

1 **Effect of daily thermo- and photo-cycles of different light spectrum on**  
2 **the development of Senegal sole (*Solea senegalensis*) larvae.**

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23 **Abstract.**

24           This paper investigates the impact of different thermo- and photo-cycles of  
25 distinct wavelengths on *Solea senegalensis* larvae from day 1 to 30 post-hatching. In  
26 experiment 1, larvae were exposed to 12 h light:12 h dark (12L:12D) cycle and (A)  
27 constant temperature (20.7°C), (B) thermocycle of 12h thermophase: 12h cryophase,  
28 22.1°C day: 19.0°C night (referred to as TC) or (C) 12h cryophase: 12h thermophase,  
29 19.2°C day: 22.0°C night (referred to as CT). In experiment 2, larvae were kept under  
30 constant temperature (20.8°C) and exposed to (A) continuous light (LL), (B) continuous  
31 darkness (DD), and LD 12L:12D cycles of (C) white light (LD<sub>W</sub>), (D) blue light (LD<sub>B</sub>)  
32 or (E) red light (LD<sub>R</sub>). The sole larvae achieved the best performance, and showed  
33 fastest development and lowest degree of deformity under natural thermocycle  
34 conditions (TC) with a deformity percentage of 31.1% and LD<sub>B</sub> cycles with 27.7% of  
35 malformation, conditions which were nearest their natural aquatic environment. Larvae  
36 reared under TC started eye migration at 9 day post-hatching (DPH), while larvae  
37 exposed to CT started eye migration at 11 DPH. In larvae under the LD<sub>B</sub> treatment the  
38 migration of the eye started earlier than in the other treatments (9 DPH), while larvae  
39 reared under LL and DD photoperiods died before metamorphosis. These findings  
40 highlight the importance of light and temperature cycles during the early development  
41 of *Solea senegalensis* larvae, which should be taken into consideration in experimental  
42 or rearing protocols.

43 **Keywords:** Temperature, Light spectrum, Metamorphosis, Senegal sole larvae.

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## 47 **1. Introduction**

48           In the natural environment, daily temperature and light cycles are key  
49 synchronising factors for fish. The Earth's rotation imposes a day/night light cycle  
50 which fish have to cope with in order to survive. Moreover, the cyclic infrared radiation  
51 from the Sun generates a thermocycle: during the day the temperature rises (the  
52 thermophase, or phase of higher temperature), while during the night the temperature  
53 drops (the cryophase, or phase of lower temperature). Thus, transitions from cold to  
54 warm temperature are roughly associated with dawn, and transitions from warm to cold  
55 temperature with dusk (Johnson et al., 2004). However, under artificial rearing  
56 conditions, the environmental conditions are set by fish farmers to try to ensure fish  
57 survival and growth, which has been documented in many aquaculture species  
58 (Barahona-Fernandes, 1979; Tandler and Helps, 1985; Batty, 1987; Downing and  
59 Litvak, 1999). Little attention has been paid to the influence of these "unnatural"  
60 environmental conditions (constant lighting and temperature) and light characteristics of  
61 artificial globes (incandescent and halogen globes) in shallow tanks, which would have  
62 more red/yellow in comparison to light conditions underwater lit by natural sunlight,  
63 although they have recently been reported to affect the rhythmic development of larvae  
64 of fish models such as zebrafish (Vallone et al., 2007).

65           The water column acts as a chromatic filter, so that the aquatic environment  
66 changes the spectral composition of incident light. There is rapid attenuation with depth,  
67 so blue wavelengths become predominant in all but the most shallow or turbid waters  
68 (Jerlov, 1968). However, standard lighting systems (bulbs and fluorescent lamps)  
69 commonly used in hatcheries are shifted towards the red wavelengths and create bright  
70 points, light sources that are not environment-specific and could compromise fish

71 welfare. Besides, inappropriate light intensity can affect fish by provoking skeletal  
72 abnormalities (Villamizar et al., 2009).

73 The metamorphosis in flatfish is related to the change from the pelagic to  
74 benthic habitat, and it implies important changes in fish physiology (Fernandez-Díaz et  
75 al., 2001). The transformation occurs at a wide range of sizes depending on species and  
76 environmental circumstances (Policansky, 1982; Ottesen and Bolla, 1998). Whether age  
77 or size are key factors in starting metamorphosis is a question that has been considered  
78 in several studies with other species, such as Atlantic halibut larvae (Ottesen and Bolla,  
79 1998). In addition, this question is important in laboratory populations in which growth  
80 and therefore, the size of larvae and successful metamorphosis depend on the rearing  
81 conditions (Fernandez-Díaz et al., 2001).

82 Temperature has long been reported to influence the growth and development of  
83 fish larvae both in the wild and laboratory (Methot and Kramer, 1979; Pepin, 1991;  
84 Green and Fisher, 2004; Johnston et al., 2004), and there are reports that have studied  
85 thermal effects on embryonic development in flatfish larvae and juveniles, although  
86 without temperature control (Buckley, 1982; Daniels et al., 1996). Moreover, we are not  
87 aware of any published information about the effect of daily thermocycles on fish  
88 larvae.

89 The Senegal sole *Solea senegalensis* Kaup (1858) is a flatfish adapted to  
90 temperate waters of around 20-21°C (Drake et al., 1984). It is extensively exploited in  
91 aquaculture, mostly in Spain and Portugal (Dinis et al., 1999). Substantial progress has  
92 been achieved in its domestication, though suitable reproduction and zootechnical  
93 optimal conditions are still to be determined (Porta et al., 2007). Due to the interest in  
94 this species for aquaculture and its characteristic embryonic development

95 (metamorphosis process), this species was selected as our experimental model to  
96 investigate the effects that temperature and light cycles may induce in larval survival,  
97 growth, development and on the onset/offset of metamorphosis. To this end, sole larvae  
98 were subjected to different temperature cycles and they were exposed to different  
99 photoperiods and light spectra.

## 100 **2. Material and methods**

### 101 *2.1. Animals and housing*

102 Fertilised eggs of Senegal sole were provided by the Spanish Oceanographic  
103 Institute (IEO) at Santander (Cantabria) and sent to the Aquaculture Institute in Torre la  
104 Sal (IATS-CSIC, Castellón) and the University of Murcia (UMU, Fish Chronolab,  
105 Murcia). Wild broodstock were distributed into 14 m<sup>3</sup> tanks, in a female:male  
106 proportion of 1:1. The tanks were covered with a shadow net that provided 0.21 W m<sup>-2</sup>  
107 (50 lux) on the surface of water. The fish were fed five days a week, three days with  
108 mussels and two days with small cuttlefish; one of the days on which they were fed with  
109 mussels, frozen polychaetes were added (Sebait Ltd., UK). Water temperature varied  
110 between 16 and 19°C (imitating nature fluctuations of temperature) and the photoperiod  
111 was 12L:12D. An egg collector was placed at the water outflow of the tank. Once  
112 fertilised eggs were collected, they were incubated in 70 L incubators under continuous  
113 darkness until hatching. On 1 DPH they were transported under darkness conditions,  
114 and the transport had duration of 12 hours approximately.

115 To feed the larvae, *Brachionus plicatilis* rotifers were cultured and enriched with  
116 commercially available freeze-dried green algae *Nannochloropsis* sp. (Phytobloom  
117 Prof® Necton, Portugal) in a proportion of 300,000 cells/ml/day from day 3 to day 7.

118 These enriched rotifers were added to tanks daily as an early live food at a density of 20  
119 individuals mL<sup>-1</sup> from 3 to 7 DPH. *Artemia* sp. nauplii at a density of 2–3 nauplii mL<sup>-1</sup>  
120 day<sup>-1</sup> were introduced from 8 to 30 DPH. Before being fed to the larvae, the nauplii  
121 were enriched with a mixture (INVE DC DHA Selco®). Three to five metanauplii  
122 mL<sup>-1</sup> day<sup>-1</sup> were added from 27 to 30 DPH. Before being fed to the larvae, the  
123 metanauplii were enriched with a mixture (ORI-GO, ORI-PRO®, Skretting AS, Spain)  
124 of phytoproteins and highly unsaturated fatty acids (HUFA) during 24 hours.

## 125 ***2.2. Experimental design***

126 The experiments performed in the present research followed the Spanish  
127 legislation on Animal Welfare and Laboratory Practices. The experimental protocol was  
128 approved by both the National Committee on Animal Welfare and the Committee of the  
129 University of Murcia.

### 130 ***2.2.1. Experiment 1. Effect of temperature.***

131 This trial was performed at IATS-CSIC. Spawed egg volume was 120 mL, with  
132 a fertilization rate of 73% and hatching rate of 78.1%. The larvae were distributed into  
133 six 500 L cylinder-conical tanks at 1 DPH and at a density of 50 larvae L<sup>-1</sup> to study the  
134 effect of temperature. The water system was semi-closed with a flow of 50 L/h  
135 exchanged seawater in each tank per day. Thermal cycles had a duration of 12 hours,  
136 the photoperiod was 12L:12D and light intensity was 0.84 W m<sup>-2</sup> (200 lux) supplied by  
137 a lamp (PHILIPS, HPL N 250W). The spectral analysis of the lights was performed  
138 using a spectroradiometer (FieldSpec® Hand Held spectroradiometer UV/VNIR from  
139 ASD Colorado, USA, with a wavelength range of 325 nm to 1075 nm at 1.6 nm

140 intervals) and a lux meter (MX Elektronik Minilux, Germany, with 6 measuring ranges  
141 from 2 lux to 200 klux.).

142 From 1 to 30 DPH, three temperature regimes with two replicates were applied:  
143 TC cycle ( $22.1 \pm 0.6^\circ\text{C}$  day/ $19.0 \pm 0.4^\circ\text{C}$  night, mean $\pm$ S.D, n=60, here and throughout);  
144 CT cycle ( $19.2 \pm 0.5^\circ\text{C}$  day/ $22.0 \pm 0.3^\circ\text{C}$  night) and constant temperature ( $20.7 \pm 0.4^\circ\text{C}$ ),  
145 Water temperature was modified by means of water heaters (Themal Compact 100 W,  
146 Askoll, Italy) or coolers (Cubigel, E-500, Spain). Temperature was continuously  
147 recorded with an underwater sensor and data logger (HOBO PENDANT<sup>®</sup> Onset  
148 Computer Corporation, Massachusetts, USA) placed in each of the tanks (Fig. 1TC,  
149 1CT).

### 150 ***2.2.2. Experiment 2. Effect of photoperiod and light spectrum.***

151 This experiment was carried out at UMU. The fertilised eggs were collected in  
152 complete darkness to ensure fish larvae did not receive any light before treatment.  
153 Spawned egg volume was 160 mL, with a fertilization rate of 98.1% and hatching rate  
154 of 77.6%. The larvae were distributed into ten 80 L aquaria with flat bottoms at 1 DPH  
155 and at a density of 18 larvae L<sup>-1</sup> to study the effects of different light spectra. The larvae  
156 were kept under constant temperature ( $20.8 \pm 1.3^\circ\text{C}$ ). The static closed water system used  
157 filtered artificial seawater by means of a biological filter EHEIM 2227 and using a bio-  
158 balls filter system, filtering surface ratio being approximately 1 L bio-balls / 10 L of  
159 water.

160 From 1 to 30 DPH, five lighting regimes with three replicates were applied:  
161 12L:12D cycle with red (LD<sub>R</sub>, half-peak bandwidth 592-668 nm), blue (LD<sub>B</sub>, half-peak  
162 bandwidth 435–500 nm) and white (LD<sub>W</sub>), 24L:0D white (LL), and 0L:24D (DD). The

163 white light had a broad spectrum, with 95% irradiance within the range of 367–757 nm.  
164 The spectral analysis of the lights was performed using a spectroradiometer  
165 (FieldSpec®, ASD, Colorado, USA) (Fig. 1A, 1B, 1C). To avoid the effects of any  
166 background light on the experiments, the experimental tanks were covered with a light-  
167 proof, black screen. For the different spectral trials, lamps were constructed using light  
168 emitting diodes (LEDs) mounted on fibreglass plaques (160×232 mm). Each red and  
169 blue lamp had 17 homogeneously distributed LEDs. White lamps had 18 white and 4  
170 red LEDs to produce a broader range wavelength. Each lamp was encased in a  
171 waterproof container which hung approximately 50 cm above the water surface. The  
172 lamps were powered by a 3 V DC supply connected to a variable resistor (0–2 kΩ) that  
173 allowed the light intensity to be adjusted to  $0.42 \text{ W m}^{-2}$ , which is low but well above the  
174 light threshold ( $0.053 \text{ W m}^{-2}$ ) required to modify melatonin contents in both the eye and  
175 plasma in Senegal sole (Oliveira et al., 2007). In addition, previous observations have  
176 shown that sole are capable of feeding in the dark from the very early hatching stage  
177 (Blaxter, 1969).

### 178 *2.3. Data collection and analysis*

179 Every two days from 1 to 30 DPH, 60 larvae from each treatment were  
180 arbitrarily chosen to measure only total length (making a straight measurement from the  
181 tip of the head to the tip of the longest lobe of caudal fin) and evaluate metamorphosis  
182 (the criteria for defining the beginning and the finishing of metamorphosis was the eye  
183 migration) and to detect physical abnormalities focusing on jaw elongation (jaw  
184 malformations). In addition, at 3, 5, 7, 9 and 11 DPH, yolk sac dimension were  
185 measured in these larvae. Yolk volume was calculated using the formula  $V=\pi/6 \text{ l h}^2$ ,  
186 where l is yolk sac length and h is yolk sac height (Bagarinao, 1986). Larvae were killed

187 on ice and fixed with 5% formaldehyde and were measured using a digital camera  
188 mounted on a stereoscopic microscope EZ4D. Samples were photographed and  
189 calibrated measurements were made using image processing and analysis software  
190 (Leica Microsystems Imaging Solutions Ltd, Cambridge, UK.).

191 To find statistical differences in growth, development and jaw malformation  
192 among treatments, all data were tested for normality (Student's *t*-test) by examining  
193 residual values and all data were found to be normal. Then, a one-way ANOVA was  
194 used to analyse all treatments. Yolk sac volumes were analysed by one-way ANOVA to  
195 establish statistical differences among treatments and time. Significant ANOVA  
196 analyses were followed by the Duncan's post hoc test to determine differences between  
197 means, with  $P < 0.05$  taken as the statistically significant threshold (SPSS 15.0 for  
198 Windows). Data are expressed as mean $\pm$ S.D. values.

### 199 **3. Results.**

#### 200 *3.1. Experiment 1. Effect of temperature.*

##### 201 *3.1.1. Growth.*

202 Initially, there were no significant differences in growth in larvae from different  
203 treatments until the day on which metamorphosis started. At 13 DPH however, larvae  
204 under CT (19°C day/22°C night) showed lower growth than those under the other  
205 treatments. Towards the end of the metamorphosis (23 DPH) and in first few days post-  
206 metamorphosis, larvae under TC showed significantly higher than those under constant  
207 temperature ( $F=5.37$ ,  $df= 2$ ,  $p=0.033$ ) and turn significantly higher ( $F=4.55$ ,  $df= 2$ ,  
208  $p=0.026$ ) than those under CT ( $F=4.67$ ,  $df= 2$ ,  $p=0.031$ ). At 30 DPH, TC larvae reached  
209 a size of  $8.5\pm 0.5$  mm, significantly higher than those under constant temperature at

210 7.7±0.2 mm which were in turn significantly than those under CT 7.2±0.2 mm (F=4.35,  
211 df= 2, p=0.023, Fig. 2).

### 212 **3.1.2. Yolk sac.**

213 The larvae which most quickly absorbed the yolk sac were those exposed to TC  
214 cycles and constant temperature. By 5 DPH they had already absorbed large part of the  
215 yolk sac, unlike the CT (F=5.36, df=2, p=0.036, Fig. 3). On 9 DPH larvae exposed to  
216 TC and those under constant temperature had already completely absorbed the sac. In  
217 contrast, larvae kept under a CT cycle showed slower yolk sac absorption, completed at  
218 11 DPH.

### 219 **3.1.3. Metamorphosis.**

220 Significant differences in the onset and finish of metamorphosis were detected  
221 among treatments. Larvae reared under TC started eye migration at 9 DPH and 100%  
222 finished the process at 17 DPH, larvae under constant temperature started at 11 DPH  
223 and 100% finished at 17 DPH, while larvae exposed to CT started eye migration at 11  
224 DPH and 100% finished at 19 DPH (F=5.4, df=2, p=0.022, Fig. 4A).

225 Larvae from the TC group which finished eye migration earliest (23 DPH)  
226 (F=6.28 df=2, p=0.028, Fig. 4B). In the other two treatments the process did not  
227 conclude until 25 DPH (F=9.51, df=2, p=0.035, Fig. 4B).

### 228 **3.1.4. Jaw malformation.**

229 From day 9 DPH jaw malformation was significantly higher in CT than in the  
230 other treatments. The larvae with the lowest percentage of malformation at 30 DPH  
231 were those exposed to TC (45.2±3.2%) and larvae submitted to constant temperature

232 (48±3.3%), but those under CT had a malformation percentage of 67±2.8% (F=4.55,  
233 df=2, p=0.026, Fig. 5).

## 234 **3.2. Experiment 2. Effect of photoperiod and light spectrum.**

### 235 **3.2.1. Growth.**

236 Larvae exposed to different lights showed differences in size before  
237 metamorphosis began, those exposed to blue light (LD<sub>B</sub>) being longer than the larvae  
238 from the other two light colour treatments (LD<sub>W</sub> and LD<sub>R</sub>) from 9 DPH onwards, while  
239 the larvae under complete darkness showed a lower growth during the experiment (from  
240 2 mm at 3 DPH to about 3.5 mm at 13 DPH). The larvae reaching the largest size at 30  
241 DPH were those under blue light (LD<sub>B</sub>), which reached 9.9±0.4 mm long, followed by  
242 those exposed to white light (LD<sub>W</sub>) which reached 8.3±0.6 mm, and finally the larvae  
243 under red light (LD<sub>R</sub>) that reached 7.2±0.4 mm (F=3.72, df= 2, p=0.028, Fig. 6).

### 244 **3.2.2. Yolk sac.**

245 The larvae which most quickly absorbed the yolk sac were those exposed to  
246 LD<sub>B</sub>, which showed significantly lower volume than LL and DD, but not other light  
247 colour treatments. By 7 DPH they had already absorbed a large part of the yolk sac,  
248 unlike the other treatments (F=4.05, df=4, p=0.041, Fig. 7). Larvae exposed to LL and  
249 DD absorbed the sac more slowly, and the yolk sac had not disappeared by 9 DPH.

### 250 **3.2.3. Eye migration and completed metamorphosis.**

251 In larvae under the LD<sub>B</sub> treatment the migration of the eye started earlier than in  
252 the other treatments, beginning at 9 DPH, while in the others groups, the migration  
253 started at 11 DPH. (F=5.12, df=4, p=0.015, Fig. 8A).

254 The groups of larvae under continuous darkness (DD) and continuous light (LL)  
255 died 15 and 17 DPH respectively at the beginning of metamorphosis. At 25 DPH, 100%  
256 of larvae in the LD<sub>B</sub> group had completed metamorphosis (F=5.03, df=2, p=0.039, Fig.  
257 8B), earlier than the other groups, where metamorphosis was not complete until 27 DPH  
258 (F=4.23, df=2, p=0.021, Fig. 8B).

#### 259 **3.2.4. Jaw malformation.**

260 Jaw malformation in larvae at 30 DPH from LD<sub>B</sub> (27.7±2.9%) was significantly  
261 lower than LD<sub>W</sub> (45.6±3%), in turn significantly lower than LD<sub>R</sub> (70.6±2.3%) (F=4.68,  
262 df=2, p=0.031, Fig. 9). Larvae under LL and DD treatments had 43.3±3.3% and  
263 26.7±2.3% of malformation respectively from early in the experiment (5 DPH),  
264 reaching 57.8±3.9% and 66.6±2.8% a few days before they died.

## 265 **4. Discussion**

266 Different conditions of temperature, photoperiod and light spectrum affected the  
267 development of Senegal sole larvae, driving growth performance and probably  
268 provoking malformations. Larvae exposed to TC (experiment 1) and LD<sub>B</sub> (experiment  
269 2) cycles provided the best results in terms of survival, growth and malformations.  
270 These experimental conditions closely approached natural environmental conditions: in  
271 the open sea, the water column acts as a chromatic filter, quickly absorbing red  
272 wavelengths, so that, temperature decreases and blue wavelengths become predominant  
273 as depth increases (Jerlov, 1968). In flatfish larvae the environmental factors may be  
274 crucial in determining recruitment to nursery grounds via their effects on growth and  
275 mortality during the metamorphosis-settlement period (Yamashita et al., 2001).

276           Some studies have demonstrated the existence of diel rhythms in temperature  
277 selection in fish maintained in wild conditions. In these studies, fish showed diel  
278 migrations as they searched for a preferred temperature for physiological activity and  
279 growth (Gibson et al., 1998; Sims et al., 2006). The present paper revealed that Senegal  
280 sole larvae showed higher growth, lower jaw malformation and faster absorption of the  
281 yolk sac under TC, and worse development under CT or constant temperature. Ottesen  
282 and Bolla (1998) reported that some physical parameters, including inappropriate  
283 temperature or salinity, may be associated with jaw malformation due to mechanical  
284 damage in cultured larvae. Under TC metamorphosis started earlier when the migration  
285 of the eye began, and were first to finish. In the other two treatments, the process of  
286 metamorphosis was slower, specially in larvae exposed to CT. Most studies about  
287 biological development and temperature in Senegal sole have used a constant  
288 temperature of 20°C (Martínez et al., 1999; Parra and Yúfera, 1999; Ribeiro et al., 1999;  
289 Yúfera et al., 1999; Cañavate et al., 2006), but little attention has been paid to the  
290 effects that temperature fluctuations in natural environment may cause in *Solea*  
291 *senegalensis*. The first study on temperature selection in fish was conducted in goldfish,  
292 in which the animals showed a diel pattern of temperature selection (Reynolds et al.,  
293 1978), which seemed to be related to body weight gain and gonadal growth (Spieler et  
294 al., 1977).

295           Through the rhythmic secretion of its hormone melatonin, the pineal organ of  
296 fish transducers photoperiod information (Ekström and Meissl, 1997). However,  
297 different light spectra do not affect melatonin production equally. As first evidenced in  
298 sea bass, the application of blue light during the dark phase inhibits nocturnal  
299 melatonin, while red light does not (Bayarri et al., 2002). Later, other studies have

300 reported that different light thresholds were required for sea bass (between  $3.8 \times 10^{-3} \mu\text{W}/\text{cm}^2$   
301  $^3 \mu\text{W}/\text{cm}^2$  and  $3.8 \times 10^{-4} \mu\text{W}/\text{cm}^2$ ) and Atlantic salmon (between  $3.8 \times 10^{-2} \mu\text{W}/\text{cm}^2$  and  
302  $3.8 \times 10^{-3} \mu\text{W}/\text{cm}^2$ ) to be perceived as night, measured by a significant increase in  
303 melatonin concentrations during the dark phases (Migaud et al., 2006) and it was  
304 demonstrated that light sensibility varies with the species. In Senegal sole, a recent  
305 paper reported a different effect of the light spectrum (red, violet and white lights) on  
306 plasma melatonin, with shorter wavelengths having the biggest impact (Oliveira et al.,  
307 2007). These observations agree with our results that larvae responded best to blue light  
308 ( $\text{LD}_B$ ), which appeared to be the most efficient light spectrum on the development on  
309 Senegal sole larvae.

310 Several studies have found a correlation between extended photoperiod and  
311 increased growth in teleost fish larvae such as *Sparus aurata* (Tandler and Helps, 1985),  
312 *Gadus morhua* (Puvanendran and Brown, 2002), *Latris lineata* (Trotter et al., 2003) and  
313 *Solea senegalensis* (Cañavate et al., 2006), and the same occurred in our experiment.  
314 Other studies, such as those carried out by Villamizar et al. (2009) with European sea  
315 bass, found no significant differences in total length between larvae exposed to either  
316 LL or  $\text{LD}_B$  cycles. Contrarily, we observed significant differences in size in our study.  
317 However, our results are supported by those obtained by Barahona- Fernandes (1979),  
318 which showed that continuous light in sea bass larvae did not induce the best growth.  
319 Downing (2002) found no relationship between growth and spectral composition of  
320 light in haddock larvae, while our experiment showed the highest growth in terms of  
321 length in those larvae reared under  $\text{LD}_B$  treatment.

322 The relationship between light conditions and malformations has been studied in  
323 recent experiments. For instance, Cobcroft and Battaglione (2009) found a relationship

324 between the colour of the rearing tank and the appearance of jaw malformations in  
325 *Latris lineata*, the larvae reared in a red tank showing the highest incidence of severe  
326 jaw malformations. Walling behaviour has been reported in Atlantic halibut larvae,  
327 where high light conditions are associated with walling and abrasion of larval jaw  
328 tissues on the tank wall (Morrison and MacDonald, 1995). This finding is in agreement  
329 with of our results in which LD<sub>R</sub>, together with LL and DD, showed the highest  
330 proportion of jaw malformation. Moreover, Villamizar et al. (2009) also reported jaw  
331 malformation and swim bladder hypertrophy in sea bass larvae under LL. The exact  
332 mechanism by which light affects malformations is unknown, but there are links  
333 between larval behavioural changes in response to light and other factors that implicate  
334 contact with walls (Morrison and MacDonald, 1995; Cobcroft and Battaglione, 2009).

335         Studies carried out by Geay et al. (2009) reported that an excess of retinoic acid  
336 increases craniofacial malformation in fish during development and Villeneuve et al.  
337 (2006) observed skeletal deformities in larvae fed high levels of vitamin A, which has  
338 an essential role in retinal function, together with carotenoids. Other study in Senegal  
339 sole related light intensity and diets supplemented with  $\beta$ -carotene, observing effects in  
340 antioxidant biomarkers (Cañavate et al., 2007). In addition, light intensity may provoke  
341 high proportion of malformations due to the mainly visual feeding activity of marine  
342 fish larvae (Blaxter, 1986). Recently, Villamizar et al. (2009) reported that light  
343 spectrum and photoperiod influenced feeding behaviour of sea bass larvae. For this  
344 reason, the fact that LD<sub>R</sub>, LL and DD treatments led to a higher level of malformations  
345 in our experiment may therefore have been due to impaired photoperiod and light  
346 spectrum. Further studies, however, are necessary to confirm whether a single culture

347 parameter (including subnutrition) or an interaction of physical parameters can  
348 contribute to the appearance of jaw malformation.

349         Since there seems to be a relationship between light and the exhaustion of  
350 endogenous sources, commercial hatcheries apply continuous darkness or dim light  
351 conditions to decrease the use of endogenous reserves and delay the start of exogenous  
352 feeding (Downing and Litvak, 1999). This agree with our results: larvae exposed to DD  
353 and LD<sub>R</sub> exhibited a delay in growth and thus the yolk sac was visible, while in the  
354 LD<sub>B</sub>, LD<sub>W</sub> and LL treatments the yolk sac was completely absorbed. These results are  
355 also consistent with those obtained by Villamizar et al. (2009) using a similar  
356 experimental design in sea bass larvae, in which the yolk sac in DD and LD<sub>R</sub> was still  
357 visible up to 48 h after its complete absorption in the LL, LD<sub>W</sub> and LD<sub>B</sub> groups.  
358 Parameters such as stocking density, salinity, feeding or light have been observed to  
359 affect the development and metamorphosis process in flatfish (Bolla and Holmefjord,  
360 1988; Daniels et al., 1996). In the present study, the LD<sub>B</sub> treatment provoked faster  
361 development in larvae than the other treatments, and these larvae completed the  
362 metamorphosis process several days before those under different light spectra.

363         The circadian system of fish develops quickly during the first stages of larval  
364 development, so the pineal organ becomes light sensitive before the lateral eyes and  
365 participates in photoperiod-controlled events such as hatching (Kazimi and Cahill,  
366 1999). In zebrafish, the key enzyme controlling melatonin production, AA-NAT, is  
367 expressed at the blastula stage from the first day post-fertilisation onwards (Ziv and  
368 Gothilf, 2006). Furthermore, in zebrafish larvae, lighting conditions were seen to  
369 influence the development of circadian clock gene expression, the clock starting ticking  
370 under LD cycles but not under constant darkness (Kaneko and Cahill, 2005). In short,

371 light and temperature cycles are required so that the rhythmic circadian clock works  
372 properly. In fish, the circadian clock mature extremely early during larval development  
373 and are thought to regulate temporal co-ordination of many physiological processes  
374 (Vallone et al., 2007). Differences in the development of the circadian system of sole in  
375 our experimental conditions may explain the differences found in the experimental  
376 treatments. Ongoing research aims to characterise clock gene expression in sole larvae,  
377 which seems to appear also very early (Muñoz-Cueto, personal communication) and is  
378 most likely influenced by light and temperature cycles during sole ontogeny.

379 In conclusion, our experiments revealed that the early development of Senegal  
380 sole larvae is affected by both, temperature and lighting conditions, larvae under TC and  
381 LD<sub>B</sub> cycles showing the best survival and performance, and the lowest incidence of jaw  
382 malformations. “Unnatural” or inappropriate light and temperature conditions seriously  
383 compromised the welfare of sole larvae and negatively affected their development.  
384 Thus, natural temperature and light cycles of blue wavelength appeared optimal  
385 conditions for the culture of this species, since they were nearest those of the natural  
386 underwater environment; such conditions seem to be a prerequisite for the proper  
387 maturation of the biological clock and the physiology of the larvae of this flatfish  
388 species. The potential benefits of TC and LD<sub>B</sub> during larval development, however,  
389 remain to be tested in other fish species.

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547 **Figure Captions.**

548 Figure 1. Treatment conditions in experiment to assess performance of Senegal sole  
549 larvae. Daily mean temperature of each treatment during the experiment: TC)  
550 thermocycle of 12h thermophase: 12h cryophase; CT) 12h cryophase: 12h thermophase.  
551 Spectral composition of each LED lamp expressed as the percentage of irradiance: A)  
552 LD<sub>B</sub> ( $\lambda=435\text{--}500$  nm); B) LD<sub>R</sub> ( $\lambda=592\text{--}668$  nm); C) LD<sub>W</sub> ( $\lambda=384\text{--}728$  nm).

553

554 Figure 2. Total length (mm) of Senegal sole larvae with age under different  
555 thermocycles. Data are expressed as mean $\pm$ SD. Different letters indicate means within  
556 age significantly different from each other.

557

558 Figure 3. Yolk sac absorption in Senegal sole larvae reared under different temperatures  
559 from 3 DPH. Data are expressed as mean $\pm$ S.D. Different letters indicate statistically  
560 significant differences from each other.

561

562 Figure 4. Evolution of the metamorphosis process in Senegal sole larvae exposed to  
563 different thermocycles. A) Percentage of larvae that started metamorphosis between 9  
564 DPH and 19 DPH. Inset: larva premetamorphosis. B) Percentage of larvae that  
565 concluded metamorphosis from 17 DPH to 30 DPH. Inset: metamorphosed larva. The

566 scale bar corresponds to 1 cm. Data are expressed as mean±SD. Different letters  
567 indicate means within age significantly different from each other.

568

569 Figure 5. Effect of temperature on jaw malformation of Senegal sole larvae with larval  
570 age. Black arrow in the picture indicates jaw malformation. Data are expressed as  
571 mean±S.D. Different letters indicate statistically significant differences from each other.  
572 The scale bar corresponds to 0.5 cm.

573

574 Figure 6. Total length (mm) of Senegal sole larvae with age under different light spectra  
575 and photoperiods. Data are expressed as mean±SD. Different letters indicate statistically  
576 significant differences from each other.

577

578 Figure 7. Yolk sac absorption in Senegal sole larvae with age reared under different  
579 light spectra and photoperiods from 3 DPH. Data is expressed as mean±S.D. Different  
580 letters indicate statistically significant differences from each other.

581

582 Figure 8. Evolution of the metamorphosis process in Senegal sole larvae with age  
583 exposed to different photoperiods and light spectra. A) Percentage of larvae that started  
584 metamorphosis between 9 DPH and 19 DPH. Inset: larva premetamorphosis. B)  
585 Percentage of larvae that concluded metamorphosis from 17 DPH to 30 DPH. Inset:  
586 metamorphosed larva. The scale bar corresponds to 1 cm. Data are expressed as  
587 mean±SD. Different letters indicate means within age significantly different from each  
588 other.

589 Figure 9. Effect of light spectrum and photoperiod on malformation of Senegal sole  
590 larvae during the experiment. Black arrow in the picture indicates jaw malformation.  
591 Data are expressed as mean±S.D. Different letters indicate statistically significant  
592 differences from each other. The scale bar corresponds to 0.5 cm.

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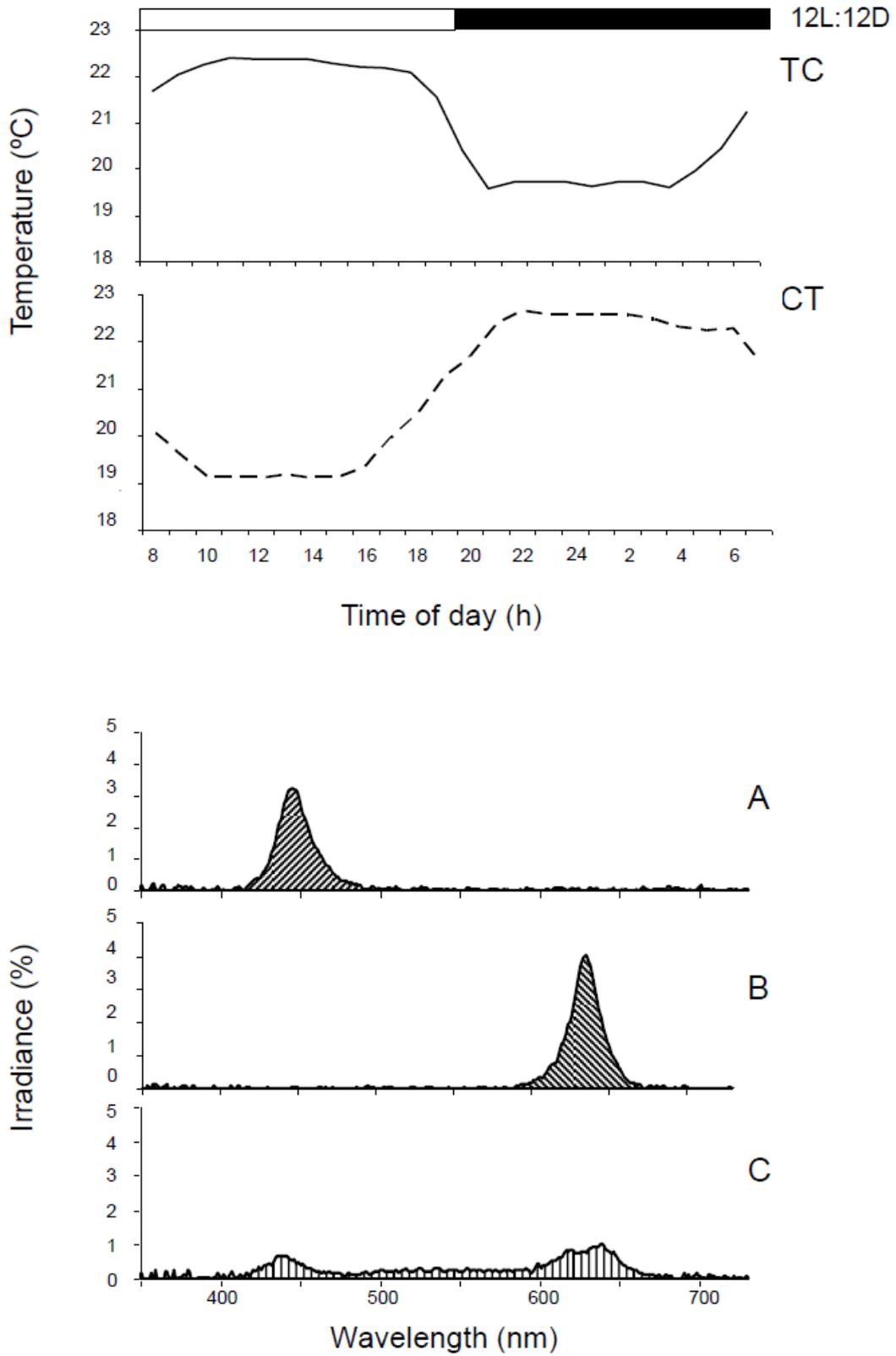
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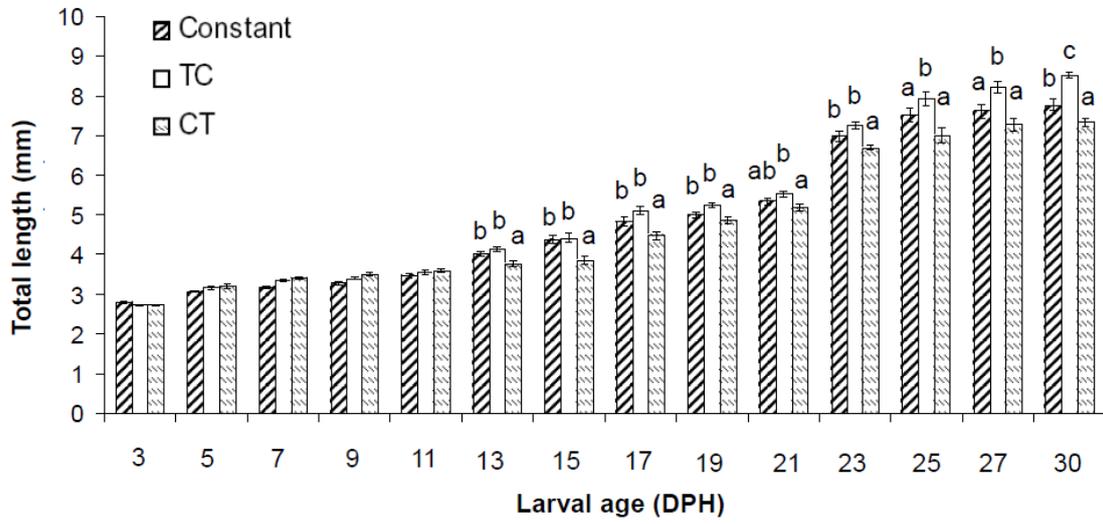
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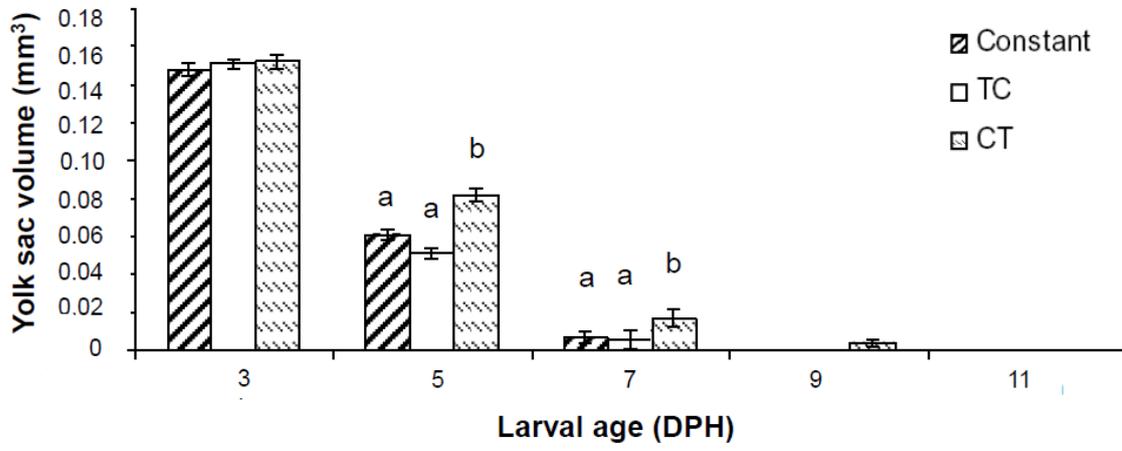
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623 Figure 3.



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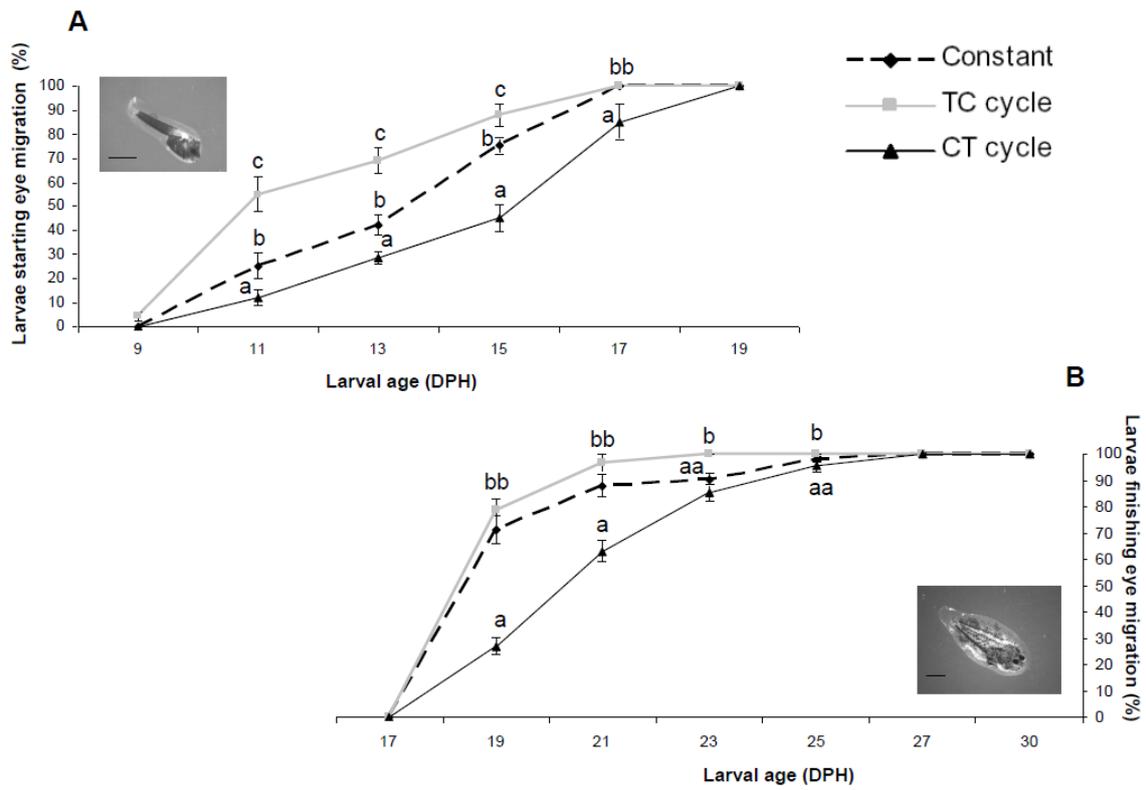
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637 Figure 4.



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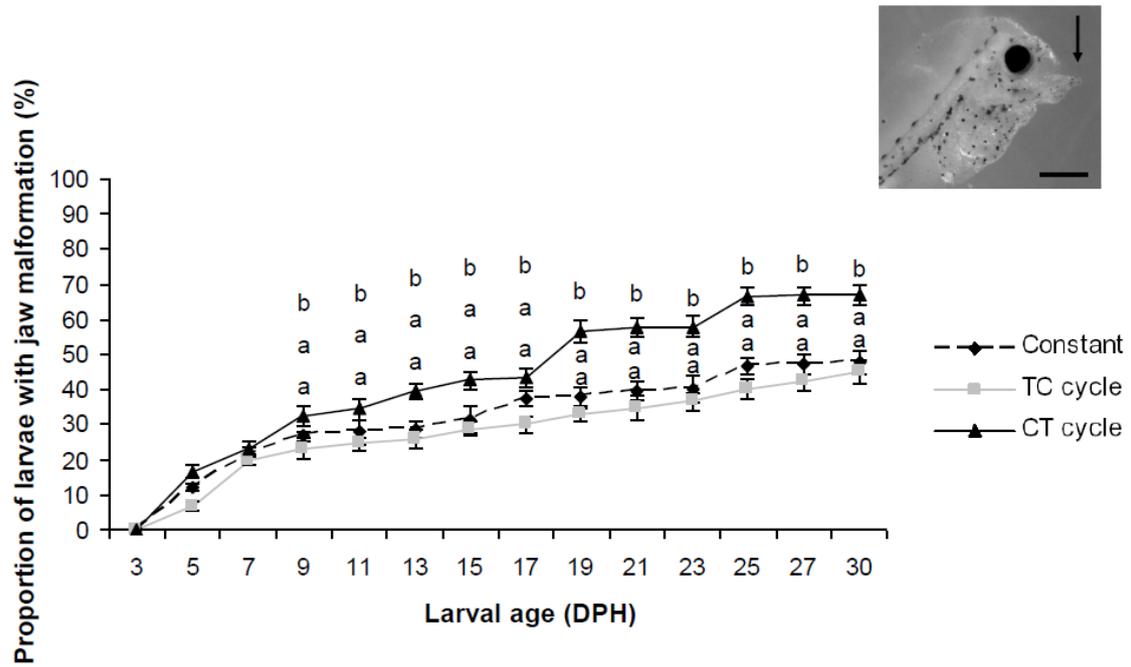
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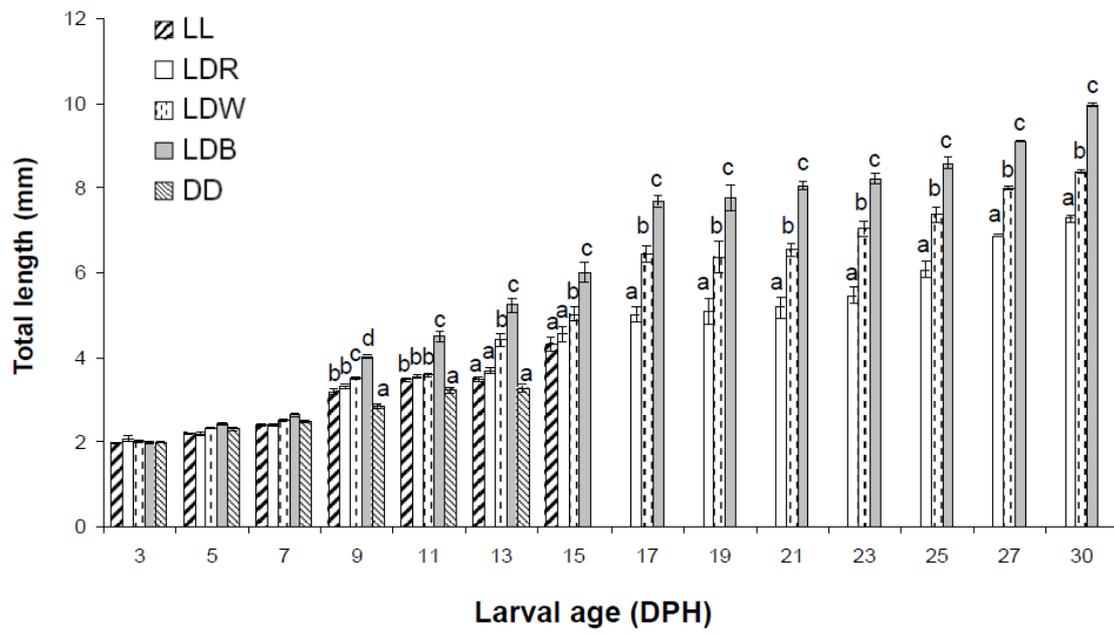
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660 Figure 6.



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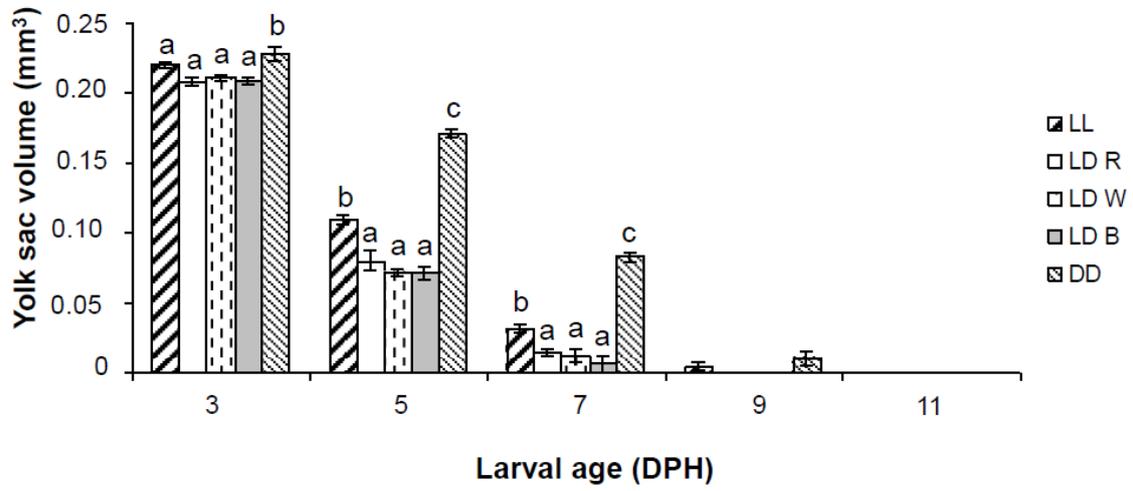
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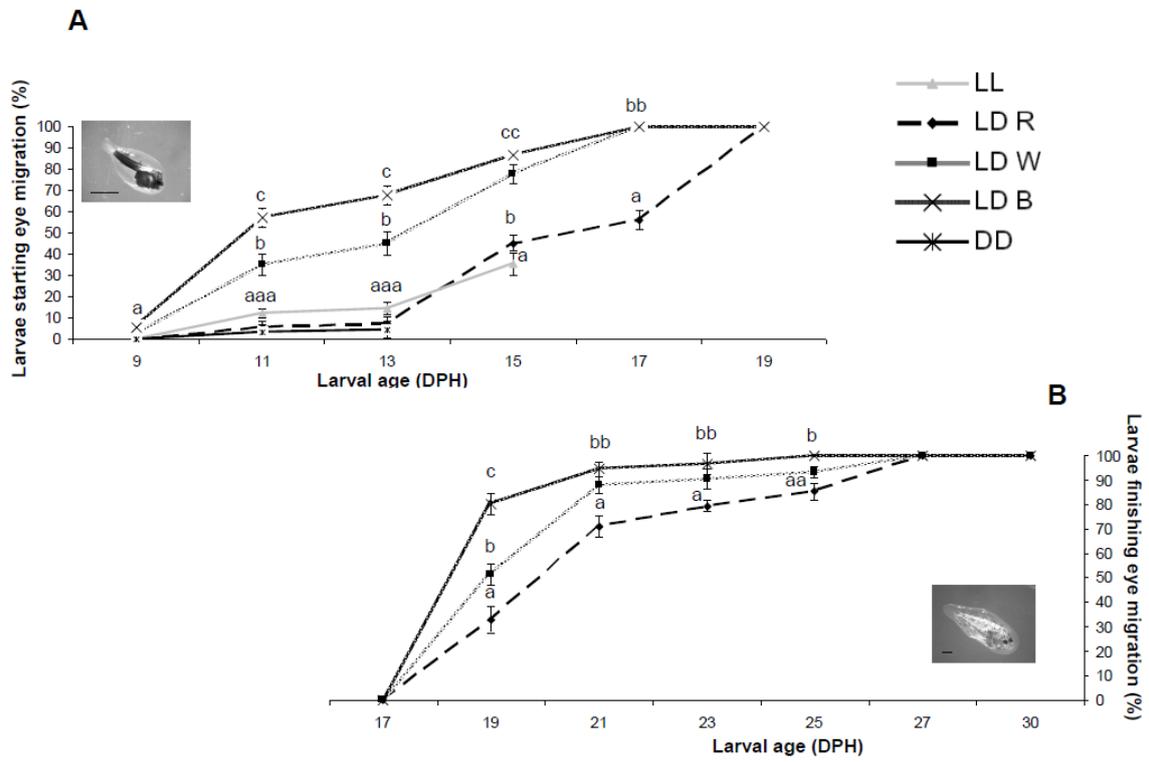
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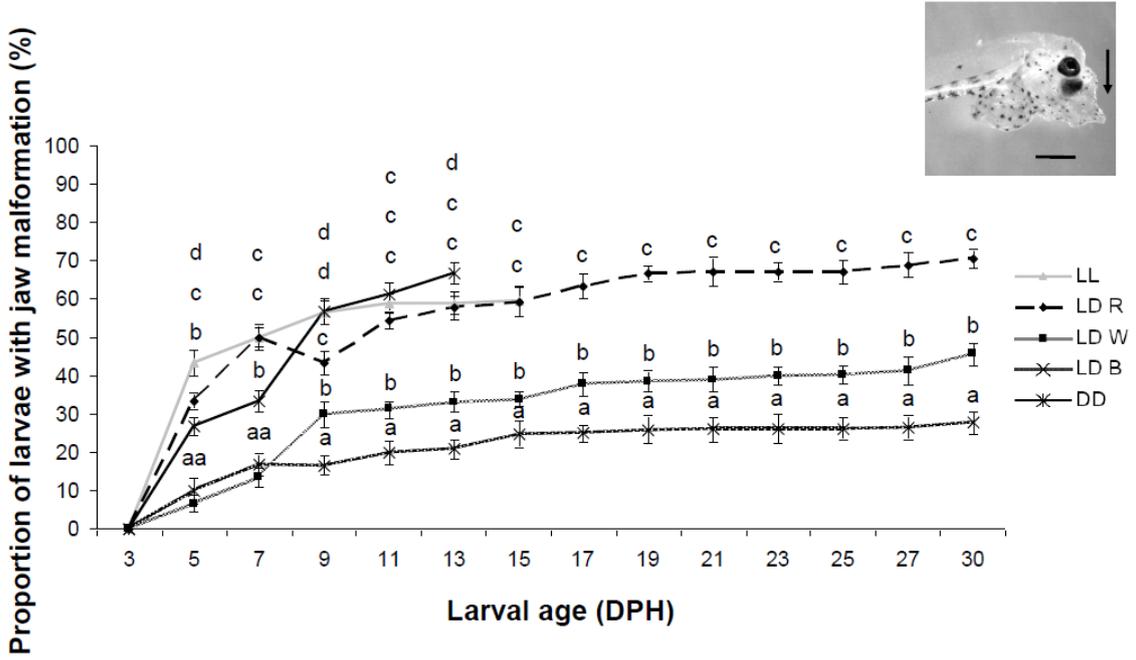
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697 Figure 9.



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