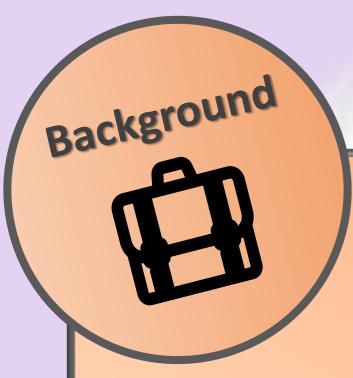
Virulence of entomopathogenic nematodes against two aerial pests: Frankliniella occidentalis (thysanoptera: thripidae) and *Tuta absoluta* (lepidoptera: gelechiidae): intra- and interspecific variability CSIC DE LA RIOJA Gobierno DE LA RIOJA BIOLOGICAL SYSTEMS

Vicente-Díez, Ignacio¹, M. González-Trujillo¹, M. Galeano², M. Chelkha¹, J.E. Belda², J. Calvo², and R. Campos-Herrera¹.

¹Instituto de Ciencias de la Vid y del Vino (Gobierno de La Rioja, CSIC, Universidad de La Rioja), La Grajera, Crta. Burgos Km. 6, Salida 13 Lo-20, 26007 Logroño (Spain). ² R&D Department of Koppert España, S.L. Paraje Piedra Rodada, 470, Vícar, Almería 04738 (Spain).



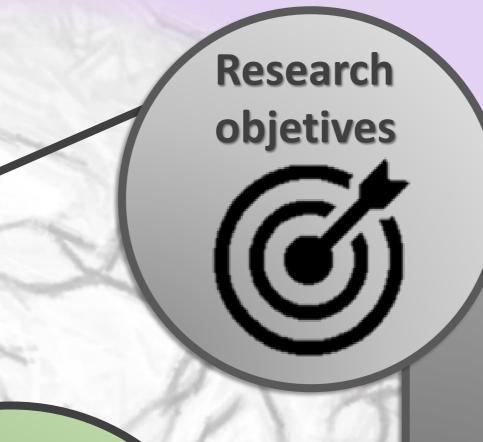
Infective juveniles (IJs) of Steinernema feltiae

Instituto de

Ciencias de la

Vid v del Vino

Entomopathogenic nematodes (EPNs), well-known as excellent biological control agents, can now be applied against aerial pests thanks to novel formulations [1]. Exploring the intra- and inter-specific variability on EPN virulence can support selecting the best candidates for a particular aerial insect pest.



F. occidentalis

<u>HYPOTHESIS</u>: populations of the same species but with different origins (habitat, geo-region) might differ in their ability to kill the same hosts.

<u>AIM</u>: to evaluate the virulence (mortality and time to kill) of various EPN species/populations against the last larval instar of two aerial pests: Frankliniella occidentalis (Thysanoptera: Thripidae) and Tuta absoluta (Lepidoptera: Gelechiidae).

Table 1. EPN species populations and their origin

Populations

Koppert

RS-5

Origin

Commercial

Switzerland

Commercial

Switzerland

Commercial

Switzerland

Portugal

Spain

Portugal

Spain

Material 8 Methods

Ten EPN populations belonging to three EPN species (Table 1) were cultured in *Galleria mellonella* (Lepidoptera: Pyralidae) larvae [2]. EPN populations were tested at two IJ concentrations: 80/160 and 4/21 IJs/cm² for *F. occidentalis* and *T. absoluta* trials, respectively (including) distilled water as controls). The *F. occidentalis* trials were evaluated in round containers (3,14 cm²), while *T. absoluta* trails in 5,5 cm diam. Petri dishes (Fig 1). Each treatment was replicated 5-6 replicates with 10 and 8 last instar larvae of *F. occidentalis* and *T. absoluta*, respectively. Both experiments were performed twice under a controlled environment chamber (60%, RH 22 °C, and 16L: 8D). The larval mortality was revised daily up to six days post-inoculation.

)	
)	C

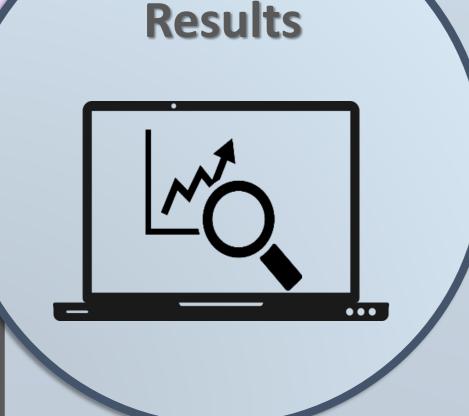
and T. absoluta (B) set up

		AM-25
		RM-107
B	S. carpocapsae	Koppert
		MG-596ª
Fig. 1. <i>F. occidentalis</i> (A)	H. bacteriophora	Koppert
		MG-618b
		AM-203
and <i>T. absoluta</i> (B) set up		RM-102

EPN species

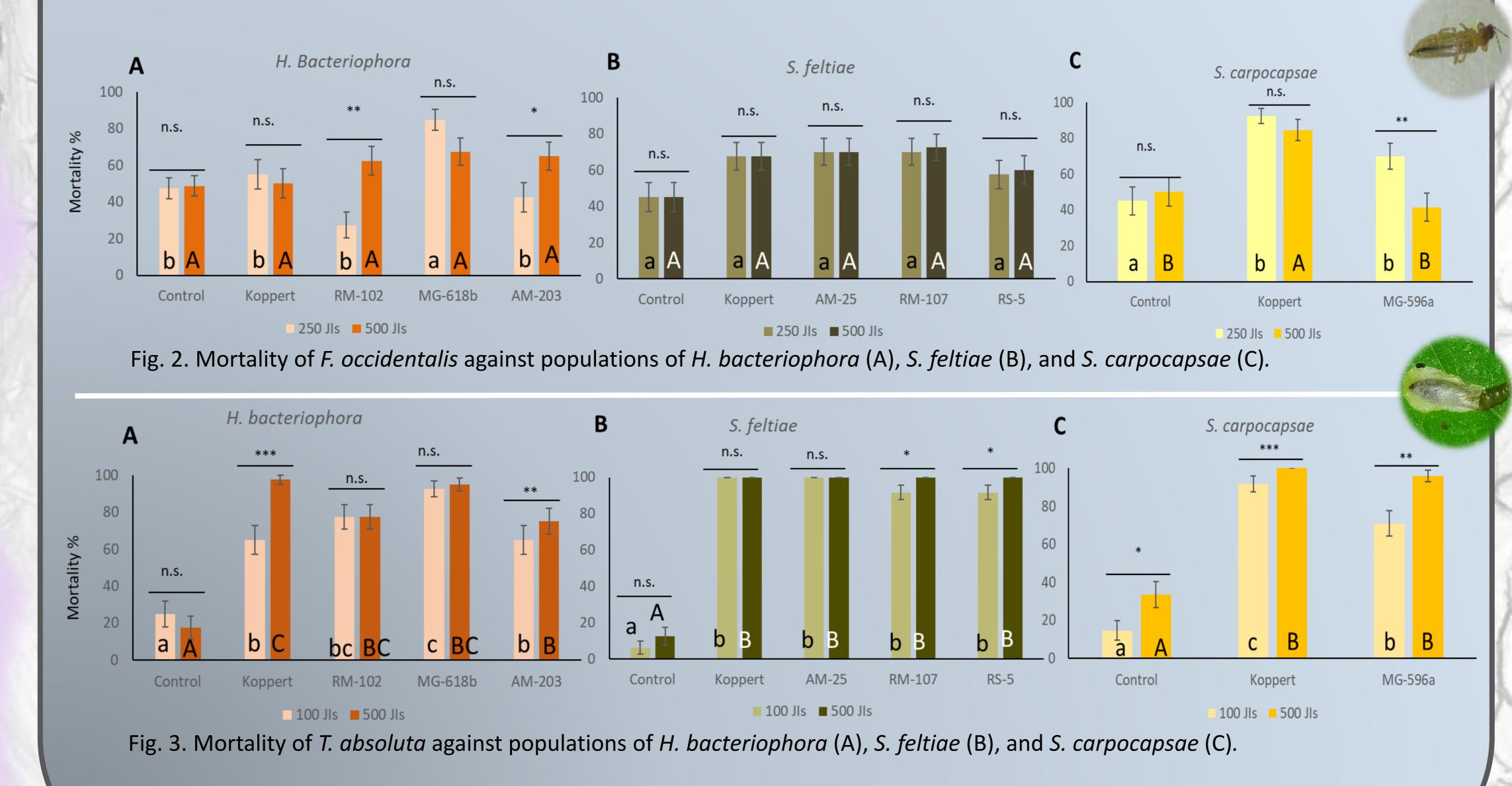
S. feltiae

T. absoluta



For F. occidentalis trails, we reported significantly different mortality rates for S. carpocapsae populations, particularly for Koppert's, which, in agreement with a previous study [3], reached values over 90% mortality after six days of exposure for both IJ concentrations (Fig.2).

Moreover, all EPN populations reached values over 65% mortality of *T. absoluta* larvae after six days of exposure for both IJ concentrations. Similar to previous reports [4,5], the four S. feltiae populations evaluated achieved over 90% mortality even for very low IJ inocula (4 IJs/cm²), but comparable values were also reported for *S. carpocapsae* Koppert strain (Fig. 3).



Overall, different EPN species populations, except few exceptions, did not highly differ in their virulence against F. occidentalis and T. absoluta larvae. We consider the EPN species *S. feltiae* and *S.* carpocapsae as very promising for their potential aerial application to control

Conclusions

Asterisks indicate significant differences within pair-treatment comparisons (t-Student) at *** P < 0.001, ** P < 0.01, * P < 0.05, and n.s. not significant. Different letters indicate significant differences in Tukey's test (HSD). Values are least-square means \pm SE.



these two serious pests. [1] Shapiro-Ilan & Dolinski 2015. In: Campos-Herrera (Ed.), Nematode Pathogenesis of Insects and Other Pests

[2] Woodring & Kaya 1988). Steinernematid and Heterorhabditid nematodes: a handbook of biology and techniques [3] Ebssa et al. 2001. J Invertebr Pathol [4] Batalla-Cabrera et al. 2010. BioControl [5] Dlamini et al. 2019. J Nematology