2004; Warren *et al.* 2008). A lack of association between niche overlap and genetic differentiation is expected if environmental niches are highly labile (Knouft *et al.* 2006; Warren *et al.* 2008), which has been previously documented in trees and can result from phenotypic plasticity and/or some level of interspecific gene exchange (Cornuault *et al.* 2015).

Methods

Sampling

Between 2010 and 2014, we sampled 812 scrub white oaks from a total of 87 localities in California (Table S1, Supporting Information; Fig. 1). Sampling was designed using occurrence records available for each taxon at the species level in the Calflora database (<http://www.calflora.org/>). We aimed to collect samples from populations located across the entire distribution range of all the species and make a more exhaustive sampling in southern California, where more taxa are present and distribution boundaries of each putative species are not well defined (Roberts 1995; eFloras 2015). Our sampling only excluded a small portion of the range of three species (*Q. berberidifolia*, *Q. dumosa* and *Q. cornelius-mulleri*) at their southernmost distribution limit in northern Baja California (Mexico) (Fig. 1). Most populations from the endangered coastal scrub oak (*Q. dumosa*) have been extirpated due to human development in southern California and only a few stands of this species could be found during our surveys across its former distribution range (IUCN 2010; Fig. 1).

SSR genotyping

We ground about 50 mg of frozen leaf tissue in tubes with a tungsten bead using a mixer mill and DNA extraction and purification was performed with the cetyl trimethyl ammonium bromide (CTAB) protocol (Doyle & Doyle 1990). To genotype scrub white oaks, we used 16 nuclear (nuSSR) and 5 chloroplast (cpSSR) SSR markers previously developed for other species (Table S2, Supporting Information). Analyses of neutrality based on the F_{ST} -outliers tests implemented in LOSITAN (100 000 simulations assuming a stepwise mutation model; Beaumont & Nichols 1996; Antao *et al.* 2008) indicated that no locus consistently deviated from neutral expectations in the studied species (data not shown). Amplifications were carried out in 10- μ L reaction mixtures containing 2.5 μ L of DNA ($\sim$ 20 ng) and 7.5 μ L of PCR mix, which contained the following reagents: 3.5 μ L of Type-it Multiplex PCR Master Mix (QIAGEN), 0.2 μ L of BSA 10 $\times$, 2.5 μ L of dye-labelled primer mix (see Table S2 for specific primer concentrations), and 1.3 μ L of ddH₂O. The PCR profile consisted of an initial denaturing of 15 min at 95 $^{\circ}$ C, followed by 44 cycles of 30-s denaturing at 95 $^{\circ}$ C, 90-s annealing at 56 $^{\circ}$ C, and 30-s extension at 72 $^{\circ}$ C, followed by a final extension step of 56 $^{\circ}$ C for 30 min. Amplification products were electrophoresed using an ABI PRISM 3700 capillary sequencer (Applied Biosystems) and genotypes were scored using GENEMAPPER 3.7 (Applied Biosystems).

Nuclear SSR genotypes were tested for departure from Hardy-Weinberg equilibrium within each sampling population/species at each locus using an exact test (Guo & Thompson 1992) based on 900 000 Markov chain iterations as implemented in the program ARLEQUIN 3.1 (Excoffier *et al.* 2005). We also used ARLEQUIN 3.1 to test for linkage equilibrium between each pair of loci for each sampling population/species

using a likelihood-ratio statistic, whose distribution was obtained by a permutation procedure (Excoffier *et al.* 2005). Sequential Bonferroni corrections were applied to account for multiple comparisons (Rice 1989).

Genetic structure

For nuSSR, we analyzed patterns of genetic structure using the Bayesian Markov chain Monte Carlo clustering analysis implemented in the program STRUCTURE 2.3.3 (Pritchard *et al.* 2000; Falush *et al.* 2003; Hubisz *et al.* 2009). We ran STRUCTURE assuming correlated allele frequencies and admixture and without using prior population information (Hubisz *et al.* 2009). We conducted ten independent runs for each value of $K = 1-10$ to estimate the “true” number of clusters with 200000 MCMC cycles, following a burn-in step of 100000 iterations. The number of populations best fitting the data set was defined using log probabilities $[\Pr(X|K)]$ (Pritchard *et al.* 2000) and the ΔK method (Evanno *et al.* 2005), as implemented in STRUCTURE HARVESTER (Earl & vonHoldt 2012). We used CLUMPP 1.1.2 and the Greedy algorithm to align multiple runs of STRUCTURE for the same K value (Jakobsson & Rosenberg 2007) and DISTRUCT 1.1 (Rosenberg 2004) to visualize as bar plots the individual’s probabilities of population membership. Clustering analyses for nuSSR were complemented with an individual-based principal component analysis (PCA), which does not rely on Hardy-Weinberg or linkage equilibrium and can be useful to detect genetic clines or complex patterns of genetic structure. We employed a MANOVA to compare simultaneously the PCA scores obtained for the first two principal component (PC) axes across all the species. *Post hoc* Tukey tests were used to examine differences between each species

pair. PCA was performed using the R 3.0.2 package ADEGENET (Jombart 2008) and MANOVA in SPSS 22.0.

We examined population genetic structure for cpSSR markers using the “clustering with linked loci” analysis in BAPS 6.0 (Corander *et al.* 2008). We conducted four independent runs for $K = 1-40$ to determine the K value with the highest likelihood. Results from BAPS were also visualized as bar plots using DISTRUCT 1.1. Complementarily, we constructed a median-joining haplotype network colored by species using NETWORK 4.6 (Bandelt *et al.* 1999).

The distribution of genetic variation in both nuclear and chloroplast loci was assessed using AMOVAs as implemented in ARLEQUIN 3.1 (Excoffier *et al.* 2005). Genetic variation was hierarchically partitioned into among species, among populations within species and among individuals within populations. Significance was tested using 10 000 permutations of the original data.

Phylogenetic analyses

To visualize the phylogenetic relationship among all taxa and populations, we reconstructed a population-based neighbour-joining (NJ) tree in POPULATIONS 1.2.30 (Langella 2007) using Cavalli-Sforza and Edwards' (1967) chord distances based on nuSSR and cpSSR allele frequency data. This genetic distance has been suggested to be the most effective in recovering the correct tree topology for SSR markers under a variety of evolutionary scenarios without making assumptions regarding constant population size or mutation rates among loci (Takezaki & Nei 1996). Levels of confidence of phylogenetic groupings were estimated using bootstrapping methods, resampling across loci 1000 times (Langella 2007). Only populations with five or more

genotyped individuals were included in NJ analyses. These analyses were performed both considering all sampled individuals and excluding hybrids/introgressed individuals from the dataset ($q < 0.90$; e.g. Ortego & Bonal 2010).

Demographic history: Approximate Bayesian Computation

We used an approximate Bayesian computation (ABC; Beaumont 2010; Bertorelle *et al.* 2010) statistical framework to compare three plausible scenarios of divergence among Californian scrub white oaks and estimate timing of divergence and effective population sizes of each taxa (Fig. 2a). Scenarios were built considering previous morphology-based knowledge on the taxonomy of the group (Nixon 2002) and STRUCTURE analyses (see results section; e.g. Liu *et al.* 2014; Tsuda *et al.* 2015), which point to an older divergence between *Q. berberidifolia*/*Q. durata* and the rest of the species than among taxa within these two groups. The first scenario (“null model”) consisted of a simultaneous divergence of all the species at t_5 . The second scenario predicts that *Q. durata* and *Q. berberidifolia* diverged from the rest of the species at t_5 . Afterwards, *Q. durata* and *Q. berberidifolia* diverged at t_4 and the other four species diverged simultaneously at t_3 . The third scenario, the most likely according with hierarchical STRUCTURE analyses, is similar to the previous one but predicts that *Q. dumosa* and *Q. pacifica* diverged from *Q. cornelius-mulleri* and *Q. john-tuckeri* at t_3 and these pairs of taxa subsequently split at t_1 and t_2 , respectively (Fig. 2a). We also performed further ABC analyses exclusively focusing on *Q. dumosa* and the island endemic *Q. pacifica* to test in more detail different scenarios of divergence among the populations of these two closely related taxa (Fig. 2b) for which we have found a complex pattern of genetic structure and admixture (see results section). In this case, we considered three

populations, one including all sampling localities for *Q. dumosa*, another for the populations of *Q. pacifica* from Santa Catalina island, and a third one including the populations of *Q. pacifica* from both Santa Cruz and Santa Rosa Islands (Fig. 2b).

Simulations for each scenario and ABC analyses were conducted using DIYABC 2.0 (Cornuet *et al.* 2014). We generated one million simulated datasets per scenario considering a 1:1 female to male sex ratio, a generalized stepwise-mutation model and uniform priors with default values for all parameters. We considered the following constraints on temporal parameters: $t_5 > t_4$, $t_5 > t_3$, $t_5 > t_2$, $t_5 > t_1$, $t_3 > t_2$, $t_3 > t_1$ for all species (Fig. 2a) and $t_2 > t_1$ for analyses focused on *Q. dumosa* and *Q. pacifica* (Fig. 2b). We used five summary statistics (SS): the mean values of expected heterozygosity (H_E) and number of alleles (A) for each population (i.e. species) and H_E , A , classification index and F_{ST} for each pair of taxa. To reduce the computational demands and avoid potentially confounding effects of contemporary hybridization (e.g. Ortego & Bonal 2010; Ortego *et al.* 2014a), we excluded hybrids/introgressed individuals from the datasets ($q < 0.90$; e.g. Besnard *et al.* 2014; Tsuda *et al.* 2014) and for analyses including all species we selected up to a maximum of 50 individuals per taxon representative of all sampled populations ($n = 281$ individuals in total). We ran ABC analyses considering only nuSSR and both nuSSR and cpSSR markers to evaluate the impact of using different kind of loci on scenario choice and posterior parameter estimation. The posterior probability of scenarios was assessed using a weighted polychotomous logistic regression on the 1% of simulated datasets closest to the observed data (Fontaine *et al.* 2013; Cornuet *et al.* 2008, 2014). We simulated 500 pseudo-observed datasets (PODs) under each scenario to estimate Type I and Type II error rates and assess confidence in scenario choice (Robert *et al.* 2011). For the best supported scenario, we estimated the posterior distribution of all parameters using local

linear regressions on the 1% of the simulations closest to the observed data after a logit transformation of parameter values (Beaumont *et al.* 2002; Cornuet *et al.* 2008). The performance of parameter estimation was determined by calculating the median of the relative median of absolute errors (RMAE) from 500 PODs (Cornuet *et al.* 2010). Finally, we evaluated the goodness of fit of the best supported scenario simulating 10 000 PODs from the posterior distribution of all parameters. As recommended by Cornuet *et al.* (2010), we used summary statistics that had not been used for model selection or parameter estimation in previous ABC treatments. We applied a principal component analysis (PCA) on test statistics vectors to visualize the fit between simulated and observed datasets. Finally, we used ranking of SS (i.e. % of simulated data < observed data) to evaluate whether the best supported model can successfully reproduce observed data.

Species distribution modeling

We used species distribution modeling (SDM) to predict the geographic distribution of suitable habitats for each studied species both in the present and during the last glacial maximum (LGM). We modelled current distributions using MAXENT 3.3.3 (Phillips *et al.* 2006; Phillips & Dudik 2008). Species occurrence data were obtained from sampling points as well as from herbarium records available at the Global Biodiversity Information Facility (<http://www.gbif.org/>), Consortium of California Herbaria (<http://ucjeps.berkeley.edu/consortium/>), Consortium of Pacific Northwest Herbaria (<http://www.pnwherbaria.org/>) and University of Arizona Herbarium (<http://ag.arizona.edu/herbarium/>). Prior to modeling, all records were mapped and examined to identify and exclude those having obvious georeferencing errors and

misidentifications. For models, we only retained a single record among those falling within the same grid cell (see final sample sizes in Table S3, Supporting Information). Initially we selected 26 variables that may potentially determine the current distribution of our species. These included growing degree days, soil organic carbon, soil pH, and soil moisture obtained from the Center for Sustainability and the Global Environment (<http://www.sage.wisc.edu/atlas/index.php>); ground frost frequency, vapour pressure, and wet day frequency obtained from the Intergovernmental Panel on Climate Change (http://www.ipcc-data.org/observ/clim/cru_ts2_1.html); and the 19 bioclimatic layers from the WorldClim dataset (<http://www.worldclim.org/>) (Hijmans *et al.* 2005). The 19 bioclimatic layers were downloaded from WorldClim at 30-arcsec (c. 1-km) resolution. The other layers were downloaded at a spatial resolution of 0.5° and resampled to 30-arcsec resolution via bilinear interpolation using ARCMAP 10.2.1 (ESRI, Redlands, CA, USA). We assessed the correlation between all the layers using ENMTOOLS (Warren *et al.* 2010). When two layers were highly correlated ($r > 0.7$), we discarded the layer with the highest number of correlations with other layers. Accordingly, we selected a final set of seven layers to construct the models: mean diurnal range (Bio2), isothermality (Bio3), mean temperature of the wettest quarter (Bio8), precipitation of the wettest month (Bio13), precipitation of the driest month (Bio14), precipitation seasonality (Bio15), and precipitation of the warmest quarter (Bio18). Model evaluation statistics were produced from ten cross-validation replicate model runs and overall model performance was assessed using the area under the receiving operator characteristics curve (AUC).

We estimated the distribution of each scrub white oak species at the Last Glacial Maximum (LGM; c. 21 000 years BP) projecting contemporary species-climate relationships to this period. We used the same seven bioclimatic layers available at

WorldClim for two paleoclimate models of the last glacial period: the Model for Interdisciplinary Research on Climate (MIROC; Hasumi & Emori 2004) and the Community Climate System Model (CCSM3; Collins *et al.* 2006). The predicted probabilities of occurrence obtained for both paleoclimate models were averaged using the R 3.0.2 package RASTER (R Core Team 2015).

Niche divergence/conservatism

We used identity and background tests in ENMTOOLS to analyze whether ecological niches are differentiated or conserved between each pair of species (Warren *et al.* 2008; e.g. Nakazato *et al.* 2010; Mao & Wang 2011). For this purpose, we first calculated actual niche overlap between each pair of species using two alternative statistics: *I* (Warren *et al.* 2008) and *D* (Schoener 1968), which are compared and described in detail in Warren *et al.* (2008). Then, we performed niche identity tests to compare the overlap of a species pair's actual niches to a distribution of niche overlaps obtained from pairs of pseudoniches ($n = 100$ pseudoreplicates) constructed based on randomly reshuffled occurrence points of the two species (Warren *et al.* 2008). Thus, this test examines the null hypothesis that a given pair of species is distributed in an identical environmental space. We also used background tests to analyze whether the ecological niches of a given pair of species overlap more or less than would be expected from the differences in the environmental backgrounds of the regions where they occur (Warren *et al.* 2008). This test compares the observed niche overlap of a given pair of species to a null distribution ($n = 100$ random samplings) of overlap values generated by comparing the ecological niche model of one taxon to an ecological niche model created from random points drawn from the geographic range of the other taxon (Warren *et al.*

2008). This process is repeated for both taxa in the comparison so that two null distributions are generated. The background area should include accessible areas for the species, not just the observed niche or an area tightly delimited by species occurrence (McCormack *et al.* 2010; Nakazato *et al.* 2010; Mao & Wang 2011). Given that the delimitation of the background area can influence the results of the analyses, we considered different background areas defined by buffer zones of 1, 5, and 10 km around the actual distribution delimited by occurrence points. These background areas should include all the accessible habitats consistent with restricted scale of acorn dispersal in oaks (Grivet *et al.* 2005, 2006, 2009; Gugger *et al.* 2013). Background areas were obtained using ARCMAP 10.2.1.

The environmental space of the studied species was also characterized using a principal component analysis (PCA) applied to scaled data of the same environmental variables employed for SDM and extracted from occurrence points. We employed a MANOVA to compare simultaneously the PCA scores obtained for the first two principal component (PC) axes (68% of variance explained) across all the species and determine whether their sites of occurrence differ in bioclimatic conditions. *Post hoc* Tukey tests were used to examine differences between each species pair. PCA was performed in R 3.0.2 package PSYCH and MANOVA in SPSS 22.0.

Niche overlap and genetic differentiation

We analyzed the correlation between genetic differentiation and the two estimates of niche overlap (*I* and *D*) using Mantel tests (e.g. Warren *et al.* 2008; Nakazato *et al.* 2010). We calculated different estimates of genetic differentiation for both nuSSR and cpSSR markers, including F_{ST} , Reynold's distance, G'_{ST} and Jost's *D* (Meirmans &

Hedrick 2011). Pairwise F_{ST} and Reynold's distances were calculated using ARLEQUIN 3.1 (Excoffier *et al.* 2005) and G'_{ST} and Jost's D were calculated using the R 3.0.2 package MMOD (R Core Team 2015). Significance of F_{ST} -values was tested using Fisher's exact tests after 10 000 permutations and a sequential Bonferroni adjustment. Mantel tests were performed using ZT software with 10 000 permutations (Bonnet & Van de Peer 2002).

Results

SSR data

All nuSSR markers were highly polymorphic and observed heterozygosity at each locus ranged from 0.21 to 0.91, with 10-35 alleles per locus (Table S2). Chloroplast SSR markers were less polymorphic than nuclear loci and had 4-6 alleles per locus (Table S2). After applying sequential Bonferroni corrections to compensate for multiple statistical tests, no nuSSR locus consistently deviated from HWE in all the studied species/populations. We did not find any evidence of genotypic linkage disequilibrium at any pair of nuSSR loci in any species/population (exact tests; all P -values > 0.05).

Genetic structure

STRUCTURE analyses and the statistic ΔK indicated an "optimal" value of $K = 2$ when populations from all the species were included in the same analysis (Fig. S1a, Supporting Information), grouping *Q. berberidifolia*-*Q. durata* in one cluster and the rest of the species in another cluster (Fig. 3a). Subsequent hierarchical analyses only

including samples from *Q. berberidifolia* and *Q. durata* showed that these two species split in two different clusters (Fig. S1b, Fig. 3b). The rest of the species also split in two other clusters, one containing the populations of *Q. cornelius-mulleri* and *Q. john-tuckeri* and another including the populations of *Q. dumosa* and *Q. pacifica* (Fig. S1c, Fig. 3d). Further, hierarchical analyses within these clusters showed that *Q. cornelius-mulleri* and *Q. john-tuckeri* split in two clusters (Fig. S1d, Fig. 3e) whereas *Q. dumosa* and *Q. pacifica* split in five clusters (Fig. S1e, Fig. 3f). All analyses revealed considerable rates of hybridization, particularly between *Q. berberidifolia* and *Q. cornelius-mulleri* in southern California and between *Q. berberidifolia* and *Q. durata* in northern California (Fig. 3). Further analyses within each of the species did not show any further genetic subdivision, with the exception of *Q. berberidifolia*, which showed two genetic clusters with a latitudinal cline (Fig. 3c). However, STRUCTURE analyses for *Q. berberidifolia* excluding individuals with admixed ancestry with other species ($q < 0.9$; e.g. Ortego & Bonal 2010) indicated an optimal value of $K = 1$ (data not shown), suggesting that the latitudinal cline of genetic differentiation within this species may be caused by hybridization with the different taxa with which it interbreeds in different parts of its distribution range. Further analyses excluding hybrid individuals for the groups *Q. cornelius-mulleri* and *Q. john-tuckeri* (Fig. 4a) and *Q. dumosa* and *Q. pacifica* (Fig. 4b) showed that optimal K values were the same than those yielded by global analyses including all individuals (Fig. 3). Genetic admixture within these two groups probably reflects both contemporary hybridization among nearby populations and shared ancestry due to recent species divergence (Fig. 4) (e.g. Tsuda *et al.* 2015).

The PCA on nuSSR data (PC1: 1.91 % inertia; PC2: 0.90 % inertia; Fig. 5a) and MANOVA analysis indicated that the obtained scores for each axis significantly differed among the studied species (PC1: $F_{5, 805} = 582.36$, $P < 0.01$; PC2: $F_{5, 805} =$

115.63, $P < 0.01$). *Post hoc* Tukey tests showed that the only non-significant pair-wise comparisons were those between *Q. cornelius-mulleri* and *Q. john-tuckeri* (for PC1), *Q. dumosa* and *Q. pacifica* (for PC1), and *Q. durata* and *Q. cornelius-mulleri* (for PC2). Thus, all species were genetically differentiated for at least one of the two PCs.

BAPS analyses for linked chloroplast markers showed the highest likelihood (log maximum likelihood = -788.1) for $K = 34$ (Fig. 3g). This high number of genetic clusters indicates strong local genetic structure both among and within species, a pattern that has been previously reported in other Californian oaks (Grivet *et al.* 2006; Gugger *et al.* 2013). Twenty clusters were exclusively represented in a single species (i.e. not shared with other species) and the rest were represented in populations from more than one taxon (Fig. 3g; see Table S4, Supporting Information). Accordingly, network analyses representing the relationships among the 45 haplotypes found in our study populations indicated that they are not clustered by species and nine of them were shared by two or three taxa (Fig. 6).

AMOVA analyses showed considerable differences between nuclear and chloroplast genomes in their respective distribution of genetic variation (Table 1). For both nuclear and chloroplast markers, the proportion of total variation among species was low (<10 %; Table 1). However, the percentage of total variation attributed to differences among populations within species was very low in nuclear markers (~3%) and very high in chloroplast markers (~81%). Conversely, variation among individuals within populations was low for cpSSR markers (~9%) and very high for nuSSR markers (~91%) (Table 1).

Phylogenetic analyses

Neighbor-joining trees based on nuSSR markers showed the same patterns of genetic differentiation revealed by STRUCTURE analyses. *Quercus berberidifolia* and *Q. durata* are closely related and more distant from the rest of the species, which in turn form discrete clades but with a low degree of genetic differentiation among them (Fig. 7a). According with STRUCTURE analyses, neighbor-joining trees also suggested that the pairs of species *Q. cornelius-mulleri*/*Q. john-tuckeri* and *Q. dumosa*/*Q. pacifica* are more closely related with each other than with any of the two other species within the clade (Fig. 7a). Hybridization did not have a major impact on phylogenetic analyses and only population 63, a hybrid swarm between *Q. berberidifolia* and *Q. cornelius-mulleri* identified by STRUCTURE analyses (Fig. 3a), was originally classified as *Q. cornelius-mulleri* but placed within the *Q. berberidifolia* clade (see arrow in Fig. 7a). Neighbor-joining trees based on cpSSR markers showed that populations from the same species tend not to be phylogenetically clustered (Fig. 7b), suggesting considerable interspecific gene exchange and little taxonomic value of our set of chloroplast markers (see Petit & Excoffier 2009 and references therein). We obtained similar results for analyses considering five randomly picked individuals per population, indicating that differences in sample sizes among populations (range = 5-18 genotyped individuals/population; Table S1) had no considerable impact on tree topology (data not shown). Analyses performed excluding hybrid individuals ($q < 0.90$) from the dataset also resulted in a similar tree topology (data not shown).

Demographic history: Approximate Bayesian Computation

Considering the scenarios tested for all species and any subset of loci (nuSSR or nuSSR+cpSSR), scenario 2 had the highest posterior probability and 95% confidence

intervals for this model did not overlap with those obtained for the other scenarios (Table 2). Analyses focused on populations of *Q. dumosa* and *Q. pacifica* indicated that a simultaneous split (scenario 1) is the best supported scenario and 95% confidence intervals for this model did not overlap with those obtained for the other scenarios (Table 2). Analyses to estimate confidence in scenario choice based on 500 PODs indicate that Type I and Type II errors for the best supported scenario were moderately low for analyses based on all species (Table 2). However, power analyses showed that Type I errors (false positives) for ABC tests focused on *Q. dumosa* and *Q. pacifica* were high, indicating that confidence in scenario choice is poor. Principal component analyses showed that SS calculated for the posterior simulated datasets of the best supported scenario explained the observed data well. Accordingly, none of the SS used as test statistics significantly differed between observed and simulated datasets after sequential Bonferroni corrections (all P s > 0.1). RMAE values were moderately low (<0.2 in most cases), indicating that estimates of posterior parameters are reliable (Table 3; see also Fig. S2 and S3, Supporting Information). Analyses based on either nuSSR or nuSSR+cpSSR yielded very similar parameter estimates (Table 3). Assuming an average generation time of 50 years for scrub oaks (i.e. half of the generation time generally assumed for large oaks; e.g. Gugger *et al.* 2013), suggest that divergence between *Q. berberidifolia*/*Q. durata* and the rest of the species probably predates the Wisconsin glacialation (~79-91 k years BP; Table 3). Divergence between *Q. berberidifolia* and *Q. durata* (~23-26 k years BP) and among the other four species (~22 k years BP) probably happened around the LGM. Parameter estimates indicate that the divergence among the populations of *Q. dumosa* and *Q. pacifica* probably also took place around the LGM (~19-26 k years BP) (Table 3). ABC analyses exclusively based on cpSSR loci resulted in very low support for any tested scenario, probably as a result

of the scarce resolution provided by only five markers with low variability from a single locus (Table S2) and the considerable haplotype sharing among species (Fig. 6 and 7b and Table S4; see also Petit *et al.* 2002; Petit *et al.* 2004). These two factors strongly limit the power of our chloroplast markers to reconstruct the evolutionary history of the studied species in comparison with nuclear loci (e.g. see Petit & Excoffier 2009).

Species distribution modeling

All SDM had high AUC values (range: 0.89-0.99; Table S3, Supporting Information) and predicted well current species distributions (Fig. 1). Projected distributions into the past suggest that the ranges of some species contracted during the LGM (*Q. berberidifolia*, *Q. cornelius-mulleri*) whereas others experienced considerable expansions (*Q. john-tuckeri*, *Q. pacifica*) (Fig. S4, Supporting Information). The expansion predicted for *Q. pacifica* during the LGM suggests that the geographic range of this species probably overlapped with the inland populations of *Q. berberidifolia* and *Q. dumosa* during this period. It is also remarkable the isolated distribution of *Q. cornelius-mulleri* in both time periods, with very little range overlap with *Q. berberidifolia* and *Q. john-tuckeri* even though they live in close geographical proximity. Finally, it should be noted that the predicted distribution of *Q. durata* in both the present and the LGM must be interpreted with caution given that this species is associated with serpentine soils that were not considered in our SDMs (Whittaker 1954; Forde & Faris 1972; Harrison 1999). For instance, the predicted southward expansion of this species into southern California during the LGM is highly unlikely due to the lack of serpentine soils in the region (Kruckeberg 1984). Serpentine soils are very patchily

distributed in California, represent less than ~1% of land surface of this state (Kruckeberg 1984) and no detailed maps are available for SDM.

Niche divergence/conservatism

All niche identity tests were significant for the two estimates of niche overlap (*I* and *D*), indicating that the studied species are not distributed in an identical environmental space (all $P_s < 0.01$). Background tests showed mixed results depending on the species' pair and the direction of the test (Table 4). Niche overlap between most pairs of species was lower than expected based on the differences in the environmental background in which they occur, particularly when the larger background area was considered (Table 4). Exceptions generally involved comparisons with the island endemic *Q. pacifica* and the widespread *Q. berberidifolia*, which often showed either non-significant differences or higher niche overlap with other species than expected from differences in their environmental backgrounds. Contrarily, the comparisons involving *Q. cornelius-mulleri* and *Q. john-tuckeri* generally revealed that niche overlap with the other species is lower than expected from their environmental backgrounds (Table 4). As indicated above for SDM, comparisons involving *Q. durata* must be interpreted with caution given that this species is associated with serpentine soils, a trait that suggest a remarkable ecological niche differentiation with respect to the other studied taxa.

The first two axes of the PCA jointly explained 68.47% of the variance (PC1: 48.91 %; PC2: 19.57 %; Fig. 5b) and MANOVA analyses indicated that the obtained scores for each axis significantly differed among the studied species (PC1: $F_{5, 2607} = 835.46$, $P < 0.01$; PC2: $F_{5, 2607} = 308.44$, $P < 0.01$). *Post hoc* Tukey tests showed that the only non-significant pair-wise comparisons were those between *Q. john-tuckeri* and

Q. durata (for PC1), and *Q. berberidifolia* and *Q. pacifica* (for PC2). Thus, all species occupied a different environmental space for at least one of the two PCs.

Niche overlap and genetic differentiation

All pair-wise F_{ST} values estimated for nuSSR markers between species were highly significant (all P s < 0.01) and ranged between 0.03 for *Q. dumosa*-*Q. pacifica* and 0.16 for *Q. durata*-*Q. john-tuckeri* (Table S5, Supporting Information). For cpSSR markers, all pair-wise F_{ST} values were also highly significant (all P s < 0.01) and ranged between 0.10 for *Q. berberidifolia*-*Q. cornelius-mulleri* and 0.48 for *Q. durata*-*Q. dumosa* (Table S5). Genetic differentiation estimated at nuSSR markers was not significantly correlated with genetic differentiation estimated at cpSSR markers (Mantel tests, F_{ST} : $r = 0.30$; $P = 0.16$; Reynold's distance: $r = 0.37$; $P = 0.10$; G'_{ST} : $r = -0.01$; $P = 0.54$; Jost's D : $r = 0.09$; $P = 0.33$). For nuSSR markers, Mantel tests showed that niche overlap was not significantly correlated with pair-wise genetic differentiation estimated with F_{ST} (I : $r = -0.17$; $P = 0.28$; D : $r = -0.20$; $P = 0.25$), Reynold's distance (I : $r = -0.17$; $P = 0.28$; D : $r = -0.20$; $P = 0.26$), G'_{ST} (I : $r = -0.05$; $P = 0.46$; D : $r = -0.08$; $P = 0.43$) or Jost's D (I : $r = -0.04$; $P = 0.47$; D : $r = -0.07$; $P = 0.44$). For cpSSR markers, Mantel tests did not show either a significant correlation between niche overlap and pair-wise genetic differentiation estimated with F_{ST} (I : $r = -0.26$; $P = 0.18$; D : $r = -0.27$; $P = 0.18$), Reynold's distance (I : $r = -0.25$; $P = 0.18$; D : $r = -0.27$; $P = 0.19$), G'_{ST} (I : $r = -0.28$; $P = 0.15$; D : $r = -0.29$; $P = 0.14$) or Jost's D (I : $r = -0.21$; $P = 0.22$; D : $r = -0.23$; $P = 0.22$). We obtained qualitatively similar results excluding from the analyses the soil-specialist *Q. durata* or when genetic distances were calculated excluding from the dataset

individuals with different degrees of admixed ancestry with other species (i.e. hybrids or introgressed individuals; see next section) (data not shown).

Discussion

Species evolutionary and demographic history

Demographic, environmental niche and SDM analyses suggest that different factors have probably driven the speciation process within the Californian scrub white oak complex. ABC analyses indicate that the most plausible demographic scenario is the one considering the divergence between the *Q. berberidifolia*/*Q. durata* clade and the rest of the species during the early Wisconsinan glaciation (~79-91 k years BP) followed by a split *Q. berberidifolia* and *Q. durata* around the LGM (~23-26 k years BP). Hierarchical Bayesian clustering analyses, PCA, and SSR-based phylogenetic analyses suggest that *Q. cornelius-mulleri* and *Q. john-tuckeri*, on one hand, and *Q. dumosa* and *Q. pacifica*, on the other hand, diverged more recently than did both species pairs. The geographical ranges of these four species also suggest such a pattern of sequential divergence: species within both pairs of taxa have non-overlapping or only partly overlapping distribution ranges but they are distributed immediately adjacent to each other (Fig. 1). However, ABC analyses suggest a simultaneous speciation among these four taxa around the LGM (~22 k years BP), a scenario also supported by pilot runs exclusively focused on these four species (data not shown). These discrepancies could have resulted from the low power of ABC analyses and our set of nuclear and chloroplast SSR markers to detect subtle differences in divergence times among taxa that have very recently split (Papadopoulou & Knowles 2015).

The ABC analyses suggest a recent split among the studied taxa, but our estimates of divergence time must be interpreted with caution due to the wide 95% CIs obtained for the inferred demographic parameters (Table 3), long overlaps between generations in tree species, and considerable uncertainty in average generation time (see Tsuda *et al.* 2015). Regarding the latter, we considered a generation time of 50 years for scrub oaks, which is half of the generation time generally assumed for large trees (e.g. oaks: Gugger *et al.* 2013; birches: Tsuda *et al.* 2015), but our confidence around this value is low due to the lack of direct estimates and the general controversy around its calculation in long-lived trees (Petit & Hampe 2006; Tsuda *et al.* 2015). So, actual generation times may be longer if Generation time may also potentially vary over time (e.g. in cold-dry vs. warm-wet periods) and among taxa inhabiting different environments (e.g. desert margins from southern California vs. mixed forest from northern California). Moreover, inter-specific hybridization (see discussion below) is expected to have resulted in our DIYABC analyses, which do not accommodate gene flow after divergence (Cornuet *et al.* 2014), have underestimated divergence times even though we excluded from the dataset introgressed individuals (e.g. Tsuda *et al.* 2015).

Different lines of evidence suggest that the divergence between *Q. berberidifolia* and *Q. durata* probably took place in peripatry or sympatry. These two species share a similar distribution range and are the only two taxa within the complex that often form mixed stands or grow in very close geographical proximity, with populations separated by a few hundreds of meters (Forde & Faris 1962). *Quercus durata* is an edaphic specialist of serpentine soils, where *Q. berberidifolia* is unable to form stable populations (Forde & Faris 1962). These soils represent less than ~1% of California's land surface and are patchily distributed across the landscape (Whittaker 1954; Kruckeberg 1984; Harrison 1999; Harrison & Rakaharuna 2010), which is likely to

647 have favored the presence of populations of both species at a dispersal distance of each
648 other and resulted in divergent selection in peripatry or sympatry (*sensu lato*, i.e.
649 absence of physical barriers to gene flow; Papadopulus *et al.* 2011, 2013) (Anacker
650 2014). Our data support a monophyletic origin of populations of *Q. durata*, indicating
651 that populations of this species do not represent soil-adapted ecotypes evolved
652 repeatedly from adjacent non-serpentine types as has been found in other Californian
653 endemics (Rajakaruna *et al.* 2003; Brady *et al.* 2005). Thus, it is possible that a single
654 divergence event associated with the adaptation to serpentine soils took place in
655 allopatry and subsequent range expansion after divergence has resulted in current
656 sympatry/peripatry with *Q. berberidifolia* (Harrison & Rakaharuna 2010; Moyle *et al.*
657 2012; Anacker 2014).

658 More intricate is the evolutionary history of the four putative species within the
659 *Q. dumosa* group (Fig. 3-6). Results of SDMs and analyses of niche overlap suggest
660 different modes of speciation within the *Q. dumosa* group after the divergence of its
661 common ancestor from the *Q. berberidifolia*/*Q. durata* clade. Parapatric divergence
662 followed by significant ecological differentiation has probably shaped the split between
663 the closely related *Q. cornelius-mulleri* and *Q. john-tuckeri*. The distributions of these
664 two species are adjacent to each other but scarcely overlapping (Fig. 1), not even during
665 the LGM when *Q. john-tuckeri* seemed to experience a considerable range expansion
666 (Fig. S4). Thus, their divergence is likely to have been favored by the Transverse
667 Ranges, which separate most populations of both taxa and have been previously
668 identified as an important geographical barrier contributing to genetic splits in oaks
669 (Gugger *et al.* 2013; Ortego *et al.* 2015) and many other taxa (Calsbeek *et al.* 2003;
670 Chatzimanolis & Caterino 2007; Vandergast *et al.* 2008). *Quercus dumosa* and *Q.*
671 *cornelius-mulleri* have probably diverged in allopatry, with no range overlap and

showing a remarkable niche differentiation. *Quercus dumosa* and *Q. john-tuckeri* show significant niche differentiation and distinct distributions in the present but SDM predict a considerable overlap during the LGM when the split was estimated to have taken place, suggesting that ecological divergence in parapatry is the most plausible scenario under the simultaneous divergence suggested by ABC analyses.

Speciation in allopatry is *a priori* the most likely scenario of divergence for the island endemic *Q. pacifica*, but the relationship between this taxa and its most likely sibling species *Q. dumosa* was fairly complex. STRUCTURE analyses showed the presence of five genetic clusters, three mostly represented in *Q. dumosa* and two in *Q. pacifica*, with considerable degree of admixture (Figs. 4g and 5b). ABC analyses focused on these two species revealed that a simultaneous split of *Q. dumosa* and *Q. pacifica* populations is the most supported scenario, suggesting a polyphyletic origin of *Q. pacifica* (Table 2b; Fig. 2b). SDMs indicate that the predicted distributions of both species considerably overlapped during the LGMs, which may have been favored by the fact that lower sea levels during the last glaciation almost connected all Northern Channel Islands with the mainland (Johnson 1978). Accordingly, ABC analyses indicate that divergence among populations of *Q. dumosa* and *Q. pacifica* may have taken place around the LGM (~19-26 k years BP), which could reflect population fragmentation as consequence of the progressively reduced connectivity among California Channel islands and the mainland due to the sea level rising during the early Holocene (Johnson 1978). Thus, the complicated demographic history and patterns of genetic structure observed in these two putative species may be related with the multiple episodes of population reconnections and admixture characteristics of species inhabiting aggregated island complexes linked to the sea level oscillations that took place during the Pleistocene (Esselstyn & Brown 2009; Papadopoulou & Knowles 2015).

The different modes of speciation evidenced by demographic analysis in conjunction with SDM and niche divergence tests are also supported by analyses of the association between genetic differentiation and niche overlap. We found no significant relationship between pair-wise species genetic differentiation and environmental niche overlap (i.e. a lack of phylogenetic niche conservatism *sensu* Losos 2008), which indicates that related species do not show a tendency to occur sympatrically, that environmental niches are highly labile (e.g. Cornuault *et al.* 2015) and, most likely, that different mechanisms (e.g. geographic isolation in vicariance *vs.* ecological divergence in sympatry or peripatry) have driven speciation within the Californian scrub white oak complex (Warren *et al.* 2008; Losos 2008; Nakamoto *et al.* 2010). However, given that these analyses cannot tease apart the geographic signals of evolutionary and ecological processes, our results must be interpreted with caution in terms of the underlying factors that may have driven the observed biogeographic patterns (Warren *et al.* 2014).

Distribution of genetic variation in nuclear and chloroplast genomes

Our comparative analyses on chloroplast and nuclear genomes revealed different demographic histories and highly contrasting patterns in the spatial distribution of genetic variation for each, which is expected given that they have different forms of inheritance, mutation rates, and dispersal vectors (Petit *et al.* 2005). Inter-specific genetic differentiation was not significantly correlated between both genomes (Table S5) and AMOVA analyses showed that most variation for cpSSR markers was explained by differences among populations whereas the largest proportion of total genetic variation for nuSSR markers was attributed to differences among individuals within populations (Table 1). AMOVA analyses also showed that only a low proportion

of variance was explained by differences among species (<10 % for both genomes; Table 1), which contrasts with previous studies comparing other closely related tree species (e.g. Du *et al.* 2015). The low degree of genetic differentiation among populations and the high levels of genetic diversity within populations for nuSSR loci relative to cpSSR loci probably reflect high rates of pollen-mediated gene flow via wind *versus* the more restricted gene flow through seed dispersal by animals, a pattern well documented in previous studies analyzing contemporary processes of pollen and seed dispersal (e.g. Dow & Ashley 1998; Ortego *et al.* 2014b; Sork *et al.* 2015). Conversely, the high genetic differentiation among populations for cpSSR markers is probably reflecting the fact that most populations have been monopolized by one or a few chloroplast haplotypes (see haplotype network and BAPS analyses; Fig. 3a and Fig. 6), which has been reported in previous studies on the Californian Valley oak (*Quercus lobata*) and is congruent with low rates of acorn dispersal (Grivet *et al.* 2005, 2006, 2009). Thus, our results are in agreement with previous studies showing strong local genetic structure for maternally inherited chloroplast genomes (e.g. Grivet *et al.* 2005, 2006; Gugger *et al.* 2013) and pervasive haplotype sharing among sympatric populations of different species that yet maintain their taxonomic, morphological, and nuclear genetic identity in presence of frequent interspecific hybridization (e.g. Whittemore & Schaal 1991; Petit *et al.* 2002, 2004; Lexer *et al.* 2006).

Interspecific hybridization

Oaks represent a paradigmatic example of the complexity around the biological species concept due to their high levels of interspecific gene flow (Coyne & Orr 2004; e.g. Howard *et al.* 1997; Curtu *et al.* 2007; Cavender-Bares & Pahlich 2009; Ortego &

Bonal 2010), which in turn has made the systematics of the genus challenging (Van Valen 1976; Manos *et al.* 1999; Hipp *et al.* 2014). Our studied species are not an exception and Bayesian clustering analyses have shown considerable rates of hybridization among different taxa within the complex (Fig. 3). Introgression was mostly detected in zones of sympatry/parapatry between two or more taxa, as is well exemplified in the co-distributed populations of *Q. berberidifolia* and *Q. cornelius-mulleri* in southern California (Riordan *et al.* 2015) and *Q. berberidifolia* and *Q. durata* in central-northern California. Although we found some populations that constitute hybrid swarms (e.g. site 63; Fig. 3b and Fig. 6), all the putative species maintain their taxonomic and genetic identity across their distribution ranges (Muir *et al.* 2000; Ortego *et al.* 2014a). Differences in flowering times among taxa or selection against hybrids due to their lower viability or performance in either parental environment may be responsible of maintaining the distinctness of the studied species in the presence of gene flow (Forde & Faris 1962; Muir *et al.* 2000; Ortego *et al.* 2014a). With the exception of *Q. dumosa* and *Q. pacifica*, our analyses also indicate a lack of intraspecific genetic structure across the entire ranges of the different taxa, even when some species (*Q. berberidifolia*, *Q. durata*; Fig. 1) are distributed over a large geographic area for which previous studies have found genetic subdivision in other co-distributed oaks (Ortego *et al.* 2012; Gugger *et al.* 2013; Ortego *et al.* 2015; Fig. 1). This suggests that widespread gene flow in these wind-pollinated trees is a remarkable homogenizing factor across populations within each studied species, but selection against hybrids or assortative mating is probably preventing all species from converging in a hybrid swarm (Muir *et al.* 2000).

Conclusions

772

773 Overall, our study exemplifies the complex processes that can underlie early stages of
774 species formation and suggest that different processes leading to divergence are likely to
775 be involved in speciation within the Californian scrub white oak complex. With the
776 exception of the island endemic *Q. pacifica* and the closely related *Q. dumosa*, our
777 analyses support the taxonomic distinctiveness of all the species from either a molecular
778 and/or an ecological perspective. Further studies taking advantage of high throughput
779 sequencing technology can provide in the future a higher resolution to address in more
780 detail the demographic history of the complex (Emerson *et al.* 2010; Papadopoulou &
781 Knowles 2015) and understand if some loci under selection related with adaptation to
782 particular environmental conditions (e.g. serpentine soils in *Q. durata* and xeric
783 conditions in *Q. cornelius-mulleri*) are involved in species divergence, which may
784 explain the subtle genetic differentiation observed among the different taxa when
785 examined at neutral markers (Funk *et al.* 2012; Shafer & Wolf 2013; e.g. Hohenlohe *et*
786 *al.* 2010; Gompert *et al.* 2013; Papadopoulos *et al.* 2013).

787

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789

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Data accessibility

Simple sequence repeat data, climate layers and occurrence data used for MAXENT and ENMTOOLS analyses, neighbor-joining phylogenetic trees, and input files used for PCAs, Mantel tests and analyses in POPULATIONS, STRUCTURE and BAPS are stored and accessible through the Dryad data repository: doi:10.5061/dryad.52504.

Supporting information

Additional supporting information may be found in the online version of this article.

Table S1 Geographical location of scrub white oak sampling sites in California.

Table S2 Nuclear and chloroplast SSR loci used to genotype Californian scrub white oaks.

Table S3 Summary of species distribution modeling (SDM) results.

Table S4 Number of individuals of each species assigned to each genetic cluster identified by BAPS analyses on chloroplast SSR markers.

Table S5 Pairwise F_{ST} , Reynold's distances, G'_{ST} and Jost's D between the studied Californian scrub white oak species for nuclear and chloroplast SSR markers.

Figure S1 Results of Bayesian clustering analyses in STRUCTURE.

Figure S2 Prior and posterior probability distribution of parameters calculated in DIYABC analyses based on nuclear SSR markers.

Figure S3 Prior and posterior probability distribution of parameters calculated in DIYABC analyses based on both nuclear and chloroplast SSR markers.

Figure S4 Predicted distribution for the six studied scrub white oaks from California during the Last Glacial Maximum (LGM; c. 21 000 BP).

1172 **Table 1** Results of AMOVAs for nuclear (nuSSR) and chloroplast (cpSSR) markers. All comparisons were highly significant ($P < 0.001$ in all
1173 cases).

1174						
	nuSSR			cpSSR		
	Sum of squares	Variance components	Percentage of variation	Sum of squares	Variance components	Percentage of variation
Among species	464.83	0.38	5.88	155.47	0.15	9.98
Among populations within species	768.98	0.20	3.13	910.04	1.23	81.41
Among individuals within populations	8954.31	5.83	90.99	93.08	0.13	8.61

Table 2 Posterior probability of each scenario and 95% confidence intervals (CI) based on the logistic regression approach for approximate Bayesian computation analyses (ABC) considering (a) all species and (b) only populations of coastal (*Q. dumosa*) and island (*Q. pacifica*) scrub oaks. Type I and Type II errors for the best supported scenario (in bold) are indicated. Simulations and ABC analyses were performed only including nuclear markers (nuSSR) and considering both nuclear and chloroplast markers (nuSSR+cpSSR).

Scenario	nuSSR				nuSSR+cpSSR			
	Posterior probability	95% CI	Type I error	Type II error	Posterior probability	95% CI	Type I error	Type II error
(a) All species								
1	0.001	<0.001-0.300			0.008	<0.001-0.213		
2	0.884	0.854-0.914	0.194	0.046	0.888	0.868-0.909	0.192	0.088
3	0.115	<0.001-0.347			0.104	<0.001-0.268		
(b) <i>Q. dumosa</i> and <i>Q. pacifica</i>								
1	0.563	0.555-0.571	0.450	0.150	0.429	0.421-0.438	0.510	0.122
2	0.050	0.046-0.054			0.125	0.119-0.131		
3	0.089	0.084-0.094			0.173	0.166-0.180		
4	0.006	0.003-0.010			0.076	0.071-0.080		
5	0.291	0.284-0.299			0.197	0.190-0.204		

Table 3 Posterior parameter estimates (median and 95% confidence intervals) for the best supported scenarios considering (a) all species (scenario 2) and (b) only populations of coastal (*Q. dumosa*) and island (*Q. pacifica*) scrub oaks (scenario 1). Estimates are based on 1% of simulated datasets closest to the observed values. Simulations and approximate Bayesian computation analyses were performed only including nuclear markers (nuSSR) and considering both nuclear and chloroplast markers (nuSSR+cpSSR). Relative median absolute errors (RMAE) based on 500 pseudo-observed data sets are also indicated for each parameter.

Parameter	nuSSR				nuSSR+cpSSR			
	Median	q [2.5]	q [97.5]	RMAE	Median	q [2.5]	q [97.5]	RMAE
(a) All species								
N1 (<i>Q. berberidifolia</i>)	7950	4980	9800	0.151	8010	5110	9790	0.148
N2 (<i>Q. durata</i>)	3610	1200	8740	0.175	3980	1300	8900	0.195
N3 (<i>Q. cornelius-mulleri</i>)	5980	2660	9530	0.147	6450	3210	9550	0.157
N4 (<i>Q. john-tuckeri</i>)	6370	2750	9640	0.171	6480	2990	9600	0.167
N5 (<i>Q. dumosa</i>)	5440	2200	9410	0.164	5070	2120	9180	0.165
N6 (<i>Q. pacifica</i>)	6990	3440	9680	0.175	7580	4460	9770	0.159
t3	423	146	914	0.199	416	157	943	0.187
t4	412	133	1140	0.248	460	136	1250	0.234
t5	1440	512	6020	0.118	1230	406	5570	0.122
μ (nuSSR)	7.8×10^{-04}	4.7×10^{-04}	9.9×10^{-04}	0.165	7.7×10^{-04}	4.6×10^{-04}	9.9×10^{-04}	0.190
μ (cpSSR)	-	-	-	-	1.4×10^{-04}	1.0×10^{-04}	3.5×10^{-04}	0.204
(b) <i>Q. dumosa</i> and <i>Q. pacifica</i>								
N1	4310	1630	9170	0.178	4690	1930	8920	0.179
N2	7690	3830	9850	0.182	6210	2930	9450	0.197
N3	6980	3100	9780	0.176	7000	3540	9660	0.191
t2	376	136	769	0.207	519	179	1910	0.206
μ (nuSSR)	7.1×10^{-04}	3.8×10^{-04}	9.8×10^{-04}	0.203	8.2×10^{-04}	4.8×10^{-04}	1.0×10^{-03}	0.214
μ (cpSSR)	-	-	-	-	1.6×10^{-04}	1.0×10^{-04}	3.5×10^{-04}	0.243

N = effective population sizes for each taxa; t = time in generations; μ = mutation rate

1201 **Table 4** Results of background tests considering different indexes of niche overlap
1202 (Warren's *I*: left; Schoener's *D*: right) and background areas obtained using different
1203 distance buffers around occurrence points. Table indicates whether actual values of
1204 niche overlap of two species are more or less similar than expected based on the
1205 differences in the environmental background in which they occur.

Species for observed distribution	Species for background	<i>I</i>	<i>D</i>	1-km buffer	5-km buffer	10-km buffer
<i>Q. berberidifolia</i>	<i>Q. durata</i>	0.849	0.618	More**/More**	NS/NS	Less*/Less***
<i>Q. berberidifolia</i>	<i>Q. cornelius-mulleri</i>	0.280	0.088	NS/NS	NS/NS	Less*/Less***
<i>Q. berberidifolia</i>	<i>Q. john-tuckeri</i>	0.717	0.437	NS/Less**	Less*/Less***	Less**/Less***
<i>Q. berberidifolia</i>	<i>Q. dumosa</i>	0.607	0.318	NS/NS	NS/NS	Less***/*Less**
<i>Q. berberidifolia</i>	<i>Q. pacifica</i>	0.126	0.042	NS/NS	NS/NS	NS/NS
<i>Q. durata</i>	<i>Q. berberidifolia</i>	0.849	0.618	Less***/*Less***	Less***/*Less***	Less***/*Less**
<i>Q. durata</i>	<i>Q. cornelius-mulleri</i>	0.081	0.020	Less*/NS	Less***/*Less**	Less***/*Less***
<i>Q. durata</i>	<i>Q. john-tuckeri</i>	0.538	0.316	Less***/*Less**	Less***/*Less**	Less***/*Less***
<i>Q. durata</i>	<i>Q. dumosa</i>	0.439	0.202	NS/NS	NS/NS	NS/NS
<i>Q. durata</i>	<i>Q. pacifica</i>	0.079	0.022	NS/NS	NS/NS	NS/NS
<i>Q. cornelius-mulleri</i>	<i>Q. berberidifolia</i>	0.280	0.088	More*/More**	NS/More**	NS/NS
<i>Q. cornelius-mulleri</i>	<i>Q. durata</i>	0.081	0.020	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. cornelius-mulleri</i>	<i>Q. john-tuckeri</i>	0.424	0.152	Less**/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. cornelius-mulleri</i>	<i>Q. dumosa</i>	0.041	0.010	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. cornelius-mulleri</i>	<i>Q. pacifica</i>	0.002	0.001	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. john-tuckeri</i>	<i>Q. berberidifolia</i>	0.717	0.437	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. john-tuckeri</i>	<i>Q. durata</i>	0.538	0.316	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. john-tuckeri</i>	<i>Q. cornelius-mulleri</i>	0.424	0.152	More**/*NS	NS/NS	NS/Less**
<i>Q. john-tuckeri</i>	<i>Q. dumosa</i>	0.357	0.148	NS/NS	Less**/*NS	Less**/*Less**
<i>Q. john-tuckeri</i>	<i>Q. pacifica</i>	0.059	0.014	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. dumosa</i>	<i>Q. berberidifolia</i>	0.607	0.318	More***/*More***	More***/*More***	More***/*More***
<i>Q. dumosa</i>	<i>Q. durata</i>	0.439	0.202	More***/*More***	Less*/Less*	Less***/*Less***
<i>Q. dumosa</i>	<i>Q. cornelius-mulleri</i>	0.041	0.010	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. dumosa</i>	<i>Q. john-tuckeri</i>	0.357	0.148	Less***/*Less***	Less***/*Less***	Less***/*Less***
<i>Q. dumosa</i>	<i>Q. pacifica</i>	0.384	0.162	Less***/*NS	NS/NS	NS/NS
<i>Q. pacifica</i>	<i>Q. berberidifolia</i>	0.126	0.042	More***/*More***	NS/NS	Less**/*NS
<i>Q. pacifica</i>	<i>Q. durata</i>	0.079	0.022	More***/*More**	NS/NS	Less***/*Less***
<i>Q. pacifica</i>	<i>Q. cornelius-mulleri</i>	0.002	0.001	NS/NS	NS/NS	NS/NS
<i>Q. pacifica</i>	<i>Q. john-tuckeri</i>	0.059	0.014	Less**/*Less**	Less***/*Less***	Less***/*Less***
<i>Q. pacifica</i>	<i>Q. dumosa</i>	0.384	0.162	More**/*NS	More*/NS	More**/*NS

1206

1207 NS = Not significant; * $P < 0.1$; ** $P < 0.05$; *** $P < 0.01$

Figure legends

Figure 1 Distribution modeling for the six scrub white oak study species from

California. Color scales refer to logistic probability of occurrence and black dots indicate sampling sites for each species.

Figure 2 Scenarios tested using Approximate Bayesian Computation (ABC)

considering (a) all studied species and (b) only populations of coastal (*Q. dumosa*) and island (*Q. pacifica*) scrub oaks. In these scenarios, $t\#$ represents time in generations, $N\#$ effective population sizes of the different populations and $r\#$ immigration rates from donor populations.

Figure 3 Results of genetic assignments based on the Bayesian methods implemented

in the programs (a-f) STRUCTURE (nuSSR markers) and (g) BAPS (cpSSR markers; $K = 34$). Panels a-f show hierarchical analyses in STRUCTURE considering (a) the six scrub white oak species ($K = 2$), (b, c) *Q. berberidifolia* and *Q. durata* ($K = 2$ and $K = 3$), (d) *Q. cornelius-mulleri*, *Q. john-tuckeri*, *Q. dumosa* and *Q. pacifica* ($K = 2$), (e) *Q. cornelius-mulleri* and *Q. john-tuckeri* ($K = 2$), and (f) *Q. dumosa* and *Q. pacifica* ($K = 5$). Each individual is represented by a vertical bar, which is partitioned into K colored segments showing the individual's probability of belonging to the cluster with that color. Thin vertical black lines separate individuals from different sampling localities, which are arranged according to their geographical location from southeast (left) to northwest (right) as presented in Table S1. The only two individuals collected from locality 6 failed to amplify for most chloroplast markers and were not included in BAPS analyses.

Figure 4 Genetic assignment of the focal populations of (a) *Quercus cornelius-mulleri* and *Q. john-tuckeri* ($K = 2$), and (b) *Q. dumosa* and *Q. pacifica* ($K = 5$) according to the Bayesian method implemented in the program STRUCTURE after excluding individuals with some degree of admixed ancestry ($q < 0.9$) with other species. Admixture proportions generated by STRUCTURE were represented using pie charts, with each color indicating a different genotypic cluster. Pie chart size is proportional to the number of genotyped individuals at each location.

Figure 5 Principal component analyses (PCA) for (a) genetic (16 nuSSR loci) and (b) environmental (7 bioclimatic variables) data for the six studied scrub white oaks from California. PCA on genetic data includes all the genotyped individuals ($n = 812$) and PCA on environmental variables considered all occurrence points used for species distribution modeling ($n = 2610$). Different species are represented with different colors.

Figure 6 Haplotype network for cpSSR markers. Pie chart size is proportional to the number of individuals and colors indicate the proportion of individuals of each species with each haplotype.

Figure 7 Unrooted neighbour-joining trees based on Cavalli-Sforza's distance for (a) nuclear and (b) chloroplast SSR markers. Only populations with five or more genotyped individuals have been included in the analyses. The arrow indicates a hybrid swarm population between *Q. berberidifolia* and *Q. cornelius-mulleri* and the scale bars show Cavalli-Sforza's genetic distance between samples. Bootstrap values below 0.50 are omitted.

(a)

(b)