Dietary resveratrol prevents Alzheimer's markers and increases lifespan in SAMP8

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Abstract
Resveratrol is a polyphenol mainly found in grapes and red wine that has been reported as a caloric restriction (CR) mimetic through Sirtuin 1 (SIRT1) activation. Resveratrol increases metabolic rate, insulin sensitivity, mitochondrial biogenesis, physical endurance and reduces fat accumulation in mice. In addition, resveratrol may be a powerful agent to prevent age-associated neurodegeneration and to improve cognitive deficits in Alzheimer's disease (AD). Moreover, several findings support that longevity in mice could be promoted by CR .
In this work, we studied the role of dietary resveratrol in SAMP8 mice, a model of age related AD. Here we found that resveratrol supplementation increases mean life expectancy and maximal life span in SAMP8 and in its control-related strain SAMR1. In addition, we also examined the resveratrol-mediated neuroprotective effects in several specifically AD hallmarks. We found that not only activates AMPK pathways and prosurvival routes such as SIRT1 in vivo, but also reduces cognitive impairment and has a neuroprotective role decreasing amyloid burden and reducing tau hyperphosphorylation.

Key words: Senescence, resveratrol, Sirtuin 1, AMPK, Alzheimer's disease, β-amyloid, Tau, memory impairment

1. **Introduction**

Resveratrol (trans-3,4',5-trihydroxystilbene), a naturally occurring polyphenol mainly found in grapes and red wine, has been reported as a caloric restriction (CR) mimetic with potential anti-aging and anti-diabetogenic properties. Resveratrol increases metabolic rate, insulin sensitivity, mitochondrial biogenesis, physical endurance and reduces fat accumulation in mice (Lagouge et al., 2006; Baur et al., 2006). However, the most accepted mechanistic hypothesis is that resveratrol effects, similar to CR, are driven through Sirtuin 1 (SIRT1) regulation. CR and resveratrol lead to increases in the cellular AMP/ATP ratio which results in activation of AMPK that initiates a signaling process that recruit mediators of oxidative metabolism and mitochondrial biogenesis including PGC1α, PPARδ and others. Of note, resveratrol also activates AMPK, which regulates insulin sensitivity and mitochondrial biogenesis (Um et al., 2010; Ruderman et al., 2010).

Several findings support that longevity can be promoted by CR (Weindruch et al., 1988, Selman et al., 2008) jointly with its broad anti-aging activity (Park et al., 2009). Similarly, resveratrol treatment has a range of beneficial effects in mice but up to now has failed to increase the longevity of ad libitum-fed animals when started midlife either alone or in combination with other anti-aging strategies such as CR (Baur and Sinclair, 2006; Pearson et al., 2008). In addition, dietary resveratrol mimics the effects of CR in insulin mediated glucose uptake in muscle in aged animals and gene expression profiling suggests that both CR and resveratrol may retard some aspects of aging through alterations in chromatin structure and transcription (Halagappa et al., 2007; Barger et al., 2008).

Several *in vitro* and *in vivo* studies also support the hypothesis that resveratrol may be a powerful agent to prevent age-associated neurodegeneration (Vingtdeux et al., 2008). In *in vitro* models, resveratrol markedly lowers the levels of secreted and intracellular amyloid-beta (Aβ) peptides (Marambaud et al., 2005). Similarly, a grape seed polyphenolic extract administered orally to Tg2576 mice, a murine model of AD (Hsiao et al., 1996) improves cognitive deficits and these effects correlate with reductions in the amounts of high molecular weight Aβ assemblies in the brain (Wang et al., 2008). Similar findings are observed in animals after moderate consumption of red wines (Wang et al., 2006; Ho et al., 2008). Recently it has been showed that resveratrol selectively remodels soluble oligomers, fibrillar intermediates, and amyloid fibrils into alternative aggregated species that are non-toxic (Ladiwala et al., 2010). These studies and other support the theory that resveratrol or polyphenol derivatives could be useful therapeutic agents for AD (Ono et al., 2008, JBC). Nevertheless, it is unknown whether resveratrol has similar effects in age-related models of AD such as the age-accelerated mouse (SAMP8), a rodent strain increasingly being recognized as a model of age-related AD (Takeda 2009; Pallàs et al., 2008). To this end, the SAMP8 strain is characterized by deficits in learning and memory, emotional disorders (reduced anxiety-like behavior), impaired immune response, etc (Yagi 1988, Flood et al., 1998). More importantly, SAMP8 mice also show AD-related pathology such as increases in Aβ (del Valle et al., 2010) and other protein aggregates (Manich et al., 2011), alterations in APP processing by secretases (Morley et al., 2000 and 2002), cerebral amyloid angiopathy (del Valle et al., 2011) and increases in tau hiperphosphorylation (Canudas et al., 2005).

Therefore, in this work we sought to elucidate the role of dietary resveratrol in the SAMP8 mouse. To this end, previous results in SAMP8 demonstrated that low doses and short-term administration of pterostilbene (poliphenolic derivative of resveratrol) show positive effects on behavior, reductions in tau phosphorylation (Chang et al., 2011) and regulation of cascades associated with PPAR alpha. Based on these encouraging findings we thus determined the effects of long-term administration of resveratrol on longevity and signaling cellular processes activated by this polyphenol, namely: the SIRT1 pathway and AMPK system. We also extended these studies by

examining the resveratrol-mediated neuroprotective mechanism in several specifically AD hallmarks present in SAMP8 as Aβ accumulation and tau phosphorylation.

2. Methods

2.1. Animals and resveratrol diet

110 SAMP8 and SAMR1 animals were used for the survival study. The animals received a standard diet (2018 Teklad Global 18% Protein Rodent Maintenance Diet, Harlan) or the same diet supplemented with trans-resveratrol (1g/Kg) starting at two months of age resulting in four groups of 50 to 60 individuals: SAMR1 control, SAMR1 resveratrol, SAMP8 control and SAMP8 resveratrol. For the neurodegeneration studies, two groups of 10-12 SAMP8 mice were fed with the standard diet or the resveratrol diet, starting the supplementation at two months and sacrificing the animals so as to obtain tissue samples at 9 months of age. All the animals had food and water *ad libitum* and were kept in standard conditions of temperature (22 ± 2°C) and 12:12-h light-dark cycles (300 lux/0 lux). Studies were performed in accordance with the institutional guidelines for the care and use of laboratory animals established by the Ethical Committee for Animal Experimentation at the University of Barcelona.

2.2. Object Recognition Test

9-months SAMP8 control (P8ctl) and SAMP8 resveratrol (P8rsv) animals were placed in a 90º two arm, 25 cm long 20 cm high 5 cm wide, black maze. The 20 cm high walls could be lifted off for easy cleaning. The light intensity in the middle of the field was 30 lux. The objects to be discriminated were made of plastic (5.25 cm high, object A and 4.75 cm high, object B). The first three days mice were individually habituated to apparatus for 10 min. The 4th day, the animals were submitted to a 10 min acquisition trial (first trial) during which they were placed in the maze in the presence of two identical novel objects (A+A or B+B) placed at the end of each arm. A 10 min retention trial (second trial) occurred 2 h later. During this second trial objects A and B were placed in the maze and the time that the animal explored the new object (tn) and the old object (to) were recorded. A discrimination index was defined as (tn-to)/(tn+to). In order to avoid object preference biases, objects A and B were counterbalanced so that half of the animals in each experimental group were first exposed to the object A and then to the object B whereas the other half saw first the object B and then the object A was presented. The maze as well as the objects were cleaned with ethanol 96º between different animals so as to eliminate olfactory cues.

2.3. Brain processing

One day after the object recognition test had been conducted, 9-month animals were intracardially perfused after being anaesthetized with 80 mg/Kg of sodium pentobarbital. Afterwards, brains were dissected and separated sagitally in two hemispheres, one for immunohistochemistry and the other for protein extraction. Immunohystochemistry brains were frozen by immersion in isopentane, chilled in dry ice and stored at -80 ºC until sectioning. Thereafter, frozen brains were embedded in OCT cryostat-embedding compound (Tissue-Tek, Torrance, CA), cut into 20-µm-thick sections on a cryostat (Leyca Microsystems, Germany) at -18ºC, and placed on slides. Slides containing brain sections were fixed with acetone for 10 min at 4º C, allowed to dry at room temperature and then frozen at -20ºC until further staining. The cortex and hippocampus of the other hemisphere were dissected and stored at -80ºC until protein extraction (see below).

2.4. Immunohistochemistry

Slides were allowed to defreeze at room temperature and then rehydrated with PBS for 5 min. Then, brain sections were blocked and permeabilized with PBS containing 1%

bovine serum albumin (BSA, Sigma–Aldrich) and 0.1% Triton-X-100 (Sigma–Aldrich) for 20 min. After two 5-min washes in PBS, slides were incubated with the primary antibody for $A\beta_{40}$, $A\beta_{42}$, (see list of antibodies and dilutions below) overnight at 4ºC. They were then washed again and incubated for 1 h at room temperature in the dark with AlexaFluor secondary antibody (see below). After washing again, nuclear staining

was performed by incubating slides in Hoechst (H-33258, Fluka, Madrid, Spain) at 2 µg/ml in PBS for 10 min at room temperature in the dark. Finally, slides were washed, mounted using Prolong Gold (Invitrogen) anti-fade medium, allowed to dry overnight at room temperature and stored at 4ºC. Image acquisition was performed with a fluorescence laser microscope (BX41, Olympus, Germany).

2.5. Protein extraction

Cortex and hippocampus were micronized through freezing with liquid nitrogen and grinding with a mortar. For total protein extraction, lysis buffer (50mM Tris HCl, 150mM NaCl, 5mM EDTA, 1% Triton X-100, pH 7.4) containing complete, Mini, EDTA-free Protease Inhibitor Cocktail (Roche, Mannheim, Germany) and Phosphatase Inhibitor Cocktail 1 (Sigma-Aldrich, St. Louis, MO, USA) were added to micronized tissue and left on ice for 30 min. Then, samples were centrifuged at 10000 g for 10 min and supernatant with total protein content was collected. All the protein extraction steps were carried out at 4ºC. Protein concentration was determined by the Bradford protein assay.

2.6. Western Blot

For Western Blot analysis, 20 ug of protein were denatured at 95 ºC for 5 min in sample buffer (0.5 M Tris-HCl, pH 6.8, 10% glycerol, 2% sodium dodecyl sulphate (SDS), 5% β-mercaptoethanol, 0.05% bromophenol blue) and separated by SDSPAGE on 10% polyacrylamide gels and transferred to Immobilon polyvinylidene difluoride membranes (Millipore, Billerica, MA, USA). The membranes were incubated overnight at 4°C with the primary antibodies (see table 1) diluted with Tris-buffered saline containing 0.1% Tween 20 (TBS-T) and 5% bovine serum albumin (BSA). Membranes were then washed and incubated with secondary antibodies (see table 1) with TBS-T for 1 hour at room temperature. Protein bands were visualized using a chemiluminescence detection kit (Amersham Biosciences). Band intensities were quantified by densitometric analysis, and values were normalized to β-actin.

2.7. List of antibodies

See table 1

2.8. Statistical analysis

Results were subjected to statistical analysis using GraphPad Prism software. Kaplan-Meier survival curve comparison was performed using the Log-Rank (Mantel-Cox) test. The other data are presented as mean ± SEM and means were compared with twotailed,
unpaired Student's t-test or ANOVA following Tukey's Multiple Comparison Test when necessary. In the ORT one-sample t test was used to examine whether single columns were different from zero. Statistical significance was considered when P values were <0.05.

3. Results

3.1. Increase of life expectancy by resveratrol.

The survival curves were plotted using the Kaplan Meier plot. A shift to the right for the resveratrol groups reveals an increased expectancy of life for animals that had been

eating the resveratrol diet. The comparison of the groups using the Mantel-Cox log rank test indicates that there is a significant difference between the survival curves of the control group vs. the resveratrol group not only in SAMP8 mice (Fig. 1A, $P<0.0001$ among groups Mantel-Cox log-rank test) but also in SAMR1 animals (Fig. 1B, $P<0.01$ among groups Mantel-Cox log-rank test). In addition, the median life expectancy of our control mice was of 10.4 months for SAMP8 mice, significantly lower than the 17,8 months of SAMR1 mice (Fig. 1C), in accordance to previous studies (Takeda 2009). However the SAMP8 resveratrol group showed a life expectancy of approximately 14 months, showing an increase in life expectancy of over a 33 % over the control SAMP8 mice (Fig. 1C). Furthermore, SAMR1 mice fed with resveratrol also presented a median lifespan of 21,8 months showing an increase of a 22 % in comparison with control SAMR1 mice (Fig. 1C).

3.2. Resveratrol decreases cognitive impairment in SAMP8.

We investigated the effect of a 7-months resveratrol food supplementation in 9-monthold
SAMP8 mice. This is an age when several alterations such as amyloid deposition or cognitive impairment have been reported (Pallàs et al., 2008). We found that, in the ORT, control mice had an impaired memory as their DI was close and not different from zero (fig. 2, $P=0.4665$, one-sample t-test) revealing there was no preference for the novel object. On the other hand, resveratrol mice had a positive DI different from zero (fig. 2, $P<0.05$, one-sample t-test) revealing that their memory was not impaired as they showed higher preference for the novel object than the already presented one. Furthermore, comparison of the two groups revealed a protective effect of resveratrol in the memory of animals in comparison with age matched SAMP8 mice (Fig. 2).

3.3. Resveratrol increases both SIRT1 and AMPK levels while decreases P53 acetylation.

Western blot analysis of cortex and hippocampus of the two groups revealed higher levels of SIRT1 (Fig. 3A, B) in the animals that had been eating a diet supplemented with resveratrol in comparison with animals with standard food (control group). In accordance with this observation, the substrate of SIRT1, p53 shows a decrease in its acetylation in these brain areas (Fig. 3C, D). In addition, higher levels of phosphorylated AMPK (p-AMPK) were found in the cortex of the resveratrol group (fig. 3E) while no modifications were seen in the AMPK levels (Fig. 3G). However, while no increment of p-AMPK levels was found in the hippocampus of the resveratrol mice (Fig. 3F) an increase in AMPK basal levels appeared in these animals in comparison with control SAMP8 mice (Fig. 3H).

3.4. Resveratrol reduces amyloid deposition and favors the non-amyloidogenic pathway in the hippocampus of SAMP8 mice.

Immunohystochemistry was performed to brain sections with specific antibodies directed against the $A\beta_{42}$ and $A\beta_{40}$ to asses if there were differences between the two groups. Visual analysis revealed amyloid clusters limited only to the hippocampal area as described before (del Valle et al., 2010). In Figure 4 it can be seen that almost no Aβ granules were present in the resveratrol group while several clusters of $A\beta_{42}$ and $A\beta_{40}$ granules appeared in the control group (Fig. 4). Furthermore, we quantified the amount of amyloid clusters that were present in the hippocampus of the two groups. We found that resveratrol decreased the amount of both $A\beta_{42}$ and $A\beta_{40}$ accumulations in SAMP8 animals in comparison with control SAMP8 mice (Fig. 4B, C). In addition, western blot analysis was performed so as to quantify the levels of two enzymes responsible the amyloidogenic/non-amyloidogenic processing of APP, the α- (ADAM10) and β- (BACE) secretases. We found that while no alterations were seen in the pro-amyloidogenic BACE enzyme (Fig. 5 A-B), an increase in the nonamyloidogenic

ADAM-10 enzyme was found both in the cortex (fig. 5C) and the hippocampus (fig. 5D) of the resveratrol group.

3.5. Resveratrol lowers Tau hyperphosphorylation at serine 396 and has a differential effect on Kinases of the cortex and the hippocampus.

The levels of phosphorylated tau (pTau) at Ser_{396} have been described as a reliable marker of the severity of AD (Hu et al., 2002). Thus, we evaluated the effect of resveratrol on Tau phosphorylation levels in cortex and hippocampus extracts by Western blot using a tau antibody that detects only the pTau at Ser_{396}. As can be seen in the figure 6, not only the cortex but also the hippocampus of animals fed with resveratrol showed lower levels of pTau (Fig. 6 A-B). In addition, we investigated the levels of CDK5 and the ratio of its activator p25 with the precursor p35 as well as the phosphorylated levels of GSK3β, CDC2 and JNK. With regard to the cortex of resveratrol animals, a diminution in CDK5 protein levels (Fig. 6C) altogether with a decrease in the p25/p35 ratio (Fig. 6E), revealing inactivation of this kinase, In addition, an increase in the levels of phosphorylated GSK3β at Ser_{9} can also be seen (Fig. 7A), a fact that also correlates with the reduced pTau levels as this enzyme is deactivated when phosphorylated at this residue. However, no modifications can be detected on the levels of phosphorylated CDC2 (Fig. 7C) nor in the levels of phosphorylated JNK (Fig. 7E). Conversely, there are no changes in resveratrol-treated-SAMP8 hippocampus in comparison with age-matched SAMP8 control mice in studied kinases (Fig. 6 D, F and 7 B, D, F), but it was observed a significant diminution in pTau at Ser_{396} (Fig. 6B).

4. Discussion

The results stated here have confirmed the positive effect of resveratrol on extending lifespan, memory and neurodegenerative markers in the SAMP8 mice.
SIRT1 activation by resveratrol increases the lifespan of *S. Cerevisiae* (Howitz et al., 2003), *C. Elegans* (Viswanathan et al., 2005), *D. Melanogaster* (Wood et al., 2004) and the short-lived seasonal fish *Nothobranchius furzeri* (Valenzano et al., 2006). Here we demonstrate for the first time that resveratrol can extend lifespan in mice. Resveratrol supplementation in the diet resulted in a significant increase in mean life expectancy and in maximal life span, both in SAMP8 and SAMR1. At present, resveratrol was reported to prevent early mortality in mice fed with a high-fat diet (Baur et al., 2006) but failed to provide significant effects on survival in old mice (Miller et al., 2011). In SAMP8 mice only a growth hormone releasing hormone antagonist has been shown to extend their median lifespan (Banks 2010) and this action was associated with decreased brain oxidative stress. Melatonin has also been reported to increase life span and longevity in SAMR1 and SAMP8 mice (Rodriguez et al., 2008). These authors conclude that the underlying effects of this indoleamine relay on mitochondrial physiology improvement, then involving a decrease of reactive oxygen species generation. As old rodents produce more reactive oxygen species than young ones and the rate of mitochondrial reactive oxygen species production is inversely proportional to species maximum life span, it would be reasonable to expect that an agent that lowered reactive oxygen species might extend lifespan (Sohal et al, 1989). Sirtuins are deacetylases that show anti-aging properties in several animal models and can protect from stress (Donmez et al., 2010). SIRT1 plays a role in regulating different cellular processes through deacetylation of important substrates such as p53, FOXO transcription factors, PGC-1α, NFκB and others, which are closely linked to some age related diseases (Saunders et al., 2010). It is well established that CR is able to extend lifespan with a result of SIRT1 activation (Cohen et al., 2004) and it has been postulated that resveratrol mimics the effect of CR. In this study we demonstrate an increase in SIRT1 levels in SAMP8 treated with resveratrol in the two brain areas

studied, that correlated with a diminution in acetylated forms of p53, one of the main substrates of the deacetylase. In addition, SIRT1 pathways are closely related with AMPK signaling as a sensor of energy availability. AMPK is activated by phosphorylation of Thr-172 by LKB1 complex in response to increase in the AMP/ATP ratio and by calmodulin-dependent protein kinase kinase-beta (CamKKβ) in response to elevated Ca^{2+} levels, which contributes to regulating Aβ generation. It was reported that activation of deacetylase and AMPK are linked through LKB, and when SIRT1 is activated, AMPK is phosphorylated and also activated. Moreover, it has been recently demonstrated that resveratrol effects on SIRT1 activation are mediated via the CamKKβ-AMPK pathway by inhibiting cAMP-specific phosphodiesterases (PDE) (Park et al., 2012). Our results showed that resveratrol activation of SIRT1 in SAMP8 mice correlated with changes in the levels or in the phosphorylation of AMPK, demonstrating again that resveratrol is able to modify the SIRT11 pathway.

Furthermore, a link between SIRT1 activation, AMPK and AD is increasingly evident (Gan, 2007). Tau phosphorylation and β-amyloid production are sensitive to AMPK inhibition (Greco et al., 2011; Park et al., 2012) as well as SIRT1 activation can prevent several signs of neurodegeneration (Bayod et al., 2012), protects against axonal degeneration (Araki et al., 2004), reduces poly-glutamine toxicity (Parker et al., 2005) and diminishes microglia-mediated Aβ toxicity (Chen et al., 2005). Resveratrol has been reported to improve memory alterations as it preserved cognitive function in aging mice (Oomen et al., 2009) and in transgenic AD mice (Kim et al., 2007). SAMP8 mice begin to show memory impairments at 6 months of age and memory related deficits get worse at 9 months of age (del Valle et al., JAD *in press*), here we found that resveratrol was able to revert the memory impairment previously reported.

Part of the beneficial effects described for SIRT1 on Aβ accumulation is the modulation of α-secretases. Transcription of ADAM10 is positively controlled by retinoic acid receptors (RAR), which are activated by their ligand retinoic acid or through deacetylation by SIRT1. Using SIRT1-transgenic and SIRT1-deficient mice, this protein was found to activate the RARb transcription factor, which in turn increased ADAM10 expression (Lichtenthaler 2011). In addition, SIRT1 activation reduced amyloid pathology in a mouse model of AD and crossing SIRT1 knockout mice with these mice dramatically increased Aβ burden (Donmez et al., 2010). Moreover, decreased SIRT1 expression has been found in patients with AD and this decrease correlates with tau and Aβ levels (Julien et al., 2009). Modulation of ADAM10 expression by SIRT1 was demonstrated previously (Gutiérrez-Cuesta et al., 2008; Donmez et al., 2010). To this end, in our experimental paradigm, we found that resveratrol reduces Aβ burden in treated SAMP8 brain concomitantly with increases in ADAM10 expression. This effect can be considered specific because no changes were observed in the expression of other secretases, such as BACE (Donmez et al., 2010). Thus, resveratrol, through SIRT1 activation, specifically induced the non-amiloidogenic processing of nonmutated APP, reducing the presence of previously described amyloid deposits (del Valle et al., 2010).

On the other hand, tau hypephosphorylation, another hallmark of AD, is mediated by several kinases in brain. We and others have demonstrated the aberrant phosphorylation of tau in brain of SAMP8 that is accomplished by activation of several tau kinases such as CDK5, GSK3β or JNK (Canudas 2005; Chang et al., 2011). Our data shows that in cortex of SAMP8 mice a diminution in CDK5 and GSK3β activity, both main tau kinases in AD, is induced by resveratrol treatment; and the inhibition of these tau kinases prevented tau phosphorylation in Ser_{396}.

On the other hand, no evident changes in JNK were found. Conversely, with low doses and only 2 months of treatment with pterostilbene, a resveratrol derivative, an inhibition of JNK was observed in SAMP8 but no changes in tau hyperphosphorylation (measured through PHF antibody) were observed in cortex (Chang et al., 2011). All these discrepancies are probably due to the different resveratrol doses and also to the long term treatment by resveratrol that we applied in the present study.

With regard to the hippocampus, although resveratrol was able to prevent tau phosphorylation, we were unable to find changes in the studied kinases. It is plausible to hypothesize that although long-term treatment by resveratrol prevents tau hyperphosphorylation detectable by using specific phospho-antibodies, the inhibition of intermediate signals under these conditions is lost because of the chronicity of the treatment. On the other hand, oxidative stress is a well-established pathogenic factor in AD (Smith et al., 1995; Markesbery 1997; Perry et al., 1998) and the association of oxidative stress with tau abnormalities is well known. As such, the resveratrol-driven reductions on tau phosphorylation in hippocampus could be mediated by the wellknown antioxidant effects of this polyphenol rather than through its inhibitory effect on tau kinases. Therefore, our results allow us to conclude that resveratrol is able to inhibit tau phosphorylation both in cortex and hippocampus.
Taken together, in this study we demonstrate that resveratrol not only increases life expectancy, favors AMPK pathways and prosurvival routes such as SIRT1 activation but also has a neuroprotective role, reducing cognitive impairment in AD and other neurodegenerative parameters as neuronal cell death, amyloid burden and Tau hyperphosphorylation.

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6. Disclosure statement

Drs. Casadesus, Canudas, Vilaplana, Pelegri, Sanfeliu, Camins, Pallas, and del Valle have no conflict of interest or disclosures to provide. David Porquet Alberto Vicente and Sergi Bayod are students with no conflicts or disclosures to provide

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Table 1

Antibody (Clone)	Catalog reference	Dilution (1:)	Provider
Acetyl-P53 (acetyl-K382)	ab37318	500	Abcam, Cambridge, UK
ADAM-10	ab39177	1000	Abcam, Cambridge, UK
Beclin-1	ab16998	1000	Abcam, Cambridge, UK
Cdc2 p34 (17)	sc-54	1000	Santa Cruz, Santa Cruz, CA, USA
Cdk5 (C-8)	sc-173	1000	Santa Cruz, Santa Cruz, CA, USA
GSK-3β (27C10)	#9315	1000	Cell Signaling, Danvers, MA, USA
LC3B	#2775	1000	Cell Signaling, Danvers, MA, USA
p35/p25 (C64B10)	#2680	1000	Cell Signaling, Danvers, MA, USA
p53 (1C12)	#2524	1000	Cell Signaling, Danvers, MA, USA
Phospho-cdc2 (Tyr15)	#9111	1000	Cell Signaling, Danvers, MA, USA
Phospho-GSK-3β (Ser9)	#9336	1000	Cell Signaling, Danvers, MA, USA
Phospho-SAPK/JNK (Thr183/Tyr185)	#9251	1000	Cell Signaling, Danvers, MA, USA
Phospho-Tau (pS396)	44752G	1000	Invitrogen, Carlsbad, CA, USA
SAPK/JNK	#9252	1000	Cell Signaling, Danvers, MA, USA
SIRT1 (SIR11)	ab50517	1000	Abcam, Cambridge, UK
Tau (Tau-5)	AHB0042	1000	Biosource, Camarillo, CA, USA
β-Actin (AC-15)	A5441	20000	Sigma-Aldrich, St. Louis, MO, USA
Aβ40	ab10147	50	Abcam, Cambridge, UK
Aβ42 (12F4)	SIG-39142	100	Covance, CA, USA
Alexa Fluor 488 donkey anti-mouse IgG	A-11001	250	Invitrogen, Carlsbad, CA, USA
Alexa Fluor 546 donkey anti-rabbit IgG	A-11035	250	Invitrogen, Carlsbad, CA, USA
Donkey ECL anti-Rabbit IgG, HRP linked	NA934V	1000	GE Healthcare, UK
Goat Anti-Mouse HRP Conjugate	#170-5047	1000	Biorad, Hercules, CA, USA

FIGURES

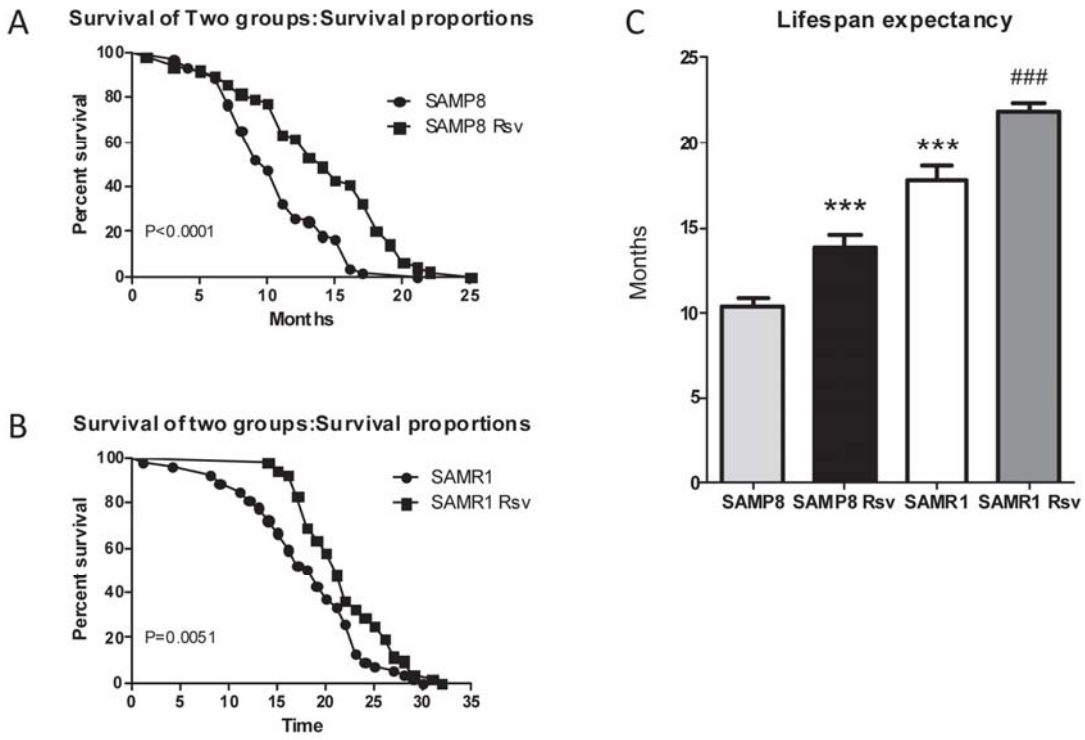

Figure 1: Kaplan Meier plot with data expressed as % of alive individuals (A, B) and median lifespan of the four groups studied (C). Mantel-Cox log rank test analysis reveals a shift to the right for the resveratrol group in SAMP8 (A, P<0.0001) and SAMR1 (B, P=0.0051). In the median life-span comparison (C), results are expressed as mean ± SEM; ANOVA followed by Tukey's post test, ***P<0.001 vs. SAMP8, ###P<0.001 vs. SAMR1.

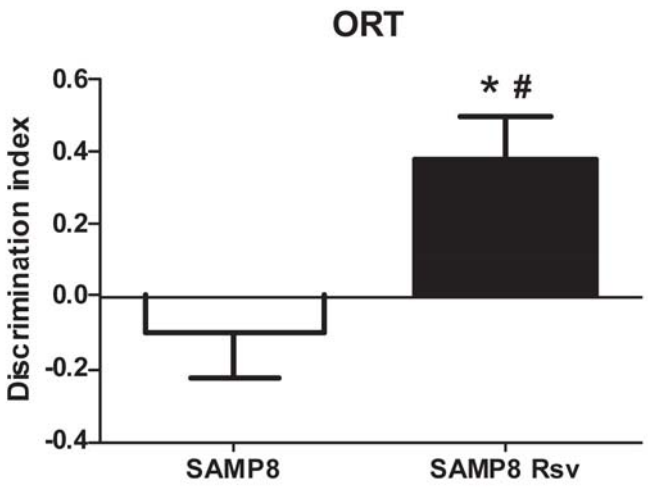

Figure 2: Discrimination index (DI) of both groups of SAMP8 animals. Only Rsv group values are positive and different from zero (* P<0.05). There is a higher DI of Rsv animals in comparison with control SAMP8 mice (# P<0.05 vs. SAMP8 mice). Bars represent mean ± SEM.

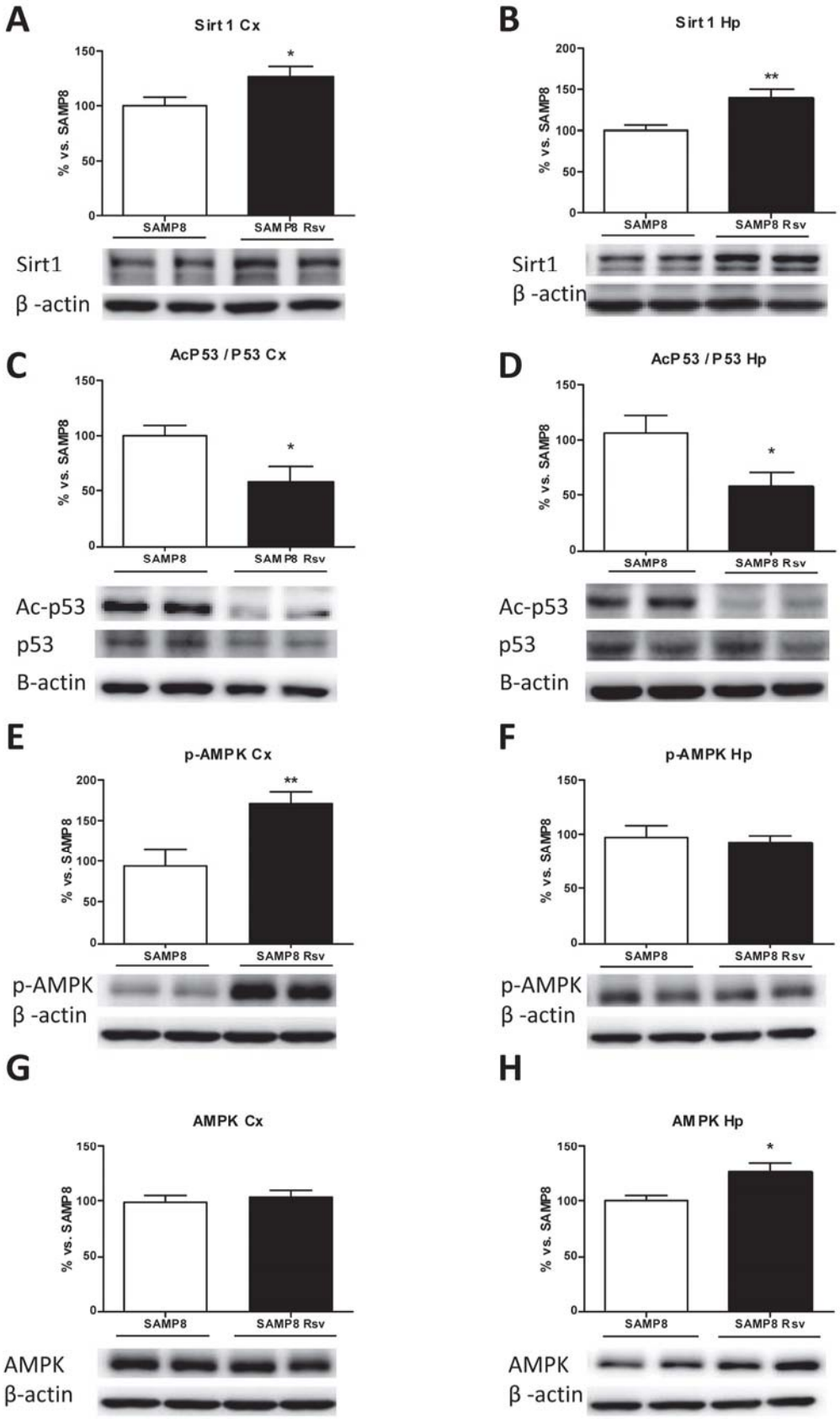

Figure 3: Levels of Sirtuin (A, B), its acetylated substrate p53 (C, D), p-AMPK (E,F) and AMPK (G, H). Bars represent mean ± SEM and values are adjusted to 100% for levels of control SAMP8 mice. Student's paired t-test; *p<0.05; **p<0.01 vs. SAMP8. Cx: Cortex, Hp: Hippocampus.

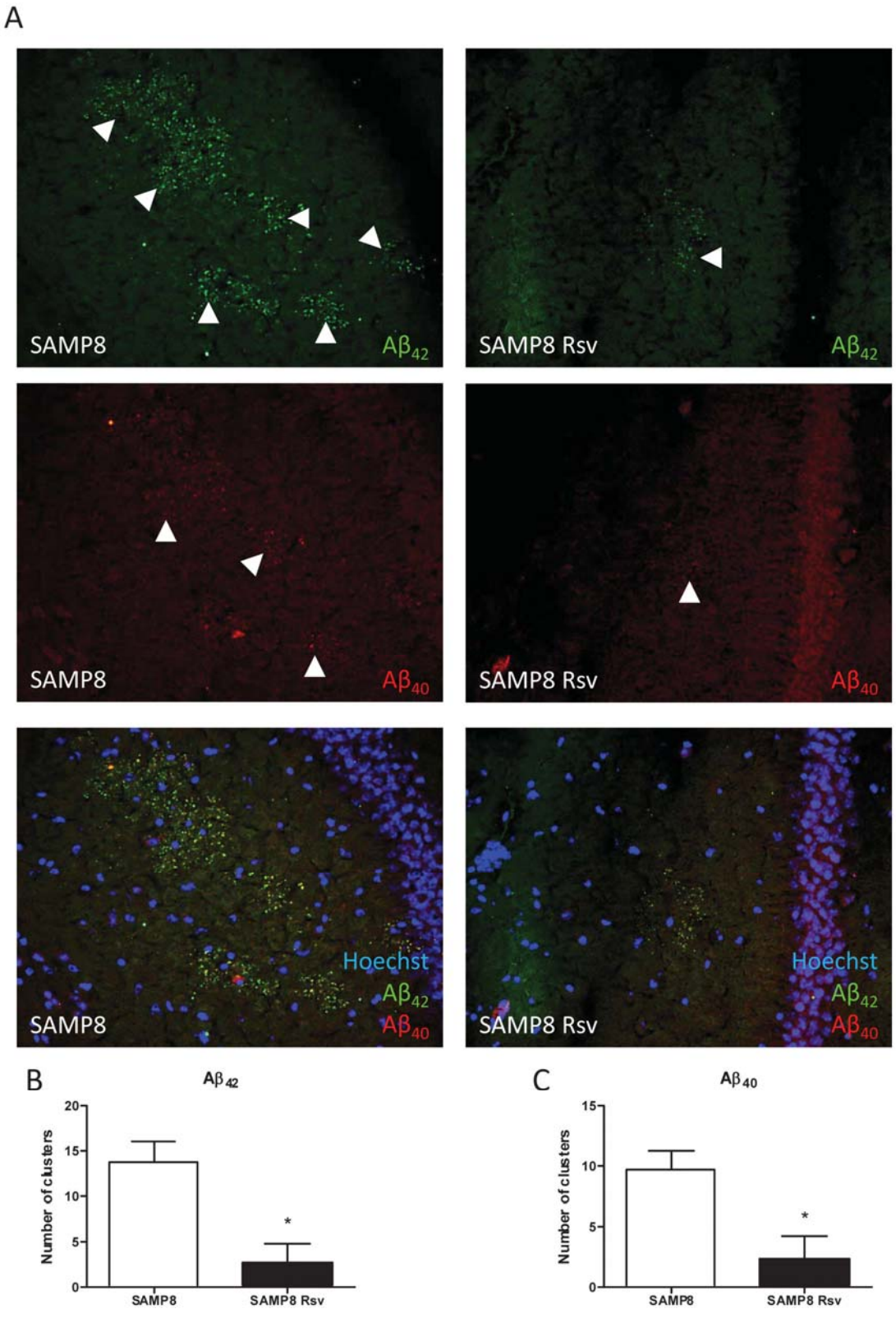

Figure 4: Representative hippocampal images of SAMP8 and SAMP8 Rsv animals (A), arrows (Aβ42) and arrowheads (Aβ40) indicate some clusters of amyloid granules in both groups. Quantification of the amount of Aβ42 (B) and Aβ40 (C) clusters in the hippocampus of the two groups. Bars represent mean ± SEM, values in D-G are adjusted to 100% for levels of control SAMP8 mice. Student's paired t-test; *p<0.05; vs. SAMP8. Cx: Cortex, Hp: Hippocampus.

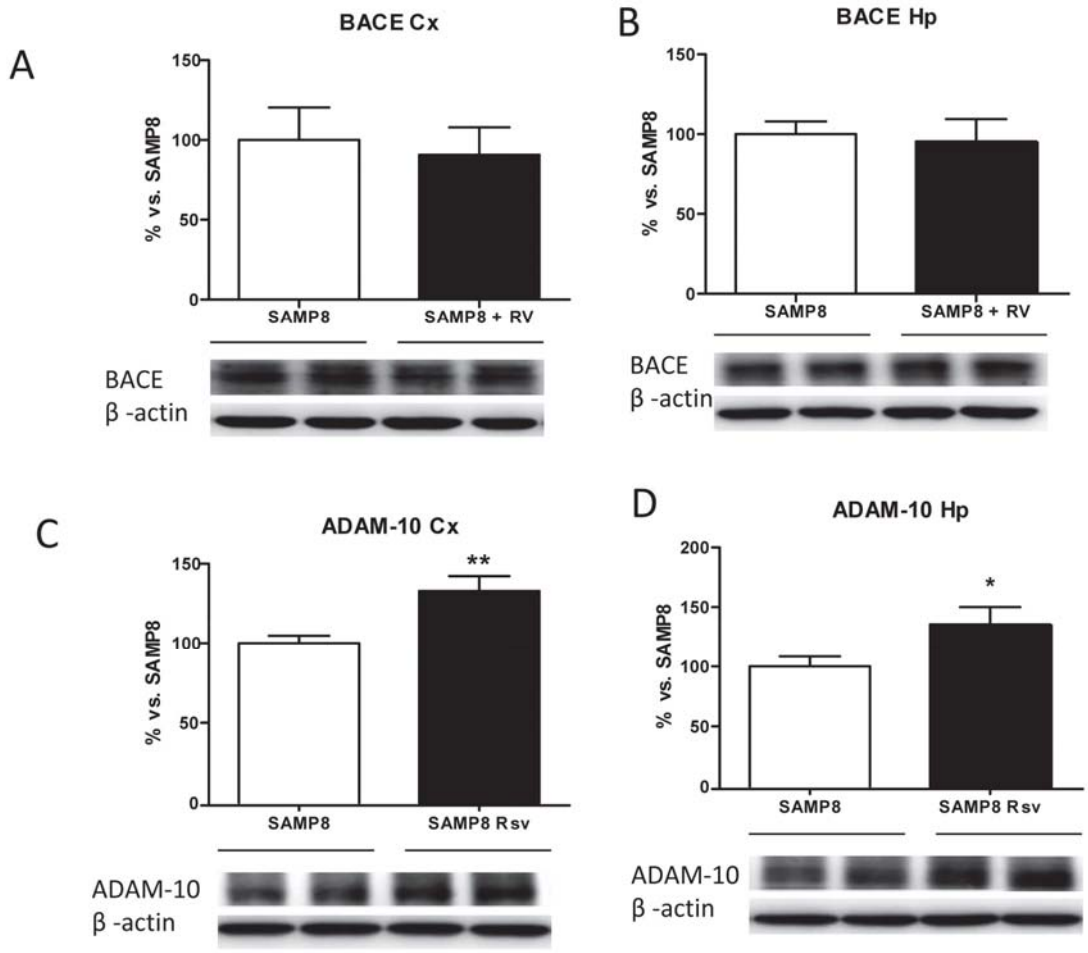

Figure 5: Cortex and hippocampal levels of BACE (A, B) and ADAM-10 (C, D) of SAMP8 and SAMP8 Rsv animals. Bars represent mean ± SEM, values in A-D are adjusted to 100% for levels of control SAMP8 mice. Student's paired t-test; *p<0.05; **p<0.01 vs. SAMP8. Cx: Cortex, Hp: Hippocampus.

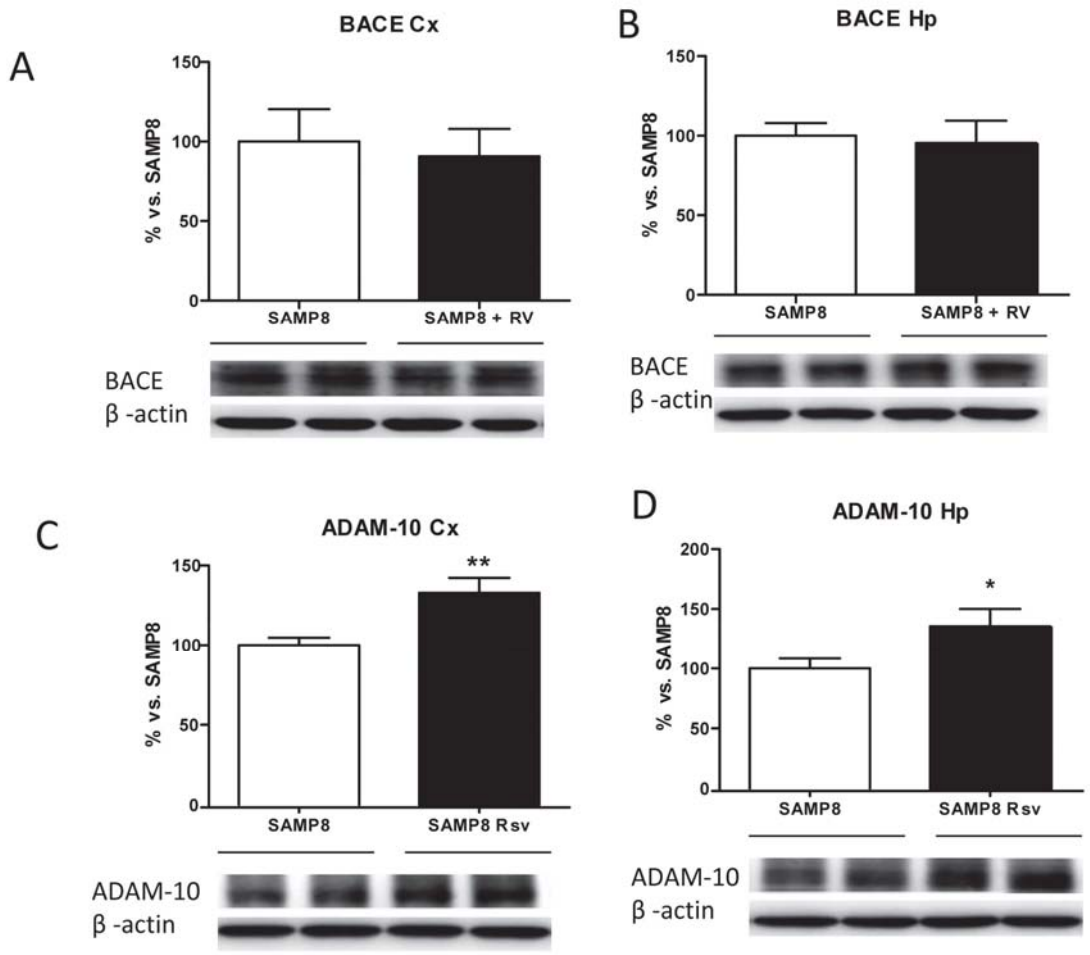

Figure 6: Levels of phosphorylated tau (pTau) at Ser_{396} in cortex (A) and hippocampus (B) of SAMP8 and SAMP8 Rsv groups. Cortex and hippocampal levels of CDK5 (C, D), P25/P35 ratio (E, F). Bars represent mean ± SEM and values are adjusted to 100% for levels of control SAMP8 mice. Student's paired t-test; *p<0.05; **p<0.01 vs. SAMP8. Cx: Cortex, Hp: Hippocampus.

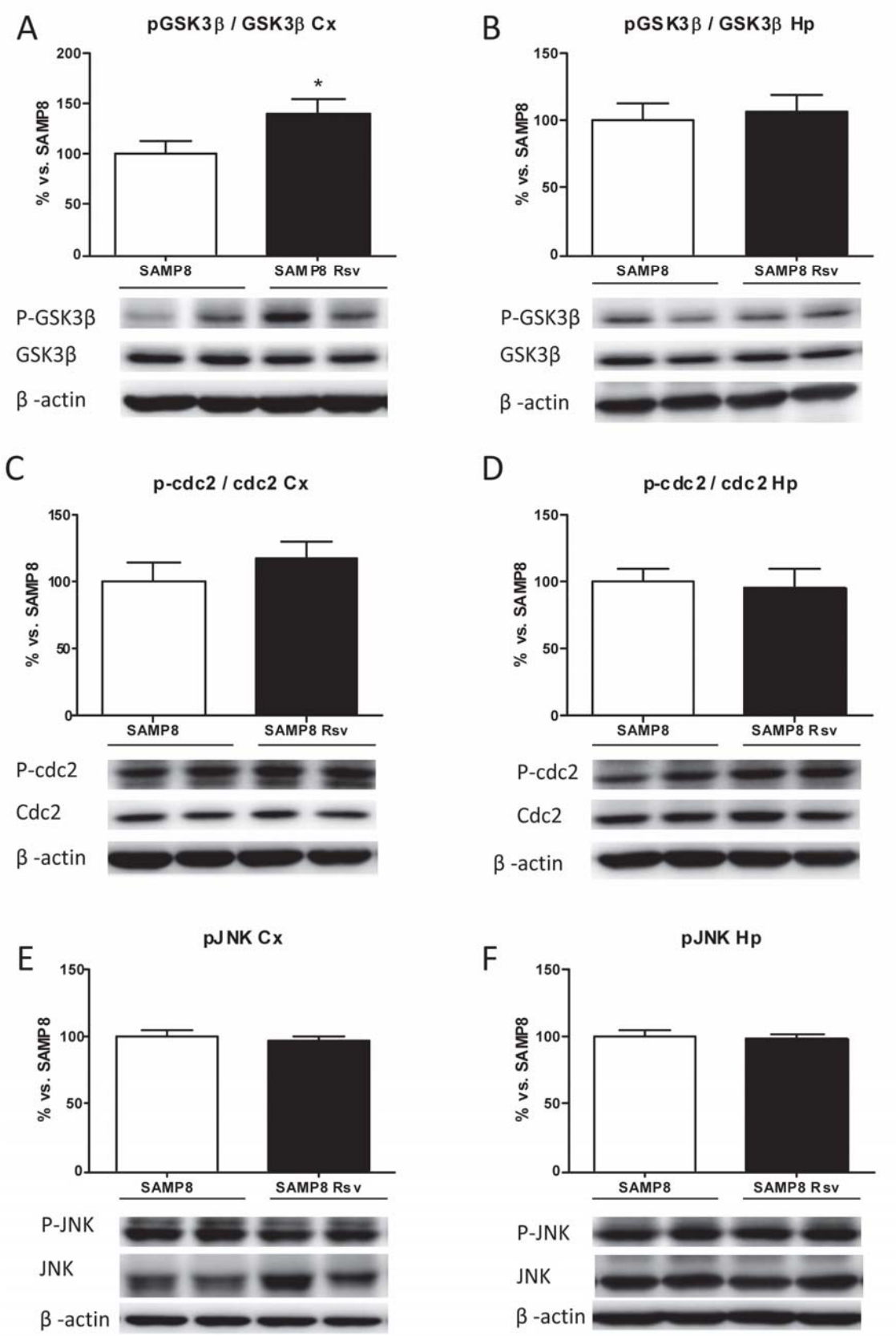

Figure 7: Cortex and hippocampal levels of p-GSK3ß (phosphorylated in Ser_9) (A, B). p-cdc2 (phosphorylated in Tyr_{15}) (C, D) and JNK (phosphorylated in Thr_{183}/Tyr_{185}) (E, F). Bars represent mean ± SEM and values are adjusted to 100% for levels of control SAMP8 mice. Student's paired t-test; *p<0.05 vs. SAMP8. Cx: Cortex, Hp: Hippocampus.