

Active behaviours produced by antidepressants and opioids in the mouse tail suspension test

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Abstract

Most classical preclinical tests to predict antidepressant activity were initially developed to detect compounds that influenced noradrenergic and/or serotonergic activity, in accordance with the monoaminergic hypothesis of depression. However, central opioid systems are also known to influence the pathophysiology of depression. While the tail suspension test (TST) is very sensitive to several types of antidepressant, the traditional form of scoring the TST does not distinguish between different modes of action. The present study was designed to compare the behavioural effects of classical noradrenergic and/or serotonergic antidepressants in the TST with those of opioids. We developed a sampling technique to differentiate between behaviours in the TST, namely, curling, swinging and immobility. Antidepressants that inhibit noradrenaline and/or serotonin re-uptake (imipramine, venlafaxine, duloxetine, desipramine and citalopram) decreased the immobility of mice, increasing their swinging but with no effect on their curling behaviour. No differences were observed between antidepressants that act on noradrenergic or serotonergic transmission. While opioid compounds also decreased the immobility of the mice [morphine, codeine, levorphanol, (–)-methadone, (±)-tramadol and (+)-tramadol], they selectively increased curling behaviour. Blocking opioid receptors with naloxone prevented the antidepressant-like effect of codeine, and μ -opioid receptor knockout decreased normal curling behaviour and blocked (±)-tramadol-induced curling, further demonstrating the reliability and validity of this approach. These results show that at least two behaviourally distinct processes occur in the TST, highlighting the antidepressant-like effects of opioids evident in this test. Furthermore, our data suggest that swinging and curling behaviours are mediated by enhanced monoamine and opioid neurotransmission, respectively.

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Introduction

There is substantial evidence implicating the opioid system in depression (Hegadoren *et al.* 2009), suggesting that compounds that enhance opioid

neurotransmission may exert genuine antidepressant effects (Berrocoso & Mico, 2009a; Jutkiewicz, 2006; Tejedor-Real *et al.* 1998). Since the description of the ‘opium cure’ (Kraepelin, 1901), clinical reports have described the effectiveness of μ -opioid receptor (MOR) agonists in patients suffering depression, such as oxycodone, oxymorphone, tramadol and buprenorphine, especially in cases of refractory depression (Bodkin *et al.* 1995; Fanelli & Montgomery, 1998; Shapira *et al.* 2001; Spencer, 2000; Stoll & Rueter, 1999). We have studied the antidepressant-like effects of opioids and,

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in accordance with other studies (reviewed in (Berrocoso *et al.* 2009), we demonstrate the antidepressant-like activity of several opioids in animal paradigms of depression (Rojas-Corrales *et al.* 1998, 2004; Tejedor-Real *et al.* 1998). Indeed, we previously described a functional interaction between opioids and selective serotonin reuptake inhibitors (SSRIs) or noradrenaline (NA) reuptake inhibitors in the tail suspension test (TST: Berrocoso & Mico, 2009a), whereby both opioids and the more classic antidepressants decrease the time that the mice spend immobile (consistent with an antidepressant-like effect).

The mouse TST is a predictive behavioural test of antidepressant activity (Cherem *et al.* 1986; Steru *et al.* 1985, 1987) that is being increasingly used with the advent of transgenic mice. When mice are suspended by the tail they are subjected to short-term inescapable stress and they adopt an immobile posture. However, if antidepressant treatments are administered prior to the test, the mice will actively pursue escape-directed behaviours over longer periods of time. The increase in such activity (i.e. the decrease in immobility) in the TST is strongly correlated with antidepressant effects in humans (Cryan *et al.* 2005a). This clinical predictive value, together with the high degree of reliability in different laboratories, has led to the inclusion of the TST in almost all batteries used to screen new antidepressant drugs (Bravo *et al.* 2009), even though the TST in its traditional form cannot reliably detect specific modes of drug action. In our laboratory, previous studies with the TST suggested that while opioids decrease the time mice spend immobile they induce a pattern of activity that differs from that seen with classical monoaminergic antidepressants. To further investigate this effect, we have systematically analysed the behaviour of mice in the TST to compare the active behaviours induced by classical antidepressants and opioids.

Materials and method

Animals

Experiments were performed using male albino CD1 mice, male and female wild-type mice or heterozygous and homozygous MOR knockout (MOR-KO) littermates obtained by crossing heterozygous/heterozygous MOR-KO mice on a C57BL/6J genetic background (Sora *et al.* 2001). Animals were maintained under standard conditions: 12-h light/dark cycle (lights on 08:00 hours), *ad libitum* access to food and water and a constant temperature ($21 \pm 1^\circ\text{C}$).

Animals were housed in groups of 10 in standard polypropylene cages (1000 cm²) and male and female mice shared the same room. All animal handling and procedures were performed in accordance with the European Communities directive 86/609-EEC and Spanish Law (RD 1201/2005) regulating animal research. The experimental protocols were approved by the Committee for Animal Experimentation of the University of Cádiz. All mice were experimentally naive, they weighed 25–30 g at the time of testing and they were only used once.

Drugs

The following drugs were used in this study: imipramine, desipramine, codeine and naloxone (Sigma-Aldrich-Química, UK); venlafaxine (Wyeth, USA); duloxetine (Eli Lilly, USA); citalopram, (\pm)-tramadol and (+)-tramadol (Grünenthal, Germany and Spain); morphine (Agencia Española de Medicamentos y Productos Sanitarios, Spain); (–)-methadone and levorphanol (RBI, USA). The selectivity of these compounds for opioid receptors (μ , δ and κ) and monoamine transporters (5-HT and NA) is summarized in Table 1.

All the drug solutions were prepared immediately before each trial and they were injected *i.p.*, with the exception of naloxone, which was administered subcutaneously. All drugs were dissolved in physiological saline (NaCl 0.9%), with the exception of duloxetine, which was dissolved in distilled water, and the control animals received saline alone (NaCl 0.9%). All the solutions were injected in a volume of 10 ml/kg body weight 30 min prior to testing and the treatments were administered under blind conditions.

Tail suspension test

We used a modified form of the TST that was previously validated for NMRI mice (Steru *et al.* 1985). Accordingly, 30 min after injection the mice were individually suspended by the tail from an aluminium hook raised 20 cm above the floor using adhesive tape placed 2 cm from the tip of tail. The mice were positioned such that the base of their tail was aligned with the horizontal plane. Typically, mice demonstrated escape-oriented behaviour interspersed with successively longer bouts of immobility. Test sessions lasted for 6 min and they were videotaped and subsequently scored by a trained observer.

Behavioural scoring

The procedure used to analyse the test sessions was similar to that described previously for the forced

Table 1. Summary of the inhibitory effects of antidepressants and opioids on monoamine uptake and opioid receptor binding

Drugs	μ	δ	κ	NA	5-HT	Reference
Antidepressants						
Imipramine	3700	12 700	1800	6.6	21	Raffa <i>et al.</i> (1992)
Venlafaxine	–	–	–	1260	74	Beique <i>et al.</i> (1998)
Duloxetine	–	–	–	3	1.8	Beique <i>et al.</i> (1998)
Desipramine	–	–	–	0.31	129	Owens <i>et al.</i> (1997)
Citalopram	–	–	–	3042	0.75	Owens <i>et al.</i> (1997)
Opioids						
With no reuptake-inhibiting activity						
Morphine	0.34	92	570	IA	IA	Raffa <i>et al.</i> (1992)
Codeine	160	5130	5970	IA	IA	Raffa <i>et al.</i> (1992)
With reuptake-inhibiting activity						
Levorphanol	0.42	3.61	4.2	1220	86.3	Codd <i>et al.</i> (1995)
(–)-Methadone	0.945	371	1860	702	14.1	Codd <i>et al.</i> (1995)
(±)-Tramadol	2100	57 600	42 700	709	990	Raffa <i>et al.</i> (1992)
(+)-Tramadol	1300	62 400	54 000	2510	530	Raffa <i>et al.</i> (1993)

NA, Noradrenaline; IA, inactive at 10 μ M.

The assays were performed on rat brain samples and the data represent K_i values \pm S.E.M. (nM).

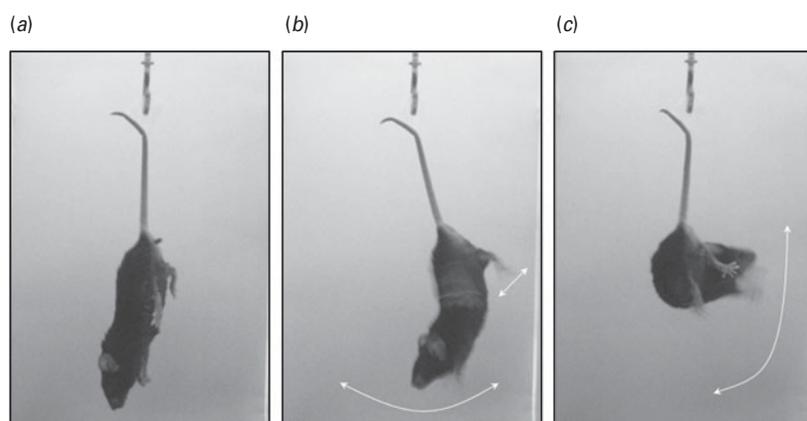


Fig. 1. Photographs illustrating the behaviours scored in the tail suspension test. (a) Immobility – the mouse hangs without engaging in any activity; (b) swinging – keeping its body straight, the mouse continuously moves its paws in a vertical position and/or moves its body from side to side; (c) curling – the mouse engages in active twisting movements.

swimming test (FST; Detke *et al.* 1995). A time-sampling technique was employed, whereby the predominant behaviour in each 5-s period of the 360 s test was recorded. The behaviours rated were: (1) immobility – a mouse was judged to be immobile when it hung by its tail without engaging in any active behaviour; (2) swinging – a mouse was judged to be swinging when it continuously moved its paws in the vertical position while keeping its body straight and/or it moved its body from side to side; (3) curling – a mouse was judged to be curling when it engaged in active twisting movements of the entire body (Fig. 1). The behavioural scoring was performed by a single

experienced observer who was blind to the treatments. Several test sessions ($n = 30$ subjects) were then scored a second time by the observer to determine the test–retest reliability (i.e. two consecutive ratings by the same observer were compared). These sessions were then scored by a second blind observer to determine the inter-rater reliability.

Statistical analysis

The data were expressed as the mean \pm S.E.M. of the parameter measured and they were analysed by one-way analysis of variance (ANOVA) followed by the

Table 2. Effects of antidepressants and opioids on immobility and active behaviours in the tail suspension test

Drugs	Immobility	Swinging	Curling
Antidepressants			
(a) Imipramine	$F_{4,44}=3.34, p<0.05$	$F_{4,44}=3.55, p<0.05$	$F_{4,44}=0.69, \text{n.s.}$
(b) Venlafaxine	$F_{4,42}=7.82, p<0.001$	$F_{4,42}=3.30, p<0.05$	$F_{4,42}=0.13, \text{n.s.}$
(c) Duloxetine	$F_{6,63}=3.77, p<0.01$	$F_{6,63}=2.33, p<0.05$	$F_{6,63}=0.71, \text{n.s.}$
(d) Desipramine	$F_{3,32}=6.43, p<0.01$	$F_{3,32}=4.13, p<0.05$	$F_{3,32}=2.79, \text{n.s.}$
(e) Citalopram	$F_{5,51}=4.25, p<0.01$	$F_{5,51}=2.70, p<0.05$	$F_{5,51}=1.64, \text{n.s.}$
Opioids			
(f) Morphine	$F_{3,36}=5.36, p<0.01$	$F_{3,36}=12.02, p<0.001$	$F_{3,36}=26.78, p<0.001$
(g) Codeine	$F_{3,35}=3.13, p<0.05$	$F_{3,35}=3.70, p<0.05$	$F_{3,35}=9.35, p<0.001$
(h) Levorphanol	$F_{5,51}=3.14, p<0.05$	$F_{5,51}=7.15, p<0.001$	$F_{5,51}=13.52, p<0.001$
(i) (-)-Methadone	$F_{3,36}=3.44, p<0.05$	$F_{3,36}=3.61, p<0.05$	$F_{3,36}=8.61, p<0.001$
(j) (\pm)-Tramadol	$F_{3,36}=5.01, p<0.01$	$F_{3,36}=0.72, \text{n.s.}$	$F_{3,36}=5.85, p<0.01$
(k) (+)-Tramadol	$F_{3,33}=6.60, p<0.01$	$F_{3,33}=4.01, p<0.05$	$F_{3,33}=12.38, p<0.001$
Pharmacological opioid receptor blockade			
(l) Codeine	$F_{1,54}=7.93, p<0.01$	$F_{1,54}=2.19, \text{n.s.}$	$F_{1,54}=3.20, \text{n.s.}$
(m) Naloxone	$F_{2,54}=0.70, \text{n.s.}$	$F_{2,54}=12.23, p<0.001$	$F_{2,54}=16.26, p<0.001$
(n) Codeine \times naloxone	$F_{2,54}=3.25, p<0.05$	$F_{2,54}=4.09, p<0.05$	$F_{2,54}=6.11, p<0.01$
MOR knockout			
(o) Gender	$F_{1,57}=0.32, \text{n.s.}$	$F_{1,57}=1.23, \text{n.s.}$	$F_{1,57}=0.02, \text{n.s.}$
(p) Genotype	$F_{2,57}=0.28, \text{n.s.}$	$F_{2,57}=1.23, \text{n.s.}$	$F_{2,57}=3.50, p<0.05$
(q) Gender \times genotype	$F_{2,57}=0.73, \text{n.s.}$	$F_{2,57}=0.86, \text{n.s.}$	$F_{2,57}=0.12, \text{n.s.}$
(r) Genotype	$F_{2,60}=0.14, \text{n.s.}$	$F_{2,60}=1.50, \text{n.s.}$	$F_{2,60}=3.75, p<0.05$
(s) (\pm)-Tramadol	$F_{2,180}=45.44, p<0.001$	$F_{2,180}=8.78, p<0.001$	$F_{2,180}=21.31, p<0.001$
(t) Genotype	$F_{2,180}=0.54, \text{n.s.}$	$F_{2,180}=4.93, p<0.01$	$F_{2,180}=14.18, p<0.001$
(u) (\pm)-tramadol \times genotype	$F_{4,180}=0.25, \text{n.s.}$	$F_{4,180}=1.19, \text{n.s.}$	$F_{4,180}=2.48, p<0.05$
(v) (\pm)-tramadol-WT	$F_{2,50}=17.95, p<0.001$	$F_{2,50}=2.84, \text{n.s.}$	$F_{2,50}=6.32, p<0.01$
(w) (\pm)-tramadol-HET	$F_{2,68}=24.07, p<0.001$	$F_{2,68}=0.61, \text{n.s.}$	$F_{2,68}=16.70, p<0.001$
(x) (\pm)-tramadol-KO	$F_{2,62}=9.77, p<0.001$	$F_{2,62}=6.12, p<0.01$	$F_{2,62}=1.59, \text{n.s.}$

MOR, μ -opioid receptor; WT, wild-type; HET, heterozygous; KO, knockout.

p values and *F* values from one- and two-way analysis of variance tests, respectively, are shown.

Dunnett's (for dose-response studies) or Tukey's test. For the mechanistic studies, the data were analysed using a two-way ANOVA followed by the Bonferroni *post-hoc* test. The factors evaluated (between subjects) were codeine/(\pm)-tramadol treatment and naloxone treatment. A Pearson's correlation test was used to determine test-retest and inter-rater reliability and $p<0.05$ was considered statistically significant.

Results

In the TST employed here, the reliability of the scoring for each of the three behaviours contemplated was very high. Moreover, the test-retest reliabilities were: $r=0.89$ for immobility; $r=0.93$ for swinging; $r=0.88$ for curling. The concordance between raters was: $r=0.92$ for immobility; $r=0.86$ for swinging; $r=0.81$ for curling ($p<0.0001$).

Antidepressants

All the antidepressants administered here induced similar behavioural changes in the TST, reducing the immobility in conjunction with an increase in swinging, exerting no effect on curling behaviour [see Table 2(a-e) for one-way ANOVA data]. For example, the tricyclic antidepressant imipramine (2.5–20.0 mg/kg) induced a dose-dependent decrease in immobility while increasing swinging behaviour at both 10 and 20 mg/kg ($p<0.05$ in all cases), without affecting curling behaviour (Fig. 2a). The 5-HT and NA reuptake inhibitors venlafaxine (2.5–20.0 mg/kg) and duloxetine (1.25–40.0 mg/kg) induced similar behavioural changes, significantly decreasing immobility and increasing swinging behaviour in a dose-dependent manner. Moreover, like imipramine, neither venlafaxine nor duloxetine altered curling

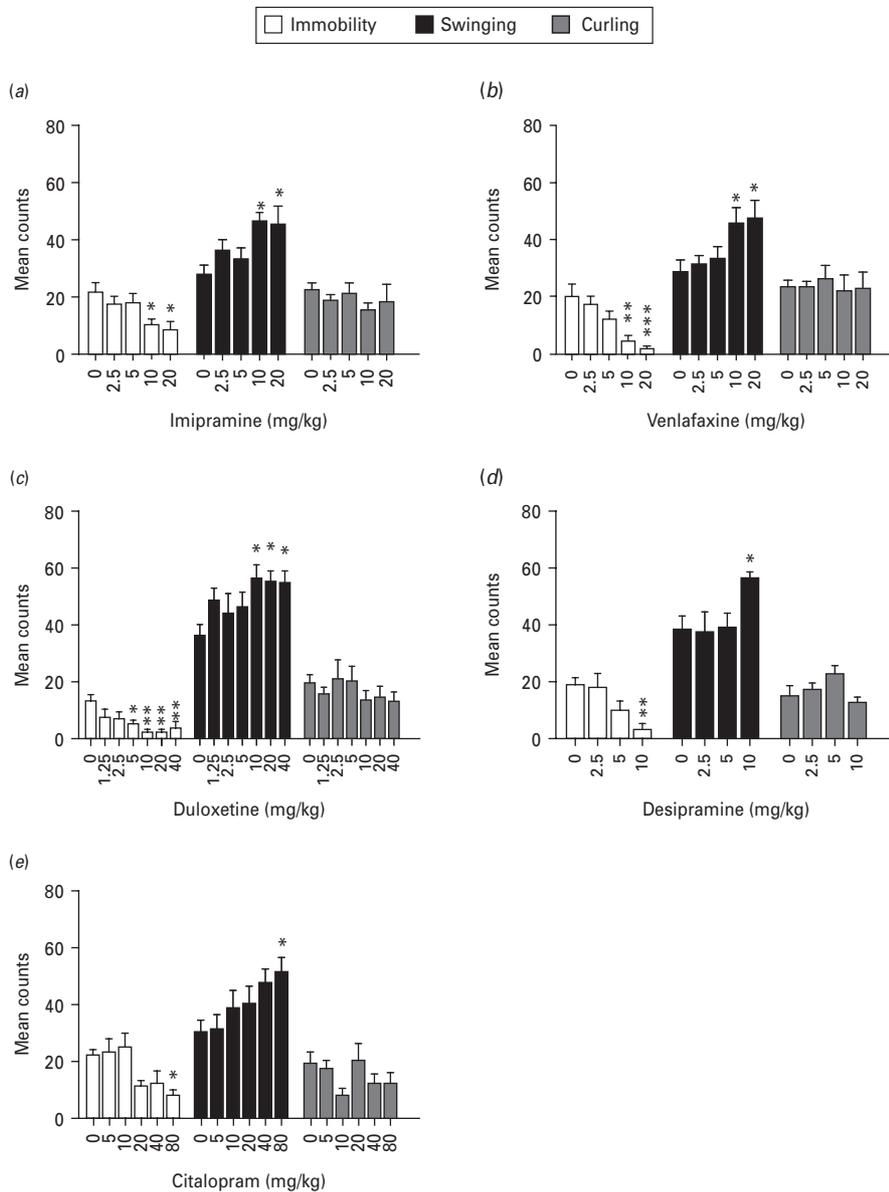


Fig. 2. Effects of (a) imipramine (2.5–20 mg/kg i.p.), (b) venlafaxine (2.5–20 mg/kg i.p.), (c) duloxetine (1.25–40 mg/kg i.p.), (d) desipramine (2.5–10 mg/kg i.p.) and (e) citalopram (5–80 mg/kg i.p.) on immobility and active behaviours in the tail suspension test. Drugs were administered 30 min before testing and the data represent the mean counts + s.e.m. from 8–11 animals per group. There were significant differences when compared to saline-treated mice (Dunnett's *post-hoc* test: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

behaviour (Fig. 2*b,c*). In a dose–response study to assess the antidepressant-like effects of the NA uptake inhibitor desipramine (2.5–10 mg/kg), 10 mg/kg desipramine reduced the time spent immobile ($p < 0.01$) while increasing swinging ($p < 0.05$), producing no effect on curling behaviour (Fig. 2*d*). Finally, the selective 5-HT uptake inhibitor citalopram (5–80 mg/kg) induced a behavioural pattern similar to that of the other antidepressants tested, decreasing

immobility at 80 mg/kg ($p < 0.05$) while increasing swinging behaviour ($p < 0.05$), with no effect on curling behaviour (Fig. 2*e*).

Opioids

All the opioids tested induced a similar behavioural pattern in the TST, decreasing immobility while increasing curling behaviour [see Table 2(*f–k*) for

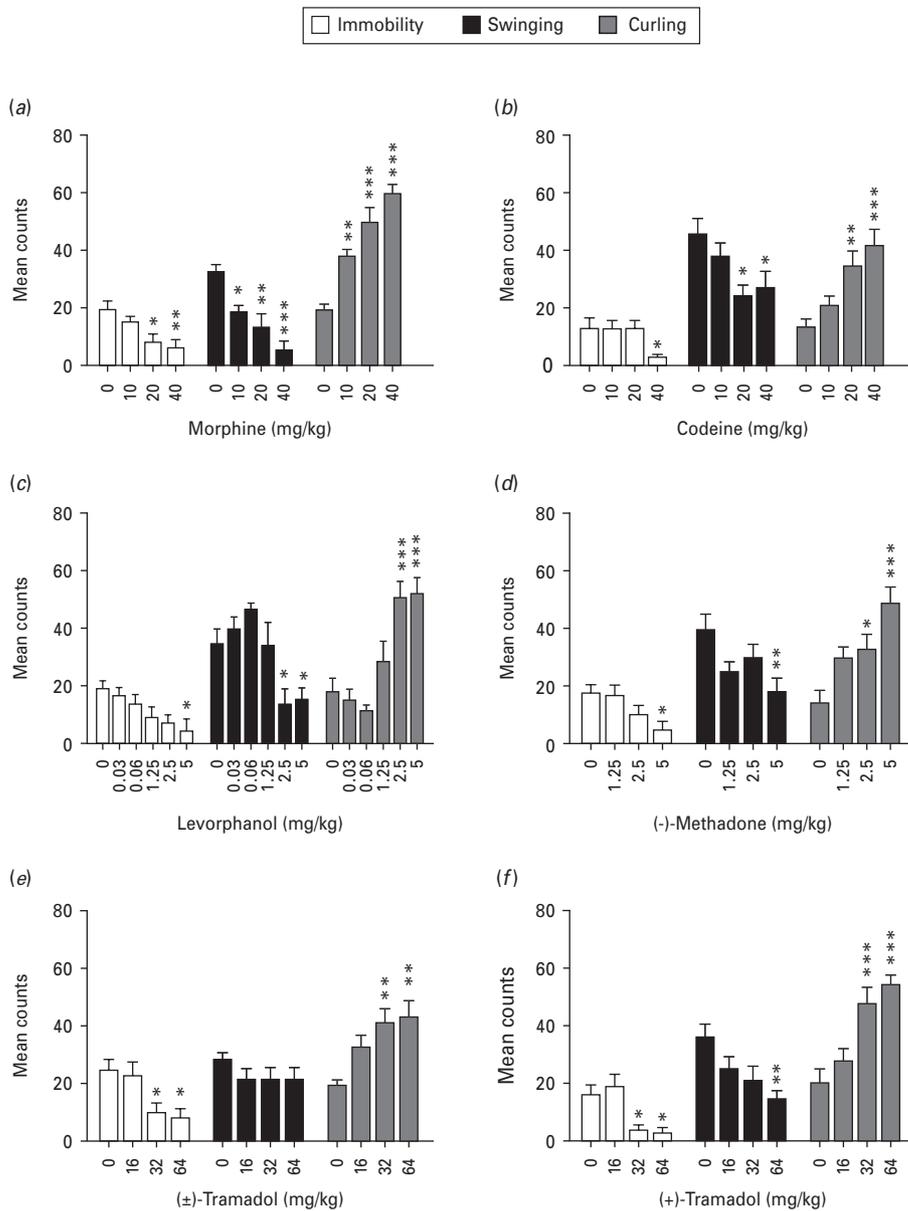


Fig. 3. Effects of (a) morphine (10–40 mg/kg i.p.), (b) codeine (10–40 mg/kg i.p.), (c) levorphanol (0.03–10 mg/kg i.p.), (d) (–)-methadone (1.25–5 mg/kg i.p.), (e) (±)-tramadol (16–64 mg/kg i.p.) and (f) (+)-tramadol (16–64 mg/kg i.p.) on immobility and active behaviour in the tail suspension test. Drugs were administered 30 min before testing and the data represent the mean counts \pm S.E.M. from 8–10 animals per group. There were significant differences when compared to saline-treated mice (one-way analysis of variance followed by Dunnett's *post-hoc* test: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

one-way ANOVA data]. Opioids that do not inhibit monoamine reuptake, such as morphine (10–40 mg/kg), attenuated immobility and swinging behaviour in a dose-dependent manner, while robustly increasing curling behaviour (Fig. 3a). Similar effects were observed for codeine (10–40 mg/kg), which as reported elsewhere (Berrocoso & Mico, 2009a), reduced immobility at 40 mg/kg ($p < 0.05$), decreased swinging ($p < 0.05$) and increased curling behaviour when

administered at 20 or 40 mg/kg ($p < 0.01$ and $p < 0.001$, respectively; Fig. 3b). Opioids with monoamine uptake-inhibiting activity such as levorphanol (0.03–10.00 mg/kg) and (–)-methadone (1.25–5.00 mg/kg) induced a behavioural pattern similar to that seen for morphine and codeine, significantly reducing immobility ($p < 0.05$ in both cases) and swinging behaviour and increasing curling behaviour (Fig. 3c,d). The behaviour elicited in response to (±)-tramadol

(16–64 mg/kg) differed slightly from that provoked by the other opioids (Fig. 3e) and, specifically, a significant reduction in immobility ($p < 0.05$) and an increase in curling behaviour ($p < 0.01$) at 32 and 64 mg/kg was not coupled with any effect on swinging behaviour. By contrast, the dextro enantiomer (+)-tramadol, which binds to the MOR and inhibits 5-HT reuptake more strongly than (\pm)-tramadol (Raffa *et al.* 1993), produced similar effects to the other opioids studied (Fig. 3f), decreasing immobility and increasing curling behaviour at 32 and 64 mg/kg and decreased swinging at 64 mg/kg ($p < 0.01$; Fig. 3f).

Role of opioid receptors in antidepressant-like effects

Pharmacological blockage

To confirm the role of the opioid system in curling behaviour, an effective dose of codeine (40 mg/kg) was co-administered with the opioid receptor antagonist, naloxone [0.5–2.0 mg/kg; Fig. 4, Table 2(*l–n*)]. Two-way ANOVA revealed a significant effect of codeine on immobility ($p < 0.01$), while swinging and curling behaviour remained unchanged. Two-way ANOVA also revealed significant effects of naloxone on swinging ($p < 0.0001$) and curling behaviours ($p < 0.001$). Furthermore, a significant interaction between factors was observed for all three behaviours: immobility ($p < 0.05$); swinging ($p < 0.05$); curling ($p < 0.01$). Indeed, a Bonferroni's analysis revealed that naloxone significantly blocked the effects of codeine on immobility, swinging and curling ($p < 0.01$ in all cases; Fig. 4).

MOR-KO study

To determine whether curling behaviour involves the activation of MORs, we evaluated the behaviour of C57BL/6J MOR-KO mice in the TST, both males and females. Initially, we investigated the effects of gender and genotype [Table 2(*o–q*)] and while a two-way ANOVA revealed no significant effect of gender and no gender \times genotype interaction, a significant effect of genotype was evident [$p < 0.05$, Table 2(*p*)]. As expected, a subsequent unpaired Student's *t* test revealed no significant difference in behaviour between male and female knockout mice (data not shown). Thus, the data from the male and female mice were pooled for the subsequent studies. While we were unable to detect any differences in the behaviour of heterozygous MOR-KO mice from their wild-type littermates, the homozygous knockout mice displayed significantly less curling behaviour compared to their wild-type littermates [$p < 0.05$, Tukey's test; Fig. 5,

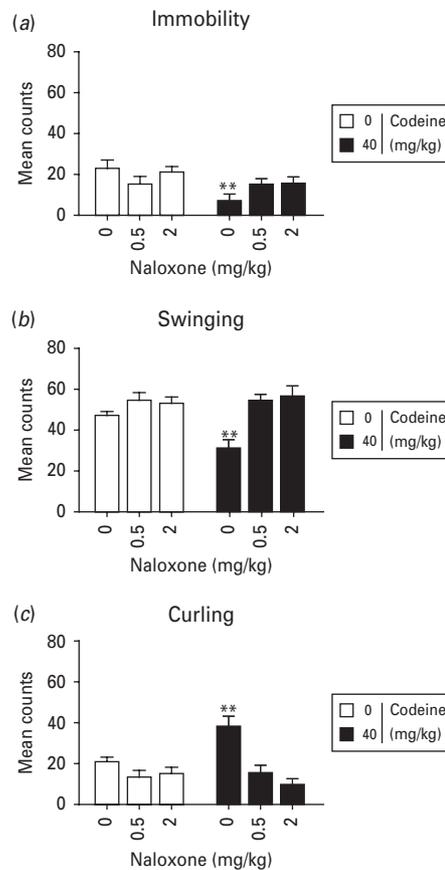


Fig. 4. Opioid receptor involvement in the effects of codeine on immobility, curling and swinging behaviour in the tail suspension test in mice. Codeine (40 mg/kg i.p.) and the opioid receptor antagonist naloxone (0.5–2 mg/kg s.c.) were administered 30 min before testing and the data represent the mean counts \pm S.E.M. from 10 animals per group. There were significant differences when compared to saline-treated mice (two-way analysis of variance followed by Bonferroni *post-hoc* test: ** $p < 0.01$).

Table 2(*r*)). This result supports the hypothesis that curling behaviour involves MOR activation.

The opioid (\pm)-tramadol acts through both MOR and 5HT/NA transporters and, hence, we explored its effects on C57BL/6J MOR-KO mice over a range of active doses (32–64 mg/kg). Two-way ANOVA revealed a significant effect of (\pm)-tramadol on all the behaviour studied [Table 2(*s–u*); $p < 0.001$] and a significant effect of genotype on swinging ($p < 0.01$) and curling ($p < 0.001$). Interestingly, the interaction of both factors ((\pm)-tramadol \times genotype) was only significant for curling ($p < 0.05$). To study the effect of (\pm)-tramadol on different behaviours, we performed a one-way ANOVA, followed by the Dunnett's test [Table 2(*v–x*)]. Results showed that (\pm)-tramadol

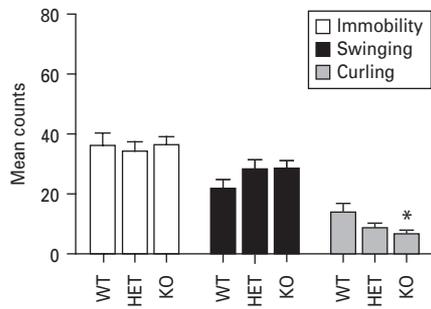


Fig. 5. Immobility, curling and swinging behaviour in wild-type (WT), heterozygous (HET) and homozygous μ -opioid receptor knockout (KO) mice in the tail suspension test. Data represent the mean counts + s.e.m. from 16–26 animals per group. There were significant differences when compared to saline-treated mice (one-way analysis of variance followed by Tukey's *post-hoc* test: * $p < 0.05$). Data from male and female mice were pooled.

significantly decreased immobility to a similar degree in all three genotypes (Fig. 6), although the effect of (\pm)-tramadol on active behaviour varied according to the genotype. Thus, at both doses (\pm)-tramadol significantly increased swinging behaviour in knockout mice alone ($p < 0.05$ and $p < 0.01$, respectively), while curling behaviour was significantly increased at 64 mg/kg in wild-type and heterozygous mice ($p < 0.01$ and $p < 0.001$, respectively; Fig. 6). Note that the effect of (\pm)-tramadol was similar in CD1 and C57BL/6J wild type mice (Figs. 3e, 6).

Discussion

The current study describes a modified means of scoring the TST, which differentiates between two active behaviours, swinging and curling. Using this approach, it was evident that antidepressants and opioids induce distinct active behaviours in the TST. While antidepressants increased swinging behaviour but had no effect on curling, opioids increased curling behaviour. Importantly, both antidepressants and opioids diminish the immobility of the mice, the traditional measure of antidepressant-like activity in the TST.

A modified form of the rat FST previously described a behavioural sampling technique that could distinguish between antidepressants with noradrenergic and serotonergic modes of action (Cryan *et al.* 2002). Hence, we employed a similar sampling technique to quantify and distinguish active behaviours induced by antidepressant treatment in the TST. While the traditional method of scoring the TST only measures the duration of immobility, we scored the frequency of the

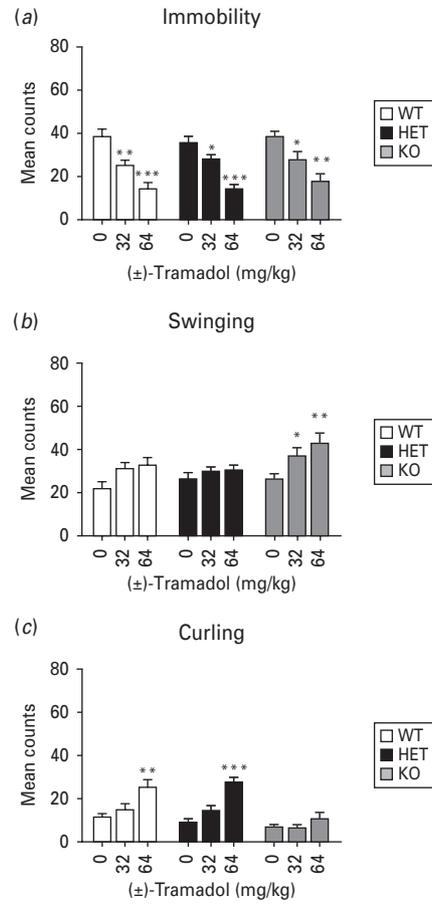


Fig. 6. Effects of (\pm)-tramadol (32–64 mg/kg i.p.) on immobility, curling and swinging behaviour in wild-type (WT), heterozygous (HET) and homozygous μ -opioid receptor knockout (KO) mice in the tail suspension test. (\pm)-Tramadol was administered 30 min before testing and the data represent the mean counts + s.e.m. from 15–24 animals per group. There were significant differences when compared to the corresponding saline-treated controls (one-way analysis of variance followed by Dunnett *post-hoc* test: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$). Data from male and female mice were pooled.

distinct behaviours in 5-s intervals throughout the test session. The results obtained using this approach do not differ from those obtained when each of the behaviours of interest are timed (Berrocoso & Mico, 2009a), yet they provided important information on the active behaviours that may be adopted when immobility is reduced. Significantly, our data are consistent with the strain differences previously observed in the TST (Cryan *et al.* 2005a; Liu & Gershenfeld, 2001) as CD1 mice were less immobile than C57BL/6J mice. Along similar lines, the immobility we observed in the dose-response experiments performed was in accordance

with the effects of antidepressants observed in other such screening tests. For example, the decrease in immobility following 10 and 20 mg/kg venlafaxine administration here is similar to its effect on mice in the FST (Berrocoso & Mico, 2009b). These findings validate the behavioural sampling technique and confirm that immobility is not to be confused with active behaviours.

The present study demonstrates that antidepressant compounds that differentially block the reuptake of NA and/or 5-HT induce a stereotypic behavioural profile that involves decreased immobility and increased swinging, with no effect on curling behaviour. These results implicate enhanced noradrenergic and/or serotonergic neurotransmission in swinging behaviour. Interestingly, and in agreement with previous data regarding the poor effect of SSRIs in animal models of depression (Berrocoso & Mico, 2009a; Lucki *et al.* 2001; Petit-Demouliere *et al.* 2005), a much higher dose of citalopram than that of other classes of antidepressants (noradrenergic or dual) is required to significantly decrease immobility and increase swinging in CD1 mice. However, this behavioural sampling technique was unable to differentiate between antidepressants that selectively increase 5-HT or noradrenergic neurotransmission, unlike the modified version of the rat FST (Detke *et al.* 1995). It should be noted that, to date, all behavioural patterns in mouse tests of depression (including the FST) have been attributed to either serotonergic or catecholaminergic signalling. The reasons for this restriction have not yet been elucidated but it suggests that differences in the coping strategies employed by mice and rats may be reflected in distinct active behaviours.

One of the main findings in this study is the clear demonstration of the antidepressant-like effect exerted by opioids, as witnessed by a decrease in immobility. These findings are consistent with previous studies using the TST or other behavioural tests to screen for depression or antidepressant effects using similar doses of these opioids (Fichna *et al.* 2007; Rojas-Corrales *et al.* 2002; Tejedor-Real *et al.* 1993, 1998). In the learned helplessness paradigm, morphine exerts an antidepressant-like effect and is antagonized by naloxone, indicating that it is indeed mediated by the opioids (Besson *et al.* 1996; Tejedor-Real *et al.* 1995). Antidepressant effects of other MOR agonists, such as (–)-methadone and levorphanol, have also been reported in the rat learned helplessness test (Rojas-Corrales *et al.* 2002). Thus, our findings validate the use of the TST as a test to screen for antidepressant-like activity of compounds that modulate opioid neurotransmission. Furthermore, the characteristic

reduction in immobility indicative of antidepressant-like activity was accompanied by a consistent increase in curling behaviour with all the opioids evaluated. The pharmacological blockade of this behaviour by naloxone suggests that the antidepressant-like effect of opioid drugs is mediated by the opioid system and not other neurotransmission systems. Furthermore, the increase in curling behaviour seems to be accompanied by a decrease in swinging. However, it is important to note that the observed dose-dependent reduction in swinging may be a consequence of the increase in curling, i.e. if the animals spend a substantial amount of the time curling then the time left for other behaviours is reduced. While opioids caused a 3- to 6-fold decrease in immobility at the highest doses, swinging was only reduced ~2-fold (with the exception of morphine), indicating that many opioids induce a shift towards relatively more swinging in the 'non-curling' periods. This would suggest a contribution (although less relevant) of the monoaminergic system in the antidepressant-like effect of opioids.

It could be argued that curling behaviour reflects an opioid-induced increase in spontaneous motor activity or, alternatively, the induction of Straub tail, an S-shaped dorsiflexion of the mouse tail, produced by contraction of the sacro-coccygeal dorsalis muscles. Such effects would suggest that curling behaviour is not an escape-oriented behaviour but, rather, that it can be considered as a false positive score in the TST. However, we previously showed that codeine (40 mg/kg) did not modify spontaneous motor activity or coordination (Berrocoso & Mico, 2009a). In addition, spontaneous locomotion is not modified in the genetic knockout of the MOR (Ide *et al.* 2010), ruling out any possible increase in spontaneous motor activity. While Straub tail may be produced by high doses of opiates (Bilbey *et al.* 1960; Narita *et al.* 1993), genetic blockade of MORs significantly and specifically decreases curling behaviour without modifying immobility, indicating that Straub tail and curling are modulated by different mechanisms. Therefore, it seems unlikely that these events are confounded in the analysis of active behaviours in the TST.

While many studies have described antidepressant-like effects of δ (Tejedor-Real *et al.* 1998; Torregrossa *et al.* 2005, 2006) and κ (Mague *et al.* 2003; Pliakas *et al.* 2001; Shirayama *et al.* 2004) receptor antagonists, the specific roles of these opioid receptors on opioid-induced behaviour was not evaluated here. Hence, further studies with opioids that specifically act through these receptors will be necessary to determine their individual contributions. Indeed, since δ -opioid receptors are thought to be critical for the analgesic

activity of tricyclic antidepressants (Benbouzid *et al.* 2008), it would be interesting to evaluate their possible contribution to the effect observed in the TST. Finally, in the light of experiences with the FST (Cryan *et al.* 2005b), it would also be very interesting to test other effective or potential antidepressants that do not directly target monoamines (such as ketamine, AMPA potentiators or CRF1 antagonists) in order to determine how they affect the active behaviours described above.

We evaluated our modified scoring method using (\pm)-tramadol, a compound that has both opioid and monoaminergic effects. This compound is a weak agonist of the MOR and, like many antidepressant drugs, it inhibits the reuptake of 5-HT and NA. When compared to its parental compound, (+)-tramadol binds more potently to MOR and it inhibits the reuptake of 5-HT (Raffa *et al.* 1993; see Table 1). Indeed, (\pm)-tramadol decreases the immobility of CD1 mice and is suggestive of an antidepressant-like action (Berrocoso *et al.* 2006; Rojas-Corrales *et al.* 2002, 2004; Yalcin *et al.* 2007), further validating our sampling method. While (\pm)-tramadol significantly increased curling behaviour (consistent with an increase in MOR activity) it had no effect on swinging behaviour, despite its monoaminergic mode of action. Interestingly, (+)-tramadol, which possesses greater affinity for MOR, acted like a typical opioid, decreasing immobility and swinging behaviour and increasing curling. These data are consistent with the stronger affinity of (\pm)-tramadol and (+)-tramadol for MOR than for 5-HT/NA transporters (Table 1).

The administration of (\pm)-tramadol to MOR-KO mice provided further evidence of its specific mode of action. Thus, (\pm)-tramadol showed similar antidepressant-like effects in all three genotypes (wild-type, heterozygous and knockout), as indicated by the decrease in immobility. However, the increase in curling behaviour in wild-type mice suggests that (\pm)-tramadol acts through the opioid system, although the increase in swinging induced by (\pm)-tramadol in MOR-KO mice suggests that it enhances monoaminergic neurotransmission. Heterozygous mice displayed a profile similar to that of wild-type mice, indicative of a predominant opioid-mediated effect. It is also noteworthy that (\pm)-tramadol does not apparently affect swinging in wild-type mice, despite its well-known influence on monoamine transporters. The decrease in immobility but not swinging indicates a shift towards more swinging in the 'non-curling' periods. However, (\pm)-tramadol-induced swinging is more pronounced in MOR-KO mice, indicating that curling induced by MOR activation might mask the

effects on swinging. These findings demonstrate the utility of this approach to explore the mode of action of monoaminergic/opioidergic compounds. Furthermore, we show that (\pm)-tramadol has both a central monoaminergic and opioidergic activity and that it can elicit antidepressant-like effects, even when one of these signalling mechanisms is blocked. This finding may be particularly relevant in pathological conditions in which both receptor profiles are modified.

Depression displays remarkable inter-individual variation in terms of symptoms and drug response. For example, opioid therapy has been successful in treating some refractory cases of depression such as when using the MOR agonists, oxycodone and oxymorphone, the partial agonist, buprenorphine, and the atypical opioid (\pm)-tramadol (Bodkin *et al.* 1995; Fanelli & Montgomery, 1998; Shapira *et al.* 2001; Spencer, 2000; Stoll & Rueter, 1999). Accordingly, alterations to the opioid system may underlie the neuroendocrine abnormalities observed in some groups of patients with this illness (Kennedy *et al.* 2006). Indeed, significant alterations in MOR activation have been observed in several brain areas (e.g. rostral region of the anterior cingulate) in patients with a major depressive disorder who did not respond to SSRI treatment. Interestingly, these alterations were correlated with corticotropin and cortisol plasma levels. Thus, although it remains to be confirmed, it is possible that the behaviour's affect in the TST could serve to predict different symptom clusters or therapeutic effects.

In summary, using a novel approach to scoring the TST, two specific active behaviours can be characterized that enable serotonergic/noradrenergic antidepressants to be distinguished from opioid compounds: swinging and curling. While traditional antidepressants that inhibit serotonin and/or NA reuptake decrease immobility and increase swinging behaviour, opioids, having decreased immobility, increase curling behaviour. Analysing these active behaviours may be useful to evaluate the mode of action of opioids and of opioids that also display monoaminergic properties, providing an important means of analysing the antidepressant effects of opioid compounds.

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Statement of Interest

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References

- Beique JC, Lavoie N, de Montigny C, Debonnel G** (1998). Affinities of venlafaxine and various reuptake inhibitors for the serotonin and norepinephrine transporters. *European Journal of Pharmacology* **349**, 129–132.
- Benbouzid M, Gaveriaux-Ruff C, Yalcin I, Waltisperger E, et al.** (2008). Delta-opioid receptors are critical for tricyclic antidepressant treatment of neuropathic allodynia. *Biological Psychiatry* **63**, 633–636.
- Berrocoso E, Mico JA** (2009a). Cooperative opioid and serotonergic mechanisms generate superior antidepressant-like effects in a mice model of depression. *International Journal of Neuropsychopharmacology* **12**, 1033–1044.
- Berrocoso E, Mico JA** (2009b). Role of serotonin 5-HT1A receptors in the antidepressant-like effect and the antinociceptive effect of venlafaxine in mice. *International Journal of Neuropsychopharmacology* **12**, 61–71.
- Berrocoso E, Rojas-Corales MO, Mico JA** (2006). Differential role of 5-HT1A and 5-HT1B receptors on the antinociceptive and antidepressant effect of tramadol in mice. *Psychopharmacology (Berlin)* **188**, 111–118.
- Berrocoso E, Sanchez-Blazquez P, Garzon J, Mico JA** (2009). Opiates as antidepressants. *Current Pharmaceutical Design* **15**, 1612–1622.
- Besson A, Privat AM, Eschalier A, Fialip J** (1996). Effects of morphine, naloxone and their interaction in the learned-helplessness paradigm in rats. *Psychopharmacology (Berlin)* **123**, 71–78.
- Bilbey DL, Salem H, Grossman MH** (1960). The anatomical basis of the Straub phenomenon. *British Journal of Pharmacology and Chemotherapy* **15**, 540–543.
- Bodkin JA, Zornberg GL, Lukas SE, Cole JO** (1995). Buprenorphine treatment of refractory depression. *Journal of Clinical Psychopharmacology* **15**, 49–57.
- Bravo L, Berrocoso E, Mico JA** (2009). Animal models in psychiatry: conceptualization and preclinical models of depression. *European Journal of Psychiatry* **23** (Suppl.), 111–122.
- Cherbat R, Thierry B, Mico JA, Steru L, et al.** (1986). Adaptation of the tail suspension test to the rat. *Journal of Pharmacology* **17**, 348–350.
- Codd EE, Shank RP, Schupsky JJ, Raffa RB** (1995). Serotonin and norepinephrine uptake inhibiting activity of centrally acting analgesics: structural determinants and role in antinociception. *Journal of Pharmacology and Experimental Therapeutics* **274**, 1263–1270.
- Cryan JF, Markou A, Lucki I** (2002). Assessing antidepressant activity in rodents: recent developments and future needs. *Trends in Pharmacological Sciences* **23**, 238–245.
- Cryan JF, Mombereau C, Vassout A** (2005a). The tail suspension test as a model for assessing antidepressant activity: review of pharmacological and genetic studies in mice. *Neuroscience and Biobehavioral Reviews* **29**, 571–625.
- Cryan JF, Valentino RJ, Lucki I** (2005b). Assessing substrates underlying the behavioral effects of antidepressants using the modified rat forced swimming test. *Neuroscience and Biobehavioral Reviews* **29**, 547–569.
- Detke MJ, Rickels M, Lucki I** (1995). Active behaviors in the rat forced swimming test differentially produced by serotonergic and noradrenergic antidepressants. *Psychopharmacology (Berlin)* **121**, 66–72.
- Fanelli J, Montgomery C** (1998). Use of the analgesic tramadol in antidepressant potentiation. *Psychopharmacology Bulletin* **32**, 442.
- Fichna J, Janicka A, Piastreniewicz M, Costentin J, et al.** (2007). Antidepressant-like effect of endomorphin-1 and endomorphin-2 in mice. *Neuropsychopharmacology* **32**, 813–821.
- Hegadoren KM, O'Donnell T, Lanius R, Coupland NJ, et al.** (2009). The role of beta-endorphin in the pathophysiology of major depression. *Neuropeptides* **43**, 341–353.
- Ide S, Sora I, Ikeda K, Minami M, et al.** (2010). Reduced emotional and corticosterone responses to stress in mu-opioid receptor knockout mice. *Neuropharmacology* **58**, 241–247.
- Jutkiewicz EM** (2006). The antidepressant-like effects of delta-opioid receptor agonists. *Molecular Interventions* **6**, 162–169.
- Kennedy SE, Koeppe RA, Young EA, Zubieta JK** (2006). Dysregulation of endogenous opioid emotion regulation circuitry in major depression in women. *Archives of General Psychiatry* **63**, 1199–1208.
- Kraepelin E** (1901). *Einführung in Die Psychiatrische Klinik*. Leipzig: Ambrosius Barth-Verlag.
- Liu X, Gershenfeld HK** (2001). Genetic differences in the tail-suspension test and its relationship to imipramine response among 11 inbred strains of mice. *Biological Psychiatry* **49**, 575–581.
- Lucki I, Dalvi A, Mayorga AJ** (2001). Sensitivity to the effects of pharmacologically selective antidepressants in different strains of mice. *Psychopharmacology (Berlin)* **155**, 315–322.
- Mague SD, Pliakas AM, Todtenkopf MS, Tomasiewicz HC, et al.** (2003). Antidepressant-like effects of kappa-opioid receptor antagonists in the forced swim test in rats. *Journal of Pharmacology and Experimental Therapeutics* **305**, 323–330.

- Narita M, Suzuki T, Misawa M, Nagase H** (1993). Antagonism of the morphine-induced Straub tail reaction by kappa-opioid receptor activation in mice. *Psychopharmacology (Berlin)* **110**, 254–256.
- Owens MJ, Morgan WN, Plott SJ, Nemeroff CB** (1997). Neurotransmitter receptor and transporter binding profile of antidepressants and their metabolites. *Journal of Pharmacology and Experimental Therapeutics* **283**, 1305–1322.
- Petit-Demouliere B, Chenu F, Bourin M** (2005). Forced swimming test in mice: a review of antidepressant activity. *Psychopharmacology (Berlin)* **177**, 245–255.
- Pliakas AM, Carlson RR, Neve RL, Konradi C, et al.** (2001). Altered responsiveness to cocaine and increased immobility in the forced swim test associated with elevated cAMP response element-binding protein expression in nucleus accumbens. *Journal of Neuroscience* **21**, 7397–7403.
- Raffa RB, Friderichs E, Reimann W, Shank RP, et al.** (1992). Opioid and nonopioid components independently contribute to the mechanism of action of tramadol, an 'atypical' opioid analgesic. *Journal of Pharmacology and Experimental Therapeutics* **260**, 275–285.
- Raffa RB, Friderichs E, Reimann W, Shank RP, et al.** (1993). Complementary and synergistic antinociceptive interaction between the enantiomers of tramadol. *Journal of Pharmacology and Experimental Therapeutics* **267**, 331–340.
- Rojas-Corrales MO, Berrocoso E, Gibert-Rahola J, Mico JA** (2002). Antidepressant-like effects of tramadol and other central analgesics with activity on monoamines reuptake, in helpless rats. *Life Sciences* **72**, 143–152.
- Rojas-Corrales MO, Berrocoso E, Gibert-Rahola J, Mico JA** (2004). Antidepressant-like effect of tramadol and its enantiomers in reserpinized mice: comparative study with desipramine, fluvoxamine, venlafaxine and opiates. *Journal of Psychopharmacology* **18**, 404–411.
- Rojas-Corrales MO, Gibert-Rahola J, Mico JA** (1998). Tramadol induces antidepressant-type effects in mice. *Life Sciences* **63**, PL175–PL180.
- Shapira NA, Verduin ML, DeGraw JD** (2001). Treatment of refractory major depression with tramadol monotherapy. *Journal of Clinical Psychiatry* **62**, 205–206.
- Shirayama Y, Ishida H, Iwata M, Hazama GI, et al.** (2004). Stress increases dynorphin immunoreactivity in limbic brain regions and dynorphin antagonism produces antidepressant-like effects. *Journal of Neurochemistry* **90**, 1258–1268.
- Sora I, Elmer G, Funada M, Pieper J, et al.** (2001). Mu opiate receptor gene dose effects on different morphine actions: evidence for differential *in vivo* mu receptor reserve. *Neuropsychopharmacology* **25**, 41–54.
- Spencer C** (2000). The efficacy of intramuscular tramadol as a rapid-onset antidepressant. *Australian and New Zealand Journal of Psychiatry* **34**, 1032–1033.
- Steru L, Chermat R, Thierry B, Mico JA, et al.** (1987). The automated tail suspension test: a computerized device which differentiates psychotropic drugs. *Progress in Neuropsychopharmacology and Biological Psychiatry* **11**, 659–671.
- Steru L, Chermat R, Thierry B, Simon P** (1985). The tail suspension test: a new method for screening antidepressants in mice. *Psychopharmacology (Berlin)* **85**, 367–370.
- Stoll AL, Rueter S** (1999). Treatment augmentation with opiates in severe and refractory major depression. *American Journal of Psychiatry* **156**, 2017.
- Tejedor-Real P, Mico JA, Maldonado R, Roques BP, et al.** (1993). Effect of mixed (RB 38A) and selective (RB 38B) inhibitors of enkephalin degrading enzymes on a model of depression in the rat. *Biological Psychiatry* **34**, 100–107.
- Tejedor-Real P, Mico JA, Maldonado R, Roques BP, et al.** (1995). Implication of endogenous opioid system in the learned helplessness model of depression. *Pharmacology Biochemistry and Behavior* **52**, 145–152.
- Tejedor-Real P, Mico JA, Smadja C, Maldonado R, et al.** (1998). Involvement of delta-opioid receptors in the effects induced by endogenous enkephalins on learned helplessness model. *European Journal of Pharmacology* **354**, 1–7.
- Torregrossa MM, Folk JE, Rice KC, Watson SJ, et al.** (2005). Chronic administration of the delta opioid receptor agonist (+)BW373U86 and antidepressants on behavior in the forced swim test and BDNF mRNA expression in rats. *Psychopharmacology (Berlin)* **183**, 31–40.
- Torregrossa MM, Jutkiewicz EM, Mosberg HI, Balboni G, et al.** (2006). Peptidic delta opioid receptor agonists produce antidepressant-like effects in the forced swim test and regulate BDNF mRNA expression in rats. *Brain Research* **1069**, 172–181.
- Yalcin I, Aksu F, Bodard S, Chalou S, et al.** (2007). Antidepressant-like effect of tramadol in the unpredictable chronic mild stress procedure: possible involvement of the noradrenergic system. *Behavioural Pharmacology* **18**, 623–631.