



Preview p38β and Cancer: The Beginning of the Road

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Abstract: The p38 mitogen-activated protein kinase (MAPK) signaling pathway is implicated in cancer biology and has been widely studied over the past two decades as a potential therapeutic target. Most of the biological and pathological implications of p38MAPK signaling are often associated with p38 α (MAPK14). Recently, several members of the p38 family, including p38 γ and p38 δ , have been shown to play a crucial role in several pathologies including cancer. However, the specific role of p38 β (MAPK11) in cancer is still elusive, and further investigation is needed. Here, we summarize what is currently known about the role of p38 β in different types of tumors and its putative implication in cancer therapy. All evidence suggests that p38 β might be a key player in cancer development, and could be an important therapeutic target in several pathologies, including cancer.

Keywords: p38MAPK; MAPK11; p38β; cancer

1. Introduction

Mitogen-activated protein kinases (MAPKs) are an evolutionarily conserved family of enzymes that link extracellular signals to the intracellular machinery in order to control a plethora of cellular processes including proliferation, cell survival, differentiation and apoptosis, among others. In fact, its deregulation is associated with many human diseases including inflammation, neurodegenerative disorders and cancer [1].

In mammals, four conventional MAPK subfamilies have been identified: extracellular signalregulated protein kinases 1/2 (ERK1/2), c-Jun N-terminal kinases 1-3 (JNK1/2/3), p38MAPKs (α , β , γ and δ), and the most recently discovered and least characterized ERK5 [1]. Each MAPK has its own activators, inhibitory phosphatases, substrates and scaffold proteins that allow the correct function of the different MAPK signaling pathways [2,3]. The diversity and specificity of MAPKs in cellular responses are achieved with a linear architecture, consisting of a module of three protein kinases: a MAPK kinase kinase (MAP3K or MKKKs) at the top, which phosphorylates a MAPK kinase (MAP2K, MKKs or

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MEKs) on specific serine (S) and threonine (T) residues. Eventually, there is a dual phosphorylation of the T and tyrosine (Y) residues of the conserved T-X-Y motif, located in a loop close to the active site of the terminal MAPK [4].

In addition, there is the group of atypical MAPKs, including ERK3/4, ERK7, ERK8 and Nemo-like kinase (NLK) [5–8], whose regulation and activation is not related to the module of the three kinases described for the conventional ones.

2. The p38MAPK Family

In mammalian cells, the p38MAPK family includes four members: p38 α (MAPK14), p38 β (MAPK11), p38 γ (MAPK12) and p38 δ (MAPK13), which have a high degree of sequence homology at the amino acid level (>60%) [9]. p38MAPKs differ in their expression patterns and substrate specificities, suggesting diverse functions. The p38MAPKs are S/T proline-directed kinases with an activation motif, T-G-Y, in which the substrate specificity is not only determined by the targeted amino acids, but also by specific docking domains present on the substrate and by a specific substrate binding motif in the MAPK (for a recent review of the p38MAPK-mediated signaling see [10]). The primary MAP2Ks for the p38MAPKs modules are MKK3 and MKK6 [11], although initially it was also considered the activation through MKK4 [12]. Activation of MAP2Ks occurs by phosphorylation of two conserved S and T residues on their activation loop by a broad range of MAPK3s. The MAP3Ks of this pathway include ASK1 (apoptosis signal-regulating kinase 1), DLK1 (dual-leucine-zipper bearing kinase 1), TAK1 (transforming growth factor β -activated kinase 1), TAO (thousand and one amino acid) 1 and 2, TPL2 (tumor progression loci 2), MLK3 (mixed-lineage kinase 3), MEKK (3 and 4), and ZAK1 (leucine zipper and sterile- α motif kinase 1) [13]. However, it has also been reported activation/inactivation of this signaling pathway by non-canonical mechanisms as in the case of T-cell receptor [14] or GRK2 [15].

The p38MAPK family can be divided into two subsets: on the one hand, p38 α and p38 β , and on the other hand p38 δ and p38 γ . This classification is based on their homology and their susceptibility to be inhibited by pyridinyl imidazoles (SB203580 and SB202190 compounds) at low concentrations. p38 α and p38 β have a higher homology between them (75%) and both can be inhibited by pyridinyl imidazoles, whereas p38 δ and p38 γ are 61% and 62% identical to p38 α , respectively, and are not susceptible to be inhibited by SB203580 and SB202190 [16,17]. The number of specific inhibitors for p38MAPK is rapidly growing, allowing for a better understanding of the biological role of each p38 family member [18]. Important substrates in the p38MAPK signaling pathway include downstream kinases such as MK2/3, PRAK MSK1 or MNK1/2, as well as various transcription factors including ATF1/2/6, c-Myc, c-Fos, GAT4A, MEF2A/C, SRF, STAT1, p53 and CHOP among others [19]. This diversity of factors associated with the p38MAPK signaling pathway gives a glimpse of the plethora of biological processes implicated in this pathway.

3. p38ß

p38 β (also known as Stress Activating Protein Kinase 2 (SAPK2), Stress Activating Protein Kinase 2b (SAPK2b), MAPK11 or P38 β 2) was described in 1996 by Jiang and coworkers [20], and is encoded by the *MAPK11* gene that maps to chr22:50,263,713-50,270,380 in the human genome (UCSC genome browser/GRCh38/hg38), and comprises 12 exons (NCBI reference sequence NM_0022751.7). The protein is 364 amino acids long, and has a kinase domain (amino acids 24-308) that includes a T-G-Y (amino acids 180-182) dual phosphorylation motif, which is required for its kinase activity [16,21]. The 3D structure of p38 β resembles that of a typical kinase with a smaller β -sheet N-terminal domain and a larger C-terminal domain. The ATP-binding site is located between the two domains, which are linked by a single polypeptide chain (residues T107-G110). This structure is also very similar to that of p38 α , although they differ in the relative orientation of the N- and C-terminal domains [22]. This orientation causes a reduction in the size of the ATP-binding pocket of p38 β compared to p38 α . The difference in size between the two pockets could play a role in their different substrate specificity [20], and could be exploited in order to design selective compounds able to inhibit each p38 protein independently [22].

p38 β is ubiquitously expressed, but at lower levels than p38 α . p38 β is expressed in the human brain, heart, placenta, lung, liver, skeletal muscle, kidney, spleen, testis, ovary, prostate, thymus and pancreas [20,21]. Moreover, p38 β is abundant in endothelial cells, but undetectable in other lineages as macrophages or monocytes [23]. Similar to p38 α , p38 β is activated by pro-inflammatory cytokines and environmental stress, such as IL-1 β , TNF, sorbitol, arsenite, anisomycin, high osmolarity, H₂O₂ and UV light [16,24]. The MAP2K that activates p38 β is MKK6, whereas p38 α is activated by MKK3 and MKK6 [20,25,26]. A unique characteristic of p38 β is the ability to modulate its basal activity by autophosphorylation events. p38 β is capable of self-activation by cis autophosphorylation of the residue T180 located in the activation loop. This activation occurs spontaneously in vitro, but can be regulated in mammalian cell cultures [27]. Moreover, p38 β also autophosphorylates in trans residues T241 and S261 in vivo. Indeed, phosphorylation of S261 reduces the activity of T180-phosphorylated p38 β , whereas, T241 phosphorylation reduces its phosphorylation in trans, although these two phosphorylation events do not seem to affect the activity of dually phosphorylated (T180/Y182) p38 β [28].

The functions of p38 β are mostly redundant with those of p38 α . For instance, it has been shown that p38 β cannot perform specific functions of p38 α during development [29]. In fact, p38 α knockout mice are lethal due to placental defects [30], while p38 β knockout mice are fertile and viable [31].

The substrates attributed to $p38\beta$ are mainly based on the use of SB compounds, which inhibit both $p38\alpha$ and $p38\beta$, not allowing to determine if they are bona fide substrates of $p38\beta$ (p38MAPK substrates are reviewed in [10]). However, there are p38 β targets that have been confirmed with other approaches. Among the several p38ß substrates, there are protein kinases, transcription factors, and transcriptional regulators. Regarding protein kinases, the MAPK-activated protein kinases MAPKAPKs are a group of proteins downstream of MAPKs. A subgroup of MAPKAPKs is composed of MK2, MK3 (also known as 3pK), and MK5 (also designated as p38-regulated/activated protein kinase (PRAK)). These three kinases regulate key cellular processes such as gene expression at the transcriptional and post-transcriptional level, control cytoskeletal architecture and cell cycle progression, and play an important role in pathological processes such as inflammation and cancer (reviewed in [32]). p38 α and p38 β inhibit mitotic entry through MK2/3 phosphorylation in vivo [33] and MK2/MK3 activation is blocked by the inhibitor SB203580 in vitro [34]. MK5 is activated by p38 α and p38 β in vitro and in vivo [35], regulating the shuttling of this protein from the nucleus to the cytoplasm [36]. Another substrate of p38 β is Protein kinase C $\mathcal{C}(PKC\mathcal{C})$, a serine/threonine kinase involved in the regulation of cytokinesis in mitotic cells. This protein is primed to bind 14-3-3 by a series of phosphorylation events initiated by p38MAPK (in S350), GSK3 (in S346) and PKC itself (in S368). In vitro studies have shown that p38 α and p38β phosphorylate S350 creating a GSK3 recognition site for the phosphorylation of S346, and that chemical inhibition by SB203580 prevents S346 phosphorylation in cells stimulated by UV-C [37].

Other studies have shown that p38 β is also associated with several transcription factors. For example, MEFs (Myocite Enhancer Factors) are a family of transcription factors composed of MEF2A-D that regulates cell differentiation, proliferation, apoptosis, migration, and metabolism [38]. MEF2A and MEF2C are phosphorylated by p38 α and p38 β in vitro through a MAP kinase docking domain that is specific to these MAPKs, and activates their transcriptional activity in vivo [39]. Moreover, SB202190 inhibits the transcriptional activity of MEF2C induced by LPS or MKK6 in monocytic cells [40]. Another transcription factor targeted by p38 β is NFATc4 (Nuclear Factor of activated T cells 4). NFATc4 belongs to the NFAT family of transcription factors, and is involved in cardiac development, mitochondrial function, and in activation of adipocyte specific genes during differentiation [41,42]. NFATC4 is phosphorylated by p38 α , β , γ and δ in the presence of an activated MKK6 mutant (MKK6-GLu) in vitro and in vivo, p38 α phosphorylates NFATc4 at S168 and S170 in the NFAT homology domain regulating the subcellular distribution of the transcription factor, promoting cytoplasmic localization of the NFATc4, and blocking adipocyte formation under differentiation conditions [43]. Moreover, phosphorylation of S168 and S170 of endogenous NFATc4 by p38MAPK is sensitive to SB203580 [44].

AP-1, a dimeric complex that is composed of members of the JUN, FOS, ATF or MAF protein families, regulates a wide range of cellular processes including cell proliferation, death, survival and differentiation, and has also been shown to be a downstream target of p38 β [45]. C-FOS and ATF2 were also shown to be phosphorylated in vitro and in vivo by the four p38MAPKs, increasing its transcriptional activity [16,21,46,47]. Furthermore, it was reported that histone deacetylase 3 interacts specifically with p38 β in LPS-stimulated cells, diminishing its phosphorylation, and leading to a repression of ATF-2 transcriptional activity as in the case of TNF gene expression [48]. Another transcription factor targeted by p38 β is MafA, a member of the MAF family of basic leucine zipper proteins, that act as an important regulator of development and differentiation in many organs/tissues, and is a key player in Insulin regulation (for a review see [49]). MafA is also phosphorylated by the four p38MAPK isoforms in vitro and in vivo, and this phosphorylation might control MafA function, as it was shown previously in lens differentiation in primary cultures of chicken neuroretinal cells [50].

Other substrates of p38 β with different functions that are shared with p38 α , have been also reported. This group of miscellaneous substrates includes the BAF 60 protein BAF60c [51,52], E47 [53], P18 (Hamlet) [54,55], Cyclin D3 [56], the variant of the histone H2A, H2AX [57], KH-type splicing regulatory protein (KSRP) [58], and the membrane associated metalloprotease TACE [59]. However, there are two proteins, Glycogen Synthase (GS) and Raptor that seem to be specific substrates of p38 β , and are not phosphorylated by any of the other p38MAPK proteins. p38 β binds specifically to GS in skeletal muscle, brain and liver, and its efficient phosphorylation allows GSK3 to phosphorylate other residues of GS, causing partial inhibition of its activity [60]. In the case of Raptor, a regulatory-associated protein of mTOR, activated p38 β by arsenite interacts with Raptor resulting in the phosphorylation of Raptor on S863 and S771, enhancing mTORC1 activity [61]. Therefore, the search for new specific substrates based on genetic evidence rather than on SB compounds, is a key step in the further understanding of biological functions mediated by p38 β .

4. p38β and Cancer

Although p38 β has been related to several pathological conditions like Huntington disease [62] and cardiac hypertrophy [63,64], this review will be focused on the role of p38 β in cancer.

Since the mid-90s, when the p38MAPK signaling pathway was initially related to the cellular response to DNA damage agents including antitumor treatments [65], up to recent evidence indicating its use as a potential therapeutic target [13], the role of the p38MAPK signaling pathway in cancer has been deeply studied. However, most of the work has been focused on p38 α , which has been repeatedly shown to play an important role in cancer biology. Consistent data from experimental models in different pathological conditions [66,67], have allowed us to consider p38 α as a biomarker [24,68,69], and also as a putative target for cancer therapy (for a review see [70]). Conversely, much less is known about the role of the other p38 proteins (p38 $\beta/\gamma/\delta$) in cancer, although recent studies have shown an important role of p38 γ/δ in cancer [71,72], however, further studies are needed to elucidate the definitive role of these two p38 proteins in cancer pathology (for a review see [73]).

Little is known about the role of p38 β in cancer, although this protein has been associated with key molecules in this disease. For example, p38 β has been proposed as a key target of the proto-oncogene Pokemon, a transcription factor known to be implicated in tumorigenesis and metastasis in hepatic cells [74]. Also, it has been reported that p38 β could be a critical step in tumor formation through regulation of lipocalin 2 (LCN2) expression, a direct target of Plakophilin 3 (PKP3). In this sense, it has been shown that in different types of tumors, high LCN2 expression correlates with increased invasion, tumor formation and metastasis (for a review see [75]). Interestingly, in the absence of PKP3, p38 β is able to control the expression of LCN2, indicating a potential role of p38 β in tumor formation [76]. p38 β has also been associated with integrin- αv , known to maintain cellular proliferation in keratinocytes by controlling c-Myc translation through FAK, p38 β and p90RSK1. Chemical inhibition of p38 β or genetic interference of *MAPK11* in keratinocytes promotes a marked decrease in c-Myc levels [77]. It was proposed that p38 β could play a key role in biological processes for tumor progression and angiogenesis. For instance, TGF- β 1 was shown to induce endothelial cell apoptosis by changing VEGF signaling from p38 β , with survival function, to p38 α with a pro-apoptotic function [78], in agreement with previous observations in cardiomyocytes [79]. Other studies have shown a direct connection of p38 β with VEGF in a murine retinal model [80], further highlighting the importance of p38 β in neovascularization and hypoxia-induced cell proliferation. Altogether, these studies suggest that p38 β could be a potential target for an anti-angiogenic approach.

Also, p38 β has been related to other aspects associated to cancer disease, with important implications in the patient's quality of life. For example, p38 β has been related to cachexia through the control exerted onto the autophagic protein ULK1 in both in vitro and in vivo muscle wasting models [81]. Indeed, it is known that p38 β functions upstream of FoxO–BNIP3 signaling axis to mediate an energy stress response [82], supporting the role of this MAPK in energy sensing. Another interesting aspect of p38 β is its relationship with cancer-associated pain. In an experimental model of rats, pain associated to intra-tibial injection of mammary gland carcinoma cells, showed a marked reduction by intrathecal administration of a p38 β antisense oligonucleotide [83]. Furthermore, the reduction of cancer-associated pain by music therapy was also attributed to low expression of p38 β and p38 α [84].

In addition to the connection with key proteins and biological processes in cancer, there are several examples showing the implication of p38 β in different types of tumors. In pancreatic cancer, Singh and coworkers reported that p38β could be a potential biomarker [85]. Furthermore, the authors showed that peptide inhibitors for $p38\beta$ are able to induce toxicity in pancreatic cell lines such as PANC-1, suggesting a potential therapeutic implication [85]. In hepatocellular carcinoma, recent data showed that p38 β is a target of miR-516a-5p, which is controlled by a novel circular RNA, circ-0001955, that increases the expression of $p38\beta$, facilitating hepatocellular tumorigenesis [86]. In bladder cancer, p38 β has been reported to be a critical player in cell motility through the signaling axis ILK-p38 β -Hsp27 [87]. In prostate cancer, p38 β has been related to metastases through the control exerted on the Wnt inhibitor Dickkopf-1, indicating the possibility of being considered as a therapeutic target [88]. Another study considered that the $p38\alpha/\beta$ inhibitor SB202190 could be used as a putative therapy in this type of tumor, in which STK11 could be a critical biomarker for this p38-based therapy, but no genetic evidence supported a critical role for $p38\beta$ [89]. Therefore, further investigation is necessary to clarify the role of this particular MAPK in the biology and therapy of prostate cancer. In endometrial cancer, p38β has been shown to mediate the proliferation of tumor cells by inhibiting apoptosis. In this case, the anti-apoptotic ability of p38ß seems to be controlled by the long non-coding RNA 1220, that controls p38β expression [90]. Interestingly, in lung cancer it has been recently reported that $p38\alpha$, but not the rest of p38MAPK members, could be a potential biomarker of chemotherapy response [68]. However, overexpression of p38 β was shown to be related to a specific subset of lung cancer in non-smokers in China [91]. Other reports indicate that a single-nucleotide polymorphism in p38 β (rs2076139) is a potential biomarker associated with progression-free survival in metastatic non-small-cell lung cancer patients receiving platinum-based chemotherapy [92]. In addition, in lung cancer of non or light smokers it was shown that p38 β and p38 α , could be predictors of the expression levels of the DNA excision repair protein ERCC1, a key protein in DNA damage reparation with implications for the response to platinum compounds [93]. Indeed, chemical inhibition of $p38\alpha/\beta$ decreased viability of lung cancer cell lines, but genetic interference showed that most of this effect relies on p38 β [93]. However, in terms of response to cisplatin, the effect of p38 β was not applicable to all the experimental models [93], suggesting a more prominent role for $p38\alpha$. Nonetheless, further studies are required to fully elucidate the role of $p38\beta$ in lung cancer and its therapy. In breast cancer, the only connection with p38 β has been related to bone metastases, through the up-regulation of the expression and secretion of monocyte chemotactic protein-1, which activates osteoclast differentiation and activity. Interestingly, the authors show how targeting $p38\beta$ in breast cancer cells could be a novel approach to treat bone destruction associated with bone metastasis [94]. In silico evidence connected triple-negative breast cancer with epirubicin response and p38 β overexpression, among other MAPKs, but no experimental data have been provided so far [95]. In Head and Neck Squamous Cell Carcinoma (HNSCC) patients, high expression levels of all p38MAPK isoforms, including p38 β , have been detected in sera. Interestingly, these levels are downregulated after therapy, except for p38 δ , suggesting that all p38MAPKs could be potential biomarkers in this disease. In addition, the authors indicated a potential role of p38 δ as a putative target for HNSCC therapy that cannot be extrapolated to p38 β [96].

 $p38\beta$ has also been related to leukemic pathology, for example, in acute myeloid leukemia (AML) and in Sézary syndrome. In AML, an aggressive hematologic malignancy, the overexpression of the SET oncoprotein, able to inhibit the protein phosphatase PP2A, is a key event that correlates with poor prognosis [97]. In this regard, p38 β has been associated with the inhibitory effect of SET onto PP2A by two different mechanisms: first, by promoting SET cytoplasmic translocation through CK2 phosphorylation, and second, by direct binding to and stabilization of the SET protein [98]. Therefore, and considering the anticancer activity of several PP2A-activating drugs [99], p38 β could be a potential novel target in AML, especially in those cases with SET over-expression. In the Sézary syndrome, a leukemic variant of cutaneous T-cell lymphomas, it has been reported that the overexpression of p38ß could be a potential driver gene or a novel biomarker [100]. Indeed, in Sézary syndrome-derived cell lines, inhibition of PKC β and GSK3 with the small molecules Enzastaurin and AR-A014418 promote a marked decrease in p38 β expression without changing p38 α levels [100]. Also, SB203580 and SB202190 promote cell death in those cell lines as well as in primary samples from Sézary syndrome patients. However, the genetic interference of p38 β does not show any effect in cell viability [100], suggesting that further studies are necessary to fully evaluate the potential therapeutic implications of $p38\beta$ in Sézary syndrome.

Nevertheless, in other types of tumors, p38 β appears not to have any implication or, if so, a marginal role. For example, in melanoma, preliminary evidence in cell lines discard this MAPK, but not p38 α , as a key player in this pathology [101]. Another example could be colorectal cancer, in which the 1628A>G (rs2235356) genetic variation in the p38 β promoter region may contribute to the susceptibility to colorectal cancer in a Chinese population [102]. However, recent reports discard specifically this result in a Swedish population [103], suggesting that maybe p38 β is not a universal biomarker for colorectal cancer. Indeed, other screening study discards p38 β and indicates that p38 α could be considered as a potential diagnostic marker and a putative therapeutic target for colorectal cancer [104]. In fact, this last observation is in agreement with previous reports using patient-derived xenografts [105]. Altogether, all this evidence suggests a marginal role for p38 β in colorectal cancer.

Finally, regarding the implications of p38 β in cancer biology, it is important to mention that Stress Activated Protein Kinases signaling pathways, JNK and p38MAPK, have been shown to play a dual role in cancer, both as an oncogene and as a tumor suppressor gene (for a review see [106,107]). This dual role seems to be dependent on several factors, including the experimental model and the stage of cell transformation, among others. Colorectal cancer is a paradigmatic example of this duality, showing that p38 α could behave as an oncogene or a tumor suppressor depending on the stage of the carcinogenesis process [66]. It is likely that similar to p38 α , p38 β may have a dual role in cancer, also playing a tumor suppressor role. For instance, p38 MAPK signaling has been proposed to act as a tumor suppressor gene by controlling oncogenic properties of key molecules such as Ras (reviewed in [108]), Wip1 [109], EGFR [110]), and urokinase plasminogen activator [111] among others. However, most of these studies address the function of p38 α specifically, with no reference to p38 β , or if so, discarding its implication in the tumor suppressor activity. Future work investigating the potential tumor suppressing activity of p38 β is needed to fully understand if p38 β can potentially act both as a proto-oncogene and as a tumor suppressor gene in cancer pathology.

From the therapeutic point of view, the implication of p38MAPK signaling pathway in the mechanism of action of several anti-cancer drugs has been widely studied, but most of these studies have focused on the role of p38 α [112]. In fact, p38 α has been connected to DNA damage-response [113] through its relation with key proteins in DNA damage such as ATM or p53 [114]. p38 α has been proposed as a master regulator of the apoptotic effects triggered by genotoxic drugs [115], and also as a

central part of the cellular response to ionizing radiation [116]. However, no data involving specifically p38 β has been published. There are only few examples suggesting a role for p38 β in response to cancer therapy. For instance, in leukemia-derived cell lines, both p38 α and p38 β have been linked to interferon- α , leading to an inhibition of the cellular growth [117]. Moreover, p38 β has been proposed as a key molecule in the stimulation of cell death triggered by the p38 α / β inhibitor SB202190, UV, and FasL indicating a role in cytotoxicity [118]. Regarding other commonly used cancer treatments, such as chemo/radiotherapy or immunotherapy, there are no studies addressing the implications of p38 β in response to these treatments, except for the one mentioned above in lung cancer [93], and for thymoquinone, a natural compound, in which its antitumor effect has been related to down regulation of p38 β [119]. The relevance "per se" of p38 β in cancer treatments, as a putative target, has been demonstrated in the previously mentioned experimental model of pancreatic cancer by using specific p38 β inhibitory peptides [85] but, unfortunately, no other examples of specific targeted therapy based on p38 β have been reported so far.

5. Future Directions

Although p38 β is the least studied member of the p38MAPK family, possibly due to its functional redundancy with p38 α , recent evidence shows that it may play a differential role with biological and pathological implications, as in the case of cancer. The lack of specific inhibitors for this MAPK has greatly complicated its study, since it involves the use of genetic approaches almost on a mandatory basis. Undoubtedly, the development of specific inhibitors for p38 β could accelerate the research of this MAPK. However, there are still aspects to be investigated in the coming years such as the role of p38 β in transcriptional regulation, its specific substrates, its involvement in the process of cell transformation and cancer (Figure 1), its implication in the cellular response to chemo and radiotherapy treatments, or even its use as a putative therapeutic target. Our knowledge of p38 β is increasing every day, similarly to other members of the p38 family, allowing us to unravel the complexity of p38MAPK signaling, and to further elucidate the specific roles of each p38 family member. However, as most of the current research is focused on p38 α , further studies on other p38 proteins, including p38 β are needed to fully understand the importance of the p38MAPK signaling in human pathology.

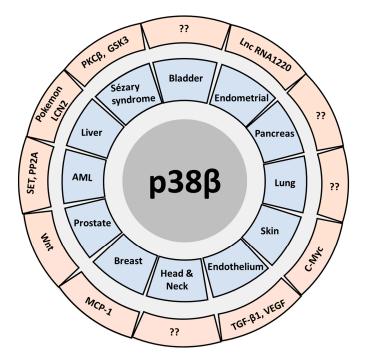


Figure 1. Schematic representation of the involvement of $p38\beta$ in different types of tumors (blue) and the related molecules (red).

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References

- 1. Kim, E.K.; Choi, E.-J. Pathological roles of MAPK signaling pathways in human diseases. *Biochim. Biophys. Acta* **2010**, *1802*, 396–405. [CrossRef]
- 2. Morrison, D.K.; Davis, R.J. Regulation of MAP kinase signaling modules by scaffold proteins in mammals. *Annu. Rev. Cell Dev. Biol.* **2003**, *19*, 91–118. [CrossRef] [PubMed]
- 3. Peti, W.; Page, R. Molecular basis of MAP kinase regulation. Protein Sci. 2013, 22, 1698–1710. [CrossRef]
- 4. Cargnello, M.; Roux, P.P. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol. Mol. Biol. Rev.* **2011**, *75*, 50–83. [CrossRef] [PubMed]
- 5. Coulombe, P.; Meloche, S. Atypical mitogen-activated protein kinases: Structure, regulation and functions. *Biochim. Biophys. Acta* 2007, 1773, 1376–1387. [CrossRef] [PubMed]
- Kuo, W.-L.; Duke, C.J.; Abe, M.K.; Kaplan, E.L.; Gomes, S.; Rosner, M.R. ERK7 expression and kinase activity is regulated by the ubiquitin-proteosome pathway. *J. Biol. Chem.* 2004, 279, 23073–23081. [CrossRef] [PubMed]
- Ishitani, T.; Ishitani, S. Nemo-like kinase, a multifaceted cell signaling regulator. *Cell. Signal.* 2013, 25, 190–197. [CrossRef]
- Abe, M.K.; Saelzler, M.P.; Espinosa, R.; Kahle, K.T.; Hershenson, M.B.; Le Beau, M.M.; Rosner, M.R. ERK8, a new member of the mitogen-activated protein kinase family. *J. Biol. Chem.* 2002, 277, 16733–16743. [CrossRef]
- 9. Yokota, T.; Wang, Y. p38 MAP kinases in the heart. *Gene* 2016, 575, 369–376. [CrossRef]
- 10. Han, J.; Wu, J.; Silke, J. An overview of mammalian p38 mitogen-activated protein kinases, central regulators of cell stress and receptor signaling. *F1000Res* **2020**, *9*, 653. [CrossRef]
- 11. Remy, G.; Risco, A.M.; Iñesta-Vaquera, F.A.; González-Terán, B.; Sabio, G.; Davis, R.J.; Cuenda, A. Differential activation of p38MAPK isoforms by MKK6 and MKK3. *Cell. Signal.* **2010**, *22*, 660–667. [CrossRef] [PubMed]
- 12. Dérijard, B.; Raingeaud, J.; Barrett, T.; Wu, I.H.; Han, J.; Ulevitch, R.J.; Davis, R.J. Independent human MAP-kinase signal transduction pathways defined by MEK and MKK isoforms. *Science* **1995**, 267, 682–685. [CrossRef] [PubMed]
- 13. Martínez-Limón, A.; Joaquin, M.; Caballero, M.; Posas, F.; de Nadal, E. The p38 Pathway: From Biology to Cancer Therapy. *Int. J. Mol. Sci.* 2020, *21*, 1913. [CrossRef] [PubMed]
- 14. Salvador, J.M.; Mittelstadt, P.R.; Guszczynski, T.; Copeland, T.D.; Yamaguchi, H.; Appella, E.; Fornace, A.J.; Ashwell, J.D. Alternative p38 activation pathway mediated by T cell receptor-proximal tyrosine kinases. *Nat. Immunol.* **2005**, *6*, 390–395. [CrossRef] [PubMed]
- 15. Peregrin, S.; Jurado-Pueyo, M.; Campos, P.M.; Sanz-Moreno, V.; Ruiz-Gomez, A.; Crespo, P.; Mayor, F.; Murga, C. Phosphorylation of p38 by GRK2 at the docking groove unveils a novel mechanism for inactivating p38MAPK. *Curr. Biol.* **2006**, *16*, 2042–2047. [CrossRef] [PubMed]
- Kumar, S.; McDonnell, P.C.; Gum, R.J.; Hand, A.T.; Lee, J.C.; Young, P.R. Novel homologues of CSBP/p38 MAP kinase: Activation, substrate specificity and sensitivity to inhibition by pyridinyl imidazoles. *Biochem. Biophys. Res. Commun.* 1997, 235, 533–538. [CrossRef] [PubMed]

- 17. Young, P.R.; McLaughlin, M.M.; Kumar, S.; Kassis, S.; Doyle, M.L.; McNulty, D.; Gallagher, T.F.; Fisher, S.; McDonnell, P.C.; Carr, S.A.; et al. Pyridinyl imidazole inhibitors of p38 mitogen-activated protein kinase bind in the ATP site. *J. Biol. Chem.* **1997**, *272*, 12116–12121. [CrossRef] [PubMed]
- 18. Haller, V.; Nahidino, P.; Forster, M.; Laufer, S.A. An updated patent review of p38 MAP kinase inhibitors (2014–2019). *Expert Opin. Ther. Pat.* **2020**, *30*, 453–466. [CrossRef]
- 19. Trempolec, N.; Dave-Coll, N.; Nebreda, A.R. SnapShot: p38 MAPK substrates. *Cell* **2013**, *152*, 924–924.e1. [CrossRef]
- 20. Jiang, Y.; Chen, C.; Li, Z.; Guo, W.; Gegner, J.A.; Lin, S.; Han, J. Characterization of the structure and function of a new mitogen-activated protein kinase (p38beta). *J. Biol. Chem.* **1996**, 271, 17920–17926. [CrossRef]
- Stein, B.; Yang, M.X.; Young, D.B.; Janknecht, R.; Hunter, T.; Murray, B.W.; Barbosa, M.S. p38-2, a novel mitogen-activated protein kinase with distinct properties. *J. Biol. Chem.* 1997, 272, 19509–19517. [CrossRef] [PubMed]
- Patel, S.B.; Cameron, P.M.; O'Keefe, S.J.; Frantz-Wattley, B.; Thompson, J.; O'Neill, E.A.; Tennis, T.; Liu, L.; Becker, J.W.; Scapin, G. The three-dimensional structure of MAP kinase p38beta: Different features of the ATP-binding site in p38beta compared with p38alpha. *Acta Crystallogr. D Biol. Crystallogr.* 2009, 65, 777–785.
 [CrossRef] [PubMed]
- Hale, K.K.; Trollinger, D.; Rihanek, M.; Manthey, C.L. Differential expression and activation of p38 mitogen-activated protein kinase alpha, beta, gamma, and delta in inflammatory cell lineages. *J. Immunol.* 1999, 162, 4246–4252. [PubMed]
- Johnston, S.J.; Ahmad, D.; Aleskandarany, M.A.; Kurozumi, S.; Nolan, C.C.; Diez-Rodriguez, M.; Green, A.R.; Rakha, E.A. Co-expression of nuclear P38 and hormone receptors is prognostic of good long-term clinical outcome in primary breast cancer and is linked to upregulation of DNA repair. *BMC Cancer* 2018, 18, 1027. [CrossRef]
- 25. Enslen, H.; Raingeaud, J.; Davis, R.J. Selective activation of p38 mitogen-activated protein (MAP) kinase isoforms by the MAP kinase kinases MKK3 and MKK6. *J. Biol. Chem.* **1998**, 273, 1741–1748. [CrossRef] [PubMed]
- 26. Keesler, G.A.; Bray, J.; Hunt, J.; Johnson, D.A.; Gleason, T.; Yao, Z.; Wang, S.W.; Parker, C.; Yamane, H.; Cole, C.; et al. Purification and activation of recombinant p38 isoforms alpha, beta, gamma, and delta. *Protein Expr. Purif.* **1998**, *14*, 221–228. [CrossRef]
- Beenstock, J.; Ben-Yehuda, S.; Melamed, D.; Admon, A.; Livnah, O.; Ahn, N.G.; Engelberg, D. The p38β Mitogen-activated Protein Kinase Possesses an Intrinsic Autophosphorylation Activity, Generated by a Short Region Composed of the α-G Helix and MAPK Insert. J. Biol. Chem. 2014, 289, 23546–23556. [CrossRef]
- Beenstock, J.; Melamed, D.; Mooshayef, N.; Mordechay, D.; Garfinkel, B.P.; Ahn, N.G.; Admon, A.; Engelberg, D. p38β Mitogen-Activated Protein Kinase Modulates Its Own Basal Activity by Autophosphorylation of the Activating Residue Thr180 and the Inhibitory Residues Thr241 and Ser261. *Mol. Cell. Biol.* 2016, *36*, 1540–1554. [CrossRef]
- del Barco Barrantes, I.; Coya, J.M.; Maina, F.; Arthur, J.S.C.; Nebreda, A.R. Genetic analysis of specific and redundant roles for p38alpha and p38beta MAPKs during mouse development. *Proc. Natl. Acad. Sci. USA* 2011, 108, 12764–12769. [CrossRef]
- Adams, R.H.; Porras, A.; Alonso, G.; Jones, M.; Vintersten, K.; Panelli, S.; Valladares, A.; Perez, L.; Klein, R.; Nebreda, A.R. Essential role of p38alpha MAP kinase in placental but not embryonic cardiovascular development. *Mol. Cell* 2000, *6*, 109–116. [CrossRef]
- Beardmore, V.A.; Hinton, H.J.; Eftychi, C.; Apostolaki, M.; Armaka, M.; Darragh, J.; McIlrath, J.; Carr, J.M.; Armit, L.J.; Clacher, C.; et al. Generation and characterization of p38beta (MAPK11) gene-targeted mice. *Mol. Cell. Biol.* 2005, 25, 10454–10464. [CrossRef] [PubMed]
- 32. Gaestel, M. MAPKAP kinases-MKs-two's company, three's a crowd. *Nat. Rev. Mol. Cell Biol.* **2006**, *7*, 120–130. [CrossRef] [PubMed]
- 33. Llopis, A.; Salvador, N.; Ercilla, A.; Guaita-Esteruelas, S.; Barrantes, I.D.B.; Gupta, J.; Gaestel, M.; Davis, R.J.; Nebreda, A.R.; Agell, N. The stress-activated protein kinases p38α/β and JNK1/2 cooperate with Chk1 to inhibit mitotic entry upon DNA replication arrest. *Cell Cycle* 2012, *11*, 3627–3637. [CrossRef] [PubMed]
- 34. Clifton, A.D.; Young, P.R.; Cohen, P. A comparison of the substrate specificity of MAPKAP kinase-2 and MAPKAP kinase-3 and their activation by cytokines and cellular stress. *FEBS Lett.* **1996**, *392*, 209–214. [CrossRef]

- 35. New, L.; Jiang, Y.; Zhao, M.; Liu, K.; Zhu, W.; Flood, L.J.; Kato, Y.; Parry, G.C.; Han, J. PRAK, a novel protein kinase regulated by the p38 MAP kinase. *EMBO J.* **1998**, *17*, 3372–3384. [CrossRef]
- New, L.; Jiang, Y.; Han, J. Regulation of PRAK subcellular location by p38 MAP kinases. *Mol. Biol. Cell* 2003, 14, 2603–2616. [CrossRef]
- 37. Saurin, A.T.; Durgan, J.; Cameron, A.J.; Faisal, A.; Marber, M.S.; Parker, P.J. The regulated assembly of a PKCepsilon complex controls the completion of cytokinesis. *Nat. Cell Biol.* **2008**, *10*, 891–901. [CrossRef]
- Pon, J.R.; Marra, M.A. MEF2 transcription factors: Developmental regulators and emerging cancer genes. Oncotarget 2015, 7, 2297–2312. [CrossRef]
- Yang, S.-H.; Galanis, A.; Sharrocks, A.D. Targeting of p38 Mitogen-Activated Protein Kinases to MEF2 Transcription Factors. *Mol. Cell. Biol.* 1999, 19, 4028–4038. [CrossRef]
- 40. Han, J.; Jiang, Y.; Li, Z.; Kravchenko, V.V.; Ulevitch, R.J. Activation of the transcription factor MEF2C by the MAP kinase p38 in inflammation. *Nature* **1997**, *386*, 296–299. [CrossRef]
- Ho, I.C.; Kim, J.H.; Rooney, J.W.; Spiegelman, B.M.; Glimcher, L.H. A potential role for the nuclear factor of activated T cells family of transcriptional regulatory proteins in adipogenesis. *Proc. Natl. Acad. Sci. USA* 1998, 95, 15537–15541. [CrossRef] [PubMed]
- 42. Bushdid, P.B.; Osinska, H.; Waclaw, R.R.; Molkentin, J.D.; Yutzey, K.E. NFATc3 and NFATc4 are required for cardiac development and mitochondrial function. *Circ. Res.* **2003**, *92*, 1305–1313. [CrossRef] [PubMed]
- 43. Yang, T.T.C.; Xiong, Q.; Enslen, H.; Davis, R.J.; Chow, C.-W. Phosphorylation of NFATc4 by p38 mitogen-activated protein kinases. *Mol. Cell. Biol.* 2002, 22, 3892–3904. [CrossRef] [PubMed]
- Yang, T.T.C.; Yu, R.Y.L.; Agadir, A.; Gao, G.-J.; Campos-Gonzalez, R.; Tournier, C.; Chow, C.-W. Integration of Protein Kinases mTOR and Extracellular Signal-Regulated Kinase 5 in Regulating Nucleocytoplasmic Localization of NFATc4. *Mol. Cell. Biol.* 2008, 28, 3489–3501. [CrossRef] [PubMed]
- 45. Shaulian, E.; Karin, M. AP-1 as a regulator of cell life and death. Nat. Cell Biol. 2002, 4, E131–E136. [CrossRef]
- Tanos, T.; Marinissen, M.J.; Leskow, F.C.; Hochbaum, D.; Martinetto, H.; Gutkind, J.S.; Coso, O.A. Phosphorylation of c-Fos by members of the p38 MAPK family. Role in the AP-1 response to UV light. *J. Biol. Chem.* 2005, 280, 18842–18852. [CrossRef] [PubMed]
- 47. Cuenda, A.; Cohen, P.; Buée-Scherrer, V.; Goedert, M. Activation of stress-activated protein kinase-3 (SAPK3) by cytokines and cellular stresses is mediated via SAPKK3 (MKK6); comparison of the specificities of SAPK3 and SAPK2 (RK/p38). *EMBO J.* **1997**, *16*, 295–305. [CrossRef]
- Mahlknecht, U.; Will, J.; Varin, A.; Hoelzer, D.; Herbein, G. Histone deacetylase 3, a class I histone deacetylase, suppresses MAPK11-mediated activating transcription factor-2 activation and represses TNF gene expression. *J. Immunol.* 2004, 173, 3979–3990. [CrossRef]
- Kaneto, H.; Matsuoka, T. Role of pancreatic transcription factors in maintenance of mature β-cell function. *Int. J. Mol. Sci.* 2015, *16*, 6281–6297. [CrossRef]
- 50. Sii-Felice, K.; Pouponnot, C.; Gillet, S.; Lecoin, L.; Girault, J.-A.; Eychène, A.; Felder-Schmittbuhl, M.-P. MafA transcription factor is phosphorylated by p38 MAP kinase. *FEBS Lett.* **2005**, *579*, 3547–3554. [CrossRef]
- 51. Simone, C.; Forcales, S.V.; Hill, D.A.; Imbalzano, A.N.; Latella, L.; Puri, P.L. p38 pathway targets SWI-SNF chromatin-remodeling complex to muscle-specific loci. *Nat. Genet.* **2004**, *36*, 738–743. [CrossRef] [PubMed]
- Forcales, S.V.; Albini, S.; Giordani, L.; Malecova, B.; Cignolo, L.; Chernov, A.; Coutinho, P.; Saccone, V.; Consalvi, S.; Williams, R.; et al. Signal-dependent incorporation of MyoD–BAF60c into Brg1-based SWI/SNF chromatin-remodelling complex. *EMBO J.* 2012, *31*, 301–316. [CrossRef] [PubMed]
- 53. Page, J.L.; Wang, X.; Sordillo, L.M.; Johnson, S.E. MEKK1 signaling through p38 leads to transcriptional inactivation of E47 and repression of skeletal myogenesis. *J. Biol. Chem.* **2004**, *279*, 30966–30972. [CrossRef]
- 54. Cuadrado, A.; Lafarga, V.; Cheung, P.C.F.; Dolado, I.; Llanos, S.; Cohen, P.; Nebreda, A.R. A new p38 MAP kinase-regulated transcriptional coactivator that stimulates p53-dependent apoptosis. *EMBO J.* **2007**, *26*, 2115–2126. [CrossRef]
- Cuadrado, A.; Corrado, N.; Perdiguero, E.; Lafarga, V.; Muñoz-Canoves, P.; Nebreda, A.R. Essential role of p18Hamlet/SRCAP-mediated histone H2A.Z chromatin incorporation in muscle differentiation. *EMBO J.* 2010, 29, 2014–2025. [CrossRef] [PubMed]
- 56. Casanovas, O.; Jaumot, M.; Paules, A.-B.; Agell, N.; Bachs, O. P38SAPK2 phosphorylates cyclin D3 at Thr-283 and targets it for proteasomal degradation. *Oncogene* **2004**, *23*, 7537–7544. [CrossRef]
- 57. Lu, C.; Shi, Y.; Wang, Z.; Song, Z.; Zhu, M.; Cai, Q.; Chen, T. Serum starvation induces H2AX phosphorylation to regulate apoptosis via p38 MAPK pathway. *FEBS Lett.* **2008**, *582*, 2703–2708. [CrossRef] [PubMed]

- Briata, P.; Forcales, S.V.; Ponassi, M.; Corte, G.; Chen, C.-Y.; Karin, M.; Puri, P.L.; Gherzi, R. p38-dependent phosphorylation of the mRNA decay-promoting factor KSRP controls the stability of select myogenic transcripts. *Mol. Cell* 2005, 20, 891–903. [CrossRef] [PubMed]
- 59. Xu, P.; Derynck, R. Direct activation of TACE-mediated ectodomain shedding by p38 MAP kinase regulates EGF receptor-dependent cell proliferation. *Mol. Cell* **2010**, *37*, 551–566. [CrossRef] [PubMed]
- 60. Kuma, Y.; Campbell, D.G.; Cuenda, A. Identification of glycogen synthase as a new substrate for stress-activated protein kinase 2b/p38beta. *Biochem. J.* **2004**, *379*, 133–139. [CrossRef]
- 61. Wu, X.-N.; Wang, X.-K.; Wu, S.-Q.; Lu, J.; Zheng, M.; Wang, Y.-H.; Zhou, H.; Zhang, H.; Han, J. Phosphorylation of Raptor by p38beta participates in arsenite-induced mammalian target of rapamycin complex 1 (mTORC1) activation. *J. Biol. Chem.* **2011**, *286*, 31501–31511. [CrossRef] [PubMed]
- 62. Yu, M.; Fu, Y.; Liang, Y.; Song, H.; Yao, Y.; Wu, P.; Yao, Y.; Pan, Y.; Wen, X.; Ma, L.; et al. Suppression of MAPK11 or HIPK3 reduces mutant Huntingtin levels in Huntington's disease models. *Cell Res.* **2017**, *27*, 1441–1465. [CrossRef]
- Zhang, S.; Weinheimer, C.; Courtois, M.; Kovacs, A.; Zhang, C.E.; Cheng, A.M.; Wang, Y.; Muslin, A.J. The role of the Grb2-p38 MAPK signaling pathway in cardiac hypertrophy and fibrosis. *J. Clin. Investig.* 2003, *111*, 833–841. [CrossRef] [PubMed]
- 64. Sharma, B.; Chaube, U.; Patel, B.M. Beneficial Effect of Silymarin in Pressure Overload Induced Experimental Cardiac Hypertrophy. *Cardiovasc. Toxicol.* **2019**, *19*, 23–35. [CrossRef] [PubMed]
- Pandey, P.; Raingeaud, J.; Kaneki, M.; Weichselbaum, R.; Davis, R.J.; Kufe, D.; Kharbanda, S. Activation of p38 mitogen-activated protein kinase by c-Abl-dependent and -independent mechanisms. *J. Biol. Chem.* 1996, 271, 23775–23779. [CrossRef] [PubMed]
- 66. Gupta, J.; del Barco Barrantes, I.; Igea, A.; Sakellariou, S.; Pateras, I.S.; Gorgoulis, V.G.; Nebreda, A.R. Dual function of p38α MAPK in colon cancer: Suppression of colitis-associated tumor initiation but requirement for cancer cell survival. *Cancer Cell* 2014, 25, 484–500. [CrossRef] [PubMed]
- 67. Vitos-Faleato, J.; Real, S.M.; Gutierrez-Prat, N.; Villanueva, A.; Llonch, E.; Drosten, M.; Barbacid, M.; Nebreda, A.R. Requirement for epithelial p38α in KRAS-driven lung tumor progression. *Proc. Natl. Acad. Sci. USA* 2020, 117, 2588–2596. [CrossRef]
- 68. Sahu, V.; Mohan, A.; Dey, S. p38 MAP kinases: Plausible diagnostic and prognostic serum protein marker of non small cell lung cancer. *Exp. Mol. Pathol.* **2019**, *107*, 118–123. [CrossRef]
- 69. Roseweir, A.K.; Halcrow, E.S.; Chichilo, S.; Powell, A.G.; McMillan, D.C.; Horgan, P.G.; Edwards, J. ERK and p38MAPK combine to improve survival in patients with BRAF mutant colorectal cancer. *Br. J. Cancer* **2018**, *119*, 323–329. [CrossRef]
- Igea, A.; Nebreda, A.R. The Stress Kinase p38α as a Target for Cancer Therapy. *Cancer Res.* 2015, 75, 3997–4002.
 [CrossRef]
- 71. Del Reino, P.; Alsina-Beauchamp, D.; Escós, A.; Cerezo-Guisado, M.I.; Risco, A.; Aparicio, N.; Zur, R.; Fernandez-Estévez, M.; Collantes, E.; Montans, J.; et al. Pro-oncogenic role of alternative p38 mitogenactivated protein kinases p38γ and p38δ, linking inflammation and cancer in colitis-associated colon cancer. *Cancer Res.* 2014, 74, 6150–6160. [CrossRef] [PubMed]
- 72. Zur, R.; Garcia-Ibanez, L.; Nunez-Buiza, A.; Aparicio, N.; Liappas, G.; Escós, A.; Risco, A.; Page, A.; Saiz-Ladera, C.; Alsina-Beauchamp, D.; et al. Combined deletion of p38γ and p38δ reduces skin inflammation and protects from carcinogenesis. *Oncotarget* **2015**, *6*, 12920–12935. [CrossRef] [PubMed]
- Cuenda, A.; Sanz-Ezquerro, J.J. p38γ and p38δ: From Spectators to Key Physiological Players. *Trends Biochem. Sci.* 2017, 42, 431–442. [CrossRef] [PubMed]
- 74. Chen, Z.; Liu, F.; Zhang, N.; Cao, D.; Liu, M.; Tan, Y.; Jiang, Y. p38β, a novel regulatory target of Pokemon in hepatic cells. *Int. J. Mol. Sci.* **2013**, *14*, 13511–13524. [CrossRef] [PubMed]
- Santiago-Sánchez, G.S.; Pita-Grisanti, V.; Quiñones-Díaz, B.; Gumpper, K.; Cruz-Monserrate, Z.; Vivas-Mejía, P.E. Biological Functions and Therapeutic Potential of Lipocalin 2 in Cancer. *Int. J. Mol. Sci.* 2020, 21, 4365. [CrossRef] [PubMed]
- Basu, S.; Chaudhary, N.; Shah, S.; Braggs, C.; Sawant, A.; Vaz, S.; Thorat, R.; Gupta, S.; Dalal, S.N. Plakophilin3 loss leads to an increase in lipocalin2 expression, which is required for tumour formation. *Exp. Cell Res.* 2018, 369, 251–265. [CrossRef]
- 77. Duperret, E.K.; Dahal, A.; Ridky, T.W. Focal-adhesion-independent integrin-αv regulation of FAK and c-Myc is necessary for 3D skin formation and tumor invasion. *J. Cell Sci.* **2015**, *128*, 3997–4013. [CrossRef]

- 78. Ferrari, G.; Terushkin, V.; Wolff, M.J.; Zhang, X.; Valacca, C.; Poggio, P.; Pintucci, G.; Mignatti, P. TGF-β1 induces endothelial cell apoptosis by shifting VEGF activation of p38(MAPK) from the prosurvival p38β to proapoptotic p38α. *Mol. Cancer Res.* 2012, *10*, 605–614. [CrossRef]
- 79. Kim, J.K.; Pedram, A.; Razandi, M.; Levin, E.R. Estrogen prevents cardiomyocyte apoptosis through inhibition of reactive oxygen species and differential regulation of p38 kinase isoforms. *J. Biol. Chem.* **2006**, *281*, 6760–6767. [CrossRef]
- 80. Kumar, R.; Janjanam, J.; Singh, N.K.; Rao, G.N. A new role for cofilin in retinal neovascularization. *J. Cell. Sci.* **2016**, *129*, 1234–1249. [CrossRef]
- Liu, Z.; Sin, K.W.T.; Ding, H.; Doan, H.A.; Gao, S.; Miao, H.; Wei, Y.; Wang, Y.; Zhang, G.; Li, Y.-P. p38β MAPK mediates ULK1-dependent induction of autophagy in skeletal muscle of tumor-bearing mice. *Cell Stress* 2018, 2, 311–324. [CrossRef] [PubMed]
- Lin, A.; Yao, J.; Zhuang, L.; Wang, D.; Han, J.; Lam, E.W.-F.; TCGA Research Network; Gan, B. The FoxO-BNIP3 axis exerts a unique regulation of mTORC1 and cell survival under energy stress. *Oncogene* 2014, 33, 3183–3194. [CrossRef] [PubMed]
- 83. Dong, H.; Xiang, H.-B.; Ye, D.-W.; Tian, X.-B. Inhibitory effects of intrathecal p38β antisense oligonucleotide on bone cancer pain in rats. *Int. J. Clin. Exp. Pathol.* **2014**, *7*, 7690–7698. [PubMed]
- 84. Gao, J.; Chen, S.; Lin, S.; Han, H. Effect of music therapy on pain behaviors in rats with bone cancer pain. *J. BUON* **2016**, *21*, 466–472. [PubMed]
- 85. Singh, A.K.; Pandey, R.; Gill, K.; Singh, R.; Saraya, A.; Chauhan, S.S.; Yadav, S.; Pal, S.; Singh, N.; Dey, S. p38β MAP kinase as a therapeutic target for pancreatic cancer. *Chem. Biol. Drug Des.* **2012**, *80*, 266–273. [CrossRef]
- Yao, Z.; Xu, R.; Yuan, L.; Xu, M.; Zhuang, H.; Li, Y.; Zhang, Y.; Lin, N. Circ_0001955 facilitates hepatocellular carcinoma (HCC) tumorigenesis by sponging miR-516a-5p to release TRAF6 and MAPK11. *Cell Death Dis.* 2019, *10*, 945. [CrossRef] [PubMed]
- Yu, L.; Yuan, X.; Wang, D.; Barakat, B.; Williams, E.D.; Hannigan, G.E. Selective regulation of p38β protein and signaling by integrin-linked kinase mediates bladder cancer cell migration. *Oncogene* 2014, 33, 690–701. [CrossRef]
- 88. Browne, A.J.; Göbel, A.; Thiele, S.; Hofbauer, L.C.; Rauner, M.; Rachner, T.D. p38 MAPK regulates the Wnt inhibitor Dickkopf-1 in osteotropic prostate cancer cells. *Cell Death Dis.* **2016**, *7*, e2119. [CrossRef]
- Grossi, V.; Lucarelli, G.; Forte, G.; Peserico, A.; Matrone, A.; Germani, A.; Rutigliano, M.; Stella, A.; Bagnulo, R.; Loconte, D.; et al. Loss of STK11 expression is an early event in prostate carcinogenesis and predicts therapeutic response to targeted therapy against MAPK/p38. *Autophagy* 2015, *11*, 2102–2113. [CrossRef]
- 90. Li, Y.; Kong, C.; Wu, C.; Wang, Y.; Xu, B.; Liang, S.; Ying, X. Knocking down of LINC01220 inhibits proliferation and induces apoptosis of endometrial carcinoma through silencing MAPK11. *Biosci. Rep.* 2019, 39. [CrossRef]
- 91. Wu, H.; Meng, S.; Xu, Q.; Wang, X.; Wang, J.; Gong, R.; Song, Y.; Duan, Y.; Zhang, Y. Gene expression profiling of lung adenocarcinoma in Xuanwei, China. *Eur. J. Cancer Prev.* **2016**, *25*, 508–517. [CrossRef] [PubMed]
- 92. Sullivan, I.; Riera, P.; Andrés, M.; Altés, A.; Majem, M.; Blanco, R.; Capdevila, L.; Barba, A.; Barnadas, A.; Salazar, J. Prognostic effect of VEGF gene variants in metastatic non-small-cell lung cancer patients. *Angiogenesis* 2019, 22, 433–440. [CrossRef] [PubMed]
- Planchard, D.; Camara-Clayette, V.; Dorvault, N.; Soria, J.-C.; Fouret, P. p38 Mitogen-activated protein kinase signaling, ERCC1 expression, and viability of lung cancer cells from never or light smoker patients. *Cancer* 2012, 118, 5015–5025. [CrossRef] [PubMed]
- 94. He, Z.; He, J.; Liu, Z.; Xu, J.; Yi, S.F.; Liu, H.; Yang, J. MAPK11 in breast cancer cells enhances osteoclastogenesis and bone resorption. *Biochimie* **2014**, *106*, 24–32. [CrossRef]
- 95. Huang, J.; Luo, Q.; Xiao, Y.; Li, H.; Kong, L.; Ren, G. The implication from RAS/RAF/ERK signaling pathway increased activation in epirubicin treated triple negative breast cancer. *Oncotarget* **2017**, *8*, 108249–108260. [CrossRef]
- 96. Sahu, V.; Nigam, L.; Agnihotri, V.; Gupta, A.; Shekhar, S.; Subbarao, N.; Bhaskar, S.; Dey, S. Diagnostic Significance of p38 Isoforms (p38α, p38β, p38γ, p38δ) in Head and Neck Squamous Cell Carcinoma: Comparative Serum Level Evaluation and Design of Novel Peptide Inhibitor Targeting the Same. *Cancer Res. Treat.* 2019, *51*, 313–325. [CrossRef]

- Cristóbal, I.; Garcia-Orti, L.; Cirauqui, C.; Cortes-Lavaud, X.; García-Sánchez, M.A.; Calasanz, M.J.; Odero, M.D. Overexpression of SET is a recurrent event associated with poor outcome and contributes to protein phosphatase 2A inhibition in acute myeloid leukemia. *Haematologica* 2012, 97, 543–550. [CrossRef]
- 98. Arriazu, E.; Vicente, C.; Pippa, R.; Peris, I.; Martínez-Balsalobre, E.; García-Ramírez, P.; Marcotegui, N.; Igea, A.; Alignani, D.; Rifón, J.; et al. A new regulatory mechanism of protein phosphatase 2A activity via SET in acute myeloid leukemia. *Blood Cancer J.* 2020, *10*, 3. [CrossRef]
- 99. Arriazu, E.; Pippa, R.; Odero, M.D. Protein Phosphatase 2A as a Therapeutic Target in Acute Myeloid Leukemia. *Front. Oncol.* **2016**, *6*, 78. [CrossRef]
- 100. Bliss-Moreau, M.; Coarfa, C.; Gunaratne, P.H.; Guitart, J.; Krett, N.L.; Rosen, S.T. Identification of p38β as a therapeutic target for the treatment of Sézary syndrome. *J. Investig. Dermatol.* 2015, 135, 599–608. [CrossRef] [PubMed]
- 101. Wen, S.-Y.; Cheng, S.-Y.; Ng, S.-C.; Aneja, R.; Chen, C.-J.; Huang, C.-Y.; Kuo, W.-W. Roles of p38α and p38β mitogen-activated protein kinase isoforms in human malignant melanoma A375 cells. *Int. J. Mol. Med.* **2019**, 44, 2123–2132. [CrossRef] [PubMed]
- 102. Huang, Q.; Chen, D.; Song, S.; Fu, X.; Wei, Y.; Lu, J.; Wang, L.; Wang, J. A genetic variation of the p38β promoter region is correlated with an increased risk of sporadic colorectal cancer. *Oncol. Lett.* **2013**, *6*, 3–8. [CrossRef] [PubMed]
- 103. Dimberg, J.; Olsen, R.S.; Skarstedt, M.; Löfgren, S.; Zar, N.; Matussek, A. Polymorphism of the p38β gene in patients with colorectal cancer. *Oncol. Lett.* **2014**, *8*, 1093–1095. [CrossRef] [PubMed]
- 104. Tian, X.; Sun, D.; Zhao, S.; Xiong, H.; Fang, J. Screening of potential diagnostic markers and therapeutic targets against colorectal cancer. *Onco Targets Ther.* **2015**, *8*, 1691–1699. [CrossRef] [PubMed]
- 105. Gupta, J.; Igea, A.; Papaioannou, M.; Lopez-Casas, P.P.; Llonch, E.; Hidalgo, M.; Gorgoulis, V.G.; Nebreda, A.R. Pharmacological inhibition of p38 MAPK reduces tumor growth in patient-derived xenografts from colon tumors. *Oncotarget* 2015, *6*, 8539–8551. [CrossRef] [PubMed]
- Engelberg, D. Stress-activated protein kinases-tumor suppressors or tumor initiators? *Semin. Cancer Biol.* 2004, 14, 271–282. [CrossRef]
- 107. Wagner, E.F.; Nebreda, A.R. Signal integration by JNK and p38 MAPK pathways in cancer development. *Nat. Rev. Cancer* **2009**, *9*, 537–549. [CrossRef]
- Loesch, M.; Chen, G. The p38 MAPK stress pathway as a tumor suppressor or more? *Front. Biosci. A J. Virtual Libr.* 2008, 13, 3581–3593. [CrossRef]
- 109. Bulavin, D.V.; Phillips, C.; Nannenga, B.; Timofeev, O.; Donehower, L.A.; Anderson, C.W.; Appella, E.; Fornace, A.J. Inactivation of the Wip1 phosphatase inhibits mammary tumorigenesis through p38 MAPK-mediated activation of the p16(Ink4a)-p19(Arf) pathway. *Nat. Genet.* 2004, *36*, 343–350. [CrossRef]
- 110. Frey, M.R.; Dise, R.S.; Edelblum, K.L.; Polk, D.B. p38 kinase regulates epidermal growth factor receptor downregulation and cellular migration. *EMBO J.* **2006**, *25*, 5683–5692. [CrossRef]
- 111. Han, Q.; Leng, J.; Bian, D.; Mahanivong, C.; Carpenter, K.A.; Pan, Z.K.; Han, J.; Huang, S. Rac1-MKK3p38-MAPKAPK2 pathway promotes urokinase plasminogen activator mRNA stability in invasive breast cancer cells. J. Biol. Chem. 2002, 277, 48379–48385. [CrossRef] [PubMed]
- 112. García-Cano, J.; Roche, O.; Cimas, F.J.; Pascual-Serra, R.; Ortega-Muelas, M.; Fernández-Aroca, D.M.; Sánchez-Prieto, R. p38MAPK and Chemotherapy: We Always Need to Hear Both Sides of the Story. *Front. Cell Dev. Biol.* 2016, 4, 69. [CrossRef] [PubMed]
- 113. Cánovas, B.; Igea, A.; Sartori, A.A.; Gomis, R.R.; Paull, T.T.; Isoda, M.; Pérez-Montoyo, H.; Serra, V.; González-Suárez, E.; Stracker, T.H.; et al. Targeting p38α Increases DNA Damage, Chromosome Instability, and the Anti-tumoral Response to Taxanes in Breast Cancer Cells. *Cancer Cell* **2018**, *33*, 1094–1110.e8. [CrossRef]
- Roy, S.; Roy, S.; Rana, A.; Akhter, Y.; Hande, M.P.; Banerjee, B. The role of p38 MAPK pathway in p53 compromised state and telomere mediated DNA damage response. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* 2018, 836, 89–97. [CrossRef]
- 115. Sui, X.; Kong, N.; Ye, L.; Han, W.; Zhou, J.; Zhang, Q.; He, C.; Pan, H. p38 and JNK MAPK pathways control the balance of apoptosis and autophagy in response to chemotherapeutic agents. *Cancer Lett.* **2014**, 344, 174–179. [CrossRef] [PubMed]
- Munshi, A.; Ramesh, R. Mitogen-activated protein kinases and their role in radiation response. *Genes Cancer* 2013, 4, 401–408. [CrossRef] [PubMed]

- 117. Lee, W.-H.; Liu, F.-H.; Lee, Y.-L.; Huang, H.-M. Interferon-alpha induces the growth inhibition of human T-cell leukaemia line Jurkat through p38alpha and p38beta. *J. Biochem.* **2010**, *147*, 645–650. [CrossRef] [PubMed]
- 118. Nemoto, S.; Xiang, J.; Huang, S.; Lin, A. Induction of apoptosis by SB202190 through inhibition of p38beta mitogen-activated protein kinase. *J. Biol. Chem.* **1998**, 273, 16415–16420. [CrossRef] [PubMed]
- 119. Abdelfadil, E.; Cheng, Y.-H.; Bau, D.-T.; Ting, W.-J.; Chen, L.-M.; Hsu, H.-H.; Lin, Y.-M.; Chen, R.-J.; Tsai, F.-J.; Tsai, C.-H.; et al. Thymoquinone induces apoptosis in oral cancer cells through p38β inhibition. *Am. J. Chin. Med.* **2013**, *41*, 683–696. [CrossRef]



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