

1 **Physical activity increases the bioavailability of flavanones after dietary aronia-**  
2 **citrus juice intake in triathletes**

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4 Medina, S.,<sup>a</sup> Domínguez-Perles, R.,<sup>a</sup> García-Viguera, C.,<sup>a</sup> Cejuela-Anta, R.<sup>b</sup>, Martínez-  
5 Sanz, J.M.<sup>b</sup>, Ferreres, F.,<sup>a</sup> Gil-Izquierdo, A.<sup>a,\*</sup>

6 <sup>a</sup>*Department of Food Science and Technology, CEBAS-CSIC, P.O. Box 164, Espinardo,*  
7 *30100, Murcia, Spain*

8 <sup>b</sup>*Department of Physical Education and Sport, Faculty of Education. University of*  
9 *Alicante. Campus de San Vicent del Raspeig, 03540 San Vicent del Raspeig, Alicante,*  
10 *Spain*

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20 \*Corresponding autor: Angel Gil-Izquierdo; Tel.: +34 968 396363 fax: +34 968 396213.

21 E-mail address: [angelgil@cebas.csic.es](mailto:angelgil@cebas.csic.es). Department of Food Science and Technology,  
22 CEBAS-CSIC, P.O. Box 164, Espinardo, 30100, Murcia, Spain

24 **Abstract**

25 Control and triathlete volunteers (n = 8 and n = 15, respectively) were given 400  
26 mL and 200 mL of aronia-citrus juice (AC-juice), respectively. The 24 h urine samples  
27 were hydrolysed to determine the flavanones concentration by UPLC-QqQ-MS/MS.  
28 The flavanones metabolites in both groups of volunteers were glucuronides, sulfates,  
29 and sulfo-glucuronides, and the total excretion of flavanones increased fivefold in the  
30 triathletes compared with the control volunteers. The increase of ninefold in the  
31 homoeriodictyol of triathletes compared to control volunteers may suggest the  
32 overactivation of the microbiota metabolism caused by physical exercise. No  
33 differences concerning the bioavailability were detected between men and women in  
34 controlboth groups. The AC-juice could provide synergistic effects on health due to the  
35 increase in the bioavailability of flavanones, avoiding the deleterious effects caused by  
36 the overdosage of nutritional supplements.

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39 **Keywords**

40 Aronia-citrus juice, bioavailability, dietary intervention, flavanones, triathletes

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43 **Highlights**

- 44 - Flavanones excretion increased five-fold in triathletes *versus* control
- 45 volunteers
- 46 - Physical activity increases the bioavailability of flavanones
- 47 - Changes in flavanones metabolites in 24 h hydrolyzed urine samples
- 48 - The main flavanones metabolites were glucuronides, sulfates and sulfo-
- 49 glucuronides

50

51        **1. Introduction**

52        In the past decade, a number of clinical trials based on dietary interventions have  
53        been performed to establish the bioefficiency of distinct subclasses of polyphenols  
54        (Kay, 2010). The applicability of polyphenols to the athlete's world and their health  
55        benefits in this concrete hallmark remains scarcely addressed (Trombold, Barnes,  
56        Critchley, & Coyle, 2010). Currently, sport medicine and training require additional  
57        efforts to improve the efficiency and results, taking into consideration athletes' health.  
58        In order to gain a further insight on the relationship between training, nutrition, and  
59        health, a variety of nutritional supplements have been developed to increase the physical  
60        outcome of the training programs regardless the natural option of fruit juices with  
61        bioactive components (Trombold, Barnes, Critchley, & Coyle, 2010).

62        Citrus juices are known for their high content in flavonoids, especially flavanones  
63        (Fls) (400-600 mg L<sup>-1</sup>) (Gil-Izquierdo, Gil, & Ferreres, 2002; Gil-Izquierdo, Gil,  
64        Ferreres, & Tomás-Barberán, 2001). These compounds are mostly attached to  
65        rhamnoglucosides which need to be removed by the colon microflora to be absorbed  
66        (Silberberg, Gil-Izquierdo, Combaret, Remesy, Scalbert, & Morand, 2006). Fls have  
67        shown a much permanent systemic level due to the enterohepatic cycle, which allows  
68        the re-excretion of metabolites, by bile and their reabsorption in the small intestine or  
69        colon, and therefore, a longer stay of them at physiological level (Manach, Morand, Gil-  
70        Izquierdo, Bouteloup-Demange, & Rémésy, 2003).

71        In recent years, research in this field has been focused on the augmentation of Fls  
72        bioavailability, by different mechanisms, in order to increase the health-promoting  
73        properties of citrus juices. The combination of aronia (*Aronia melanocarpa*) with citrus  
74        juices could provide synergistic effects of the Fls plus the anthocyanins, among other  
75        bioactive compounds (Habauzit, Sacco, Gil-Izquierdo, Trzeciakiewicz, Morand, Barron,

76 et al., 2011). However, as far as we are aware, the effect of physical activity on the  
77 bioavailability of the target compounds (Fls) from aronia-citrus juices (AC-juice)  
78 remains unknown.

79 The aim of the present study was to identify of the circulating Fls metabolites after  
80 the intake of AC-juice, and compare their bioavailability in triathletes with that found in  
81 control volunteers. Besides, the Fls excretion was also evaluated, during a week before  
82 and after AC-juice intake, in triathletes.

83

## 84 **2. Methods and Materials**

### 85 *2.1. Chemicals*

86 Naringenin, eriodictyol, homoeriodictyol, hesperetin, and isosakuranetin were  
87 purchased from Extrasynthèse (Genay, France). Hesperetin 7-O-glucuronide was  
88 synthesized in our lab according to the method described by Boumendjel and co-  
89 workers (2009). All LC-MS grade solvents were obtained from J.T. Baker (NJ, USA).  
90 Formic acid and chlorhydric acid were purchased from Panreac (Barcelona, Spain). The  
91  $\beta$ -glucuronidase, type H2 from *Helix pomatia* was obtained from Sigma-Aldrich (MI,  
92 USA).

### 93 *2.2. AC-juice*

94 The juice composition is based on the mixture of citrus juice (95%) with 5% of  
95 *Aronia melanocarpa* juice. The content in nutrients and caloric supply of the AC-juice  
96 that consumed control volunteers and triathletes were summarized in Tables 1 and 2,  
97 respectively detailing the percentage of contribution of the juice to the total diet.

98

### 99 *2.3. Calculation of the training loads in triathletes.*

100 The quantification of training programs is addressed to evaluate their effects on

101 physiological adaptation and subsequent performance (Borresen & Lambert, 2009).  
102 Currently, mathematical models are suitable for the quantification of the training loads.  
103 Indeed, the training impulse proposed by Banister & Calvert (1980) and its subsequent  
104 revision are recognized as valid for the quantification of training efforts. However, the  
105 evolution of sports performance has led to specific models for some exercises (Lucia et  
106 al., 2006).

107 In our work, the training load quantification was performed using the ‘Objective  
108 Load Scale’ (ECOs) developed by Cejuela & Esteve-Lanao (2011). The training load  
109 that supports a triathlete is an indication of its performance level. The training loads  
110 developed by the triathletes in the present work were similar to those found in other  
111 studies of endurance athletes (Lucia et al., 2006; Rodríguez-Marroyo et al., 2003).

112 The previously-developed method used allowed the quantification of the training  
113 loads in the sport of triathlon (swim, bike, run, and transitions), which are determined  
114 by the difficulty to maintain technique, delayed muscle soreness, typical workout  
115 density, and energy cost of each separate sport. The values of daily and weekly trainings  
116 were determined and summarized to assess the training load (ECOs) of each volunteer,  
117 depending on their physical characteristics and the intensity of the training program (the  
118 ECO data presented in this work are the average of the individual ECOs of the  
119 triathletes). Briefly, and from a general point of view, intensity is considered  
120 exponentially –not linearly– with the aim of leveling-off the total training stress for a  
121 given performance level. The volume is quantified by time and this allows a better  
122 comparison of different performance levels and terrain conditions (pavement, uneven  
123 laps) (Cejuela & Esteve-Lanao, 2011).

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#### 126 2.4. Study Design

127 Two clinical assays were developed with two intervention groups, control  
128 population (non-trained volunteers) (n=8, 4 women and 4 men) and triathletes (n=15, 5  
129 women and 10 men). Both groups were non smokers and did not receive any medication  
130 during the experimental procedure. Women were not in menstrual days during the  
131 study. Control population consumed two glasses of juice while triathletes ingested a  
132 glass of juice according to their planning supervised by the nutritionist and the training  
133 load (ECO).

134 In the first assay, the bioavailability and metabolism were compared between a  
135 control group of non-trained volunteers and the group developing a strenuous and  
136 chronic exercise in order to know the influence of the sport performance on the nature,  
137 occurrence and excretion of flavanone metabolites in urine. The control volunteers  
138 followed a diet without polyphenol-based food and products derived and the only  
139 source of flavonoids was that provided by the AC-juice. The diet for triathletes  
140 developed by the nutritionist provided the balanced products for a proper contribution  
141 of carbohydrates, proteins, fats, vitamins and microelements where the plant-foods  
142 (included those content flavanones) were ingested when were required. In detail, control  
143 volunteers and triathletes (2500 ECOs of training load (Cejuela Anta & Esteve-Lanao,  
144 2011) consumed a diet with equal intake of Fls provided by juice (86.1 mg) and by  
145 juice+diet (88.1 mg), respectively for a week (Tables 1 and 2).

146 In the other assay, the objective was to know the evolution of the bioavailability and  
147 metabolism of flavanones between a low charge of training and a period of strenuous  
148 chronic exercise during two weeks. The triathletes followed during the first week a  
149 control diet containing 45.0 mg of Fls (2400 ECOs) and 88.1 mg of Fls (provided by a

150 glass of juice+diet) (2500 ECOs) during the second week (Table 2). The study was  
151 approved by the Bioethics Committee of University Hospital of Murcia and all  
152 participants gave the written informed consent to participate in the dietary intervention  
153 study.

154 Volunteer's urines were collected during 24h starting from 8.00 a.m are the intake  
155 of the juice up to the 8.00 a.m. of the following day. They were immediately frozen at -  
156 80 °C until their analysis. Fls concentration was determined after total hydrolysis of  
157 urine with urine  $\beta$ -glucuronidase (sulfatase activity) (Habauzit, et al., 2011; Manach,  
158 Morand, Gil-Izquierdo, Bouteloup-Demange, & Rémésy, 2003).

159

#### 160 2.5. UPLC-QqQ-MS/MS analysis

161 The Fls metabolites in urines were analysed using an UPLC-MS/MS (Agilent  
162 Technol., Germany) with an ACQUITY BEH C18 column (2.1 x 150 mm; 1.7  $\mu$ m;  
163 Waters, Ireland). The solvents were water/formic acid (99.9:0.1, v/v) (A) and  
164 acetonitrile/formic acid (99.9:0.1, v/v) (B). The flow rate was 0.32 mL min<sup>-1</sup> using a  
165 linear gradient (t; %B): (0.00; 10), (3.50; 30), (8.00; 35), (8.01; 60), (10.00; 60), (10.01;  
166 100), (12.00; 100), (12.10; 10). The volume injection was 10  $\mu$ L. For the qualitative  
167 analyses, the flavanone conjugates were analysed according the MRM transition  
168 detailed in the Table 3 in order to know the original metabolites excreted in 24h-urine.  
169 For the quantitation of the total flavanones in urine, the capillary exit voltage and  
170 collision energy were previously optimized for the aglycones and hesperetin 7-O-  
171 glucuronide. The MRM transitions (negative mode) for maximum intensity after  
172 optimization of the MS parameters were at  $m/z$  271 $\rightarrow$ 119, 301 $\rightarrow$ 151, 287 $\rightarrow$ 151,  
173 301 $\rightarrow$ 151, and 285 $\rightarrow$ 164 for naringenin, hesperetin, eriodictyol, homoeriodictyol, and  
174 isosakuranetin, respectively (Table 3). The product ion scan allowed the identification



175 of aglycones by comparison of the MS2 spectra with those corresponding standards.  
176 The urinary concentration of FIs was calculated on freshly and daily prepared standard  
177 curves using MassHunter software version B.04.00 (Agilent Technol., Germany).

## 178 2.6. Statistics

179 Quantitative data are presented as mean  $\pm$  SD. Specific differences among the target  
180 compounds in samples from control volunteers and triathletes were examined by  
181 paired *t*-tests. Statistical analysis was conducted using SPSS 17.0 software package  
182 (LEAD Technol. Inc., USA), and the level of statistical significance was set at  $P < 0.05$ .

183

## 184 3. Results and discussion

### 185 3.1. Results

#### 186 3.1.1 Qualitative analyses

187 The nature of the excreted metabolites was determined in 24-h urine samples  
188 after ingestion of 200 mL and 400 mL in triathletes and control volunteers,  
189 respectively. The goal of this assay was to know if there are differences between the  
190 urinary profile of the flavanone metabolites in both groups. The characterization of  
191 metabolites from total urinary FIs in non-hydrolyzed urine samples are presented in  
192 Table 3. The analysis of the non-hydrolyzed samples showed the occurrence of free  
193 aglycones as previous reports (Habauzit, et al., 2011; Manach, Morand, Gil-Izquierdo,  
194 Bouteloup-Demange, & Rémésy, 2003). Unparallely to the other metabolites, the  
195 sulfates of isosakuranetin and diglucuronides of eriodictyol were not detected, whereas  
196 homoeriodictyol was only found as aglycone form and conjugated to two glucuronides  
197 (Table 3). Hesperetin and naringenin showed the highest variety of conjugates including  
198 glucuronides, diglucuronides, sulphates, and sulfo-glucuronides. In addition, we  
199 distinguished a 7-*O*-glucuronide of hesperetin by comparison with its authentic marker.

200 Fls metabolites showed different retention times although had the same  $m/z$   $[M-H]^-$  and  
201 MS2 fragmentation patterns undergoing a 176 amu and 80 amu losses corresponding to  
202 a glucuronide acid and sulfate moieties, respectively (Habauzit, et al., 2011). There  
203 were no differences between the patterns of Fls metabolites excreted in urine of control  
204 vs. triathletes after AC-juice intake.

### 205 3.1.2 Quantitative analyses

206 In urine, the quantification of the Fls bioavailability has been carried out after  
207 enzymatic urine hydrolysis (Habauzit, et al., 2011). The total excretion of Fls was ~5-  
208 folds higher in triathletes compared with control volunteers ( $1860 \pm 838$  and  $376 \pm 122$   
209 nmol  $24\text{ h}^{-1}$ , respectively) (Fig. 1A). This is of quite importance because of the amount  
210 of juice ingested by athletes was half than that consumed by control volunteers (200  
211 and 400 mL, respectively). Then and according to this fact, the theoretical bioavailability  
212 would be 10-fold higher respect to the ingested dose. In addition, when comparing  
213 control volunteers with triathletes, naringenin, eriodictyol, and hesperetin were the Fls  
214 that contributed more significantly to the increase of the total urinary Fls (~92%, ~88%,  
215 and ~53%, respectively) (Fig. 1B). The other two aglycones considered,  
216 homoeriodictyol and isosakuranetin, even though were at the lowest concentration  
217 among all considered Fls, presented a significant increase in bioavailability, between  
218 almost seven and nine-fold, respectively compared to that of control volunteers  
219 (theoretical 14 and 18-fold higher respect to the ingested dose, respectively) (Fig. 1B).  
220 Besides, no differences concerning to the bioavailability were detected in men and  
221 women in control and triathletes volunteers.

222 In the second clinical trial, the triathletes developing elite training during 2 weeks,  
223 (control week (1<sup>st</sup>): 2400 ECOs and AC-juice intake (200 mL) week (2<sup>nd</sup>): 2500 ECOs)  
224 the differences on total Fls bioavailability between the first and second week were not

225 so clear since no significant differences were found (Fig. 2A), whereas eriodictyol and  
226 hesperetin excretion increased thanks to the additional flavanones intake provided by  
227 the AC-juice (Fig. 2B).

### 228 3.2 Discussion

229 Flavanones have been reported to be excreted conjugated to distinct combinations  
230 of glucuronic acid and/or sulfate in human urine (Manach, Morand, Gil-Izquierdo,  
231 Bouteloup-Demange, & Rémésy, 2003). The metabolites of hesperetin and naringenin  
232 were coincident with the glucuronide, diglucuronide, and sulfoglucuronide conjugates  
233 identified by Mullen and colleagues in urine (Mullen, Archeveque, Edwards,  
234 Matsumoto, & Crozier, 2008). Our qualitative analysis on the excretion of Fls after the  
235 intake of AC-juice by triathletes and control volunteers was consistent with the  
236 detection of the sulfates of eriodictyol, naringenin, and hesperetin (Table 3) as  
237 previously described (Bredsdorff, Nielsen, Rasmussen, Cornett, Barron, Bouisset, et al.,  
238 2010). The high abundance of sulfo-glucuronides compared to the sulfates may be due  
239 to the activation of the enterohepatic cycle which favors the longer stay of the  
240 metabolites in the human's body (Manach, Morand, Gil-Izquierdo, Bouteloup-  
241 Demange, & Rémésy, 2003).

242 The first assay (control volunteers *vs* triathletes) showed that the total excretion of  
243 Fls increased five-fold in the triathletes compared with control volunteers. The  
244 absorption of Fls could be affected by a plethora of physiological factors linked to the  
245 physical activity. Among them, the triathletes may exhibit an increased intestinal  
246 motility secondary to the physical work. This entails an accelerated colorectal transit  
247 (Lippi, Banfi, Luca Salvagno, Montagnana, Franchini, & Cesare Guidi, 2008), and the  
248 modification of the bioavailability of dietary bioactive compounds, including Fls since  
249 the triathletes consumed the AC-juice 15 min after the training. During the exercise,

250 there is an increase in sympathetic and decrease in parasympathetic activity, which has a  
251 large influence on digestion because of their role in the innervation of the muscular  
252 layer within the intestine wall. Blood is usually moved away from the digestive tract in  
253 order to increase blood flow to the skeletal muscles and skin (Rao, Beaty, Chamberlain,  
254 Lambert, & Gisolfi, 1999) and the reduced blood supply may further inhibit gut  
255 function. However, after exercise, when blood flow is restored, the AC-juice was  
256 ingested by triathletes. At this point, the physiological changes may reestablish colonic  
257 motility allowing maximum exposure and absorption of nutrients including polyphenols  
258 and thus, the increase the Fls bioavailability. This augmented motility could favor the  
259 interaction of the AC-juice and food components of the diet with the gut microbiota.

260 A key point on Fls bioavailability is the cleavage of the original rhamnoglucosides  
261 forms of Fls into their aglycone, by intestinal enzymes of bacteria origin ( $\alpha$ -  
262 rhamnosidases and  $\beta$ -glucosidases) (Erlund, Meririnne, Alfthan, & Aro, 2001). In fact,  
263 the interindividual variations, in the bioavailability of Fls, could be caused by  
264 differences in gut microflora. This leads us to think that elite training, as the triathlon,  
265 could modify the activity of the colon microbiota and therefore, the concentration of  
266 hydrolytic enzymes available, which might increase the Fls bioavailability. In this way,  
267 homoeriodictyol and its metabolites are produced as result of the action of the gut  
268 microbiota (Habauzit, et al., 2011). Thus, they could be valuable ‘signalling  
269 compounds’ about the overactivation of the microbiota metabolism since they were  
270 produced in higher extent in triathletes compared to control volunteers (9-fold higher).

271 In the second assay, the AC-juice intake contributed to an increase of Fls  
272 bioavailability in a normal diet adjusted for an elite training of triathletes. However, the  
273 two-fold higher intake of Fls provided by the AC-juice (88.1 mg) compared to the  
274 control diet (45 mg) was not proportionally delivered in the same extent concerning to

275 the Fls excretion rates. This fact would indicate to a saturation point of flavanone uptake  
276 caused by the physical activity. Taking into account the first and second assay, the high  
277 rate of Fls bioavailability in triathletes after the consumption of a glass of AC-juice  
278 compared to control volunteers was only owing to the physical activity regardless the  
279 origin of the Fls source in the diet (AC-juice or control diet) since in previous studies,  
280 this bioavailability has not been increased by a dietary dose of any type of citrus product  
281 consumed in a hallmark of a normal diet (Manach, Morand, Gil-Izquierdo, Bouteloup-  
282 Demange, & Rémésy, 2003).

#### 283 **4. Conclusions**

284 In summary, the supplementation with a natural fruit juice may provide protection to the  
285 triathletes avoiding the intake of nutritional supplements, to enhance sport performance,  
286 and that eventually may entail adverse health outcomes. The total excretion of Fls  
287 increased five-fold in the triathletes compared with control volunteers (theoretical 10-  
288 fold higher respect to the ingested dose). It is of quite importance to underline the  
289 increase of 9-fold of the homoeriodictiyol in thriathletes compared to control volunteers  
290 (theoretical 18-fold higher respect to the ingested dose) since it could be considered as  
291 ‘signalling compound’ about the overactivation of the microbiota metabolism caused by  
292 physical exercise. Therefore, the effect of physical activity in triathletes consuming 1  
293 glass of juice was enough to widely overcome the bioavailability of Fls generated by  
294 control volunteers after the intake of 2 glasses of juice. Despite the high intensity of the  
295 elite training by triathletes, a glass of AC-juice was able to increase the bioavailability  
296 of Fls in a diet already containing food products able to produce Fls metabolites at  
297 systemic level.

298

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305

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365

366

367 **Figure Captions**

368

369 **Fig. 1** Content of total (A) and single Flavanones (naringenin, eriodictyol, hesperetin,  
370 isosakuranetin, and homoeriodictyol) (B) (nmol 24h<sup>-1</sup>) after enzymatic hydrolysis  
371 determined in the 24h-urine of control volunteers (n=8) and triathletes (n=15). Bars  
372 with asterisks are statistically different at \**P*<0.05 and \*\*\**P*<0.001.

373

374 **Fig. 2** Content of total (A) and single Flavanones (naringenin, eriodictyol, hesperetin,  
375 isosakuranetin, and homoeriodictyol) (B) (nmol 24h<sup>-1</sup>) after enzymatic hydrolysis  
376 determined in the 24h-urine of triathletes (n=15) before and after intake the AC-juice.  
377 Bars with asterisks are statistically different at \**P*<0.05, and \*\**P*<0.01.

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**Table 1** Dietary characteristics and caloric intake of the control volunteers group (% of contribution of the juice to the diet has been detailed between brackets).

	Daily intake	AC-juice intake (400 mL)
Carbohydrates (g)	214.0	36.0 (16.8%)
Sugars (g)	38.1	13.3 (34.9%)
Proteins (g)	129.8	1.8 (1.4%)
Fats (g)	55.7	0.1 (0.2%)
Iron (mg)	12.5	0.5 (4.2%)
Vitamin C (mg)	162.4	154.8 (95.3%)
Vitamin E (mg)	5.0	0.2 (3.0%)
Vitamin A ( $\mu$ g)	220.1	3.0 (1.4%)
Total polyphenols (mg)	115.5	115.5 (100.0%)
Flavanones (mg)	86.1	86.1 (100.0%)
Flavones (mg)	29.4	29.4 (100.0%)
Energy intake (kcal)	1857.9	152.0 (8.2%)

**Table 2** Dietary characteristics, caloric intake, and training loads of the triathlete volunteers group  
(% of contribution of the juice to the diet has been detailed between brackets).

	Diet control	Diet + juice intake	AC-juice intake (200 mL)
Carbohydrates (g)	326.1	344.1	18.0 (5.2%)
Sugars (g)	121.3	127.9	6.6 (5.2%)
Proteins (g)	133.7	134.6	0.9 (0.6%)
Fats (g)	113.7	113.7	0.1 (0.1%)
Iron (mg)	20.9	21.2	0.3 (1.2%)
Vitamin C (mg)	178.7	256.1	77.4 (30.2%)
Vitamin E (mg)	21.0	21.1	0.1 (0.4%)
Vitamin A ( $\mu$ g)	2970.0	2971.5	1.5 (0.1%)
Total polyphenols (mg)	64.6	122.4	57.8 (47.2%)
Flavanones (mg)	45.0	88.1	43.1 (48.9%)
Flavones (mg)	0.7	15.4	14.7 (95.5%)
Flavan-3-ols (mg)	8.4	8.4	-
Flavonols (mg)	8.9	8.9	-
Isoflavones (mg)	1.6	1.6	-
Energy intake (kcal)	2820.0	2896.0	76.0 (2.6%)
<i>Training loads</i>			
ECOs	2400	2500	

**Table 3** Qualitative analysis of Fls metabolites non-hydrolyzed urine after the ingestion of AC-juice.

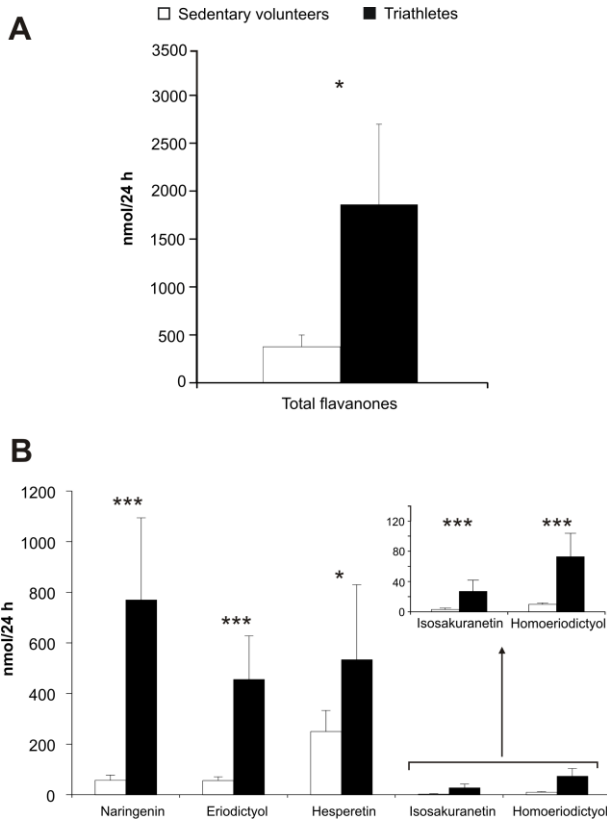
Retention time (min)	MRM transitions ( <i>m/z</i> )	Compounds
<i>Naringenin metabolites</i>		
2.48	623→271	Naringenin-diglucuronide
3.15	623→271	Naringenin-diglucuronide
3.58	527→271	Naringenin-sulfoglucuronide
3.74	623→271	Naringenin-diglucuronide
3.74	527→271	Naringenin-sulfoglucuronide
4.53	447→271	Naringenin-glucuronide
4.64	447→271	Naringenin-glucuronide
4.69	351→271	Naringenin-sulfate
7.27	271→119	Naringenin
<i>Hesperetin metabolites</i>		
2.96	653→301	Hesperetin-diglucuronide
3.52	557→301	Hesperetin-sulfoglucuronide
3.68	653→301	Hesperetin-diglucuronide
3.82	557→301	Hesperetin-sulfoglucuronide
4.16	653→301	Hesperetin-diglucuronide
4.66	477→301	Hesperetin-glucuronide
4.88	477→301	Hesperetin-7- <i>O</i> -glucuronide
5.19	381→301	Hesperetin-sulfate
7.91	301→151	Hesperetin
<i>Isosakuranetin metabolites</i>		
4.09	541→285	Isosakuranetin-sulfoglucuronide
4.28	637→285	Isosakuranetin-diglucuronide
6.89	461→285	Isosakuranetin-glucuronide
9.79	285→164	Isosakuranetin
<i>Eriodictyol metabolites</i>		
2.09	463→287	Eriodictyol-glucuronide
3.02	367→287	Eriodictyol-sulfate
3.33	367→287	Eriodictyol-sulfate
3.65	543→287	Eriodictyol-sulfoglucuronide
5.77	287→151	Eriodictyol
<i>Homoeriodictyol metabolites</i>		
3.73	477→301	Homoeriodictyol-glucuronide
4.11	477→301	Homoeriodictyol-glucuronide
7.55	301→151	Homoeriodictyol

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401 **Figure 1.**



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413 **Figure 2.**

