Murine models to investigate the influence of diabetic metabolism on the development of atherosclerosis and restenosis

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1. ABSTRACT

Atherosclerosis and related forms of cardiovascular disease (CVD) are associated with several genetic and environmental risk factors, including hypercholesterolemia, diabetes mellitus (DM), hypertension, obesity and smoking. Human DM is a multi-system disorder that results from progressive failure of insulin production and insulin resistance. Most diabetic patients die from complications of atherosclerosis and CVD, and DM is also associated with increased risk of restenosis post-angioplasty. Furthermore, the incidence of DM, particularly type 2-DM, is expected to increase significantly during the next decades owing to the unhealthy effects of modern life-style habits (e.g., obesity and lack of physical exercise). Thus, it is of utmost importance to develop novel preventive and therapeutic strategies to reduce the social and health-care burden of CVD and DM. Although a number of physiological alterations thought to promote atherosclerosis have been identified in diabetic patients, the precise molecular mechanisms that link DM and atherosclerosis are largely unknown. Thus, the aim of this review is to discuss current murine models of combined DM and atherosclerosis and to explore how these experimental systems are being utilized to gain mechanistic insights into diabetes-induced neointimal lesion development, as well as their potential use in evaluating the efficacy
of new therapies. Our discussion includes models generated by streptozotocin treatment and those resulting from naturally occurring or targeted mutations in the mouse.

2. DIABETES AND ATHEROSCLEROSIS

Despite major efforts during the last decades to identify cardiovascular risk factors (CRFs) and to elucidate the cellular and molecular mechanisms involved in the initiation and progression of the atherosclerotic plaque, atherosclerosis and associated CVD are the major cause of mortality and morbidity in Western societies, and their incidence in developing countries is increasing at alarming rates (1). Experimental and clinical studies have revealed that atherosclerosis is a chronic inflammatory disease resulting from the interaction between atherogenic stimuli and elements of the arterial wall (1,2). Native and adaptive immune mechanisms seem to play major roles in the aetiology and pathogenesis of atherosclerosis (3,4). The inflammatory process leads to the development of complex lesions and rupture of atherosclerotic plaques at advanced disease states provokes thrombus formation and acute clinical complications (i.e. stroke and myocardial infarction).

In addition to predisposing genetic factors, a number of modifiable CRFs are known to promote the development of atherosclerosis and CVD, including hyperlipidemia, hypertension, obesity, smoking, and DM (1,2,4-6). There is a wealth of clinical data demonstrating a relationship between the two major forms of DM (type 1 or insulin-dependent, and type 2 or non-insulin-dependent) and atherosclerosis and its clinical complications, with diabetic patients exhibiting a 2 to 10-fold higher risk of macrovascular disease than the general population (7-10) (see also http://www.who.int/diabetes/en). DM affects approximately 100 million persons worldwide, with 90-95% patients suffering from type 2-DM, 80% of which die from complications of diabetes (7-10). DM is often linked to the pathophysiology known as the metabolic syndrome, which combines glucose intolerance, hyperlipidemia, obesity, and hypertension (11). This coalescence of unfavorable health factors affects approximately one quarter of the population in industrialized countries and is associated with a marked increase in the risk of CVD. Remarkably, the incidence of type 2-DM is expected to increase by 165% between 2000 and 2050 (11), representing the plague of the 21st century, caused by increases of obesity and aging in the population (12). It is now estimated that the metabolic syndrome has a greater detrimental effect on our overall health as a society than cancer. Therefore, there is an enormous need for therapeutic and preventive measures to control this epidemic of metabolic diseases (13,14).

Human DM is a multi-system disorder characterized by a reduced utilization of glucose in peripheral tissues as a result of insulin resistance and/or the progressive failure of insulin production by pancreatic islets (12). Type 1-DM is a chronic autoimmune disease triggered by the autoimmune destruction of pancreatic insulin-producing β-cells (15). Human type 1-DM is frequently associated with polymorphisms in HLA class II, although environmental factors such as pathogens, toxins, drugs and especially viruses have been implicated (15-18). To some extent, it is possible to identify individuals at high risk for type 1-DM because circulating auto-antibodies are present years before the onset of clinical symptoms. By contrast, type 2-DM occurs when the pancreas fails to produce sufficient insulin to cope with an increasing metabolic demand; this may reflect an acquired defect of insulin secretion or synthesis and/or a decrease in β-cell mass (19). In adult humans, β-cell mass is plastic and the balance between insulin supply and metabolic demand is maintained by adjustments to β-cell growth and survival. However, eventually this β-cell adaptation fails since type 2-DM patients show a decrease in the β-cell mass. Type 2-DM is a condition that usually co-exists with the metabolic syndrome which is characterized by abdominal obesity, hyperglycemia, hyperinsulinemia, insulin resistance, hypertension, dyslipidemia, and microalbuminuria (11,20,21-23). Recently, dyslipidemia has also been found in a subset of patients with type 1-DM that have elevated intra-abdominal fat accumulation and obesity (24). Pre-atherogenic lipid alterations in the plasma of DM patients that are expected to accelerate atherosclerosis include: (1) increased triglyceride (TG) and free fatty acid (FFA) levels, (2) decreased content of cholesterol bound to high-density lipoproteins (HDL-cholesterol), and (3) the presence of the pro-atherogenic small dense low-density lipoproteins (sdlDLs), which has been used as a marker for the metabolic syndrome and type 2-DM (20,21,22,25). These alterations are hypothesized to produce endothelial dysfunction, which triggers atherosclerosis and CVD (26).

DM provokes other vascular complications that accelerate atherosclerosis and involve different cell types in the arterial wall, such as endothelial cells (ECs), vascular smooth muscle cells (VSMCs) and platelets (8,11,27). DM triggers EC dysfunction by impairing endothelium-dependent (nitric oxide-mediated) vasodilatation (8,28). Hyperglycemia inhibits the production of nitric oxide (NO) by blocking endothelial NO synthase (eNOS) activation, and also increases the production of reactive oxygen species (ROS) in ECs and VSMCs, which in turn activate receptors for advanced glycation end-products (RAGE). Lipid abnormalities in DM, like increased very low-density lipoprotein (VLDL) and FFA plasmatic levels, can promote growth of atherosclerotic lesions by inducing transcription factors involved in the expression of proatherogenic molecules (i.e. adhesion molecules, chemokines and other proinflammatory mediators). When nascent fatty streaks form, VSMCs migrate into the intimal lesion and stabilize the atheroma by producing collagen. Diabetic ECs display increased production of cytokines that lower in VSMCs the production of collagen an additional extracellular matrix component that confer plaque stability. Moreover, DM increases the production of metalloproteinases that lead to breakdown of collagen. Thus, the combination of reduced collagen synthesis and increased collagen breakdown predisposes diabetics to plaque rupture and higher risk of thrombus formation (8).
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Hyperglycemia may also facilitate plaque instability and rupture through direct actions on VSMCs. Glycated LDLs activate VSMC migration; however, oxidized glycated LDLs can induce VSMC apoptosis thus reducing collagen content within the lesion (8,29). Hyperglycemia also enhances thrombogenesis by activating platelets and reducing production of endogenous platelet inhibitors. Moreover, DM patients display an increase in blood clotting due to impaired fibrinolytic activity, increased levels of procoagulants (i.e. tissue factor) and coagulants (i.e. factor VII), and decreased endogenous anticoagulant levels (such as antithrombin III and protein C). In summary, DM promotes atherosclerotic lesion formation, increases plaque instability, and favors the formation and persistence of thrombi (8,29).

The next sections discuss current mouse models which have been used to investigate the relationship between DM and atherosclerosis/restenosis, including naturally occurring mouse mutants and genetically-engineered mice with phenotypes similar to human DM (Table 3 and 4). Because the mouse as a species is rather resistant to the development of atherosclerosis (30,31), most of the current models of diabetic atherosclerosis are based on the induction of DM by various means in genetically-modified mice with a propensity to atherosclerosis.

3. MOUSE MODELS OF TYPE 1-DIABETES COMBINED WITH ATHEROSCLEROSIS

3.1. Streptozotocin treatment

Numerous animal models of type 1-DM and CVD are based on treatment with streptozotocin (STZ), a monofunctional nitrosourea derivative isolated from Streptomyces achromogenes with cytotoxic effects on pancreatic β-cells (32,33). However, it is important to note that the action of STZ is not restricted to β-cells (33). Rats and dogs treated with STZ develop DM but display increased levels of protein kinase C (PKC) and diacylglycerol (DAG) in the heart and aorta; administration of insulin normalized glucose levels in plasma and prevented the concomitant increases in PKC and DAG (34,35). These findings suggest that chronic activation of the PKC-DAG pathway by hyperglycemia facilitates the development of diabetic vascular complications.

Kunjathoor et al. investigated the effect of STZ-induced diabetes on atheroma development in two different strains of mice (C57BL/6 and BALB/c) fed either rodent chow or an atherogenic promoting diet for 12-20 weeks (36). Regardless of strain and dietary regimen, STZ treatment induced sustained hyperglycemia and a modest reduction in plasma insulin levels. Whereas normoglycemic and hyperglycemic fed C57BL/6 mice displayed fatty streak aortic sinus lesions of similar size, hyperglycemic BALB/c mice fed the atherogenic diet displayed a 17-fold increase in the area of atherosclerotic lesions as compared with normoglycemic fat-fed BALB/c controls. Moreover, correlations were noted between lesion size and plasma glucose levels in the BALB/c strain and between lesion size and plasma cholesterol in C57BL/6 mice. Based on these observations, the authors of this study suggest that hyperglycemia as opposed to hyperinsulinemia contributes heavily to risk of atherosclerosis.

Several laboratories have studied the effect of STZ treatment on genetically-modified mice with a strong predisposition to atherosclerosis. Mice deficient for apolipoprotein E (apoE-KO), which develop severe hypercholesterolemia and spontaneous atherosclerosis on a normal chow diet (37,38), exhibit accelerated atherosclerosis when DM is induced by STZ (39). This is associated with increased macrophage-foam cell formation as measured by the cell lipid peroxidation and capacity to take up oxidized LDL (oxLDL). This may be caused by a direct effect of hyperglycemia on macrophages which exhibit a glucose-dependent elevation of peroxide content and increased oxLDL uptake associated with induction of the mRNA for scavenger receptor CD36 (40).

STZ-induced type 1-DM has also been superimposed on mice deficient for the LDL receptor (LDLR-KO), which develop hyperlipidemia and atherosclerosis when challenged with a Western-type diet (41). Reaven et al. (42) fed LDLR-KO mice with a diet containing 0.75% cholesterol. Mice were left untreated or received STZ and a low-dose of insulin to prevent excessive mortality and extreme elevations in TG. After six months of high-fat feeding, diabetic STZ-treated LDLR-KO mice exhibited enhanced arterial production of advanced glycation end (AGE) products and higher levels of blood glucose and VLDL cholesterol as compared with non-diabetic controls. However, the degree of aortic atherosclerosis was similar in both groups of mice, suggesting that hyperglycemia and enhanced AGE formation do not contribute significantly to atherogenesis in LDLR-KO mice. In marked contrast, subsequent studies by Keren et al. (43) and Vikramadithyan et al. (44) have reported that STZ-treated LDLR-KO mice placed for 6-12 weeks on a high-fat diet exhibit higher levels of plasma glucose, cholesterol and TG and develop larger atherosclerotic lesions in the aortic root than non-diabetic controls (43,44). These discrepancies may be related to different dietary regimens (e.g., long versus short time of fat-feeding) and/or a possible protective action of the low-dose insulin treatment used by Reaven et al. (42). Keren et al. (43) demonstrated that both humoral and cellular immune responses to heat shock protein 65 (HSP65) were more pronounced in STZ-injected LDLR-KO mice, and splenocytes from these animals displayed increased production of the T-helper (Th)-1 cytokine γ-interferon when challenged in vitro with HSP65. On the other hand, Vikramadithyan and co-workers (44) found that aortic atherosclerosis was further increased in STZ-treated LDLR-KO mice overexpressing the human aldose reductase, and peritoneal macrophages from these mice expressed more scavenger receptors and had a greater uptake of modified lipoproteins than non-transgenic controls. The results of this study confirm the notion that hyperglycemia is toxic to larger arteries and suggest that aldose reductase might modify this pathological response.
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Kako et al. crossed transgenic mice expressing human apolipoprotein B (apoB) with mice heterozygous for lipoprotein lipase (LPL1) and/or with transgenic mice expressing human cholesteryl ester transfer protein (CETP) (45). These animals were generated to create a more human-like lipoprotein profile, given that expression of apoB is low and CETP is absent in wild-type mice. HuB/LPL1 mice displayed increased TG in VLDLs whereas triple mutant (HuB/LPL1/CETP) mice had decreased HDL and increased VLDL and IDL/LDL. Treatment with STZ did not alter lipid profiles or atherosclerosis in HuB or HuB/LPL1/CETP mice. However, STZ-treated HuB/LPL1 mice were more diabetic, severely hyperlipidemic due to increased cholesterol and TG in VLDL and IDL/LDL, and displayed more atherosclerosis. Thus, these results obtained with STZ-treated mice suggest that type 1-DM without hyperlipidemia does not accelerate atherosclerosis in this animal model.

3.2. The GP transgenic:LDLR-KO mouse

Oldstone and co-workers (46) generated transgenic mice which express the lymphocytic choriomeningitis virus (LCMV) glycoprotein (GP) as a self antigen under the control of the rat insulin promoter. Infection with LCMV triggers an immune reaction that specifically destroys the GP-expressing β-cells thus provoking rapidly the onset of type 1-DM. The subsequent breeding of these mice with LDLR-KO mice produced atherosclerosis-prone mice with type 1-DM (LDLR-KO:GP mice); analysis of this model has suggested that hyperglycemia and diabetes-associated lipid abnormalities have distinct effects on initiation and progression of atherosclerotic plaques (47,48). When LDLR-KO:GP mice are fed standard chow, they develop hyperglycemia, but not lipid abnormalities, and display accelerated lesion initiation as well as increased arterial macrophage accumulation without augmented proliferation, suggesting that hyperglycemia may represent a primary stimulus for neointimal macrophage accumulation by inducing their recruitment. Interestingly, insulin treatment corrected the hyperglycemia and reduced the size of early lesions in chow-fed mice. Thus, atheroma initiation is increased by induction of DM and not from a toxic effect related to viral destruction of pancreatic islets. On the other hand, fat-fed LDLR-KO:GP mice exhibited hyperlipidemia and advanced lesions with augmented macrophage proliferation and extensive intraluminal hemorrhages in a manner largely dependent on diabetes-induced dyslipidemia, since the degree of atherosclerosis was similar between hyperlipidemic diabetic and nondiabetic mice with comparable plasma cholesterol levels. Lamharzi et al. (48) have suggested that the induction of macrophage proliferation by the concerted action of hyperglycemia and hyperlipidemia may involve glucose-oxidized LDL-mediated phosphorylation of extracellular signal-regulated kinase (ERK) and protein kinase B/Akt, and that this mitogenic effect of glucose-oxidized LDLs is mediated by CD36 and by protein kinase C- and phosphatidylinositol 3-kinase-dependent Erk activation.

3.3. The non-obese diabetic (NOD) mouse

The NOD mouse is a model of type 1-DM in which the autoimmune process culminates in spontaneous hyperglycemia caused by the destruction of β-cell islets (49,50). Keren and co-workers (51) found that two distinct formulations of Western-type diet were insufficient to elicit atherosclerosis in the aortic sinus of diabetic and nondiabetic female NOD mice even in the presence of hyperlipidemia and increased susceptibility to copper induced LDL oxidation.

4. MOUSE MODELS OF TYPE 2-DIABETES COMBINED WITH ATHEROSCLEROSIS

Normal blood glucose levels are maintained by a balance between glucose absorption in the gastro-intestinal tract, production by the liver and uptake by peripheral tissues, all of which are regulated by insulin signaling through the insulin receptor (INSR) (52). Dysregulation of these physiological processes leads to insulin resistant states and increased post-pandrial glucose and lipid levels. A detailed discussion of knockout models which have provided novel insights into insulin action and the development of insulin resistance and DM is beyond the scope of this review and can be found elsewhere (53). This section discusses available mouse models of type 2-DM and atherosclerosis which are based either on the use of ‘diabetogenic’ and ‘atherogenic’ diets, naturally occurring mutations which induce obesity and diabetes, genetically-modified mice, or a combination of the above (Table 3).

4.1. Diabetogenic diets to induce atherosclerosis in susceptible mouse strains

Several studies have suggested that both genetic and dietary factors determine susceptibility to obesity and type 2-DM. For example, while C57BL/6 mice develop obesity, hyperglycemia, hyperinsulinemia and insulin resistance when challenged with a diabetogenic diet enriched in fat and sucrose (54-56), A/J mice maintained on the same dietary regimen are resistant to the weight gain and metabolic perturbations noted in C57BL/6 mice (57). On the other hand, Schreyer et al. (58) found that maintenance on an atherogenic diet (enriched with fat and cholesterol) for 14 weeks did not induce hyperglycemia, hyperinsulinemia or obesity in strains C57BL/6, C3H/He, BALB/c nor in seven recombinant inbred strains. Moreover, this study did not reveal any significant correlation between susceptibility to atherosclerosis and fasting insulin or glucose levels, or glucose clearance following short-term insulin or glucose treatment. These authors also found that a diabetogenic diet enriched in fat and sucrose but lacking cholesterol and bile acids induced obesity, diabetes and 2-fold increases in plasma lipoprotein concentrations in C57BL/6 mice. Remarkably, 40% of C57BL/6 mice fed the diabetogenic diet exhibited small aortic fatty streaks that were not observed in mice fed standard rodent chow. Because diabetic C57BL/6 mice developed both hyperglycemia and hyperinsulinemia, it is not possible to attribute aortic atheroma development to one particular diabetic parameter. Thus, it is unclear from these studies whether atherosclerosis induced by the diabetogenic diet was a consequence of increased plasma lipids or the diabetic metabolism, or a combination of both. It is also noteworthy that the atherogenic and diabetogenic diets had...
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differential effects on the lipid profiles of C57BL/6 mice, thus suggesting that different dietary components regulate lipid transport genes in distinct ways.

Dietary studies in the atherosclerosis-prone LDLR-KO and apoE-KO mouse models have yielded conflicting results. Merat et al. (59) analyzed the effects of a fructose-rich diet, a fat-enriched (Western), or standard chow diet on male C57BL/6x129SV (5th generation backcrossed) LDLR-KO mice. Compared to the chow-fed group, mice fed either experimental diet during 5.5 months developed high hypercholesterolemia, but increased body weight, insulin resistance and hyperinsulinemia were only induced by the Western diet. Surprisingly, these insulin-resistant mice had significantly less atherosclerosis than the non-insulin-resistant, fructose-fed mice, thus indicating that insulin resistance and hyperinsulinemia do not enhance atherosclerosis in severely hypercholesterolemic male LDLR-KO mice. On the other hand, Schreyer et al. (56,60) noted that feeding a fat- and sucrose-enriched diet for 16 weeks produced the largest effects on adiposity, hyperglycemia, hypertriglyceridemia, hypercholesterolemia, and hyperleptinemia in male LDLR-KO compared with wild-type mice (both C57BL/6 strain); atherosclerotic lesions in diabeticogenic diet-fed LDLR-KO mice were increased 3.7-fold over chow fed values. These authors also included in their studies male C57BL/6 apoE-KO mice fed a diabeticogenic diet which were resistant to changes in glucose and lipid homeostasis and to accelerated atherosclerosis despite the development of obesity. Discrepancies between the studies of Merat et al. (59) and Schreyer et al. (56,60) might be related to different genetic backgrounds of the mouse strains utilized (mixed C57BL/6x129SV versus pure C57BL/6), and/or differences related to the carbohydrate composition in the experimental diets (sucrose-rich versus fructose-rich). Nevertheless, taken all together, these findings demonstrate that the metabolic alterations induced by the loss of LDLR or apoE have profound and distinct consequences on the propensity to develop dyslipemia, diet-induced obesity and type 2-DM phenotypes and on susceptibility to atherosclerosis in C57BL/6 male mice.

4.2. Lipocalin-type prostaglandin D(2) synthase (L-PGDS)-deficient mice

Upregulation of the L-PGDS protein in patients with type 2-DM and atherosclerosis has been suggested as a protective mechanism to alleviate the cardiovascular problems associated with DM. Ragoila et al. (61) found that L-PGDS-null mice become glucose intolerant and develop insulin resistance at an accelerated rate compared to the C57BL/6 control strain. L-PGDS-null mice have larger adipocytes than controls, and cultured wild-type and L-PGDS-deficient VSMCs exhibit insulin-stimulated adhesion kinase expression levels. Remarkably, feeding a diabetogenic high fat diet for 20 weeks provoked nephropathy and an aortic thickening reminiscent of the early stages of atherosclerosis only in L-PGDS-deficient mice. Thus, a thorough characterization of the L-PGDS-null mouse may yield significant insight into the mechanisms underlying insulin resistant- and DM-related atherosclerosis and nephropathy.

4.3. Gold thioglycolate (GTG)-induced obesity-related type 2-diabetes in the apoE-KO mouse model

Obesity and insulin resistance can be induced in the mouse by a single peripheral injection of GTG, a neurotoxic glucose analogue which rapidly destroys within 24 hours leptin receptor-positive hypothalamic neurons, including those that regulate energy expenditure and food intake (satiety) (62). Lyngdorf et al. (63) have analyzed the consequences of GTG injection on atherosclerosis development in female apoE-KO mice fed ad libitum standard rodent chow. Based on their body weight progression after injection, mice were classified into two groups: non-responders (n=10) which followed a growth curve similar to that of control mice, and responders (n=12) which displayed a marked long-term weight gain. Additional features of type 2-DM in the GTG-sensitive mice included insulin resistance, hyperinsulinemia, hyperglycemia and hypertriglyceridemia. Paradoxically, obese and diabetic GTG-responding apoE-KO mice developed less atherosclerosis than either lean GTG-non-responders or saline-injected ad libitum fed apoE-KO mice. Based on these findings, Lyngdorf et al. have raised concerns regarding the usefulness of mouse models in studying the relation of obesity-related type 2-DM to atherosclerosis. However, although the destruction of hypothalamic neurons is the only known action of GTG, the possibility that diminished atherosclerosis in GTG-responding mice reflects systemic effects of this drug cannot be excluded.

4.4. Leptin receptor-deficient (Lepr<sup>ob/db</sup>) and leptin-deficient (Lepr<sup>ob/ob</sup>) mouse models

The spontaneous autosomal recessive diabetes mutation (db) was first detected in progeny of the C57BL/KsJ strain at the Jackson Laboratory (64), and animals with the same phenotype were subsequently described and named obese (ob) mice (65). It has been established that ob and db encode for leptin and its receptor, respectively (65-67). Mice homozygotes for db (Lepr<sup>ob/db</sup>) and ob (Lepr<sup>ob/ob</sup>) mutations display similar obesity-diabetes phenotypes, including early-onset obesity, extreme insulin resistance, more efficient conversion of food to lipid and a slower rate of catabolism on fasting (64,65,68). These spontaneous mutations convey a selective advantage for heterozygous db/+ and ob/+ mice, as they survived a prolonged fast significantly longer than normal homozygotes (+/+), and did not display pathological manifestations when food was abundant in populations subjected to alternating periods of supply and deprivation (68). It has been recently shown that ablation of ghrelin improves the diabetic but not obese phenotype of Lepr<sup>ob/db</sup> mice (69).

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Hasty and co-workers (70) were the first to report augmented aortic atherosclerosis in the double mutants Lepr<sup>ob/db</sup>-LDLR-KO maintained on normal chow as compared with different control groups: wild-type LDLR-KO, Lepr<sup>ob/db</sup>-LDLR<sup>-/+</sup>- mice. This phenotype of Lepr<sup>ob/db</sup>-LDLR-KO mice was accompanied by severe hypercholesterolemia and hypertriglyceridemia caused by a large increase in apoB-containing broad-β remnant lipoprotein fraction. These authors provided

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evidence to suggest that hypertriglyceridemia and hypercholesterolemia in the double mutant Lep<sup>ob/ob</sup>:LDLR-KO mice are caused by distinct mechanisms and that leptin might have some impact on plasma cholesterol metabolism, possibly through an LDLR-independent pathway. They concluded that LDL cholesterol is a stronger atherogenic risk factor than are the combined effects of obesity, DM and insulin resistance when mice are fed a normal diet.

Liang et al. (71) found increased CD36 expression in thiglycollate-elicited peritoneal macrophages (TGEM) from Lep<sup>ob/ob</sup> mice, possibly caused by defective insulin signaling. This was associated with increased binding and uptake of oxLDL and acetylated LDL (acLDL) in vitro, thus suggesting a mechanism that might contribute to augmented foam cell formation and accelerated atherosclerosis in insulin resistant and DM pathological states. Consistent with this notion, in vivo administration of PPAR-γ activators to Lep<sup>ob/ob</sup> macrophages improved insulin resistance, decreased CD36 protein expression and diminished oxLDL uptake (71). Kjerrulf et al. (72) also reported increased CD36 expression and acLDL uptake in TGEMs using this same model. Surprisingly, these authors found diminished expression of CD36 (and SR-A) and reduced acLDL uptake and cholesterol ester accumulation in cultures of Lep<sup>ob/ob</sup> resident peritoneal macrophages, thus highlighting important differences between these two common sources of macrophages.

Gruen et al. (73) performed an exhaustive comparison of Lep<sup>ob/ob</sup>:LDLR-KO and Lep<sup>ob/ob</sup>:apoE-KO mice maintained on standard rodent chow. Compared to simple mutant control mice, body and fat pad weight and total plasma cholesterol levels were increased to the same extent in both models of double mutant mice. Interestingly, neither model showed increased plasma glucose. Lep<sup>ob/ob</sup>:apoE-KO and Lep<sup>ob/ob</sup>:1LDLR-KO mice transported cholesterol primarily on VLDL and on LDL, respectively. Other differences included lower (55%) TGs and higher non-esterified fatty acid (NEFA) (1.5-fold) levels in the plasma of Lep<sup>ob/ob</sup>:apoE-KO mice. Atherosclerosis in Lep<sup>ob/ob</sup>:apoE-KO and Lep<sup>ob/ob</sup>:LDLR-KO mice was enhanced compared to single mutant apoE-KO and LDLR-KO controls. Remarkably, both aortic atheroma size (3.2-fold) and collagen content (7.7-fold) were increased in Lep<sup>ob/ob</sup>:apoE-KO compared to Lep<sup>ob/ob</sup>:LDLR-KO mice. Lesion burden was not associated with total plasma cholesterol and TG nor with individual lipoprotein pools in either animal model. In contrast, atheroma size was positively correlated with body weight and NEFA only in Lep<sup>ob/ob</sup>:LDLR-KO mice, and with plasma insulin in both mouse models. Thus, despite the significant differences in atheroma size between leptin-deficiency in LDLR-KO vs. apoE-KO, these results support the notion that several typical features of DM (hyperinsulinemia, increased body weight and elevated plasma levels of NEFA) aggravate atherosclerosis in the mouse, even in the absence of hyperglycemia.

Wu et al. (74) reported that Lep<sup>ob/ob</sup>:apoE-KO mice exhibit features of the metabolic syndrome and type 2 DM, including obesity, hyperglycemia, hyperinsulinemia, and dyslipidemia. When fed a regular chow diet, aortic lesion size and lipid content were increased in Lep<sup>ob/ob</sup>:apoE-KO mice compared to age-matched apoE-KO littermates. Atherosclerosis in Lepr<sup>db/db</sup>:apoE-KO mice was further aggravated by fat feeding. More recently, Wendt et al. (75) have also noted accelerated aortic atherosclerosis in diabetic Lepr<sup>db/db</sup>:apoE-KO mice fed normal chow as compared with non-diabetic controls.

Lepr<sup>ob/db</sup> mice fed regular chow displayed a prominent peak in the LDL range, consisting almost exclusively of sLDL, that was absent in non-diabetic controls, although HDL was the predominant species in both groups of mice (76). Upon administration of a Western type diet, the size distribution of lipoproteins was unchanged in control mice whereas fat-fed Lepr<sup>ob/db</sup> mice develop a marked hypercholesterolemic response which was the result of a massive increase in the LDLR region, that was absent in non-diabetic controls, although HDL was the predominant species in both groups of mice (76). Upon administration of a Western type diet, the size distribution of lipoproteins was unchanged in control mice whereas fat-fed Lepr<sup>ob/db</sup> mice as a model to investigate the pathogenesis and treatment of diabetic dyslipidemia.

Gruen et al. (77) found that both Lep<sup>ob/ob</sup> and Lepr<sup>db/db</sup> mice display increased plasma HDL levels and accumulate a unique lipoprotein referred to as LDL/HDL1. By analyzing the phenotype of apolipoprotein A-I-deficient (apoA-I-KO) mice homozygous for either the db or the ob mutation, these authors demonstrated that these obese apoA-I-KO mice had dramatically decreased levels of HDL, although the LDL/HDL1 particle persisted as small LDLs and large HDLs. Moreover, they provide evidence suggesting that the maturation and removal of large HDLs depends on the integrity of a functional axis consisting of apoA-I, hepatic lipase, and scavenger receptor class B type I (SR-BI).

In a recent study, Li et al. (78) observed enhanced proatherogenic responses in both short-term cultured VSMCs and macrophages derived from obese, insulin-resistant and diabetic Lepr<sup>db/db</sup> mice, including increased expression of cyclooxygenase-2 and 12/15 lipooxygenase as well as key inflammatory cytokines and chemokines (e. g., IL-1β, IL-6, IL-8, IL-12, TNFα, MCP-1, and IP-10), increased oxidant stress, and activation of key signaling kinases (e. g., Src, ERK1/2, Akt, p38) and transcription factors involved in the regulation of atherogenic and inflammatory genes (e. g., cAMP response element-binding protein, nuclear factor-kB). Remarkably, aortas derived from Lepr<sup>db/db</sup> mice also exhibited increased expression of inflammatory genes, and VSMCs from these animals had enhanced migratory activity and adhesion to monocytes.

4.5. Defective insulin signaling and atherosclerosis: Insulin receptor and insulin receptor substrate 2-null mouse

The INSR is a cell surface receptor that belongs to the family of growth factor receptor tyrosine kinases. The propagation of the signal generated by insulin through the insulin receptor substrate (IRS)/phosphatidylinositol 3-kinase (PI3K) pathway plays a key role in glucose and lipid metabolism. Among other effects, activation of the serine-threonine kinase Akt stimulates
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glycogen synthesis and inhibits lipolysis while PKC activation has been linked to accumulation of TGs in micelles, a hallmark of insulin resistance (79). INSR-null mice exhibit growth retardation, a progressive increase in glucose levels and severe hyperinsulinemia. Within a few days of birth, pancreatic beta cell failure is followed by death of the animals caused by diabetic ketoacidosis (80). The severity of this phenotype indicates that the INSR plays a major role in postnatal glucose homeostasis (79,80).

IRS proteins mediate the majority of metabolic and growth-promoting actions of insulin and IGF-1 and play a critical role in the regulation of beta cell function and in the maintenance of glucose homeostasis (81). Analysis of mice deficient for the four IRS genes has provided important information about the signaling mechanisms regulated by insulin (82). IRS-1 deletion in mice retards somatic growth and produces mild insulin resistance without type 2-DM (83,84). Insulin signaling in liver is normal in IRS-1-KO mice suggesting that is mainly regulated by IRS-2 (85). Disruption of the IRS-2 gene produces female infertility and pathological alterations very similar to type 2–DM and metabolic syndrome, including insulin resistance, hyperinsulinemia, glucose intolerance, hypertension, and moderate hyperlipidemia (86-88). At 6-8 weeks of age, male IRS-2-KO mice exhibit severe diabetes as the reduced β-cell mass fails to produce sufficient insulin (86,87). Furthermore, deletion of IRS-2 produces female obesity owing to a dysregulation of appetite and leptin sensitivity, demonstrating that IRS-2 is required for central regulation of nutrient homeostasis and obesity (88-90). The importance of IRS-2 signals for the development of pancreatic β-cells and β-cell function has been further established by generation of RIP-IRS2 transgenic mice; overexpression of IRS-2 specifically in the pancreas of IRS-2-deficient mice and Lepob/ob mice restored β-cell mass and thus, glucose tolerance (91). In contrast to the phenotypes of IRS-1-KO and IRS-2-KO mice, deficiency for either IRS-3 or IRS-4 does not appear to alter growth or metabolism in the mouse (92-95).

Disruption of insulin signaling specifically in macrophages has been used recently to evaluate the role of this pathway in macrophage foam cell formation and diabetes-accelerated atherosclerosis. Transplantation of INSR-null bone marrow into irradiated LDLR-KO mice increased lipid uptake through the upregulation of CD36 and SRA, thereby aggravating atherosclerosis (96). In agreement with this study, previous studies in mice (71) and humans (97,98) have linked defective insulin signaling to increased lipid accumulation in macrophages and foam cell formation via enhanced CD36 receptor expression. By contrast, Baumgartl and co-workers (99) reported that macrophage-specific deficiency of either INSR or IRS-2 ameliorates modulation of glucose and insulin signaling in macrophages plays a direct role in lipid deposition and foam cell formation in the vessel wall.

Global IRS-2 deficiency in fat-fed apoE-KO mice aggravates atherosclerosis compared with apoE-KO mice with intact IRS-2, although both models displayed similar levels of hypercholesterolemia (~2500 mg/dL in Baumgartl et al. (99) and >600 mg/dL in González-Navarro et al. (100)). Clough et al. (101) also found accelerated atherosclerosis in apoE-KO mice heterozygous for IRS-2. In contrast, atherosclerotic lesions were undetectable in fat-fed hyperglycemic and mildly hypercholesterolemic (~280 mg/dL) IRS-2-KO mice with intact apoE (100). Although both Baumgartl et al. (99) and González-Navarro et al. (100) reported impaired glucose tolerance in apoE-IRS-2-KO mice, only the former found significantly elevated fasting blood glucose concentration in these animals. On the other hand, circulating insulin levels predicted atherosclerotic lesion burden in apoE-IRS-2-KO mice (100), as previously reported in Lepob/leptin-deficient and apoE-IRS-2-KO mice (73). These results suggest that hyperinsulinemia resulting from IRS-2-deficiency contributes to increased atherosclerosis when combined with severe hypercholesterolemia even in the absence of overt hyperglycemia (apoE-IRS-2-KO mice). Thus, these observations implicate IRS-2 as an important modulator of murine hypercholesterolemia-dependent atherosclerosis. Future studies are necessary to determine whether IRS-2 dysfunction may promote atherosclerosis in normoglycemic, pre-diabetic patients with clinical manifestations of hyperinsulinemia and insulin resistance.

5. ANIMAL MODELS TO STUDY THE EFFECTS OF ALTERED LEPTIN/GLUCOSE/INSULIN SIGNALING ON NEointimal Thickening INduced BY Mechanical Injury of the Vessel WALL

Patients with DM are particularly prone to vessel re-narrowing (restenosis) after successful revascularization by percutaneous transluminal angioplasty and stent implantation (102-106). This section discusses the results of animal studies designed to investigate the effects of altered leptin, glucose, and/or insulin signaling on the development of obstructive neointimal lesions as induced by endovascular mechanical injury (Table 4).

5.1. Type 1-diabetes models

Different studies have yielded seemingly controversial results on the role of type 1-DM on mechanically-induced neointimal thickening. On the one hand, studies in alloxan-induced diabetic rabbits (107) and BB Wistar diabetic rats (108) demonstrated increased neointima development after balloon angioplasty of the carotid artery and aorta, respectively, compared with nondiabetic controls. In contrast, neointimal lesion size was unchanged after endovascular injury of the femoral artery of ins2Δlox/lox mice (109) and aorta of STZ-treated Sprague-Dawley rats (110), or even reduced in the carotid artery of hyperglycemic STZ-induced diabetic Wistar rats (111).

5.2. LeprΔdb/db mice
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As indicated above, obese type 2 diabetic doubly mutant Lepr<sup>db/db</sup>:apoE-KO mice exhibit accelerated atherosclerosis [74]. Surprisingly, neointimal formation after angioplasty was attenuated by approximately 90% in Lepr<sup>db/db</sup> compared with nondiabetic wild-type mice [109]. This phenotype correlated with diminished medial VSMC death at four hours after injury, but neointimal proliferation at two weeks post-injury appeared unaffected. It is noteworthy that arterial injury studies in the obese Zucker rats carrying a mutated leptin receptor have not been consistent, with two studies showing a 2-fold increase in neointimal thickening [110,112] and another study reporting no changes in lesion development in these animals compared with lean controls [113]. Further studies are thus required to unravel the relationship between leptin function and neointimal thickening in the setting of mechanical vascular injury and atherosclerosis in dyslipidemic animals.

5.3. IRS-deficient mice

Using the femoral artery cuff placement model of vascular injury [114], Kubota et al. [115] examined the effects of IRS-1 and IRS-2 deficiency on neointimal thickening. At both 8 and 20 weeks of age, the absence of IRS-2 markedly increased neointima formation (1.5-fold and 2.3-fold increase versus wild-type mice, respectively). In contrast, neointimal thickening was unchanged in 8-week-old IRS-1-KO and augmented only by 1.6-fold at 20 weeks of age as compared with wild-type mice. Based on the results of Western blot analysis, these authors have suggested that reduced IRS-2 protein expression in vessel tissue might account for the mild increase in neointima formation in 20-week-old IRS-1-KO mice. Comparison of risk factors for atherosclerosis indicated that the observed differences in neointimal thickening may be directly related to metabolic abnormalities of IRS-2-KO versus IRS-1-KO mice at both 8 and 20 weeks, including hypertriglyceridemia, hypercholesterolemia, hyperinsulinemia, and hypertension. Moreover, hyperglycemia was only seen in 20-week-old IRS-2-KO mice. This study provides the first evidence that intact IRS-2 signaling may protect against the development of neointimal lesions induced by mechanical injury.

5.4. Role of hyperinsulinemia

Using the rat carotid artery model of balloon angioplasty, Indolfi et al. [111] reported that insulin therapy significantly increases neointima formation in hyperglycemic STZ-induced diabetic rats. This effect of exogenous insulin was abrogated upon inhibition of cellular ras by intraluminal adenovirus-mediated transfection of the N17H-ras-negative mutant gene. Hyperinsulinemia also increases neointimal thickening in balloon-injured arteries of nondiabetic rats, as revealed pancreatic islet transplantation [111] and insulin infusions [116] studies. Remarkably, VSMC migration and proliferation, two key events in neointimal thickening, are enhanced in vitro by insulin [111,117].

6. EXPERIMENTAL THERAPIES FOR TREATING DIABETIC ATHEROSCLEROSIS

In this section, the therapeutic approaches that efficiently suppress diabetic atherosclerosis in the mouse are discussed (Table 5).

6.1. Inhibition of the AGE pathway

Animal and human studies indicate that AGEs and their receptor (RAGE) are upregulated in the vasculature and atherosclerotic lesions of diabetic individuals [118]. Park et al. [39] first reported that treatment of STZ-induced diabetic apoE-KO mice with a soluble extracellular ligand-binding domain of RAGE (sRAGE) that binds up AGEs suppresses dose-dependently diabetic atherosclerosis in a glyceremia- and lipid-independent manner. Subsequently, Buccarelli et al. [119] reported that sRAGE administration decreases parameters of inflammation and mononuclear phagocyte and VSMC activation and stabilizes established atherosclerotic plaques in the same animal model. Likewise, Forbes et al. [120] found that blockade of the RAGE-AGE axis by administration of the AGE cross-link breaker ALT-711, or the inhibitor of AGE formation aminoguanidine, reduced vascular AGE accumulation and diminished both plaque area and complexity in the thoracic and abdominal aortas of STZ-treated apoE-KO mice. Moreover, STZ-treated apoE-KO mice fed an AGE-enriched diet developed larger lesions at the aortic root than controls fed a low-AGE diet, in the absence of diet-related changes in plasma glucose, TGs, or cholesterol [121]. Thus, in STZ-treated diabetic apoE-KO mice, AGE-RAGE interactions contribute to accelerated atherosclerosis independent of changes in levels of glucose, insulin or lipids. More recently, Wendt et al. [75] extended these concepts to a murine model of type 2-DM by demonstrating that treatment of diabetic Lepr<sup>db/db</sup>:apoE-KO mice with sRAGE significantly reduces atherosclerosis in a glyceremia- and lipid-independent manner. Administration of sRAGE to nondiabetic Lepr<sup>db/db</sup>:apoE-KO controls also reduced atherosclerotic lesion area at the aortic sinus, suggesting that upregulation of RAGE ligands and RAGE occurs even in the nondiabetic, hyperlipidemic state. Similarly, sRAGE administration inhibited neointimal thickening after balloon angioplasty in both Zucker diabetic and nondiabetic rats, indicating that RAGE mediates neointimal formation in response to arterial injury [113]. Interestingly, RAGE also seems to be involved in impairment of angiogenesis in diabetic mice [122].

6.2. Blockade of the renin-angiotensin system

Candido et al. [123] demonstrated that accelerated atherosclerosis in STZ-treated apoE-KO mice correlates with a significant increase in aortic angiotensin-converting enzyme (ACE) expression and activity, and connective tissue growth factor and vascular cell adhesion molecule-1 expression. Administration of the ACE inhibitor perindopril inhibited these pathological alterations. Subsequently, these researchers demonstrated that the angiotensin II subtype 1 (AT1) receptor blocker irbesartan, but not clinical doses of the calcium channel antagonist amlodipine, suppresses the upregulation of aortic AT1 receptor expression...
and diabetes-associated atherosclerosis (124). More recently, they have reported that combined ACE and neutral endopeptidase inhibition in STZ-treated apoE-KO mice by omapatrilat administration reduced atherosclerosis and protected the animals from renal structural injury and albuminuria (125). The ACE inhibitor quinapril was equally effective at inhibiting aortic ACE activity and reducing atherosclerosis, but the modest anti-hypertensive response of omapatrilat correlated with superior renal protection.

6.3. **Imatinib**

Imatinib (also known as Gleevec, STI-571, and CGP57148B) is a low molecular weight inhibitor of the protein tyrosine kinase activity of both platelet-derived growth factor receptor (PDGF-R) subtypes, as well as that of Ab1, Bcr-Abl, and c-kit. This drug is currently being used clinically to successfully treat several forms of malignancies, in particular chronic myelogenous leukemia (126). Other therapeutic applications of imatinib have been demonstrated in animal models of vascular proliferative disease, including restenosis after balloon angioplasty (127), diet-induced atherosclerosis (128) and transplant atherosclerosis (129). Lasilla et al. (130) found that imatinib treatment prevents the development of atherosclerotic lesions and diabetes-induced inflammatory cytokine overexpression in the aorta of STZ-treated apoE-KO mice, suggesting a therapeutic approach to retard the development of diabetes-associated atherosclerosis.

6.4. **17β-estradiol**

Epidemiological data and animal studies indicate that estrogens markedly attenuate the risk of cardiovascular morbidity and death in premenopausal women (131). Using STZ-treated apoE-KO mice, Tse et al. (132) found that chronic administration of 17β-estradiol via a slow release pellet significantly decreased blood glucose and TG levels and effectively prevented diabetic-related atherosclerosis and premature calcified cartilaginous metaplasia in the aorta.

6.5. **Inhibition of 11β-hydroxysteroid dehydrogenase type 1 (11β-HSD1) activity**

11β-HSD1 raises the effective glucocorticoid tone above serum levels by converting cortisone into active cortisol in cells. Hermanowski-Vosatka et al. (133) reported that pharmacologic inhibition of 11β-HSD1 with a competitive inhibitor for cortisone has salutary effects on multiple aspects of the metabolic syndrome in both Western-type diet-induced obese mice (lowered body weight gain, fat pad weight, and plasma levels of insulin, fasting glucose, TGs, and cholesterol) and type 2-DM murine models (DIO and HF/SZT mice) (lowered fasting plasma glucose, insulin, glycation, TG, and FFA levels, as well as improved glucose tolerance). Moreover, 11β-HSD1 inhibition reduced serum cholesterol and TGs, diminished aortic accumulation of total cholesterol and slowed plaque progression in treated apoE-KO mice. Whether 11β-HSD1 inhibition reduces diabetic atherosclerosis is not known.

6.6. **The thromboxane A2 receptor antagonist S18886**

DM is associated with increased thromboxane A2 (TXA2) and prostaglandin endoperoxide receptor. The orally active TP receptor antagonist S18886, which is in clinical development for use in secondary prevention of thrombosis in patients affected by CVD, has been shown to inhibit the development of atherosclerosis in rabbits (134) and apoE-KO mice (135). Moreover, S18886 lessens enhanced aortic atherogenesis caused by STZ-induced DM in female apoE-KO mice without affecting the associated increase in hyperglycemia or hypercholesterolemia (136). This salutary effect of S18886 correlated with inhibition of intimal markers of inflammation associated with DM and less deterioration of endothelial function and eNOS expression. Furthermore, S18886 abrogated the induction of vascular cell adhesion molecule-1 and prevented the decrease in eNOS expression caused by high glucose in cultured human aortic ECs.

6.7. **Peroxisome proliferator-activated receptor (PPAR) agonists**

PPAR-α, γ, and δ are nuclear receptors that play major roles in the regulation of lipid metabolism, glucose homeostasis and inflammatory processes involved in the development of DM and atherosclerosis (137). Fibrates (e.g. fenofibrate, gemfibrozil) are PPAR-α agonists which are primarily used for the treatment of dyslipidemia based on their efficacy at reducing TGs and increasing the levels of HDL-cholesterol and large buoyant, less-atherogenic LDL cholesterol particles (138). Administration of fenofibrate to diabetic Leprdb/db,apoE-KO mice fed a Western type diet from 4 to 14 weeks of age reduced aortic atherosclerosis by 2.5- and 5.3-fold, respectively. However, fenofibrate only reduced total plasma cholesterol by approximately 30%, suggesting that this PPAR-α agonist may have a direct anti-atherogenic action on the vessel wall independently of its lipid lowering capacity (74). Likewise, gemfibrozil exerts anti-atherogenic effects in STZ-treated diabetic apoE-KO mice independently of significant changes in plasma glucose, glycated haemoglobin, cholesterol and insulin (139). This therapeutic effect of gemfibrozil correlated with the following changes in aortic tissue: 1) diminished expression of various NADPH oxidase subunits and reduced superoxide production; 2) reduced expression of proinflammatory genes, RAGE, angiotensin II receptor subtype 1, and metalloproteinase 9.

Rosiglitazone, an insulin sensitizing PPAR-γ agonist currently used for the treatment of type 2-DM patients, significantly attenuates atherosclerosis in STZ-treated diabetic apoE-KO mice (140); however, there were no significant changes in plasma glucose, insulin or cholesterol levels between treated and untreated diabetic mice. This inhibitory effect of rosiglitazone was associated with attenuated superoxide production, increased expression of the reverse cholesterol transport marker ABCA1 and reduced angiotensin II receptor gene expression in aortic tissue, and attenuated neointimal macrophage accumulation. These studies in the mouse suggest that PPAR-α and PPAR-γ agonists confer vascular protection independent of their effects on...
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glucose and lipid metabolism. These agents also attenuate diabetic kidney disease in STZ-treated apoE-KO mice (141). However, it is noteworthy that rosiglitazone appears to have little effect on cardiovascular endpoints in human (e.g. DREAM trial).

Zadelaar et al. have investigated the effect of the dual PPARalpha/gamma agonist tesaglitazar on atherosclerosis in insulin resistant and hypercholesterolemic ApoE*3Leiden transgenic mice (142). In fat-fed mice, tesaglitazar decreased plasma cholesterol by 20% compared with the HC group, caused a 92% reduction in atherosclerosis beyond that expected from cholesterol lowering, and induced a shift to less severe lesions with reduced number of adhering monocytes and macrophage-rich and collagen areas. Additionally, tesaglitazar inhibited the expression of inflammatory markers.

6.8. Antioxidant therapy: polyphenols, α-lipoic acid and paraoxonase 1 (PON1)

It is becoming increasingly evident that hyperglycemia-induced oxidative stress contributes to atherosclerosis and associated cardiovascular complications in diabetic patients. Recent animal studies indicate that dietary antioxidants (e.g., polyphenols and α-lipoic acid) may be a new therapeutic avenue for reducing diabetic atherosclerosis. Dietary polyphenols are natural antioxidants which appear to have beneficial effects on dyslipidemia and atherosclerosis (143). Several polyphenols, including resveratrol, apigenin, and S17834, induce the phosphorylation of AMP-activated kinase (AMPK) and its downstream target acetyl-CoA carboxylase (ACC) in cultured HepG2 hepatocytes (144). Moreover, the polyphenols prevented lipid accumulation induced by exposure of these cells to high glucose, whereas overexpression of constitutively active and dominant-negative AMPK mutants mimicked and abrogated these effects, respectively. Remarkably, S17834 administration to STZ-treated diabetic LDLR-KO mice inhibited hyperlipidemia and the acceleration of aortic lesion development, and this correlated with maintenance of AMPK and ACC phosphorylation and reduced accumulation of lipids in the liver (144). These observations suggest that hepatic AMPK inactivation plays an important role in the pathogenesis of hyperlipidemia in DM, and that polyphenols, by ameliorating dyslipidemia through AMPK activation, may ameliorate diabetic atherosclerosis.

Chronic treatment of fat-fed STZ-treated diabetic apoE-KO mice with the potent natural antioxidant α-lipoic acid significantly lowered markers of oxidative stress and produced a reduction of plasma glucose and accelerated the recovery of pancreatic insulin-producing β-cells (145). These effects were accompanied by an attenuation of the increase in plasma total cholesterol, atherosclerotic lesions, and the general deterioration of health caused by DM.

A frequent feature of the metabolic syndrome is low level of PON1, a HDL-associated antioxidant enzyme that prevents the oxidation of LDL particles. Acetoxys-mediated overexpression of human PON1 (AdPON1) in Lepob/LDLR-KO mice, an experimental model of metabolic syndrome, resulted in a 4-fold increase in PON1 activity and inhibited the development of atherosclerosis (146). The beneficial effect of AdPON1 was accompanied by reduced macrophage and oxidized LDL content and increased number of VSMCs within atherosclerotic lesions, but did not have overall effects on total plasma cholesterol and TG levels. Thus, without ameliorating dyslipidemia, AdPON1 inhibits atherosclerosis in diabetic hypercholesterolemic mice probably by reducing the amount of oxidized LDL in plasma and within the plaque.

7. CONCLUSIONS AND FUTURE PERSPECTIVES

Clinical studies have firmly established that type 1- and type 2-DM are associated with an increased risk for CVD. Moreover, given that the incidence of DM is increasing dramatically in developed countries, it is of utmost importance to identify suitable animal models for unraveling the molecular mechanisms mediating diabetic atherosclerosis in order to facilitate the development of novel preventive and therapeutic strategies. As we discuss extensively throughout this review, accelerated atherosclerosis has been demonstrated in available mouse models of both type 1- and type 2-DM. In addition to yielding insights into the molecular connections between diabetic metabolism and CVD, these models serve as valuable tools for evaluating therapeutic strategies.

A frequent observation in murine studies is that DM per se does not promote atherosclerosis in the absence of dyslipidemia. Indeed, diabetic mice present also dyslipidemia in the majority of experimental settings, thus making it difficult to differentiate between the effects of DM and those induced by aggravated lipid abnormalities. Although this may seem a limitation, it is noteworthy that deranged carbohydrate and lipid metabolism is a common feature in human diabetic patients, thus highlighting the relevance of available murine models of diabetic atherosclerosis. Several authors have attributed to hyperglycemia and hyperinsulinemia different importance in the development of accelerated atherosclerosis in type 1- and type 2-DM. Regardless of the peculiarities of different murine models, it appears certain that several features of type 2-DM/metabolic syndrome, including hyperinsulinemia, increased body weight and elevated plasma levels of NEFA, aggravate atherosclerosis in the mouse, even in the absence of hyperglycemia. On the other hand, it has been suggested that hyperglycemia may represent a primary stimulus of accelerated atherosclerosis in type 1-DM. Precisely how these aspects of diabetic metabolism predispose for the development of neointimal lesions has yet to be determined but clearly the possibility to perform biochemical and genetic manipulations in mouse models will provide the necessary tools to unravel the complex links between these important human diseases.

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**Key Words:** Atherosclerosis, Restenosis, Diabetes Mellitus, Irs, Metabolic Syndrome, Genetically-Modified Mice, Insulin Resistance, Chronic Inflammation, Hypercholesterolemia, Review

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Table 1. Proatherogenic plasma lipid alterations in diabetic patients

<table>
<thead>
<tr>
<th>Alteration</th>
<th>Mechanisms</th>
<th>Effects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hypertriglyceridemia</td>
<td>Increased plasma FFA level caused by augmented release from adipose tissue</td>
<td>Enhanced hepatic production of TG-rich VLDLs</td>
</tr>
<tr>
<td>Increased plasma sdLDL</td>
<td>Stimulation of lipase activity on TG-rich lipoproteins</td>
<td>Increased foam cell formation caused by augmented accumulation of oxLDL within the subendothelial space</td>
</tr>
<tr>
<td>Decreased plasma HDL</td>
<td>Stimulation of lipase activity on TG-rich HDL</td>
<td>Increased foam cell formation caused by decreased reverse cholesterol transport</td>
</tr>
</tbody>
</table>

Table 2. Major proatherogenic vascular alterations in diabetic patients

<table>
<thead>
<tr>
<th>Alteration</th>
<th>Mechanism</th>
</tr>
</thead>
<tbody>
<tr>
<td>Increased plaque instability</td>
<td>Decreased neonatal collagen content caused by increased metalloproteinase expression/activity, and increased endothelial production of cytokines that decreased VSMC-dependent collagen synthesis</td>
</tr>
<tr>
<td>Increased oxidative stress</td>
<td>Decreased production of endothelialia NO</td>
</tr>
<tr>
<td>Increased thrombotic activity</td>
<td>Increased production of reactive oxygen species in ECs and VSMCs</td>
</tr>
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</table>

Table 3. Atherosclerosis in mouse models of diabetes and insulin resistance

<table>
<thead>
<tr>
<th>Diabetic phenotype</th>
<th>Mouse model</th>
<th>Diabetic agent</th>
<th>Atherogenic agent</th>
<th>Effect on atherosclerosis</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type 1-DM</td>
<td>BALB/c</td>
<td>STZ</td>
<td>Hyperlipidemia (high-fat feeding)</td>
<td>Increased</td>
<td>36</td>
</tr>
<tr>
<td></td>
<td>apol/LPL1 transgenic</td>
<td>STZ</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>45</td>
</tr>
<tr>
<td></td>
<td>LDLR-KO</td>
<td>STZ</td>
<td>Hyperlipidemia (LDLR disruption and high-fat feeding)</td>
<td>No change (mice treated with low-dose insulin)</td>
<td>42</td>
</tr>
<tr>
<td></td>
<td>LDLR-KO</td>
<td>STZ</td>
<td>Hyperlipidemia (LDLR disruption and high-fat feeding)</td>
<td>Increased</td>
<td>43</td>
</tr>
<tr>
<td></td>
<td>apol/LPL1 transgenic</td>
<td>STZ</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>43</td>
</tr>
<tr>
<td></td>
<td>GP-2Akita</td>
<td>LDLR-KO</td>
<td>Hyperlipidemia (LDLR disruption and high-fat feeding)</td>
<td>Increased</td>
<td>47</td>
</tr>
<tr>
<td></td>
<td>Leprdb/db</td>
<td>LDLR-KO</td>
<td>Hyperlipidemia (LDLR disruption and high-fat feeding)</td>
<td>Absence of atheromas</td>
<td>51</td>
</tr>
<tr>
<td></td>
<td>apol-LDLr-KO</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>61</td>
</tr>
<tr>
<td></td>
<td>apol-LDLr-KO</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (LDLR disruption)</td>
<td>Increased</td>
<td>73</td>
</tr>
<tr>
<td>Type 2-DM</td>
<td>Leperb/ob:LDLR-KO</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (LDLR disruption)</td>
<td>Increased</td>
<td>73-75</td>
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<tr>
<td></td>
<td>Leperb/ob:apol-KO</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>96</td>
</tr>
<tr>
<td></td>
<td>LDLr-KO transplanted with INSR-null bone marrow</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>96</td>
</tr>
<tr>
<td></td>
<td>apol-KO transplanted with INSR-null bone marrow</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>99</td>
</tr>
<tr>
<td></td>
<td>apol-KO transplanted with IRS-2-null fetal liver cells marrow</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>99</td>
</tr>
<tr>
<td></td>
<td>apol-KO:IRS-2KO</td>
<td>Absence of leptin function</td>
<td>Hyperlipidemia (apoB overexpression and high-fat feeding)</td>
<td>Increased</td>
<td>100</td>
</tr>
</tbody>
</table>

Table 4. Animal models to study the effects of altered leptin/glucone/insulin signaling on neointimal thickening induced by mechanical injury

<table>
<thead>
<tr>
<th>Alteration</th>
<th>Animal model</th>
<th>Type of injury and effect on neointimal</th>
<th>Lesion size</th>
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## Murine models of diabetic atherosclerosis

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First Galley