

1 **Effect of dissolved oxygen and temperature on macromolecular composition and PHB storage of**  
2 **activated sludge**

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4 PAULA REYES<sup>1</sup>, ALEJANDRA URTUBIA<sup>2</sup>, MARÍA C. SCHIAPPACASSE<sup>1</sup>, ROLANDO  
5 CHAMY<sup>1</sup>, SILVIO MONTALVO<sup>3</sup> and RAFAEL BORJA<sup>4\*</sup>

6  
7 *<sup>1</sup>Escuela de Ingeniería Bioquímica, Pontificia Universidad Católica de Valparaíso, Valparaíso, Chile*

8 *<sup>2</sup>Departamento de Ingeniería Química y Ambiental,*  
9 *Universidad Técnica Federico Santa María, Valparaíso, Chile*

10 *<sup>3</sup>Departamento de Ingeniería Química, Universidad de Santiago de Chile, Santiago de Chile, Chile*

11 *<sup>4</sup>Instituto de la Grasa (CSIC), Sevilla, Spain*

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14 \* Address correspondence to R. Borja, Instituto de la Grasa (CSIC), Avda. Padre García Tejero, 4,  
15 41012-Sevilla, Spain; Phone: +34 95 4692516, Ext. 152; Fax : +34 95 4691262; E-mail:  
16 rborja@cica.es.

24 **Abstract**

25 The macromolecular composition of activated sludge (lipids, intracellular proteins and intracellular  
26 polysaccharides) was studied together with its capacity to store macromolecules such as  
27 polyhydroxybutyrate (PHB) in a conventional activated sludge system fed with synthetic sewage water  
28 at an organic load rate of 1.0 kg COD/(m<sup>3</sup>·d), varying the dissolved oxygen (DO) and temperature. Six  
29 DO concentrations (0.8, 1.0, 1.5, 2.0, 2.5 and 8 mg/L) were studied at 20°C with a sludge retention  
30 time (SRT) of 6 days. In addition, four temperatures (10°C, 15°C, 20°C and 30°C) were assessed at  
31 constant DO (2 mg/L) with 2 days SRT in a second experimental run. The highest lipid content in the  
32 activated sludge was 95.6 mg/g VSS, obtained at 30°C, 2 mg/L of DO and a SRT of 2 days. The  
33 highest content of intracellular proteins in the activated sludge was 87.8 mg/g VSS, obtained at 20°C, 8  
34 mg/L of DO and a SRT of 6 days. The highest content of intracellular polysaccharides in the activated  
35 sludge was 76.6 mg/g VSS, which was achieved at 20°C, a SRT of 6 days and a wide range of DO.  
36 The activated sludge PHB storage was very low for all the conditions studied.

37  
38 **Keywords:** Activated sludge, dissolved oxygen, macromolecular composition, PHB, temperature.

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41 **Introduction**

42  
43 A large amount of excess waste activated sludge (WAS) from urban wastewater treatment plants is  
44 generated daily worldwide.<sup>[1, 2]</sup> In aerobic treatment systems 0.5 to 0.6 kg of mixed liquor volatile  
45 suspended solids (MLVSS) per kg of biological oxygen demand (BOD<sub>5</sub>) removed is produced. Not  
46 only that, the treatment and disposal of the excess WAS from sewage sludge originating from  
47 wastewater treatment plants accounts for as much as 60% of the total cost of wastewater treatment.<sup>[3]</sup>

48 At worldwide level, the common practices for the management of sewage sludge include: codisposal  
49 landfilling, land application and thermal processing. In Chile, sludges are arranged in monofills or  
50 codisposal landfills, with limited application to soils due to environmental restrictions. In order to  
51 stabilize sludge, the major sewage water treatment plants in **most countries** digest it anaerobically,  
52 producing biogas, which is sometimes used as energy. However, for different reasons the exploitation  
53 of biogas for energy purposes is limited. Therefore, this way of recovering part of the investment made  
54 in constructing the plant is not widely used in **some countries such as Chile**. Therefore, it is advisable  
55 to look for other alternatives for the economic valorisation of the sludge generated in wastewater  
56 treatment plants by activated sludge systems.

57 On the other hand, secondary sludge typically consists of valuable organic substances such as nucleic  
58 acids, enzymes, proteins, and polysaccharides,<sup>[4,5]</sup> and between 66-81% of total solids are volatile  
59 solids (VS). **Proteins, carbohydrates and lipids were in the ranges 0.34-0.47 g equivalent bovine serum  
60 albumin/g VS, 0.17-0.30 g equivalent glucose/g VS and 0.00-0.09 g/g VS, respectively.**<sup>[6]</sup>

61 Huynh et al.<sup>[7]</sup> discovered that the predominant fatty acids of sludge oil extracted from activated sludge  
62 are palmitic acid (19-27%), palmitoleic acid (15-20%), and octadecenoic isomers (20-33%). These  
63 same researchers observed that the amount of neutral lipids from activated sludge is 7.87% of its dry  
64 weight, using for this quantification subcritical water treatment instead of the traditional systems  
65 regularly used.

66 It is well known that under transient conditions the growth of the heterotrophic biomass becomes  
67 imbalanced because there is a faster adaptation to the changing environment. Two physiological  
68 adaptations can occur: an increase in the growth rate and/or substrate storage. The presence of  
69 biodegradable storage compounds such as polyhydroxyalkanoate (PHA) and glycogen in activated  
70 sludge has been repeatedly reported.<sup>[6, 8-10]</sup> **The forms of PHA in activated sludge are mainly  
71 polyhydroxybutyrate (PHB) and polyhydroxyvalerate (PHV).**<sup>[11]</sup>

72 Many studies have been carried out on activated sludge to produce PHA, most of them using feast-  
73 famine regimes or nutrient pulses.<sup>[12]</sup> It has been observed that its production depends on operational  
74 parameters such as the type of carbon source, C/N ratio, organic load, dissolved oxygen (DO)  
75 concentration, pH, sludge retention time (SRT), temperature, magnetic field intensity, and nutrient  
76 deficiencies such as nitrogen and phosphorus.<sup>[13-15]</sup>

77 There is currently great interest worldwide to produce various commercially usable macromolecules  
78 from the WAS from sewage treatment plants.<sup>[13, 15]</sup> With this in mind, research has been carried out to  
79 study different alternatives, such as: PHA production as an alternative to the usual petroleum-derived  
80 plastics;<sup>[9]</sup> the production of oils, chars or gases by means of pyrolysis;<sup>[16-18]</sup> the production of  
81 proteins<sup>[19]</sup> and biodiesel production from either the lipids present in sludge<sup>[7]</sup> or the lipids stored by  
82 yeasts growing on pre-treatment sewage sludge.<sup>[20]</sup>

83 As most of the large sewage water treatment plants have activated sludge systems where the  
84 heterotrophic biomass grows in transient states, the objective of this study was to evaluate in a  
85 laboratory-scale activated sludge system the effect of two of the most important above-mentioned  
86 variables such as DO and temperature on the macromolecular composition (intracellular carbohydrates,  
87 intracellular proteins and lipids) and on the PHB stored by the activated sludge. The influence of the  
88 mentioned variables on COD removal efficiency was also assessed.

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90

## 91 **Materials and methods**

92

### 93 ***Experimental set up***

94 Two laboratory-scale activated sludge systems (ASS) were used. Each one is composed of an aerobic  
95 reactor and a secondary clarifier. The useful volumes of the tank reactors and settlers were 2.56 and

96 2.18 L, respectively. The reactors had a cylinder-conical geometry. An air pump with variable flow-  
97 rate was used to control the DO in the systems. The air inlet was located at the bottom of the tanks.  
98 The DO in the reactors was measured with a DO electrode. A cooler-heating blanket connected to a  
99 thermostatic bath controlled the temperature of each reactor.

100 Sludge from a conventional full-scale activated sludge wastewater treatment plant was used as  
101 **concentrated mixed liquor** for the laboratory-scale reactors (AAS). The main characteristics of this  
102 inoculum were: pH, 7.25; total suspended solids (TSS): 4900 mg/L; and volatile suspended solids  
103 (VSS), 3900 mg/L. The influent of the AAS was a synthetic wastewater with a chemical oxygen  
104 demand (COD) concentration of approximately 600 mg/L. It was prepared with starch (400 mg/L),  
105 sunflower oil (35 mL/L), ovoalbumin (40 mg/L), urea (26 mg/L),  $\text{KH}_2\text{PO}_4$  (10.5 mg/L),  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$   
106 (44 mg/L),  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  (0.98 mg/L), KCl (42.5 mg/L),  $\text{NaHCO}_3$  (17.5 mg/L), yeast extract (125  
107 mg/L) and a trace element solution (2 mL/L). The composition of the trace element solution was based  
108 on  $\text{FeCl}_3 \cdot 4\text{H}_2\text{O}$  (1000 mg/L),  $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$  (1000 mg/L),  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$  (250 mg/L),  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$  (15  
109 mg/L),  $\text{H}_3\text{BO}_3$  (25 mg/L),  $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$  (45 mg/L),  $\text{NaSeO}_3 \cdot \text{H}_2\text{O}$  (50 mg/L),  $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$  (25  
110 mg/L), EDTA (500 mg/L), and HCl 36% (0.5 mL/L). The COD concentration of the synthetic  
111 wastewater can be considered as intermediate according to the literature, where values in the range of  
112 350-1000 mg/L are reported.<sup>[21]</sup>

113

#### 114 ***Operational conditions***

115 The ASS was operated in continuous mode at a constant organic load rate (OLR) of 1.0 kg COD/(m<sup>3</sup>·d)  
116 and a hydraulic retention time (HRT) of 14.4 h for all conditions studied. Two different operating  
117 strategies were evaluated: (1) operation of ASS at ambient temperature (20°C) with the DO varying  
118 from 0.8, 1.5, 2, 2.5 to 8 mg/L, with a SRT of 6 days; (2) operation of ASS at a constant DO of 2  
119 mg/L, varying the operational temperature from 10°C, 15°C, 20°C to 30°C, with a SRT of 2 days. A

120 recirculation ratio (recirculation flow/raw wastewater) of 0.5 was applied. All experiments were carried  
121 out in duplicate reactors and the final results averaged.

122

### 123 *Analytical methods*

124 Influent and effluent COD, total suspended solids (TSS) and volatile suspended solids (VSS) within  
125 the reactor were measured according to Standard Methods.<sup>[22]</sup> Lipids, intracellular proteins,  
126 intracellular polysaccharides and PHB content of the sludge were measured when a steady-state was  
127 reached. Steady-state was assumed when the percentage of COD removal was kept about 90% for  
128 more than 10 days. To measure the intracellular proteins and the intracellular polysaccharides, the cells  
129 were lysed by means of treatment for 30 min in boiling sodium hydroxide (2M), then cooled on ice and  
130 neutralized with 2M hydrochloric acid.<sup>[23]</sup> Finally, the samples were subjected to vortex and  
131 centrifuged, taking the supernatant for the analysis of non-cellular organic matter. The lipid, protein  
132 and polysaccharide content in the sludge was measured by Soxhlet,<sup>[24]</sup> the Lowry modified method<sup>[25]</sup>  
133 and the Dubois method,<sup>[26]</sup> respectively. The biomass PHB content was obtained by HPLC using a  
134 Biorad HPX-87H column with a DAD L-7450A detector, prior to extracting with potassium  
135 hydroxide.<sup>[23]</sup>

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## 138 **Results and discussion**

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### 140 *Effect of dissolved oxygen concentration*

141 The ASS operated at 1 kg COD/(m<sup>3</sup>·d) OLR, 20°C, a SRT of 6 days and DO of between 0.8 and 8.0  
142 mg/L, keeping an average TSS of 3 g/L in the reactor, of which VSS was about 78%. Average COD  
143 removal efficiencies reached 94% for most cases studied, as can be observed in Figure 1.

144 COD removal efficiencies of 83% and 57% were achieved in the aerobic treatment of diluted palm oil  
145 mill effluent (POME) (COD = 1000 mg/L) in AAS operating at 36h and 24h HRTs respectively, and at  
146 DO concentrations in the range of 1.8-2.2 mg/L.<sup>[27]</sup>

147 Lipids, intracellular protein and intracellular polysaccharide contents of the active sludge at different  
148 DOs are presented in Figure 2. As can be observed, high variations in lipid and intracellular protein  
149 contents were obtained. However, the intracellular polysaccharides remained steady at a value of  
150 around 76.6 mg/g VSS.

151 The highest lipid and intracellular protein contents in the activated sludge were found at 8 mg/L of DO  
152 (57.8 and 87.8 mg/g VSS, respectively). The maximum lipid value obtained corresponded to 67% of  
153 the maximum value reported by Mottet et al.<sup>[6]</sup> The maximum intracellular protein and intracellular  
154 polysaccharide values obtained from activated sludge corresponded to 56% and 47% of the values  
155 reported by Frølund et al.,<sup>[28]</sup> respectively, for sludge where the extracellular polymeric substances  
156 have been subtracted. The values obtained in the present work were considered adequate, taking into  
157 account that they did not consider the proteins and carbohydrates contained in the cell membranes and  
158 walls.

159 In the present work sludge lipid content positively correlated with DO through the following equation:

$$160 \quad L = 24.17 \ln(DO) + 8.59 \quad (1)$$

161 where  $L$  is the lipid content expressed in mg/g VSS and  $DO$  is the dissolved oxygen concentration  
162 expressed in mg/L, the regression coefficient being 0.983. An increase in the sludge intracellular  
163 protein content was also observed with increased DO, but no correlation among the values obtained  
164 was observed.

165 The PHB content of the sludge obtained at the different DOs studied (Figure 3) varied between 0.44  
166 and 2.28 mg/g VSS, very low values compared with those obtained in transient systems.<sup>[29]</sup> A positive

167 effect on the PHB intracellular storage by the biomass was not observed with limited oxygen as was  
168 also previously reported by Padian et al.<sup>[30]</sup>

169 By contrast, another previous study focussing on PHB production by activated sludge in a two-stage  
170 process revealed that the rate of substrate uptake, as well as the yield and content of PHB increased  
171 with an increase in DO concentration.<sup>[12]</sup> The studies reported by Qu and Liu<sup>[12]</sup> also showed that an  
172 enhanced F/M ratio favored PHB accumulation.

173

#### 174 *Effect of temperature*

175 The ASS operated at 1 kg COD/(m<sup>3</sup>·d) OLR, a SRT of 2 days, DO of 2 mg/L and temperatures ranging  
176 between 10°C and 30°C, keeping an average reactor TSS concentration of about 1.7 g/L, of which the  
177 VSS were approximately 80%. For all the cases studied, high COD removal values were achieved,  
178 with average values of 90% at temperatures of 20°C and 30°C, as can be seen in Figure 4.

179 It was previously reported<sup>[21]</sup> that 40% COD removal efficiencies were achieved in activated sludge  
180 systems treating synthetic wastewater (1000 ± 20 mg/L) and real domestic wastewater at 10°C. At  
181 higher temperatures (25-30°C) the efficiency of the reactor was 80%.<sup>[21]</sup> In the same way, Song et al.<sup>[31]</sup>  
182 reported that aerobic granular sludge was cultivated in sequencing batch airlift reactors (SBAR) at  
183 25°C, 30°C and 35°C. These above-mentioned results also showed that 30°C was optimum for mature  
184 granule cultivation, where the granules had a more compact structure, better settling ability and higher  
185 bioactivity with COD removal efficiency of 97% and a total phosphorus removal rate of 75%.<sup>[31]</sup>

186 Activated sludge reactors operating at temperatures in the range of 20-30°C were also used for treating  
187 saline wastewaters generated by marine-product industries. COD removal efficiencies of 88% were  
188 achieved at an OLR of 1 g COD/(L·d) when the system is inoculated with NaCl-acclimated culture.<sup>[32]</sup>

189 Finally, and also for comparative purposes, the influence of the temperature on aerobic treatment was

190 studied in a sequencing batch reactor over 40 weeks in mesophilic (35°C) operation.<sup>[33]</sup> An average  
191 COD removal percentage of 75% was achieved in this case.

192 The lipids, intracellular protein and intracellular polysaccharide contents of the biomass were strongly  
193 affected by temperature (Fig. 5). It was observed that at the highest temperature, the contents of these  
194 macromolecules increased, obtaining 95.6 mg/g VSS of lipids, 36.0 mg/g VSS of intracellular proteins,  
195 and 74.3 mg/g VSS of intracellular polysaccharides at 30°C.

196 Only the lipid content ( $L$ , in mg/g VSS) of the activated sludge could be properly correlated to  
197 temperature ( $T$  in °C) through the following equation (regression coefficient: 0.994):

$$198 \quad L = 3.90 \cdot T - 23.2 \quad (2)$$

199 Mottet et al.<sup>[6]</sup> analysed the lipid content of sludge from activated sludge systems with capacities higher  
200 than 62,000 equivalent inhabitant and their results showed that when decreasing the sludge age both in  
201 mesophilic and thermophilic systems, the lipid content increased. However, it was not possible to  
202 evaluate the effect of the operating temperature on lipid content and these authors<sup>[6]</sup> could not find a  
203 correlation between these two parameters for a fixed sludge age.

204 The lipid content of the sludge increased as DO and temperature increased, and the highest values were  
205 obtained at a temperature of 30°C and a DO of 2 mg/L. However, from the results obtained it was not  
206 possible to evaluate the interaction of both parameters.

207 It was not possible to detect PHB at any temperature, and this suggests that in conventional activated  
208 sludge systems there is no selection or acclimatizing pressure for the enrichment of microorganisms  
209 that can accumulate this type of polymer.

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211

212 **Conclusion**

213

214 The macromolecular composition of sludge can vary according to the operational conditions (such as  
215 DO and temperature) of the ASS. The lipid content of activated sludge increased when either DO or  
216 temperature was increased. On the other hand, the highest activated sludge intracellular polysaccharide  
217 and protein contents were obtained at a temperature of 20°C and a SRT of 6 days. The influence of DO  
218 on the polysaccharide content was not significant, while the maximum protein content was obtained at  
219 a DO of 8 mg/L. Temperatures of 20 °C and 30 °C and DO values between 2.0-2.5 led to achieve  
220 COD removal efficiencies of about 90%. In ASS at the above-mentioned operational conditions, the  
221 sludge generated had very low PHB contents.

222

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### FIGURE CAPTIONS

334

335 **Figure 1.** Average COD removal (%) **as a function of** DO (the dotted vertical lines indicate a change in  
336 conditions).

337 **Figure 2.** Lipid, intracellular protein and intracellular polysaccharide contents of the biomass obtained  
338 from ASS operating at 1.0 kg COD/(m<sup>3</sup>·d) OLR, 20°C, SRT of 6 days for the different DO conditions  
339 tested (the standard deviations of the plotted values were less than 5% in all cases).

340 **Figure 3.** PHB contents of the biomass obtained from ASS operating at 1.0 kg COD/(m<sup>3</sup>·d) OLR,  
341 20°C and SRT of 6 days for the different DO conditions tested (the standard deviations of the plotted  
342 values were less than 5% in all cases).

343 **Figure 4.** Average COD removal **as a function of** temperature (■: influent COD; ◆: effluent COD; ▲:  
344 COD removal efficiency) (the dotted vertical lines indicate a change in conditions).

345 **Figure 5.** Lipid, intracellular protein and intracellular polysaccharide contents of the biomass obtained  
346 from ASS operating at 1.0 kg COD/(m<sup>3</sup>·d) OLR, 2 mg/L of DO and SRT of 2 days for the different  
347 temperatures assayed (the standard deviations of the plotted values were less than 5% in all cases).

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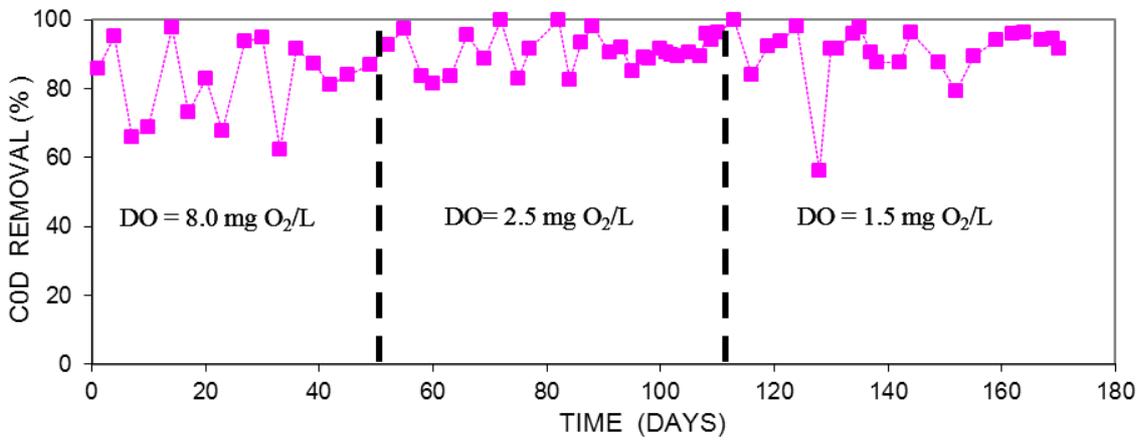
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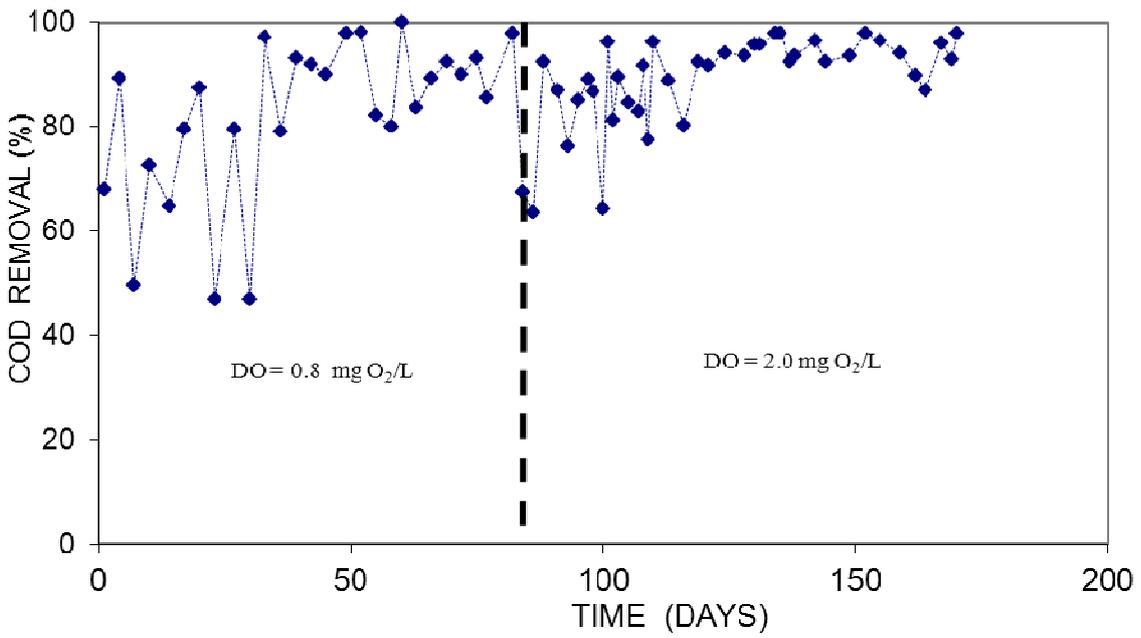
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359 **Fig. 1**

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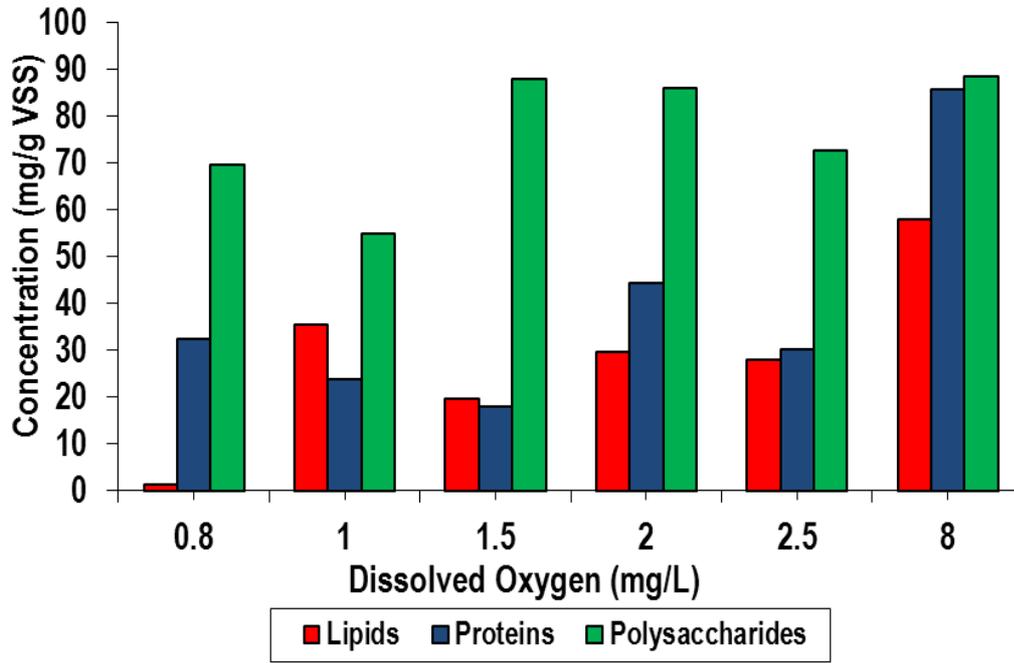
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367 **Fig. 2**

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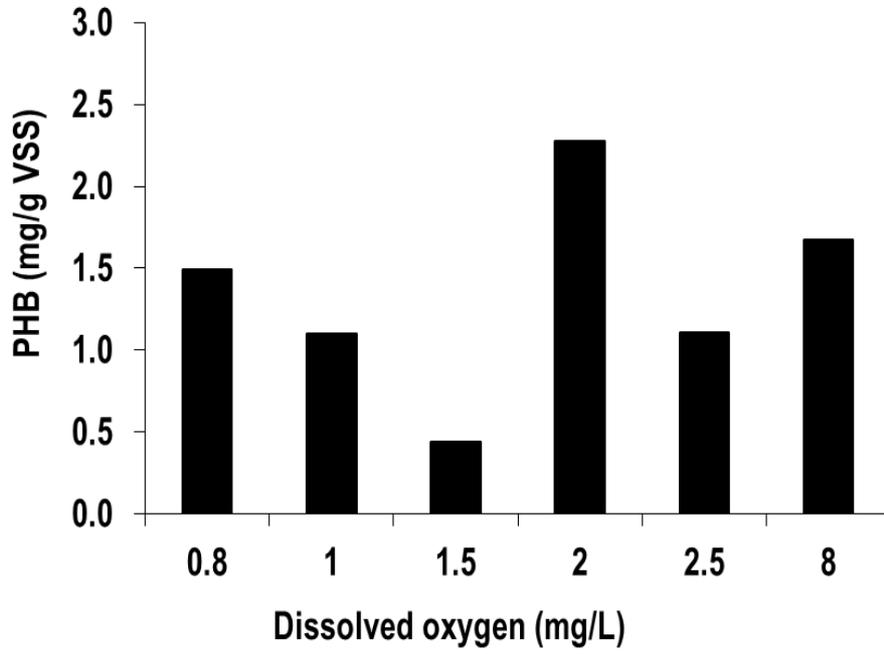
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382 **Fig. 3**

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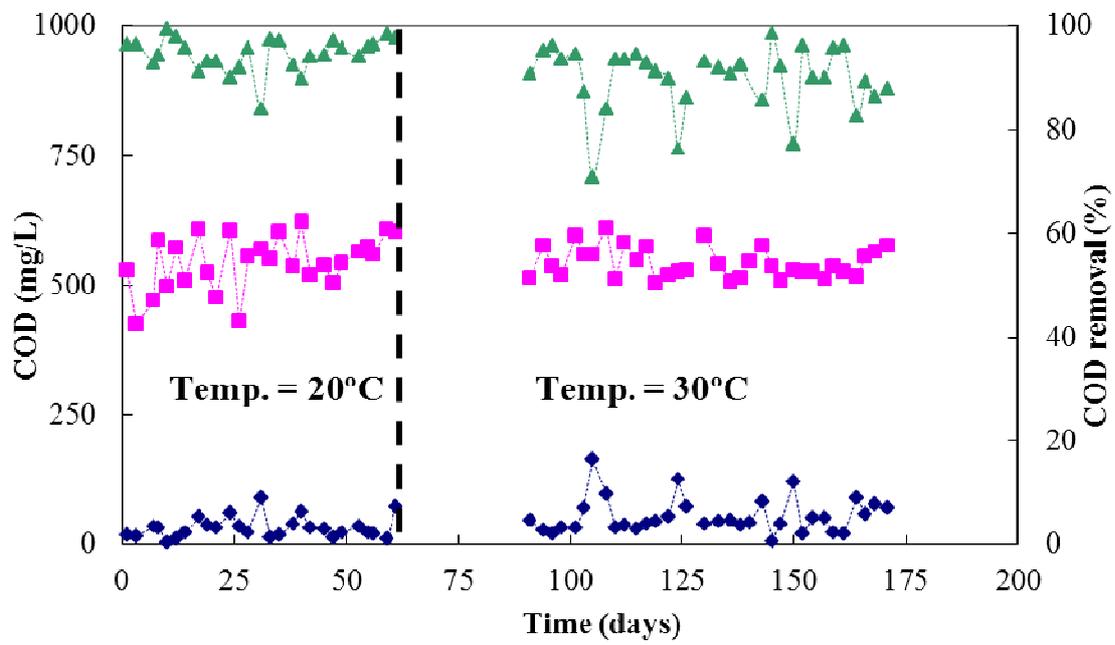
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397 **Fig. 4**

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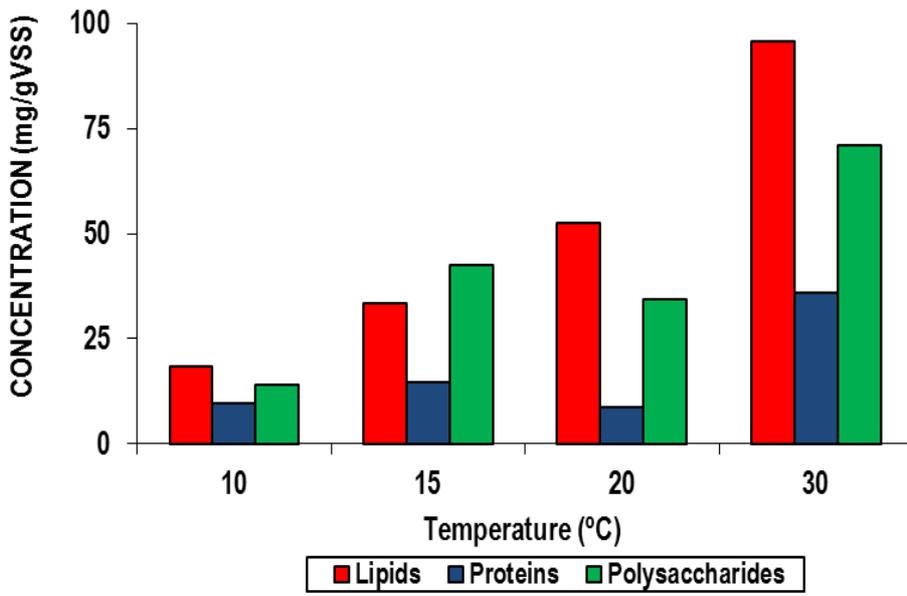
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411 **Fig. 5**

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