




REVIEW ARTICLE

A roadmap to integrate astrocytes into Systems Neuroscience

Ksenia V. Kastanenka¹ | Rubén Moreno-Bote^{2,3} | Maurizio De Pittà⁴ |
 Gertrudis Perea⁵  | Abel Eraso-Pichot⁶ | Roser Masgrau⁶  | Kira E. Poskanzer⁷ |
 Elena Galea^{3,6} 

¹Department of Neurology, MassGeneral Institute for Neurodegenerative Diseases, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts

²Center for Brain and Cognition and Department of Information and Communications Technologies, Universitat Pompeu Fabra, Barcelona, Spain

³ICREA, Barcelona, Spain

⁴BCAM Basque Center for Applied Mathematics, Bilbao, Spain

⁵Instituto Cajal, CSIC, Madrid, Spain

⁶Institut de Neurociències i Departament de Bioquímica, Universitat Autònoma de Barcelona, Barcelona, Spain

⁷Department of Biochemistry & Biophysics, Neuroscience Graduate Program, and Kavli Institute for Fundamental Neuroscience, University of California-San Francisco, San Francisco, California

Correspondence

Ksenia V. Kastanenka, Department of Neurology, MassGeneral Institute for Neurodegenerative Diseases, Massachusetts General Hospital and Harvard Medical School, Boston, MA 02129, USA.
 Email: kkastanenka@mgh.harvard.edu

Kira E. Poskanzer, Department of Biochemistry & Biophysics, Neuroscience Graduate Program, and Kavli Institute for Fundamental Neuroscience, University of California, San Francisco, San Francisco, CA 94143, USA.
 Email: kira.poskanzer@ucsf.edu

Elena Galea, Institut de Neurociències i Departament de Bioquímica, Universitat Autònoma de Barcelona, Bellaterra, 08193 Barcelona, Spain.
 Email: Elena.Galea@uab.es

Funding information

Agència de Gestió d'Ajuts Universitaris i de Recerca (Spain), Grant/Award Number: 2017 SGR547; BCAM Severo Ochoa, Grant/Award Number: SEV-2017-0718; Eusko Jaurlaritz; Fundació La Caixa (Spain); Howard Hughes Medical Institute (HHMI, USA), Grant/Award Number: 55008742; Ministerio de Economía y Competitividad (MINECO, Spain), Grant/Award Numbers: BFU2016-75107-P, BFU2016-79735-P, BFU2017-85936-P, FLAGERA-PCIN-2015-162-C02-02; Ministerio de Educación, Cultura y Deporte (Spain),

Abstract

Systems neuroscience is still mainly a neuronal field, despite the plethora of evidence supporting the fact that astrocytes modulate local neural circuits, networks, and complex behaviors. In this article, we sought to identify which types of studies are necessary to establish whether astrocytes, beyond their well-documented homeostatic and metabolic functions, perform computations implementing mathematical algorithms that sub-serve coding and higher-brain functions. First, we reviewed Systems-like studies that include astrocytes in order to identify computational operations that these cells may perform, using Ca²⁺ transients as their encoding language. The analysis suggests that astrocytes may carry out canonical computations in a time scale of subseconds to seconds in sensory processing, neuromodulation, brain state, memory formation, fear, and complex homeostatic reflexes. Next, we propose a list of actions to gain insight into the outstanding question of which variables are encoded by such computations. The application of statistical analyses based on machine learning, such as dimensionality reduction and decoding in the context of complex behaviors, combined with connectomics of astrocyte–neuronal circuits, is, in our view, fundamental undertakings. We also discuss technical and analytical approaches to study neuronal and astrocytic populations simultaneously, and the inclusion of astrocytes in advanced modeling of neural circuits, as well as in theories currently under exploration such as predictive coding and energy-efficient coding. Clarifying the relationship between astrocytic Ca²⁺ and brain coding may represent a leap forward toward novel approaches in the study of astrocytes in health and disease.

Kira E. Poskanzer and Elena Galea contributed equally to this study.

Grant/Award Number: FPU13/05377;
National Institutes of Health (NIH, USA),
Grant/Award Number: R01NS099254;
National Science Foundation (NSF, USA),
Grant/Award Number: 1604544

KEYWORDS

astrocytes, decoding, dimensionality reduction, energy-efficient coding, predictive coding

1 | SYSTEMS NEUROSCIENCE IS PRIMARILY A NEURONAL FIELD

The study of the central nervous system (CNS) encompasses different levels of analysis: Molecular, cellular, anatomical, behavioral, cognitive, and systems. Systems Neuroscience aims at integrating these former fields, which have mostly grown independently. For example, Molecular Neuroscience has traditionally focused on the smallest functional level without a connection to cognition, whereas Behavioral Psychology and Psychophysics have typically studied cognition separately from its molecular and neuronal underpinnings. The overarching goal of Systems Neuroscience is to understand how neural circuits give rise to cognitive functions, emotions, and behavior by *simultaneously* recording neuronal activity and behavior at the highest spatiotemporal resolution possible.

Systems Neuroscience is arguably a field of neurons. A proof of this can be found in recent editions (2015–2017) of the three international conferences dedicated to Systems and Computational Neuroscience—Here, we will not dwell on what is “Systems” and what “Computational” since the two fields are highly overlapping and complementary. The conferences are the “Conference and Workshop on Neural Information Processing Systems” (NIPS), the “Organization for Computational Neurosciences” (OCNS), and “Computational and Systems Neuroscience” (COSYNE). Of approximately 3000 communications, fewer than 1% included non-neuronal cells. The pervasive use of the phrase “neural circuit” in the programs most of the time refers to computational integration of information embedded in neuronal biophysical substrates. The scarce attention to non-neuronal cells is puzzling, at least from the perspective of the astrocyte field, given the evidence that astrocytes contribute to circuit-based phenomena at the synaptic (Araque et al., 2014) and network (Poskanzer & Yuste, 2016) levels. Although efforts are being made in the US Brain Initiative and the European Human Brain Project to develop studies incorporating non-neuronal cells, it seems as though progress in astrocyte biology has advanced in parallel to Systems Neuroscience, and astrocytes have been excluded from unified theories of brain function, as previously noted (Poskanzer & Molofsky, 2018). Although extensive modeling of astrocytic Ca^{2+} fluxes exists (Manninen, Havela, & Linne, 2018), and sporadic studies have explored the application of astrocyte-based computations to artificial intelligence (Alvarellós-Gonzalez, Pazos, & Porto-Pazos, 2012; Porto-Pazos et al., 2011), astrocytes are characteristically missing from advanced *in silico* modeling of neural circuits (Capone et al., 2017; Deneve, Alemi, & Bourdoukan, 2017; Gjorgjieva, Drion, & Marder, 2016; Markram et al., 2015).

Is this exclusion justified because the mechanisms underlying the well-documented impact of astrocytes on neural circuits fall within the realm of intercellular signaling, homeostasis, and metabolism, which, although essential for the maintenance of neural circuits, may

not qualify as “computing” processes? Or, are astrocytes fundamental to the computational foundations of the brain? Later we will elaborate on what is and what is not computation, but rather than struggling to define “computation” we ask, instead, whether processes that take place in astrocytes participate in the implementation by neural circuits of processes sub-serving coding, complex behaviors, and higher brain functions. In other words, if computation is an emerging property of a given neural network (Yuste, 2015), do astrocytes help to shape such property beyond providing metabolic and homeostatic support? If they do, specific questions are whether there are niche(s) in Systems Neurosciences that would particularly profit from astrocyte idiosyncrasies and whether the impressive techniques and theoretical armamentarium deployed by Systems Neuroscience could be used—and are sufficient—to unravel possible astrocyte-based computations. In an early article in Computational Neuroscience, it was argued that *anatomical features provide valuable insights about how the CNS operates because the nervous system is a product of evolution, not design. The computational solutions evolved by nature may be unlike those that humans would invent, if only because evolutionary changes are always made within the context of design and architecture that already is in place* (Sejnowski, Koch, & Churchland, 1988). It follows that the unique anatomical arrangement between astrocytes and neurons might be part of computational solutions refined by evolution that has made the brain a highly efficient task-performing system. In this article, we will explore the possible computations carried out by astrocytes. First, we will succinctly describe the fundamentals (Section 2) and current challenges (Sections 3 and 4) of Systems Neuroscience. We will continue by reviewing Systems-like studies involving astrocytes (Sections 5 and 6). We will then propose a to-do list to further integrate astrocytes in Systems Neurosciences, thus helping to dissipate the historical and perhaps no longer tenable gap between astrocytes and neurons (Section 7). We do not touch upon other glial cells because, as discussed earlier (Masgrau, Guaza, Ransohoff, & Galea, 2017), the cells grouped under this name are molecularly and morphologically distinct; hence, their contribution to higher-brain functions deserves individual attention.

2 | COMPUTATIONAL FOUNDATIONS OF THE CNS

2.1 | What is computation?

When we say that the brain computes we mean that it creates and stores representations of physical and conceptual entities, and performs operations on these representations in order to carry out discrete tasks underlying behavior. The goal of Computational/Systems Neuroscience is to describe these processes in mathematical and

computational terms. In this framework, it is believed that the mathematical treatment of the representations is possible precisely because computation implies abstraction, thus permitting generation of internal models of the world using biophysical substrates (Marr & Poggio, 1976). The action of generating representations is known as *encoding* because the brain converts physical and conceptual entities into a code, that is, a combination of symbols representing variables. Symbols can be discrete, continuous, and distributed among numerous neurons and brain areas. A prime example of what is and is not computation can be found in action potentials. Their generation is caused by fine homeostatic adjustments of membrane voltage that per se may not qualify as a computation (Stuart, Spruston, Sakmann, & Hausser, 1997), but complex combinations of action potentials constitute the “symbols” of the “alphabet” used by the brain to compute. Examples of variables encoded by the brain are the position, color, and shape features of a given object (Seymour, Clifford, Logothetis, & Bartels, 2010), sound categories (Tsunada & Cohen, 2014), the distance between the eyes in face recognition (Chang & Tsao, 2017), and the reward value of a choice during decision making (Saez et al., 2018). The information embedded in neural biophysical substrates can be *decoded* and transferred (rerouted), possibly transformed into different formats and neural substrates. Examples are the online holding of memory during decision making (Hasson, Chen, & Honey, 2015), and memory replay during memory consolidation (Foster, 2017). It is worth stressing that the current computational view of the brain is not an established truth, but a simplified framework highly influenced by information theory, computer science, and linguistics to guide experimental testing.

2.2 | Computation takes place at several hierarchically organized levels

Levels include brain areas, nuclei, maps, columns, circuits, single neurons, and subneuronal compartments, such as dendrites, spines, somas, and axons (Mesulam, 1998). Levels, moreover, interact in specific temporal and topological patterns (Betzel & Bassett, 2017; Vidaurre, Smith, & Woolrich, 2017). A hierarchical organization is, in essence, a modular organization of computation (Meunier, Lambiotte, Fornito, Ersche, & Bullmore, 2009), such that a successful general theory of the brain will have to explain how tasks performed at one module(s) give rise to tasks performed by the larger module(s). Currently, a widely assumed premise is that most components of cognition emerge from the level of transiently active circuits—some authors prefer to speak about ensembles of neurons or cell assemblies (Buzsaki, 2010)—whose dynamics arise, in turn, from complex interactions involving the three classical building blocks: neuronal intrinsic excitability, synaptic efficiency, and connectivity (Gjorgjieva et al., 2016). Simply put, circuit dynamics within the range of millisecond to minutes control fast behaviors such as perception and decision making (Khani & Rainer, 2016), whereas synaptic changes lasting hours and days control learning and memory (Sweatt, 2016). Connectivity includes two main patterns: feed-forward, supporting a unidirectional flow of information, and recurrent, composed of positive and negative feedbacks that lead to self-sustained multiple activity patterns (Duarte, Seeholzer, Zilles, &

Morrison, 2017). Connections are mostly selective but they can be random as well, giving rise to complex, slow dynamics that include chaotic interactions (Mastrogiuseppe & Ostojic, 2018). Another widely assumed premise is that local circuits, however dynamic, are too anatomically fixed to adapt their behavior to contexts that need to be globally broadcast, for instance, sleep–wake cycles, mood, reward, and attention during perception and decision making. To circumvent this problem, neuromodulation has been suggested as a solution. Neuromodulation refers to the relatively rapid (in the range of seconds) functional reconfiguration of circuits throughout the brain by acetylcholine, dopamine, noradrenaline, and serotonin, which are released by subcortical and brainstem nuclei: The *nucleus basalis* of Meynert (NBM), the striatum, the *locus coeruleus*, and the Raphe nucleus (Avery & Krichmar, 2017). Neuromodulation participates in working memory, attention, brain state, and plasticity (Meunier, Chameau, & Fossier, 2017; Sara, 2009; Thiele & Bellgrove, 2018).

2.3 | Neural substrates of brain computations

The ultimate goal of Systems/Computational Neuroscience is to explain how electrical and chemical signals are used in the brain to represent and process information (Sejnowski et al., 1988). Currently, a widely accepted assumption is, as noted, that external variables are encoded into action potentials. Theories and empirical evidence point to firing rates (average number of action potentials per unit of time; Gerstner, Kreiter, Markram, & Herz, 1997), action-potential timing (length of time between action potentials; Panzeri, Petersen, Schultz, Lebedev, & Diamond, 2001), population coding (joint activity of several neurons; Panzeri, Macke, Gross, & Kayser, 2015), and neural dynamics (the way electrical activities evolve with time and space; Shenoy, Sahani, & Churchland, 2013), as potential features of action potentials that, an infinite amount of combinations, have enough breadth to constitute the basis of the brain code(s). A key implication of the multi-level organization of the brain is that code(s) are multi-level, too. This means that external variables are encoded by the collective activity of numerous simpler elements, which carry either synergistic or complementary information (Panzeri et al., 2015). This principle is the driving premise in population and dynamic coding, and has informed the development of methods for recording from large populations of neurons, including multi-electrode arrays, which can record up to 10^3 neurons (Einevoll, Franke, Hagen, Pouzat, & Harris, 2012), Ca^{2+} imaging, which can simultaneously record over 10^4 neurons (Pachitariu et al., 2016; Sofroniew, Flickinger, King, & Svoboda, 2016), and functional magnetic resonance imaging (fMRI), which makes use of BOLD (blood-oxygen-level contrast imaging) to unravel functional connectivity among regions encompassing over 10^5 neurons (Fox & Raichle, 2007). It is worth stressing that the measurable signals in the latter two approaches are not action potentials, but single-cell Ca^{2+} rises and regional oxygen consumption, respectively. Although the premise for using large-scale Ca^{2+} imaging in neurons is that single-neuron Ca^{2+} signals represent slower nonlinear encoding of the underlying action potentials (Lutcke, Gerhard, Zenke, Gerstner, & Helmchen, 2013; Vogelstein et al., 2010), nonelectrical signals, as well

as global voltage oscillations measured with field potentials and electroencephalograms, plausibly carry additional information that is computationally relevant. For example, it has been proposed that synaptic facilitation mediated by neuronal Ca^{2+} signals sustains working memory (Mongillo, Barak, & Tsodyks, 2008). All in all, biophysical substrates of brain computations other than the ones directly or indirectly based on neuronal activity will plausibly arise in the future, including, we posit, astrocyte-based computations.

2.4 | Contemporary brain theories

According to the number of publications, one of the most influential brain frameworks is predictive coding, which aim to account for core principles underlying adaptive circuit remodeling. The key tenets of predictive coding are the following. First, representationalism, the brain operates by building models of the outer world, conceptual categories, and expected outcomes of actions. Second, evaluation of new information against embedded models is at the core of many brain operations besides decision making, including perceptual discrimination, voluntary selective attention, and learning. Third, the nature of such evaluations is probabilistic, since the underlying algorithms weigh in pros and cons and similarity of the novel information with respect to internal models. A central notion is that “*organisms care less about representing what is actually out there in the world than about how this reality conflicts with their predictions about what should be there*” (Fitch, 2014). An apparent virtue of this strategy is minimization of data storage since it takes fewer bits to represent the mean and deviations from it than to attempt de novo representations (Fitch, 2014). Fourth, the brain tries to minimize its prediction errors such that internally generated predictions are constantly optimized with external inputs in an iterative process. In predictive coding, neuromodulation is proposed as computing part of the statistics of errors made by predictions (Lau, Monteiro, & Paton, 2017; Stephan, Iglesias, Heinzle, & Diaconescu, 2015). The bulk of empirical support for predictive coding lies in the domains of perception, reward learning, and decision making, as documented in humans, monkeys, and rodents (Diederer et al., 2017; Kok & de Lange, 2014; Leinweber, Ward, Sobczak, Attinger, & Keller, 2017; Markov et al., 2014; Nasser, Calu, Schoenbaum, & Sharpe, 2017; Summerfield, Trittschuh, Monti, Mesulam, & Egnér, 2008; Wacongne et al., 2011), whereas the framework appears to be under exploration in memory consolidation (Cross, Kohler, Schlesewsky, Gaskell, & Bornkessel-Schlesewsky, 2018) and emotion (Barrett, 2017). Other general CNS frameworks worth mentioning are global workspace theory, which describes the basic circuit from which consciousness emerges (Baars, 2005), and liquid computing, which states that neural circuits have the capacity to store information of previous perturbation(s), analogous to the ripples generated on the surface of a pond when stones are thrown into it (Maass, Natschlag, & Markram, 2002). Finally, influential theoretical constructions about basic operative principles of the brain—compatible with global frameworks—include brain oscillations (Buzsáki & Draguhn, 2004), efficient coding (Chalk, Marre, & Tkacik, 2018), energy-efficient coding (Laughlin, 2001), neural integrators

(Mazurek, Roitman, Ditterich, & Shadlen, 2003), inhibitory/excitatory balance (Brunel, 2000; Litwin-Kumar & Doiron, 2012), noise (Arieli, Sterkin, Grinvald, & Aertsen, 1996), and circuit degeneracy (Sporns, 2013).

3 | CHALLENGES, OBSTACLES, AND GROWTH AREAS IN SYSTEMS NEUROSCIENCE

Despite the progress in the last decade, understanding brain computations remains a central challenge of modern Neuroscience. The readily observable behavioral variables that are used experimentally to study brain encoding, for instance, rewards, choices, and stimulus features, represent the tip of the iceberg, because the vast majority of variables used by the brain in complex behaviors and higher-brain functions are typically latent. However, this should not distract us from the impressive predictive power that analytical tools are achieving in Systems Neuroscience. Examples of success can be found in neuroprosthetics, where the electrical activity of the brain of a human user is decoded into motor commands (Cangelosi & Invitto, 2017); decision-making, in which decision outputs can be predicted from action potentials with 80% accuracy in monkeys before a response is observed (Kiani, Cueva, Reppas, & Newsome, 2014), and with 70% accuracy in rats, even before stimulus onset (Nogueira et al., 2017), and face recognition. Here, the face seen by a Rhesus monkey can be reproduced with 90% accuracy by tracking neuronal activity in the inferior temporal cortex (Chang & Tsao, 2017). Although the achievements are remarkable, there is still room to improve these numbers. In the workflow of Systems Neuroscience from signal capture to deciphering the brain code, areas of improvement include signal recording, signal processing, data analyses, and astrocyte-focused studies (Figure 1). Key issues are briefly described next.

3.1 | Data load in large-scale recordings

The trend of improving predictions by simultaneously recording more neurons has created a serious challenge: The ever-increasing size of the data seriously hampers storage, processing, and analysis. In order to simplify and reduce data size of recordings, several methods exist to extract low-dimensional mathematical representations from multi-neuronal electrical recordings (Aljadeff, Lansdell, Fairhall, & Kleinfeld, 2016; Cunningham & Yu, 2014). The obstacle is all the more complex in Ca^{2+} imaging, which has become a dominant method for recording from large populations of neurons, because special methods are necessary to extract the coarse-grained and noisy Ca^{2+} data prior to data analysis. Algorithms such as Suite2p (Pachitariu et al., 2016), and CNMF (Constrained and/or non-negative matrix factorization; Pnevmatikakis et al., 2016), represent advances in the simplification of imaging data processing prior analysis. Caveats of current Ca^{2+} imaging data processing are discussed in Stringer and Pachitariu (2018). Alternatively, shot-gun statistics unravels network connectivity information from recording at only 10% of the neurons at a given time,

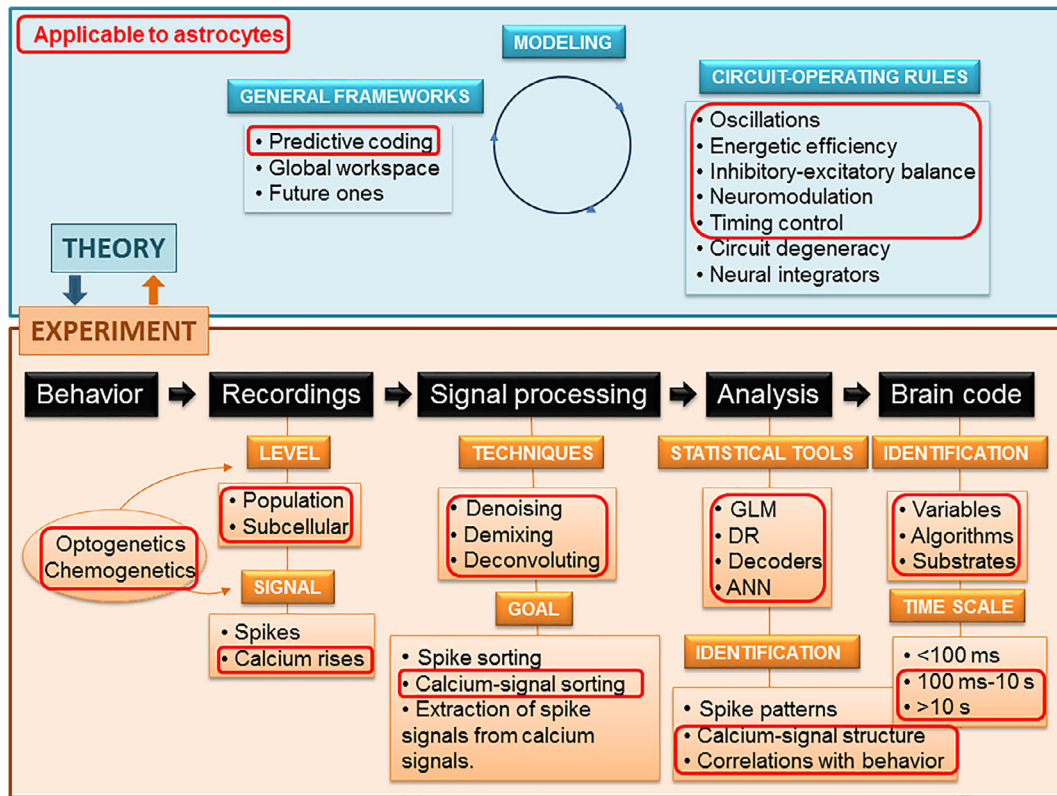


FIGURE 1 Workflow in Systems Neuroscience. A central problem in Neuroscience is to explain how electrical and chemical signals are used in the brain to represent and process information. The workflow depicts the stages and tools currently used to decipher neuronal codes. In red squares, we highlight the elements that are relevant to the study the role of astrocytic Ca^{2+} in neuronal coding

thus simplifying the experimental load of large-scale recordings (Soudry et al., 2015). Data-sharing and collaborative solutions have been proposed as well to manage the surge of data (Paninski & Cunningham, 2018).

3.2 | Statistical tools for understanding data

The standard problem is to determine how behavioral variables are encoded by neurons, and how this information is decoded, either by downstream neurons or by an external observer. Different statistical tools address encoding and decoding. For encoding, generalized linear models (GLMs), a generalization of multiple linear regression, regress neuronal activity against behavioral variables to determine the set of variables that explain more neuronal activity (Aljadeff et al., 2016; Nogueira et al., 2017). Decoding techniques, typically linear classifiers (Arandía-Romero, Nogueira, Mochol, & Moreno-Bote, 2017; Quiñero & Panzeri, 2009), as well as more recent artificial neural networks (ANNs; Paninski & Cunningham, 2018) are used to predict, trial-by-trial, values of behavioral variables from neuronal activity, either using single neuronal activity or the individual activity of large neuronal populations recorded from multielectrode-arrays or Ca^{2+} imaging. These methods are supervised machine learning tools because both behavioral and neuronal variables are preselected and labeled. Also, unsupervised tools such as dimensionality reduction have been developed, and used in parallel, in order to reduce data

complexity by identifying low-dimensional latent factors, where relevant behavioral variables could be represented (Cunningham & Ghahramani, 2015). Of note, detection of relevant subspaces of neuronal activity, and optimal selection of behavioral features to regress against neuronal data will facilitate the discovery of computational principles. An elegant example is the aforementioned study by Chang and Tsao (2017), in which successful face identification in nonhuman primates was possible with 50-dimensional data, and recordings of 200 neurons. Likewise, feature selection can be adaptively improved with artificial intelligence (Yamins & DiCarlo, 2016). As with signal processing, data load is a challenge in signal analysis, for the number of observations per condition does not necessarily grow in parallel with the growth of complexity and number of dimensions of the data. For example, recording 20 neurons for 30 min produces the same number of observations per neuron than recording 1,000 neurons during the same amount of time, but the number of dimensions increases 50-fold with the larger neuronal population. This means that encoding, decoding, and dimensionality reduction techniques need to be constrained by specific structural and anatomical knowledge of the neural substrates to be operationally useful.

3.3 | Optogenetics and chemogenetics

These anatomically precise and reversible tools allow establishing cause-effect relationships between the electrical activity of single

neurons, or neuronal populations, and behavioral parameters. Optogenetics is based on the expression of light-sensitive regulators of transmembrane conductance (ion channels and chloride pumps) coupled with fiber optic- and laser diode-based light delivery (Boyden, Zhang, Bamberg, Nagel, & Deisseroth, 2005; Li et al., 2005). Cell type specificity is accomplished by targeting the light-sensitive channels with cell-type specific promoters. Light-activation of neurons expressing channels like channelrhodopsins (ChR1, ChR2) result in neuronal depolarizations due to import of cations such as Na^+ , K^+ , and Ca^{2+} —the latter at trace levels. In contrast, optical stimulations of archaerhodopsin (Arch) and halorhodopsins (NpHR) pumps cause hyperpolarization of neurons by exporting H^+ , or by importing chloride ions, respectively. An alternative approach to classic opsins is the light-sensitive G-coupled receptor, also called OptoGq/Gs, which modulates receptor-initiated biochemical signaling pathways (Airan et al., 2009). Chemogenetics is based on the use of designer receptors exclusively activated by designer drugs (DREADDs), a family of G protein-coupled receptors (GPCRs) that are solely activated by a pharmacologically inert drug, clozapine N-oxide (CNO; Alexander et al., 2009). DREADDs can also be targeted to neurons with viral or transgenic delivery systems using neuron-specific promoters. Relevant insights into behavior, cognition, and basic brain homeostasis have been gained with neuron-targeted optogenetic and chemogenetic approaches (Deisseroth, 2015; Roth, 2016).

3.4 | Subcellular computations

Increasing the number of recorded neurons may not be the only solution for obtaining better data. Insofar each and every neuron must integrate and convert thousands of synaptic inputs into a single output (London & Hausser, 2005), concerns have been raised about the oversimplification of neurons as “integrate-and-fire” nodes in large-scale recordings and *in silico* simulations, and a plea exists to pay renewed attention to the great computational potency of single neurons (Fitch, 2014). Spine computations and biophysical substrates are reviewed in (Yuste, 2013), and a recent example of the computational relevance of dendritic shafts is the finding that nonlinear dynamics based on dendritic conductance can help sharpen time and rate codes in grid cells, thereby improving the accuracy of space representation (Schmidt-Hieber et al., 2017). In the context of imaging, voltage dyes represent a growth area allowing for recording at subcellular resolution at multiple points along dendrites and axons (Xu, Zou, & Cohen, 2017). The data, combined with whole-cell reconstructions with electron microscopy (Vishwanathan et al., 2017), will arguably improve the understanding of dendritic computations and network connectivity.

3.5 | A need for theoretical frameworks and modeling

The wealth of descriptive data will not advance knowledge unless analyses are guided by hypotheses and complemented with modeling. Computational/Systems Neuroscience is thus engaged in a virtuous cycle whereby data generate models, and models make predictions

that can be tested *ad infinitum* against new proposed experiments. The trade-offs of increasing the realism of models by incorporating more biophysical variables versus developing simplifying models, as discussed in (Sejnowski et al., 1988), are still debated (Marder, 2015). Whatever the approach, *in vivo* models, and their *in silico* counterparts, need to be informed by large-scale hypotheses combined with simpler questions, in order to advance on the outstanding question of how the brain processes information with such energetic efficiency. We discussed the remarkable production of studies informed by predictive coding and other theoretical constructions. Other theories will plausibly arise in the future.

4 | ASTROCYTE-BASED COMPUTATIONS AS A GROWTH AREA IN SYSTEMS NEUROSCIENCE

We posit that variables used in brain coding may be partially embedded in astrocyte biophysical substrates, such that the incorporation of astrocytes as computational building blocks in neural circuits may help advance Systems Neurosciences. Significant gaps of knowledge, however, exist. First, there is no evidence that astrocytes gate, transform, store, and reroute information in the brain by carrying out processes that can be described in abstract mathematical terms. Astrocytes do participate in brain state (Poskanzer & Yuste, 2016), neuromodulation (Magistretti & Morrison, 1988; Paukert et al., 2014; Srinivasan et al., 2015), and in a wide variety of naturally occurring recurrent circuits, where they have been proposed as carrying out spatiotemporal integration of multicellular inputs (Araque et al., 2014). Examples indeed exist of discrimination and integration of synaptic information by astrocytes (Perea & Araque, 2005), but the underlying algorithms and their behavioral correlates remain undetermined. Second, if astrocytes compute, are Ca^{2+} transients a biophysical substrate of astrocyte-based computations? The intuition that they already exist in the field, resting on a wealth of studies that, since the 1990s, have used Ca^{2+} imaging to assess astrocyte activation at increasing spatiotemporal resolution, thanks to the unremitting refinement of fluorescent indicators and optical imaging [reviewed in Kastanenka et al., 2016 and Bazargani & Attwell, 2016]. However, although *in silico* modeling documents that astrocytes can encode extracellular cues into variables in Ca^{2+} transients (De Pitta et al., 2008), the statistical methods currently used to encode and decode neuronal action potentials (Section 3) have not been applied to astrocyte data obtained *in vivo*. Third, it is not known whether the subcellular Ca^{2+} microdomains in astrocytes would carry out different functions within distinct circuits associated with different complex behaviors, nor whether astrocytes would perform similar computations throughout the brain, or are as functionally heterogeneous as neurons. It is worth mentioning that in the last decade controversies have arisen concerning the regulation and consequences of Ca^{2+} signaling in astrocytes. Specifically, whether Ca^{2+} comes from endoplasmic reticulum and mitochondria, or from the extracellular milieu, the very notion of Ca^{2+} -dependent gliotransmission, the role of astrocytes in long-term potentiation (LTP), and whether D-serine is a

gliotransmitter have been debated—reviewed in Bazargani and Attwell (2016) and Savtchouk and Volterra (2018). Currently, the prevailing notion reconciling these discrepancies is that Ca^{2+} responses are highly complex and context-dependent, such that the signaling leading to Ca^{2+} rises, the subcellular source of such Ca^{2+} , the speed of transients, as well as the downstream effects, are dependent on the subcellular astrocyte compartment(s), and the neural circuit (Savtchouk & Volterra, 2018). In this piece, we do not focus on mechanistic issues, but rather on whether and how astrocytes may perform computations using Ca^{2+} transients.

5 | SYSTEMS-LIKE STUDIES IN ASTROCYTES

A prototypical study in Systems Neuroscience includes three components: (a) recording of electrical activity in multiple neurons, (b) computerized analysis to decode information embedded in action-potential firings, and (c) simultaneous measurement of a cognitive or behavioral function. The statistical analyses reveal correlations and, increasingly often, causal relationships between changes in patterns of neuronal-population firing and specific behavioral or cognitive responses (Sections 2 and 3). There are no studies, to the best of our knowledge, recording the Ca^{2+} activity of multiple astrocytes, followed by analysis with GLM or decoders in the context of a behavioral paradigm defined by distinct features that can be correlated with patterns of astrocytic Ca^{2+} activity. Among studies linking astrocytes and behavior [for reviews Oliveira, Sardinha, Guerra-Gomes, Araque, & Sousa, 2015 and Santello, Toni, & Volterra, 2019], in Section 5.1, we discuss the ones closer to the neuron-focused experimental design in Systems Neuroscience, for they include recordings of Ca^{2+} -based astrocyte excitability, as well as electrical or optical recordings of neuronal activity, in the context of complex behaviors or neuromodulation. Conversely, in Section 5.2, we focus on studies showing modulation of local brain circuits associated with complex behaviors, or brain state, by *transient* optogenetic or chemogenetic astrocyte activation. In Section 6, we extract computational lessons from these studies, and identify gaps of knowledge, taking into account, when appropriate, previous, and recent studies that, although lacking any of the aforementioned components, support our computational insights. Table 1 summarizes the analysis. In Figure 1, we highlight in red approaches within the general workflow of Systems Neuroscience including signal capture, processing, and analysis that could be used with astrocytic data.

5.1 | Activation of Ca^{2+} transients in astrocytes by sensory stimulation and neuromodulation

Studies in the mouse barrel cortex have shown activation of Ca^{2+} in astrocyte somata after whisker stimulation using fluorescent Ca^{2+} dyes (Takata et al., 2011; Wang et al., 2006) and genetically encoded Ca^{2+} indicators (Stobart et al., 2018). Astrocytic Ca^{2+} increases are delayed with respect to Ca^{2+} rises in neurons (Stobart et al., 2018). Also, astrocytic Ca^{2+} rises are dependent on whisker stimulation frequency, and they are blocked by inhibitors of metabotropic glutamate

receptors, indicating that they are caused by glutamate released from neurons (Wang et al., 2006). Whisker stimulation-dependent Ca^{2+} rises in astrocytes are detected as early as at 2 s when dyes are used, and at 120 ms in the case of faster, genetically encoded indicators, although peak responses range between 3 and 12 s regardless of the Ca^{2+} indicator. Likewise, visual stimulation triggers neuron-dependent somatic Ca^{2+} transients in astrocytes in the visual cortex of ferret, with a delay of 1–3 s and a peak at 6 s (Schummers et al., 2008). Importantly, the latter study demonstrates that astrocyte activation is highly tuned to orientation maps at a single-cell resolution, and documents that astrocytes mediate hemodynamic signals in the visual cortex, which was confirmed in another study in the barrel cortex (Stobart et al., 2018). The study by Takata et al. (2011) is also relevant because it demonstrates the following. First, cholinergic neuromodulation originating in the NBM potentiates the activation of local field potentials elicited by whisker stimulation. Second, neuromodulation is strictly dependent on Ca^{2+} rises in astrocytes, as shown by the disappearance of neuronal-activity potentiation in mice lacking IP3R2-dependent signaling. Crucially, abrogation of Ca^{2+} signaling in astrocytes in these mice shifts brain state to a desynchronized mode, as assessed with local field potentials in cortex. The impact of cholinergic neuromodulation on astrocyte Ca^{2+} responses is also documented in hippocampus. Specifically, the increase in Ca^{2+} rises triggered by somatosensory stimulation in rat hippocampal astrocytes is mediated by cholinergic neurotransmission, since it is blocked by the cholinergic inhibitor atropine (Navarrete et al., 2012). Astrocyte activation, in turn, induces the long-term potentiation (LTP) of field EPSPs in CA3–CA1 synapses (Navarrete et al., 2012). These data support the notion that, in addition to setting circuit dynamics for attention in sensory processing, cholinergic neuromodulation participates in the encoding of new information during memory formation (Hasselmo & McGaughy, 2004). The importance of neuromodulation via astrocytic Ca^{2+} in sensory cortical processing has also been reported for the *locus coeruleus* (Ding et al., 2013; Paukert et al., 2014; Srinivasan et al., 2015). This brain-stem nucleus amplifies as well the effect of locomotion on Ca^{2+} rises in Bergman glia in the cerebellum (Paukert et al., 2014). Timewise, neuromodulation-elicited Ca^{2+} rises in astrocytes occur in the range of a few seconds, with regards to both onset and peak after sensory stimulation (Ding et al., 2013; Srinivasan et al., 2015).

5.2 | Modulation of behavior and brain state by optogenetic and chemogenetic stimulation of astrocytes

As in neurons, important insights into *causal* relationships between astrocytic Ca^{2+} signals and behavioral outcomes are emerging from optogenetics and chemogenetic studies. These technologies allow temporally precise and reversible modulation of astrocyte activity, in contrast to permanent loss- or gain-of-function genetic manipulations. In mice, optogenetic stimulation of astrocytes using ChR1/2, Arch and OptoGq has been reported to modulate breathing according to pH changes in the respiratory system (Gourine et al., 2010), induce

**TABLE 1** System-like studies in astrocytes

Direction of experimental manipulation	Stimulation	Neural circuits	Readouts	References	Predicted canonical computations			
BEHAVIOR TO ASTROCYTES	SENSORY STIMULATION	Barrel cortex	Astrocytic Ca ²⁺ ; LFP; local postsynaptic activity	(X. Wang et al., 2006)	Filtering			
		Barrel cortex	Astrocytic Ca ²⁺ ; LFP; brain state	(Takata et al., 2011)	Thresholding; Coincidence detection; State switching; Gain control; E/I balance			
		Visual cortex	Astrocytic Ca ²⁺ ; neuronal Ca ²⁺ ; hemodynamic responses	(Schummers et al., 2008)	Filtering			
		Visual cortex	Astrocytic Ca ²⁺ ; neuronal Ca ²⁺ ; hemodynamic responses	(Stobart et al., 2018)	Thresholding			
		Visual cortex	Astrocytic Ca ²⁺ ; EPSP; IPSP; SIC; patch-clamp recordings; visual response selectivity	(Perea et al., 2016)	Thresholding; Gain control			
		Hippocampus	Astrocytic Ca ²⁺ ; LTP; CA1 post-synaptic depolarization	(Navarrete et al., 2012)	Thresholding; Coincidence detection; Gain control			
		NEUROMODULATION		Cholinergic	Astrocytic Ca ²⁺ ; LTP; CA1 post-synaptic depolarization	(Navarrete et al., 2012)	Thresholding; Coincidence detection; Gain control	
				Cholinergic	Astrocytic Ca ²⁺ ; LFP; brain state	(Takata et al., 2011)	Thresholding; Coincidence detection; Gain control; E/I balance	
				Noradrenergic	Astrocytic Ca ²⁺ ; EcoG	(Ding et al., 2013)	Thresholding; Coincidence detection	
				Noradrenergic	Astrocytic Ca ²⁺ ; locomotion; electromyography	(Paukert et al., 2014)	Thresholding; Coincidence detection	
Cerebellum	Glutamate release; EPSP; LTD; motor behavior			(Sasaki et al., 2012)	Gain control			
Somatosensory cortex	Astrocytic Ca ²⁺ ; neuronal Ca ²⁺ ; LFP; glutamate release; brain state			(Poskanzer & Yuste, 2016)	Gain control; E/I balance; State switching			
Visual cortex	Astrocytic Ca ²⁺ ; EPSP; IPSP; SIC; patch-clamp recordings; visual response selectivity			(Perea et al., 2016)	Thresholding; Gain control			
Brain stem	Astrocytic Ca ²⁺ ; ATP release; neuronal membrane potentials; EPSC; breathing			(Gourine et al., 2010)	Gain control			
Hypothalamus	Sleep			(Peilluru et al., 2016)	Gain control			
Hypothalamus	Adenosine release; open-field behavior; food intake			(Sweeney et al., 2016)	Gain control			
ASTROCYTES TO BEHAVIOR		Hypothalamus	Astrocytic Ca ²⁺ ; patch clamp; IPSC; food intake	(Chen et al., 2016)	Gain control			
		Hypothalamus	Astrocytic Ca ²⁺ ; patch clamp; IPSC; food intake	(L Yang et al., 2015)	Gain control			
		Hippocampus	Astrocytic Ca ²⁺ ; LTP; EPSC; memory acquisition; contextual and spatial memory	(Adamsky et al., 2018)	Gain control			
		Hippocampus	Astrocytic Ca ²⁺ ; LTP; EPSC; memory acquisition; contextual and spatial memory	(Adamsky et al., 2018)	Gain control			
		Amygdala	Astrocytic Ca ²⁺ ; IPSC; EPSC; fear-expression	(Martin-Fernandez et al., 2017)	Gain control; E/I balance			
		CHEMOGENETICS		Hippocampus	Astrocytic Ca ²⁺ ; LTP; EPSC; memory acquisition; contextual and spatial memory	(Adamsky et al., 2018)	Gain control	
				Amygdala	Astrocytic Ca ²⁺ ; IPSC; EPSC; fear-expression	(Martin-Fernandez et al., 2017)	Gain control; E/I balance	
				Abbreviations: LFP, Local field potentials, LTD, long-term depression, LTP, long-term potentiation, EPSP, excitatory postsynaptic potential, IPSP, inhibitory postsynaptic potential, EcoG, electrocorticogram recordings, SIC, slow inward currents.				

Abbreviations: LFP, Local field potentials, LTD, long-term depression, LTP, long-term potentiation, EPSP, excitatory postsynaptic potential, IPSP, inhibitory postsynaptic potential, EcoG, electrocorticogram recordings, SIC, slow inward currents.

long-term depression in Purkinje cells and motor behavior (Sasaki et al., 2012), modulate response selectivity of the visual cortex (Perea et al., 2016), inhibit food intake (Sweeney et al., 2016), induce sleep (Pelluru et al., 2016), promote a switch to the slow-oscillation state by triggering the UP state of slow waves (Poskanzer & Yuste, 2016), and enhance memory acquisition (Adamsky et al., 2018).

A key issue is that the downstream consequences of optogenetic activation of astrocytes are not well understood. In the case of neurons, since they are excitable cells that can operate via all-or-nothing changes in membrane voltage driven by fast-acting voltage-gated channels (although they also have subthreshold voltage fluctuations), the probability of neuronal firing is decreased by activation of NpHR and Arch, and increased upon activation of ChR2 (Yizhar, Fenno, Davidson, Mogri, & Deisseroth, 2011). However, astrocytes are not as electrically excitable as neurons. In the first report of successful modulation of neuronal activation (with no behavioral consequences) upon optogenetic manipulation of nearby ChR2-expressing astrocytes, it was assumed, but not shown, that the response was mediated by Ca^{2+} fluxes through ChR2 (Gradinaru, Mogri, Thompson, Henderson, & Deisseroth, 2009). Two subsequent studies confirmed Ca^{2+} rises using Ca^{2+} indicator dyes (Pelluru et al., 2016; Perea, Yang, Boyden, & Sur, 2014), yet it is unclear how these rises can occur, considering that ChR2 has a relatively low Ca^{2+} permeability, is only open during a few milliseconds—decay constant is ~ 10 ms—and presents depolarization-dependent slowing of deactivation (Nagel et al., 2003; Yizhar et al., 2011). One possibility is that it is the entry of Na^+ through ChR2 that causes Ca^{2+} uptake by reverse activity of the Na^+/Ca^+ exchanger (Yang, Yu, et al., 2015). Furthermore, the possibility exists that the effects of ChR2 activation are due to undetected Ca^{2+} rises in astrocyte processes, of which somatic Ca^{2+} might be a consequence (Bernardinelli et al., 2011). In this regard, the use of Arch combined with genetically encoded Ca^{2+} indicators represents a technical refinement because this opsin induces, after 5 s of photostimulation in the mouse cortex, fast Ca^{2+} transients in astrocyte arbors reminiscent of spontaneous activity (Poskanzer & Yuste, 2016). Still, how such a brief photo-stimulation of Arch, whose decay constant is ~ 9 ms (Yizhar et al., 2011), translates into ~ 20 -s-long Ca^{2+} rises after a delay of ~ 10 s is unclear (Poskanzer & Yuste, 2016). Plausibly, Arch-elicited hyperpolarization engages voltage-sensitive elements in astrocyte processes. All in all, optogenetics clearly activates astrocytes, although clarification of underlying mechanisms will help optimize this approach for Systems-level basic and clinical studies.

A DREADD receptor that successfully triggers Ca^{2+} transients in astrocytes is hM3Dq (Bonder & McCarthy, 2014; Chen et al., 2016). Studies using hM3Dq in astrocytes have shown: (a) changes in neuronal activity, either reduced or increased firing, in the mouse arcuate nucleus with opposing effects on feeding behavior, perhaps stemming from CNO dose differences, which, in turn, might launch complex feedback loops leading to paradoxical data (Chen et al., 2016; Yang, Qi, & Yang, 2015), (b) regulation of excitatory and inhibitory neurotransmission in the amygdala, with a net effect of reduced fear expression in a fear-conditioning paradigm (Martin-Fernandez et al., 2017); and (c) potentiation of the amplitude of evoked EPSC and,

when chemogenetic activation is carried out at specific stages during learning paradigms, improvement of contextual and spatial memory acquisition (Adamsky et al., 2018). As with optogenetics, caution has to be exerted about the resemblance of the Ca^{2+} signaling elicited by chemogenetics to physiological signaling. Also, the CNO metabolite clozapine, and not CNO, might be the real activator of DREADD, as shown with radioligand receptor occupancy measurement, and in vivo positron emission tomography (Gomez et al., 2017). Since clozapine has multiple targets, this recent evidence raises doubts about the specificity of DREADD-based approaches (Gomez et al., 2017). That said, these studies offer several computational insights, to be discussed below.

6 | COMPUTATIONAL LESSONS LEARNED FROM SYSTEMS-LIKE STUDIES IN ASTROCYTES

6.1 | Time scales of Ca^{2+} responses and filtering effect

According to Ca^{2+} -based dynamics, the time scale of astrocyte activation after a physiological input ranges from hundreds of milliseconds to tens of seconds, while the earliest reported effect on nearby neurons after optogenetic stimulation of astrocytes is at 500 ms (Gradinaru et al., 2009). The onset of hemodynamic response is within 1–3 s from the onset of Ca^{2+} responses (Otsu et al., 2015). Upon sensory stimulation, astrocytes are activated *after* neurons in cortex, suggesting that neurons *reroute* information to astrocytes. The observation that Ca^{2+} response curves in astrocytes are qualitatively similar but narrower than those in neurons, as shown by local field potentials (Schummers et al., 2008; Wang et al., 2006), suggests that astrocytes *filter* neuronal activity. Filtering can be either in terms of rectification (high pass filtering), cut-off (low pass filtering), or both (band pass filtering). The latter appears to be the case since astrocytes are not responsive to the highest and lowest frequencies of neuronal input. Interestingly, adaptive modulation of breathing by pH is the only context in which astrocytes *directly* compute external stimuli, for astrocytes sense changes in pH, even if local neurons are inactivated with tetrodotoxin (Gourine et al., 2010). In other paradigms, astrocyte activation is either secondary to neuronal activation (Section 5.1), or the result of gain-of-function induced by optogenetics and chemogenetics in the context of already active circuits (Section 5.2).

6.2 | Existence of short- and long-term modalities in Ca^{2+} responses

The computational and homeostatic functions of astrocytes manifest themselves in at least two broad modalities, depending on time range, nature of inputs, and the intracellular location of Ca^{2+} rises. One modality is the fast rising Ca^{2+} signals that originate within 0.2–5 s from stimulus onset, are short-lived (up from 0.3–10 s), are usually reported in peripheral processes and end-feet (Stobart et al., 2018), and are sufficiently fast to *locally* mediate task-relevant regulation of

blood flow (Otsu et al., 2015), metabolic coupling, and neurotransmitter supply (Agarwal et al., 2017; Otsu et al., 2015; Tani et al., 2014), as well as short-term modulation of synaptic efficacy (Perea et al., 2016). The second modality corresponds to robust somatic Ca^{2+} transients that can last tens of seconds, have a slow rise time, and have been reported in the context of cholinergic (Navarrete et al., 2012; Takata et al., 2011) and noradrenergic (Ding et al., 2013; Paukert et al., 2014; Srinivasan et al., 2015) neuromodulation, as well as upon ChR2-based optogenetics and by chemogenetics (Adamsky et al., 2018). In hippocampus, the functional consequences of this modality are long-lasting effects on synaptic connections (Adamsky et al., 2018; Navarrete et al., 2012), plausibly associated with memory formation. In cortex, we reason that astrocytic Ca^{2+} rises, as reported by (Takata et al., 2011), participate in a well-accepted role of neuromodulation: Control of arousal and attention, which involves recruitment of large, spatially distributed neuronal populations (Thiele & Bellgrove, 2018). Importantly, the two modalities reveal the existence of *threshold heterogeneity* in Ca^{2+} responses in astrocytes, which might be of computational importance. Consider, for example, the relative ease with which minimal synaptic stimuli trigger Ca^{2+} transients in astrocytic processes (Haustein et al., 2014; Panatier et al., 2011), which is consistent with a relatively low threshold for activation. This suggests that, in microdomains, the number of synaptic inputs may be of little importance, so that a microdomain could invariantly get activated, either by individual synapses or by an ensemble thereof, akin to the logical OR function. Conversely, the phenomenon of *coincidence detection* in which activation of cortical sensory neurons (Paukert et al., 2014; Takata et al., 2011) and postsynaptic hippocampal neurons (Navarrete et al., 2012), needs to coincide with neuromodulation to trigger somatic Ca^{2+} transients, and, similarly, the requirement for high inter-neuronal activity to promote astrocytic Ca^{2+} -dependent facilitation of excitatory synaptic transmission in the hippocampus (Perea et al., 2016), may be regarded as examples in which the threshold for astrocytic activation is high, and astrocytes will become activated only if multiple inputs impinge *together* on them, akin to the logical AND function. Density of IP3R2 (De Pitta et al., 2008) and baseline Ca^{2+} levels (Zheng et al., 2015) may be among the factors setting thresholds of stimulation. Plausibly, the described modalities of astrocytic Ca^{2+} responses are the extremes of a context-dependent spectrum, encompassing mixed regimes in terms of number of astrocytic domains involved, and short versus long-term effects. Key questions emerge: how are different astrocytic microdomains recruited, which neural circuits are activated as a consequence of different response modalities, and, finally, do specific computations, other than thresholding, operate in different modalities? In Section 7, we propose gaining insight into these questions by treating single astrocytes as mini-circuits, and by identifying relevant patterns of Ca^{2+} responses with dynamical-systems statistics approaches such as *dimensionality reduction*.

6.3 | Regulation of neuronal gain

This appears to be a computation carried out by astrocytes throughout a variegated collection of circuits and behavioral contexts. Signal

coincidence detection of sensory stimulation and neuromodulation by cortical astrocytes is one example that may have implications in attention (Paukert et al., 2014; Takata et al., 2011). Computationally, attention consists of a gain change (in amplitude of response or contrast) that results in the prioritization of relevant inputs over irrelevant information (Thiele & Bellgrove, 2018). Input prioritization is called top-down (or inside-out) because the process is shaped by internal models and goals conveyed to the sensory areas by neuromodulators (Thiele & Bellgrove, 2018)—note the influence of predictive coding in this assumption. The modulation of gain is facilitated by a normalization mechanism whereby neurons' responses are reduced in proportion to the activity of neighboring neurons by the joint activation of inhibitory and excitatory neurons (Reynolds & Heeger, 2009). Instructed by signal coincidence detection, astrocytes might help prioritize information by regulation of gain via modulation of excitatory synaptic drive by Ca^{2+} -dependent glutamate uptake (Schummers et al., 2008), gliotransmission (Takata et al., 2011), intrinsic neuronal excitability (Sasaki et al., 2012), and comodulation of excitatory and inhibitory neurotransmission (Perea et al., 2014).

In the case of brain state, a gain change might account for the transition from an asynchronous to a synchronous mode through a change in the network's ratio of excitation versus inhibition, according to the general theory of neural networks (Brunel, 2000). Hence, a possible mechanism whereby astrocytes might synchronize brain state through gain control is regulation of excitatory-synaptic strength, either by reducing glutamate uptake (Poskanzer & Yuste, 2016), releasing ATP/adenosine and glutamate in a Ca^{2+} -dependent manner (Fellin et al., 2009; Halassa et al., 2009), or taking up GABA via GAT-3 transporters (Shigetomi, Tong, Kwan, Corey, & Khakh, 2011).

Memory-related tasks in hippocampus can also be interpreted as a phenomenon of gain control. Thus, chemogenetic and optogenetic stimulations of hippocampal astrocytes result in increased frequency and potency of mEPSCs in local neurons, leading to long-term potentiation of excitatory synaptic connections (Adamsky et al., 2018). Significantly, astrocyte-mediated NMDA-dependent long-term potentiation appears to be: (a) task-specific insofar as fear-conditioned mice, but not home-caged ones, show synaptic potentiation and (b) stage-selective, for it very precisely affects distinct phases along the memory-formation continuum, such as memory allocation. Likewise, the interneuron-induced potentiation of excitatory neurotransmission mediated by astrocytes might be one example of neuronal gain (Perea et al., 2016). Intriguingly, a dual mechanism in which astrocyte-mediated depression of excitatory synapses combines with potentiation of inhibitory ones seems at play in afferents to neurons in the medial central region of the amygdala (Martin-Fernandez et al., 2017). The ensuing net increase of inhibitory drive to these neurons (i.e., a case of negative gain) was then shown to correlate with transient reduction of fear conditioning and anxiety.

Finally, the role of astrocytes in reflex homeostatic behaviors modulating feeding and breathing can be explained in terms of use of gain modulation to adapt behavior to stimuli intensity. Thus, the presence of food modulates the synaptic efficacy of neurons in the

hypothalamus (Chen et al., 2016; Yang, Qi, & Yang, 2015), whereas pH acidification induces adaptive neuronal firing in the brain stem which, in turn, activates breathing (Gourine et al., 2010).

6.4 | Decoding and rerouting of information

Coincidence detection of sensory cortical and neuromodulatory sub-cortical neuronal inputs (Paukert et al., 2014; Takata et al., 2011), transformation of inhibitory neurotransmission into synaptic facilitation in hippocampus (Perea et al., 2016), and the transformation of neuronal inputs into potentiation or inhibition, depending on the duration and frequency of the inputs (Covelo & Araque, 2018), might be three examples of *decoding* of neuronal signals by astrocytes, and *rerouting* of decoded information to other neurons. Plausibly, the information rerouted by astrocytes is gliotransmitter-dependent (Covelo & Araque, 2018). Since neuronal action potentials and astrocytic Ca^{2+} transients have utterly different temporal resolutions, it is improbable that variables represented in trains of action potentials are represented in astrocytic Ca^{2+} without significant loss of information. Rather, we posit that what astrocytes “hear” from neurons are instructions to “tell” other neurons to modify their activity via canonical computations. In computational science, canonical computations are fundamental operations carried out in circuits in a variety of contexts. We have hitherto identified a few: signal filtration, thresholding (implicating AND/OR functions and coincidence detection), gain, and control of the balance between excitation and inhibition. It is not clear whether synaptic scaling should be added because this function might be performed by microglia rather than astrocytes (Stellwagen & Malenka, 2006). In the roadmap, we propose to use *decoding approaches* from machine learning to identify possible variables encoded by astrocyte computations.

6.5 | Astrocytes could act as switches in brain state transitions

The causal implication of astrocytes in cortical slow oscillations (<1 Hz; Takata et al., 2011; Poskanzer & Yuste, 2016) supports the relevance of astrocytes in network activity beyond tripartite synapses. Slow waves have been hypothesized to represent the default mode of cortical network activity (Sanchez-Vives, Massimini, & Mattia, 2017). During UP states, there is synchronization in beta and gamma frequencies, synaptic gain modulation, modulation of replay and memory formation, and some cortical features might inform about transitions between unconsciousness and consciousness [reviewed in Sanchez-Vives et al., 2017]. An intriguing paradox exists in that astrocytes induce a synchronized state, but also mediate cholinergic and noradrenergic neuromodulations, which are characteristically associated with asynchronous, high-rate activity that facilitates sensory processing (Lee & Dan, 2012). We posit that astrocytes might act as *switches* whose default action is to sustain UP states, whereas neuromodulation-driven attention renders astrocytes independent of the cortical oscillator, and shifts their action toward short-term plasticity related to sensory processing. Indeed, network theory predicts

that a key parameter in setting asynchronous versus synchronous network activity, as well as the frequency of eventual oscillations, is afferent synaptic activity (Brunel, 2000; Ledoux & Brunel, 2011). Coincidence detection can be thus regarded as a scenario of afferent stimulation—specifically mediated by neuromodulation—whereby astrocytes induce the network's transition to the asynchronous state. Finally, although astrocytes are particularly attuned to slow oscillations because of their internal dynamics, as judged by Ca^{2+} transients, fall within a time scale of seconds, they are also involved in the generation of faster waves such as theta (4–12 Hz) and slow gamma (30–50 Hz; Perea et al., 2016; Sardinha et al., 2017). The effect of astrocytes on fast waves may be due to cross-frequency coupling, a mechanism whereby global slow oscillations modulate local fast oscillations, usually their amplitude (Canolty & Knight, 2010), which happens to be the predominant effect of astrocytes on fast waves (Perea et al., 2016; Sardinha et al., 2017). By regulating fast waves, astrocytes will have an impact on neuronal encoding, because fast rhythms provide temporal reference frames for local and large-scale computations (Hawellek, Wong, & Pesaran, 2016). Dimensionality reduction (below) may reveal specific astrocytic Ca^{2+} regimes associated with coincidence detection, oscillations, and brain state transitions.

7 | A ROADMAP TO ADVANCE THE INTEGRATION OF ASTROCYTES INTO SYSTEMS NEUROSCIENCE

7.1 | Theoretical and conceptual improvements

Is there a minimal astrocyte–neuronal circuit?

Anatomical, molecular, and functional factors matter when considering astrocytes from a computational point of view. From an anatomical perspective, a single astrocyte can be regarded by itself as a “mini-circuit”, in light of the subcellular compartmentalization of calcium signals (Bazargani & Attwell, 2016), along with the consideration that one astrocytic anatomical domain may comprise numerous neurons, dendrites, and synapses. Estimations in the mouse hippocampus are: 1–20 neurons (Halassa, Fellin, Takano, Dong, & Haydon, 2007), 300–600 dendrites (Halassa et al., 2007), and 140,000 synapses in Bushong, Martone, Jones, and Ellisman (2002), and 50,700–75,200 in Chai et al. (2017). Recently, a FRET-based study reports dynamic interactions of astrocytic distal processes with different types of synaptic inputs (Octeau et al., 2018). Moreover, because astrocytes are characteristically territorial, they give rise to a tiled arrangement of the brain space, which can be then seen as a patchwork of mini-circuits. The function of tiling is an outstanding question. From a molecular perspective, according to single-cell gene profiling, and unbiased hierarchical clustering in mouse brains, astrocyte populations are not as functionally heterogeneous as neuronal populations (Zeisel et al., 2015). Thus, in the mouse somatosensory cortex and hippocampal CA1 region, there are 29 types of neurons including pyramidal cells, glutamatergic neurons, and interneurons, as opposed to just two types of astrocytes (Zeisel et al., 2015). This suggests that, although both neurons and astrocytes are molecularly specialized cells,



additional and extensive sub-specialization exists among neurons but not astrocytes. On the other hand, the lack of molecular definition may provide astrocytes with greater adaptive capacity to operate in a variety of circuits (Poskanzer & Molofsky, 2018), which may explain phenotypical differences of astrocytes from region to region (Chai et al., 2017; Martin et al., 2015). We thus argue that neurons imprint functional signatures on networks by encoding, for example, odors, position, images, words, abstract categories, and executive functions, whereas the size, anatomical arrangement, and molecular makeup of astrocytes suggest that they might be designed to operate canonical computations (Section 6, Table 1) in local mini-circuits within larger-scale networks—as well as homeostatic and metabolic support. Support for the hypothesis that astrocytes perform canonical computations comes from studies showing that astrocyte-based computations such as synaptic potentiation, a type of gain control, improve the performance of ANNs (Alvarellos-Gonzalez et al., 2012; Portopazos et al., 2011). Additional support comes from recent theoretical studies in computer science, and formal language theory, which showed that canonical filtering of synaptic transmission by astrocytes (described as “astrocyte-like control”) facilitates the generation of the so-called logic gates (Song et al., 2016), which are basic building blocks in neural circuits performing logic Boolean operations such as AND, OR, NOT, XOR, and NAND (Binder et al., 2007). According to these studies, simple ensembles of astrocytes and synapses reminiscent of our mini-circuits might account for all elementary logical functions and, properly combined, allow, in principle, computation of any real-world function in a scalable manner (Song et al., 2016). It should be kept in mind that multiple strategies are likely at play across species in shaping astrocytic mini-circuits, and their possible computational functions. For example, although single-cell genomics is not yet available in humans, the fact that human astrocytes are larger, more complex (including 270,000–2 million synapses), and present more morphological variants than mouse astrocytes (Oberheim et al., 2009), together with the striking observation that engraftment of human astrocytes into mouse brains enhances synaptic plasticity and learning (Han et al., 2013), suggests that more complex astrocytic mini-circuits are present in humans, possibly underpinning a larger variety of canonical computations. All in all, it appears that in order to reinforce the presence of astrocytes in Systems Neuroscience, we must zoom out at astrocyte populations as well as zoom into single-astrocyte mini-circuits. This is akin to neuron-focused studies that, as noted, should cover both systems-wide and sub-cellular computations. Indeed, the latter should be considered as part of the computations within astrocyte mini-circuits, for spines and dendrites are inextricably embedded in an astrocyte “matrix.”

Where might the “slow” spatiotemporal dynamics of astrocytic Ca^{2+} enter Systems Neuroscience?

The question of which time scales are relevant for neuronal computations has long been debated. Action potentials of individual neurons are characteristically fast and short-lived voltage depolarizations in the range of 1–2 ms. The speed and all-or-nothing nature of these responses, as well as their lack of attenuation due to axonal myelination, makes them well suited to transmitting information throughout

the brain in milliseconds. Currently, the *minimal* temporal resolution of the neuronal code appears to be on a millisecond time scale, as shown in sensory processing in the auditory system of mammals (Butts et al., 2007; Kayser, Logothetis, & Panzeri, 2010), and in basic human cognitive capabilities, including semantic abstract categorization of images (e.g., identifying an image as a “dog”; Vanmarcke, Calders, & Wagemans, 2016). This means that stimuli arriving within intervals of a few milliseconds are distinguished as individual entities by neurons that fire individual, millisecond-long spikes in response to each stimulus. Clearly, if astrocyte Ca^{2+} transients are the astrocytic substrate of neural computing—and they are the best candidate thus far—they are too slow to encode ultrafast representations. However, the brain characteristically operates in parallel on a gradient of time scales that are nested and hierarchically organized (Murray et al., 2014). Thus, attention and decision making can last seconds, emotions can arise within seconds, and mood changes in minutes. In prediction coding, the slow contextual changes in the prefrontal brain under which fast sensory representations are interpreted require seconds (Kiebel, Daunizeau, & Friston, 2008). Also, there are circadian time scales affecting sleep and global homeostasis, and very long time scales in the range of hours, weeks, or years affecting learning and memory (Hari & Parkkonen, 2015). This means that complex operations ought to exist prolonging the effect of ultrafast (up to 10 ms) and fast (<100 ms) neuronal time scales up to minutes, which precludes structural changes caused by gene expression. Working memory during decision making is a prototypical example of the need for sustained activity in the short-term scale. The question is how several discrete, millisecond-long events related are engaged in a continuum of network activities that last up to hundreds of seconds (Hasson et al., 2015). Since there is no external input during delays (time between input and action), working memory must arise from the intrinsic dynamics of neural circuits. Computational neuroscience identified this problem over 20 years ago (Seung, 1996), and has since struggled to provide answers using realistic neuronal parameters (Chaudhury and Fiete, 2016). Answers include: (a) biophysical properties of neurons such as the slow “membrane-time constant”, which reflects the time during which information can be maintained by neuronal voltage without a substantial leak, estimated to last between 5 and 20 ms, (b) intervention of NMDA receptors, which are ideally suited to enlarge “memory” capabilities of neurons beyond their membrane time constants because they are active around 100 ms after the synaptic input (X. J. Wang, 1999), (c) short-term synaptic plasticity (Abbott & Regehr, 2004), (d) an effective computational solution called long short-term memory (Hochreiter & Schmidhuber, 1997), and (e) sustained firing rate of neurons, or “persistent activity,” achieved upon the exquisite tuning of recurrent circuits such that an input re-entering a synapse exactly matches the decay of the neuron, keeping its firing rate for a prolonged time (Goldman-Rakic, 1995; Renart, Moreno-Bote, Wang, & Parga, 2007). These solutions present limitations. Slow time constants need to be reset, and, at present, slow time constants in neurons do not seem to have that capability. The time constant of the NMDA receptor is appropriate to maintain memories up to 1–5 s, but not longer. Long short-term memory works very well in current machine learning applications, but its application to natural circuits is unclear.

Finally, it is also unclear how the exact timing of feedback loops in persistent activity is achieved. Clearly, additional solutions are in order, perhaps including astrocytes.

Inclusion of astrocytes in current theoretical frameworks and circuit-operating principles

The temporal dynamics of Ca^{2+} -based excitability make astrocytes suitable to operate in circuit computations running in the subsecond to a supra-second scale, including the ones already mentioned such as short-term plasticity, neuromodulation, and slow rhythms. Interestingly, computations such as signal-coincidence detection and oscillation control imply detection of the order and interval of arrival of time-varying signals, suggesting that astrocytes might encode time. Theoretical models of timing in the brain such as oscillators (Goel & Buonomano, 2014) and liquid state (or liquid computing; Maass et al., 2002) may be useful to explore this idea. Astrocytes might also have a role in predictive coding. As shown in silico renditions (Deneve et al., 2017), the core idea of the framework is that neural circuits are error-driven, such that differences between predictions and new inputs are computed as prediction errors, which might be transformed (i.e., “rerouted”) into changes in synaptic strength by short-term plasticity. The greater the error, the more synaptic changes would be needed in order to “update” circuit information. The quality of prediction errors is computed by the variable “precision”, which is akin to the standard error in the Student's *t* test, and is hypothesized to occur in a scale of seconds, and to be encoded by neuromodulators (Friston, 2009; Stephan et al., 2015). Since astrocytes participate in neuromodulation (Navarrete et al., 2012; Takata et al., 2011; Ding et al., 2013; Paukert et al., 2014), the possibility emerges that astrocytes might encode precision, perhaps by temporally decoding prediction errors from multiple synapses in the astrocyte mini-circuit, in order to ensure sufficient statistics. It is tempting to speculate that the aforementioned canonical computations carried out by astrocytes are manifestations of computation of error-related statistics and/or time in different contexts. These computations would be canonical, for they would occur throughout the brain. Decoding analyses (below) may provide information about the specific computations carried out by astrocytes in complex behaviors where issues like timing, temporal holding of information, and error between predictions and real outcomes, are particularly prominent.

Astrocytes and energy-efficient coding

Circuit modeling and biophysical analyses support the idea that neuronal circuits are designed to produce energy-efficient codes because action potentials are energetically demanding; hence, energy supply becomes a relevant constraint in information processing (Laughlin, 2001). Three reasons justify a revision of the adjustment of coding to energy constraints from the perspective of astrocytes. First, astrocytes may lessen the metabolic constraint by facilitating lactate to neurons during task-elicited glutamatergic neurotransmission (Magistretti & Allaman, 2015). Of note, lactate qualifies as a gliotransmitter, and hence may be harvested for computational signaling tasks, because it instructs memory acquisition (Suzuki et al., 2011), and stimulates neurons by a mechanism independent of its uptake,

perhaps receptor-mediated (Tang et al., 2014). Second, as noted in (Magistretti & Allaman, 2015), the anatomical arrangement of local neurons, projections from neuromodulatory nuclei and astrocytes within cortical columns, points to optimized circuit design to facilitate energetic coupling between neurons and astrocytes. Here we extend this notion to astrocyte mini-circuits, and argue that they might represent a coding strategy to optimize energy utilization, for example, by integrating sparse coding, which is coding distributed among many synapses to reduce individual computational load, and has been described as a solution to energy limitations (Laughlin, 2001). Third, whether energy is also a constraint in Ca^{2+} -based computations in astrocytes is an outstanding question. There is currently no estimation of the energy demand of Ca^{2+} signaling in astrocytes. ATP-consuming steps are: (a) in the context of IP3R2-mediated Ca^{2+} -release, reuptake of cytosolic Ca^{2+} back into the endoplasmic reticulum via Ca^{2+} /ATPase pumps, which are crucial in dictating the period of Ca^{2+} fluctuations/oscillations, as well as their shape and duration; (b) the plasmalemma Ca^{2+} /ATPase pump involved in capacitive Ca^{2+} entry/flux; (c) Na^+ / K^+ -ATPase activity dependent on glutamate uptake (Pellerin & Magistretti, 1997), which appears to critically influence Ca^{2+} rises in sensory processing (Schummers et al., 2008); (d) V-ATPase dependent uptake of Ca^{2+} into acidic stores; and (e) neuronal-activity dependent Ca^{2+} rises in astrocytic microdomains in distal processes, as shown in mice with membrane-anchored GCaMP3 (Agarwal et al., 2017). This study documents a critical link between energy metabolism and Ca^{2+} -based excitability, because it shows that Ca^{2+} rises in microdomains are the result of Ca^{2+} efflux from mitochondria, which, in turn, is triggered by short events (mitoflashes) of superoxide production during oxidative phosphorylation. Still, the need for ATP for several critical processes is an open question, a prime example of which is gliotransmission: The exact source of gliotransmitters such as ATP, glutamate, and D-serine, and the energy expenditure involved in their production, is unknown. All in all, it is worth stressing that fatty acids are a fuel for oxidative metabolism in astrocytes (Eraso-Pichot et al., 2018). Since fatty-acid oxidation yields over 50 times more ATP molecules than glycolysis, astrocyte metabolism might be optimized to undertake costly computations from the point of view of energy requirements.

Ca^{2+} -independent computations

Although productive, the adoption of Ca^{2+} signaling as a readout of astrocyte excitability should not blind us to the possibility that similar to Ca^{2+} transients in neurons following action potentials, the astrocytic Ca^{2+} response might be a late manifestation of yet undiscovered signals. If we recover classic perspectives of biophysics (Barlow, 1996; Destexhe, 1999), many components of the astrocytic response could potentially encode stimulations and perform computations. This is the case of second messenger molecules such as IP_3 or cAMP that are conventionally associated with GPCR-mediated astrocytic Ca^{2+} signaling (DePittà, Ben-Jacob, & Berry, 2019) but also of other ion-based signals. Among the latter, Na^+ is an emerging candidate because it presents activity-dependent fluctuations, although advanced

fluorescent probes are necessary to fully establish this ion as a novel readout of astrocyte excitability (Rose & Verkhratsky, 2016).

7.2 | Technical and analytical improvements

7.2.1 | Zooming into astrocyte mini-circuits

Dimensionality reduction of Ca²⁺ data

We posit that single-astrocytes and astrocyte populations are dynamical systems governed by function-specific regimes resulting from coordinated changes in Ca²⁺ signaling. At the single-astrocyte level, the local and global activation modalities described earlier might be the extremes of a spectrum of possible regimes. Dimensionality reduction is a statistical method developed in machine learning to facilitate analysis of the characteristically multidimensional (i.e., multivariate) dynamical systems. What dimensionality reduction does is to identify key variables determining relationships within the data (the so-called latent variables), thereby reducing input data to low-dimensional representations defined by such latent variables. In systems, dimensionality reduction has been applied to neuron-population recordings in decision making, movement, odor perception, working memory, visual attention, audition, rule learning, and speech [reviewed in Cunningham & Yu, 2014]. The complex spatiotemporal patterns of spontaneous and evoked Ca²⁺ transients in single astrocytes, which now can be measured with three-dimensional Ca²⁺-imaging (Bindocci et al., 2017), represent a multidimensional data set that will benefit from dimensionality reduction techniques. Thus far, Ca²⁺ transients in astrocytes have been simplified for quantification purposes by using a single Ca²⁺ readout (Perea et al., 2014), the average of calcium signals detected in multiple ROIs pooled from a population of astrocytes (Poskanzer & Yuste, 2016), the categorization of these signals by spatial location and averaging within subcellular compartments (Chai et al., 2017), and machine-learning based identification of true signals (Agarwal et al., 2017). Although these approaches have already yielded useful insights into correlations between astrocytic and neuronal activities and behaviors—as described in Section 6—they have not revealed possible canonical spatiotemporal computations within and between astrocytes, in distinct experimental paradigms. Dimensionality reduction will thus facilitate detection of noise (stochastic Ca²⁺ transients), indicate whether some of the manually selected ROIs based on visual inspection are not independent, and hence can be considered as the same ROI, and reveal correlations and anti-correlations of distant regions belonging to the same ROI. The latter can occur when distant regions are synchronized due to oscillations or synchronous inputs that regularly occur in those regions. Thus, dimensionality reduction of single-astrocytes may help to reveal and select dimensions, that is, the minimum number of ROIs (e.g., 5–10 from up to 200 original ones), in which fluctuations are more pronounced and meaningful, thus paving the way for population analyses, which will require the simplification of Ca²⁺ signals per astrocyte with the minimal loss of relevant information. Linear methods for dimensionality reduction that can be used in astrocytes include simple principal component analysis (PCA), the prime linear method (Cunningham & Yu,

2014), as well as factor analysis, as used with neuronal Ca²⁺ (Paninski & Cunningham, 2018).

Machine learning

Nonlinear methods such as ANNs are increasingly being used to replace stages in signal processing and analysis in neuronal populations, as well as a method for dimensionality reduction (Paninski & Cunningham, 2018). Thus, ANNs could a priori uncover latent variables that best account for Ca²⁺ data from astrocyte mini-circuits, and are nonlinearly related. Current ANNs appear well-suited to extract latent variables from Ca²⁺ imaging of large populations of neurons (Paninski & Cunningham, 2018), and their application to multidimensional astrocytic Ca²⁺ data should be explored. Conversely, ANNs can be also used as generative models, that is, models that infer classes of inputs from a low number of latent variables (Dosovitskiy, Springenberg, & Brox, 2015). Another statistical tool of machine learning that holds promise is Bayesian hierarchical modeling (Bishop, 2006). The general idea is to build a graph that hierarchically and probabilistically relates relevant variables related to Ca²⁺ and to other data from connectomics. Indeed, if the graphs are well-informed about the connectome within mini-circuits, they can be used as an inverted model to infer the values of the latent variables accounting for Ca²⁺ signals. One advantage of these methods is that the number of free parameters is typically lower than in standard ANNs, which might require massive amounts of data for training.

Connectomics

Providing an accurate picture of the synaptic contacts within astrocyte mini-circuits, in rodents and humans, and in different brain regions, is necessary to help interpret and model in silico Ca²⁺-based regimes defined by dimensionality reduction, and to identify constraints that could be incorporated into machine-learning algorithms. Specific questions are the density of excitatory and inhibitory synapses (and subtypes of the latter), their functional interplay in distinct astrocyte regimes defined by Ca²⁺. For example, astrocyte mini-circuits might adopt feed-forward, recurrent or mixed patterns, depending on the behavioral task, and present hierarchical organizations between astrocytic and neuronal elements, as well as topological/functional “motifs” and wiring rules—as shown in the analysis of small neuronal networks (Schroter, Paulsen, & Bullmore, 2017). Tools for connectomics include graph theory (Fornito, Zalesky, & Bullmore, 2016), Bayesian hierarchical modeling (Bishop, 2006), and topological tools (Kanari et al., 2018; Reimann et al., 2017). In all these approaches, both morphological and functional readouts could serve as input data. Morphological readouts of the synaptic architecture of astrocyte mini-circuits at mesoscale and microscale can be obtained with array tomography, a form of light microscopy based on the serial sectioning of ultrathin (hundreds of microns) sections, which permits 3D reconstructions at a micrometer/nanometer resolution (Micheva et al., 2010). Array tomography can be complemented with automated 3D electron microscopy techniques, such as serial block-face ANNs electron microscopy (SBFSEM). Crucially, fixation methods must not distort contacts within mini-circuits (Korogod et al., 2015). Functional analyses are more challenging, for they will require the development of improved optical tools and

probes to simultaneously monitor the activities of excitatory and inhibitory neuronal populations, as well as those of astrocytes. The emerging combination of two-photon calcium imaging with SBFSEM for examining neural circuits at cellular resolution may pave the way for subcellular analyses (Vishwanathan et al., 2017). Finally, recent multiplex Ca^{2+} imaging at a single synapse-astrocyte interface (J. P. Reynolds et al., 2019), application of nanotechnology to voltage recording in neurons (Jayant et al., 2017), and FRET-based analysis of contacts between synapses and astrocytes (Oceau et al., 2018), are advances toward integrating structure and function in astrocyte mini-circuits.

7.2.2 | Zooming out to astrocyte populations

Decoding astrocytes in complex behavioral tasks

The identification of an astrocytic Ca^{2+} -based code is a prime objective that, importantly, can be started with current statistical tools developed to study neuron-based encoding and decoding. Moreover, we argue that the increased interest in neuronal Ca^{2+} as a tool to decipher the brain code (the reason being that the number of neurons recorded with optical tools is one order of magnitude higher than with multi-electrode arrays, see Section 2) benefits the analysis of Ca^{2+} -based astrocyte computations. For simplicity, here we focus on decoding approaches, which specifically seek to predict external variables from signal patterns, although tools to study encoding can be also considered (Section 3). Decoding astrocyte signals entails measuring Ca^{2+} activity populations in behavioral paradigms in which several time scales, including those in the range of action defined for Ca^{2+} -based signaling in astrocytes (hundreds of milliseconds to tens of seconds), are relevant for the task at hand. One such paradigm is reward-associated decision making over variable contexts in which an animal must associate stimuli with choices (responses) to obtain an immediate reward. The association can be abruptly reversed, as in the case of reversal learning, where in a given Context 1, stimulus A leads to reward and stimulus B does not lead to reward, whereas in another Context 2, stimulus B predicts reward (Schoenbaum, Nugent, Saddoris, & Setlow, 2002). The performance in such varying contexts involves tracking variables at both fast and slow time scales. Variables such as “immediate reward,” “confidence,” “option values,” and “choice” are fast, represented in the millisecond time scale, whereas the deliberation occurring before a decision is taken lasts hundreds of milliseconds to seconds, and even up to minutes, if this deliberation involves inference about the current context. During this time, the brain computes correlations between fast variables, and represents differences between the prediction based on previous experience and the real outcome as “error.” We argue that the precise computation of prediction error is key in the identification of a true association between stimulus and reward, such that varying contexts plausibly require more complex computations. Frontal areas are expected to track the mixture of relevant variables in the form of “cognitive maps.” In rat, the orbitofrontal cortex encodes the millisecond-long fast variables (Nogueira et al., 2017; Rolls, Critchley, Mason, & Wakeman, 1996). It is unclear, however, how transitions between contexts and associated deliberations are represented at

the much slower time scale of seconds. We posit that the network may use astrocytes as a buffer to help represent the prior history of rewards and choices, which is necessary to infer the true nature of the current context. Specifically, astrocytes may temporally integrate error signals, or somehow influence behavior based on accumulated information through canonical computations such as gain modulation. Along these lines, dopaminergic neuromodulation, which signals reward prediction error (O'Doherty, Cockburn, & Pauli, 2017), might serve to gate information from neurons to astrocytes, and vice versa.

Technical and analytical challenges associated with large-scale recordings of Ca^{2+} rises in astrocytes and neurons

The specific experimental design we propose involves the simultaneous recording of Ca^{2+} activity in astrocytes with two-photon microscopy in awake animals (Srinivasan et al., 2015), and Ca^{2+} or electrophysiological responses in neurons (Poskanzer & Yuste, 2016). From previous work indicating that with tens of neurons, it is possible to predict animal choices with high accuracy (Kiani et al., 2014; Nogueira et al., 2017), we reason that tens of astrocytes will suffice to observe statistically significant trends that can be used to guide subsequent recordings and analyses. At this time, optimal selection of paradigms and analytical methods may be more helpful to make significant leaps toward understanding astrocyte-based computations than massively increasing the number of astrocytes recorded. Data acquisition, signal processing and increased dimensionality of the data present additional challenges when there is a need to perform recordings of two cell types with different Ca^{2+} dynamics. As to data acquisition, although recent advances have pushed the boundaries of multiphoton imaging, with significant improvements that enable imaging in multiple brain areas, across *laminae*, and in nonhead-fixed configurations (Yang & Yuste, 2017), since these imaging methodologies have been developed specifically to record the activity of neuronal populations, they may not always be translatable to astrocyte populations. For example, many of the technologies used to carry out 3D two-photon imaging rely on source separation algorithms that assume the Ca^{2+} signals are nonpropagative and spatially static. While this is true for Ca^{2+} imaging of neuronal somata, astrocyte Ca^{2+} imaging data obviously do not obey these rules. Thus, new two-photon imaging methodologies born from an astrocytic perspective, particularly those that allow imaging multiple *laminae* simultaneously, are necessary to advance our understanding of these cells within larger, mesoscale circuits. Another area of improvement for large-scale Ca^{2+} recording in astrocytes and spike-recording in neurons is the development of new electrophysiological approaches, including flexible polymer probes (Chung et al., 2019) and clear electrode arrays (Thunemann et al., 2018), to solve the current problem posed by the large equipment necessary to carry out single-neuron recordings, which precludes astrocyte imaging. Despite the advances in Ca^{2+} imaging, single-neuron electrophysiological measurements are preferable, for Ca^{2+} transients lack temporal resolution to reveal single-action potentials. With regards to signal processing, we described earlier the state-of-the-art in signal processing in large-scale recordings in neurons, including methods to denoise, demix, and simplify Ca^{2+} data. As



to astrocytes, readouts to be assessed per astrocyte are Ca^{2+} signals in microdomains measured in dynamic ROIs (Agarwal et al., 2017; Wang et al., 2006), and/or processed with dimensionality reduction techniques as explained above. A priori, dimensionality reduction and decoding techniques can be used with data from astrocyte and neuronal populations. Possible experimental scenarios are paired Ca^{2+} imaging from both cell types (e.g., low-dimensional data *per* astrocyte could be paired with one optical or electrophysiological signal *per* neuron). Dimensionality reduction may reveal pools of neurons interacting with specific astrocytes. Similarly, linear and nonlinear decoders could be trained to predict relevant behavioral variables from neuron-astrocyte networks and to study which sets of neurons and astrocytes are more relevant for that decoding. Linear decoding techniques could be used even if the amount of behavioral data is not massive, such that around 10 trials per stimulus-choice condition might suffice to obtain a description of astrocyte-neuronal interactions at behaviorally relevant time scales.

7.3 | Translation: Clinical Systems Neuroscience

When it comes to treatments for CNS diseases, molecular and cellular approaches should not be abandoned, because they have successfully led to current therapeutic venues. For example, in multiple sclerosis, relapses are mitigated by immunotherapy against specific populations of immune cells (Torkildsen, Myhr, & Bo, 2016), and in Alzheimer's disease, promising anti- β -amyloid treatments are being tested in clinical trials (Kastanenka et al., 2016; Sevigny et al., 2016). However, there are no effective preventive or disease-modifying treatments for neurodegenerative and psychiatric disorders, suggesting that reductionist approaches aimed at fighting disease one molecule or one cell at a time might be insufficient. Moreover, degeneration of neuromodulatory nuclei (Kelly et al., 2017; Liu, Chang, Pearce, & Gentleman, 2015), as well as large-scale network disarrangement (Westerberg et al., 2012), are hallmarks of psychiatric and neurodegenerative diseases. Clearly, brain diseases are associated with dysfunction of neural systems. Although the outstanding question persists of whether such dysfunction is cause, consequence, or epiphenomenon, the notion that Systems-oriented research will prove more fruitful than traditional approaches to discovering, and thus manipulating, the biological underpinnings of diseases, has already been voiced for autism (Rosenberg, Patterson, & Angelaki, 2015), and motivates therapeutic approaches such as deep brain stimulation in Parkinson's disease (Ashkan, Rogers, Bergman, & Ughratdar, 2017). We anticipate that optogenetic and chemogenetic stimulations will be the most productive avenues in the emerging field of Clinical Systems Neuroscience (Kastanenka, Herlitze, Boyden, Tsai, & Bacskaï, 2017). First, these approaches offer the advantage of selective actions at the network and cellular levels—critically allowing the assessment of neuronal versus astrocytic effects—since viral vectors may be targeted at specific regions through stereotaxic surgery. Second, they enable preclinical research in rodents and primates to demonstrate *causality* between network dysfunction and disease hallmarks (Kastanenka et al., 2017). Third, advances in viral vector technology for gene transfer significantly reduce vector-associated

cytotoxicity and immune responses (Lundstrom, 2018), rendering chemogenetics and optogenetics amenable for clinical use in human patients.

8 | CONCLUDING REMARKS

We started this perspective article by posing several questions to guide the analysis of the role of astrocytes within Systems Neurosciences. We looked for initial answers in available studies that include measurements of astrocyte Ca^{2+} activity, targeted optogenetic and chemogenetic manipulations, and complex behaviors or neural networks. We asked whether astrocytes are as functionally heterogeneous as neurons. We contend that they are not. We put forth anatomical, molecular, and computational arguments in support that astrocytes may operate modules akin to mini-circuits in large scale networks, performing canonical computations throughout the brain. Mathematical analyses of *in vivo* data together with *in silico* modeling will be necessary to firmly establish the existence, and nature, of astrocytic computations, and whether they encode specific variables. We may get closer to the answer using decoding approaches in reward-associated decision making over variable contexts, a complex behavioral paradigm in which the brain needs to perform difficult computations within the slow time scale of astrocytic Ca^{2+} signals. Another question was whether astrocytes use Ca^{2+} to carry out spatiotemporal integration of multicellular signals. A first insight is that there is behavior-dependent integration in a time scale of subseconds to supra-seconds, perhaps driven by signal thresholding and timing control. We propose to use dimensionality reduction, a tool developed in the context of machine learning, to identify the minimum amount of ROIs that carry independent information in Ca^{2+} transients in different contexts. This is a mandatory step toward finding structure in these transients, with the assumption that astrocytic Ca^{2+} responses behave like a dynamical system that can adopt multiple regimes. Thus, the question of whether subcellular compartments in astrocytes perform different functions ought to be reformulated to whether there are function-specific Ca^{2+} regimes. Furthermore, we identify technical and analytical shortages in joint astrocyte- and neuron-population imaging, and ensuring data processing algorithms. Finally, we point to theoretical frameworks used by Systems Neurosciences that might benefit from the inclusion of astrocytes. Many avenues of exploration remain. To cite two, we have the role of astrocyte-based computations in long-term processes underlying memory, perhaps by intervening in memory replay in the so-called resting brain, and the failure of neural circuits including astrocytes in neurodegenerative and psychiatric diseases. Decoding astrocytes may represent a leap forward toward novel approaches in the study of astrocytes in health and disease.

ACKNOWLEDGMENTS

The authors thank Amit Agarwal of the Institute for Anatomy and Cell Biology, Heidelberg University, Germany, and Alfonso Araque of the

Department of Neuroscience, University of Minnesota, USA, for critical reading of the manuscript. The authors also thank Tom Yohannan for copy-editing. Authors declare no conflict of interest. The work was funded by the following agencies: BFU2017-85936-P and FLAGERA-PCIN-2015-162-C02-02 from MINECO (Spain) and the Howard Hughes Medical Institute (HHMI; ref 55008742) to R.M-B; Ministerio de Educacion, Cultura y Deporte (Spain), FPU13/05377 to AEP. The "Junior Leader" Fellowship Program by "la Caixa" Foundation, the Eusko Jaurlaritz through the BERC 2018-2021 program, and the Spanish Ministry of Science, Innovation and Universities by the BCAM Severo Ochoa accreditation SEV-2017-0718 to MDP; MINECO, BFU2016-75107-P to G.P. NIH R01NS099254 and NSF 1604544 to KEP; Agència de Gestió d'Ajuts Universitaris i de Recerca, 2017 SGR547 to EG. MINECO, BFU2016-79735-P to EG and RM.

ORCID

Gertrudis Perea  <https://orcid.org/0000-0001-5924-9175>

Roser Masgrau  <https://orcid.org/0000-0002-6722-5939>

Elena Galea  <https://orcid.org/0000-0003-4537-9897>

REFERENCES

- Abbott, L. F., & Regehr, W. G. (2004). Synaptic computation. *Nature*, 431(7010), 796–803. <https://doi.org/10.1038/nature03010>
- Adamsky, A., Kol, A., Kreisel, T., Doron, A., Ozeri-Engelhard, N., Melcer, T., ... Goshen, I. (2018). Astrocytic activation generates de novo neuronal potentiation and memory enhancement. *Cell*, 174, 59–71.e14. <https://doi.org/10.1016/j.cell.2018.05.002>
- Agarwal, A., Wu, P. H., Hughes, E. G., Fukaya, M., Tischfield, M. A., Langseth, A. J., ... Bergles, D. E. (2017). Transient opening of the mitochondrial permeability transition pore induces microdomain calcium transients in astrocyte processes. *Neuron*, 93(3), 587–605.e587. <https://doi.org/10.1016/j.neuron.2016.12.034>
- Airau, R. D., Thompson, K. R., Fenno, L. E., Bernstein, H., & Deisseroth, K. (2009). Temporally precise in vivo control of intracellular signalling. *Nature*, 458(7241), 1025–1029. <https://doi.org/10.1038/nature07926>
- Alexander, G. M., Rogan, S. C., Abbas, A. I., Armbruster, B. N., Pei, Y., Allen, J. A., ... Roth, B. L. (2009). Remote control of neuronal activity in transgenic mice expressing evolved G protein-coupled receptors. *Neuron*, 63(1), 27–39. <https://doi.org/10.1016/j.neuron.2009.06.014>
- Aljadeff, J., Lansdell, B. J., Fairhall, A. L., & Kleinfeld, D. (2016). Analysis of neuronal spike trains, deconstructed. *Neuron*, 91(2), 221–259. <https://doi.org/10.1016/j.neuron.2016.05.039>
- Alvarelos-Gonzalez, A., Pazos, A., & Porto-Pazos, A. B. (2012). Computational models of neuron-astrocyte interactions Lead to improved efficacy in the performance of neural networks. *Computational and Mathematical Methods in Medicine*, 2012, 1–10. <https://doi.org/10.1155/2012/476324>
- Arandia-Romero, I., Nogueira, R., Mochol, G., & Moreno-Bote, R. (2017). What can neuronal populations tell us about cognition? *Current Opinion in Neurobiology*, 46, 48–57. <https://doi.org/10.1016/j.conb.2017.07.008>
- Araque, A., Carmignoto, G., Haydon, P. G., Oliet, S. H., Robitaille, R., & Volterra, A. (2014). Gliotransmitters travel in time and space. *Neuron*, 81(4), 728–739. <https://doi.org/10.1016/j.neuron.2014.02.007>
- Arieli, A., Sterkin, A., Grinvald, A., & Aertsen, A. (1996). Dynamics of ongoing activity: Explanation of the large variability in evoked cortical responses. *Science*, 273(5283), 1868–1871.
- Ashkan, K., Rogers, P., Bergman, H., & Ughratdar, I. (2017). Insights into the mechanisms of deep brain stimulation. *Nature Reviews. Neurology*, 13(9), 548–554. <https://doi.org/10.1038/nrneurol.2017.105>
- Avery, M. C., & Krichmar, J. L. (2017). Neuromodulatory systems and their interactions: A review of models, theories, and experiments. *Frontiers in Neural Circuits*, 11, 108. <https://doi.org/10.3389/fncir.2017.00108>
- Baars, B. J. (2005). Global workspace theory of consciousness: Toward a cognitive neuroscience of human experience. *Progress in Brain Research*, 150, 45–53. [https://doi.org/10.1016/S0079-6123\(05\)50004-9](https://doi.org/10.1016/S0079-6123(05)50004-9)
- Barlow, H. (1996). Intraneuronal information processing, directional selectivity and memory for spatio-temporal sequences. *Network: Computation in Neural Systems*, 7(2), 251–259. <https://doi.org/10.1088/0954-898x/7/2/004>
- Barrett, L. F. (2017). The theory of constructed emotion: An active inference account of interoception and categorization. *Social Cognitive and Affective Neuroscience*, 12(11), 1833. <https://doi.org/10.1093/scan/nsx060>
- Bazargani, N., & Attwell, D. (2016). Astrocyte calcium signaling: The third wave. *Nature Neuroscience*, 19(2), 182–189. <https://doi.org/10.1038/nn.4201>
- Bernardinelli, Y., Salmon, C., Jones, E. V., Farmer, W. T., Stellwagen, D., & Murai, K. K. (2011). Astrocytes display complex and localized calcium responses to single-neuron stimulation in the hippocampus. *The Journal of Neuroscience*, 31(24), 8905–8919. <https://doi.org/10.1523/JNEUROSCI.6341-10.2011>
- Betzel, R. F., & Bassett, D. S. (2017). Multi-scale brain networks. *NeuroImage*, 160, 73–83. <https://doi.org/10.1016/j.neuroimage.2016.11.006>
- Binder, A., Freund, R., Oswald, M., & Volk, L. (2007). *Extended spiking neural P systems with excitatory and inhibitory astrocytes*. In: Proceedings of the Fifth Brainstorming Week on Membrane Computing (pp. 63–72). Sevilla, ETS de Ingeniería Informática, 29 de Enero-2 de Febrero, 2007, Publisher Fénix editor
- Bindocci, E., Savtchouk, I., Liaudet, N., Becker, D., Carriero, G., & Volterra, A. (2017). Three-dimensional ca(2+) imaging advances understanding of astrocyte biology. *Science*, 356(6339), eaai8185. <https://doi.org/10.1126/science.aai8185>
- Bishop, C. M. (2006). *Pattern recognition and machine learning*. New York, NY: Springer.
- Bonder, D. E., & McCarthy, K. D. (2014). Astrocytic Gq-GPCR-linked IP3R-dependent Ca2+ signaling does not mediate neurovascular coupling in mouse visual cortex in vivo. *The Journal of Neuroscience*, 34(39), 13139–13150. <https://doi.org/10.1523/JNEUROSCI.2591-14.2014>
- Boydén, E. S., Zhang, F., Bamberg, E., Nagel, G., & Deisseroth, K. (2005). Millisecond-timescale, genetically targeted optical control of neural activity. *Nature Neuroscience*, 8(9), 1263–1268. <https://doi.org/10.1038/nn1525>
- Brunel, N. (2000). Dynamics of sparsely connected networks of excitatory and inhibitory spiking neurons. *Journal of Computational Neuroscience*, 8(3), 183–208. <https://doi.org/10.1023/A:1008925309027>
- Bushong, E. A., Martone, M. E., Jones, Y. Z., & Ellisman, M. H. (2002). Protoplasmic astrocytes in CA1 stratum radiatum occupy separate anatomical domains. *The Journal of Neuroscience*, 22(1), 183–192.
- Butts, D. A., Weng, C., Jin, J., Yeh, C. I., Lesica, N. A., Alonso, J. M., & Stanley, G. B. (2007). Temporal precision in the neural code and the timescales of natural vision. *Nature*, 449(7158), 92–95. <https://doi.org/10.1038/nature06105>
- Buzsáki, G. (2010). Neural syntax: Cell assemblies, synapse ensembles, and readers. *Neuron*, 68(3), 362–385. <https://doi.org/10.1016/j.neuron.2010.09.023>
- Buzsáki, G., & Draguhn, A. (2004). Neuronal oscillations in cortical networks. *Science*, 304(5679), 1926–1929. <https://doi.org/10.1126/science.1099745>
- Cangelosi, A., & Invitto, S. (2017). Human-robot interaction and Neuroprosthetics: A review of new technologies. *IEEE Consumer Electronics Magazine*, 6(3), 24–33. <https://doi.org/10.1109/Mce.2016.2614423>



- Canolty, R. T., & Knight, R. T. (2010). The functional role of cross-frequency coupling. *Trends in Cognitive Sciences*, 14(11), 506–515. <https://doi.org/10.1016/j.tics.2010.09.001>
- Capone, C., Rebollo, B., Munoz, A., Illa, X., Del Giudice, P., Sanchez-Vives, M. V., & Mattia, M. (2017). Slow waves in cortical slices: How spontaneous activity is shaped by laminar structure. *Cerebral Cortex*, 29, 1–17. <https://doi.org/10.1093/cercor/bhx326>
- Chai, H., Diaz-Castro, B., Shigetomi, E., Monte, E., Octeau, J. C., Yu, X., ... Khakh, B. S. (2017). Neural circuit-specialized astrocytes: Transcriptomic, proteomic, morphological, and functional evidence. *Neuron*, 95(3), 531–549 e539. <https://doi.org/10.1016/j.neuron.2017.06.029>
- Chalk, M., Marre, O., & Tkacik, G. (2018). Toward a unified theory of efficient, predictive, and sparse coding. *Proceedings of the National Academy of Sciences of the United States of America*, 115(1), 186–191. <https://doi.org/10.1073/pnas.1711141115>
- Chang, L., & Tsao, D. Y. (2017). The code for facial identity in the primate brain. *Cell*, 169(6), 1013–1028 e1014. <https://doi.org/10.1016/j.cell.2017.05.011>
- Chaudhuri, R., & Fiete, I. (2016). Computational principles of memory. *Nature Neuroscience*, 19(3), 394–403. <https://doi.org/10.1038/nn.4237>
- Chen, N., Sugihara, H., Kim, J., Fu, Z., Barak, B., Sur, M., ... Han, W. (2016). Direct modulation of GFAP-expressing glia in the arcuate nucleus bi-directionally regulates feeding. *eLife*, 5, e18716. <https://doi.org/10.7554/eLife.18716>
- Chung, J. E., Joo, H. R., Fan, J. L., Liu, D. F., Barnett, A. H., Chen, S., ... Frank, L. M. (2019). High-density, long-lasting, and multi-region electrophysiological recordings using polymer electrode arrays. *Neuron*, 101(1), 21–31 e5. <https://doi.org/10.1016/j.neuron.2018.11.002>
- Covelo, A., & Araque, A. (2018). Neuronal activity determines distinct gliotransmitter release from a single astrocyte. *eLife*, 7, e32237. <https://doi.org/10.7554/eLife.32237>
- Cross, Z. R., Kohler, M. J., Schlesewsky, M., Gaskell, M. G., & Bornkessel-Schlesewsky, I. (2018). Sleep-dependent memory consolidation and incremental sentence comprehension: Computational dependencies during language learning as revealed by neuronal oscillations. *Frontiers in Human Neuroscience*, 12, 18. <https://doi.org/10.3389/fnhum.2018.00018>
- Cunningham, J. P., & Ghahramani, Z. (2015). Linear dimensionality reduction: Survey, insights, and generalizations. *Journal of Machine Learning Research*, 16, 2859–2900.
- Cunningham, J. P., & Yu, B. M. (2014). Dimensionality reduction for large-scale neural recordings. *Nature Neuroscience*, 17(11), 1500–1509. <https://doi.org/10.1038/nn.3776>
- De Pitta, M., Volman, V., Levine, H., Piozzia, G., De Rossi, D., & Ben-Jacob, E. (2008). Coexistence of amplitude and frequency modulations in intracellular calcium dynamics. *Physical Review. E, Statistical, Nonlinear, and Soft Matter Physics*, 77(3 Pt 1), 030903. <https://doi.org/10.1103/PhysRevE.77.030903>
- Deisseroth, K. (2015). Optogenetics: 10 years of microbial opsins in neuroscience. *Nature Neuroscience*, 18(9), 1213–1225. <https://doi.org/10.1038/nn.4091>
- Deneve, S., Alemi, A., & Bourdoukan, R. (2017). The brain as an efficient and robust adaptive learner. *Neuron*, 94(5), 969–977. <https://doi.org/10.1016/j.neuron.2017.05.016>
- DePittà, M., Ben-Jacob, E., & Berry, H. (2019). G-protein-coupled receptor-mediated calcium signaling in astrocytes. In M. De Pittà & H. Berry (Eds.), *Computational Glioscience* (pp. 115–150). New York, NY: Springer.
- Destexhe, A. (1999). Biophysics of computation: Information processing in single neurons. *Trends in Cognitive Sciences*, 3(11), 444–444. [https://doi.org/10.1016/S1364-6613\(99\)01380-7](https://doi.org/10.1016/S1364-6613(99)01380-7)
- Diederen, K. M., Ziauddeen, H., Vestergaard, M. D., Spencer, T., Schultz, W., & Fletcher, P. C. (2017). Dopamine modulates adaptive prediction error coding in the human midbrain and striatum. *The Journal of Neuroscience*, 37(7), 1708–1720. <https://doi.org/10.1523/JNEUROSCI.1979-16.2016>
- Ding, F., O'Donnell, J., Thrane, A. S., Zeppenfeld, D., Kang, H., Xie, L., ... Nedergaard, M. (2013). alpha1-adrenergic receptors mediate coordinated Ca²⁺ signaling of cortical astrocytes in awake, behaving mice. *Cell Calcium*, 54(6), 387–394. <https://doi.org/10.1016/j.ceca.2013.09.001>
- Dosovitskiy, A., Springenberg, J.T., Brox, A. (2015). *Learning to generate chairs with convolutional neural networks*. In: Proceedings of the 2015 IEEE Conference on Computer Vision and Pattern Recognition (CVPR) (pp 1538–1546). Boston, USA.
- Duarte, R., Seeholzer, A., Zilles, K., & Morrison, A. (2017). Synaptic patterning and the timescales of cortical dynamics. *Current Opinion in Neurobiology*, 43, 156–165. <https://doi.org/10.1016/j.conb.2017.02.007>
- Einevoll, G. T., Franke, F., Hagen, E., Pouzat, C., & Harris, K. D. (2012). Towards reliable spike-train recordings from thousands of neurons with multielectrodes. *Current Opinion in Neurobiology*, 22(1), 11–17. <https://doi.org/10.1016/j.conb.2011.10.001>
- Eraso-Pichot, A., Braso-Vives, M., Golbano, A., Menacho, C., Claro, E., Galea, E., & Masgrau, R. (2018). GSEA of mouse and human mitochondriomes reveals fatty acid oxidation in astrocytes. *Glia*, 66, 1724–1735. <https://doi.org/10.1002/glia.23330>
- Fellin, T., Halassa, M. M., Terunuma, M., Succol, F., Takano, H., Frank, M., ... Haydon, P. G. (2009). Endogenous nonneuronal modulators of synaptic transmission control cortical slow oscillations in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 106(35), 15037–15042. <https://doi.org/10.1073/pnas.0906419106>
- Fitch, W. T. (2014). Toward a computational framework for cognitive biology: Unifying approaches from cognitive neuroscience and comparative cognition. *Physics of Life Reviews*, 11(3), 329–364. <https://doi.org/10.1016/j.plrev.2014.04.005>
- Fornito, A., Zalesky, A., & Bullmore, E. (2016). *Fundamentals of brain network analysis*. Boston, MA: Academic Press.
- Foster, D. J. (2017). Replay comes of age. *Annual Review of Neuroscience*, 40, 581–602. <https://doi.org/10.1146/annurev-neuro-072116-031538>
- Fox, M. D., & Raichle, M. E. (2007). Spontaneous fluctuations in brain activity observed with functional magnetic resonance imaging. *Nature Reviews. Neuroscience*, 8(9), 700–711. <https://doi.org/10.1038/nrn2201>
- Friston, K. (2009). The free-energy principle: A rough guide to the brain? *Trends in Cognitive Sciences*, 13(7), 293–301. <https://doi.org/10.1016/j.tics.2009.04.005>
- Gerstner, W., Kreiter, A. K., Markram, H., & Herz, A. V. (1997). Neural codes: Firing rates and beyond. *Proceedings of the National Academy of Sciences of the United States of America*, 94(24), 12740–12741.
- Gjorgjieva, J., Drion, G., & Marder, E. (2016). Computational implications of biophysical diversity and multiple timescales in neurons and synapses for circuit performance. *Current Opinion in Neurobiology*, 37, 44–52. <https://doi.org/10.1016/j.conb.2015.12.008>
- Goel, A., & Buonomano, D. V. (2014). Timing as an intrinsic property of neural networks: Evidence from in vivo and in vitro experiments. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 369(1637), 20120460. <https://doi.org/10.1098/rstb.2012.0460>
- Goldman-Rakic, P. S. (1995). Cellular basis of working memory. *Neuron*, 14(3), 477–485.
- Gomez, J. L., Bonaventura, J., Lesniak, W., Mathews, W. B., Sysa-Shah, P., Rodriguez, L. A., ... Michaelides, M. (2017). Chemogenetics revealed: DREADD occupancy and activation via converted clozapine. *Science*, 357(6350), 503–507. <https://doi.org/10.1126/science.aan2475>
- Gourine, A. V., Kasymov, V., Marina, N., Tang, F., Figueiredo, M. F., Lane, S., ... Kasparov, S. (2010). Astrocytes control breathing through pH-dependent release of ATP. *Science*, 329(5991), 571–575. <https://doi.org/10.1126/science.1190721>
- Gradinaru, V., Mogri, M., Thompson, K. R., Henderson, J. M., & Deisseroth, K. (2009). Optical deconstruction of parkinsonian neural circuitry. *Science*, 324(5925), 354–359. <https://doi.org/10.1126/science.1167093>

- Halassa, M. M., Fellin, T., Takano, H., Dong, J. H., & Haydon, P. G. (2007). Synaptic islands defined by the territory of a single astrocyte. *The Journal of Neuroscience*, 27(24), 6473–6477. <https://doi.org/10.1523/JNEUROSCI.1419-07.2007>
- Halassa, M. M., Florian, C., Fellin, T., Munoz, J. R., Lee, S. Y., Abel, T., ... Frank, M. G. (2009). Astrocytic modulation of sleep homeostasis and cognitive consequences of sleep loss. *Neuron*, 61(2), 213–219. <https://doi.org/10.1016/j.neuron.2008.11.024>
- Han, X., Chen, M., Wang, F., Windrem, M., Wang, S., Shanz, S., ... Nedergaard, M. (2013). Forebrain engraftment by human glial progenitor cells enhances synaptic plasticity and learning in adult mice. *Cell Stem Cell*, 12(3), 342–353. <https://doi.org/10.1016/j.stem.2012.12.015>
- Hari, R., & Parkkonen, L. (2015). The brain timewise: How timing shapes and supports brain function. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 370(1668), 20140170. <https://doi.org/10.1098/rstb.2014.0170>
- Hasselmo, M. E., & McGaughy, J. (2004). High acetylcholine levels set circuit dynamics for attention and encoding and low acetylcholine levels set dynamics for consolidation. *Progress in Brain Research*, 145, 207–231. [https://doi.org/10.1016/S0079-6123\(03\)45015-2](https://doi.org/10.1016/S0079-6123(03)45015-2)
- Hasson, U., Chen, J., & Honey, C. J. (2015). Hierarchical process memory: Memory as an integral component of information processing. *Trends in Cognitive Sciences*, 19(6), 304–313. <https://doi.org/10.1016/j.tics.2015.04.006>
- Hausteil, M. D., Kracun, S., Lu, X. H., Shih, T., Jackson-Weaver, O., Tong, X., ... Khakh, B. S. (2014). Conditions and constraints for astrocyte calcium signaling in the hippocampal mossy fiber pathway. *Neuron*, 82(2), 413–429. <https://doi.org/10.1016/j.neuron.2014.02.041>
- Hawellek, D. J., Wong, Y. T., & Pesaran, B. (2016). Temporal coding of reward-guided choice in the posterior parietal cortex. *Proceedings of the National Academy of Sciences of the United States of America*, 113(47), 13492–13497. <https://doi.org/10.1073/pnas.1606479113>
- Hochreiter, S., & Schmidhuber, J. (1997). Long short-term memory. *Neural Computation*, 9(8), 1735–1780.
- Jayant, K., Hirtz, J. J., Plante, I. J., Tsai, D. M., De Boer, W. D., Semonche, A., ... Yuste, R. (2017). Targeted intracellular voltage recordings from dendritic spines using quantum-dot-coated nanopipettes. *Nature Nanotechnology*, 12(4), 335–342. <https://doi.org/10.1038/nnano.2016.268>
- Kanari, L., Dlotko, P., Scolamiero, M., Levi, R., Shillcock, J., Hess, K., & Markram, H. (2018). A topological representation of branching neuronal morphologies. *Neuroinformatics*, 16(1), 3–13. <https://doi.org/10.1007/s12021-017-9341-1>
- Kastanenka, K. V., Arbel-Ornath, M., Hudry, E., Galea, E., Xie, H., & Bacskaï, B. J. (2016). Optical probes for in vivo imaging. Optical probes in biology. In J. Zhang, S. Mehta, & C. Schultz (Eds.), *Optical probes in biology* (Vol. 14, pp. 355–376). Boca Raton, FL: CRC Press.
- Kastanenka, K. V., Bussiere, T., Shakerdige, N., Qian, F., Weinreb, P. H., Rhodes, K., & Bacskaï, B. J. (2016). Immunotherapy with Aducanumab restores calcium homeostasis in Tg2576 mice. *The Journal of Neuroscience*, 36(50), 12549–12558. <https://doi.org/10.1523/JNEUROSCI.2080-16.2016>
- Kastanenka, K. V., Herlitze, S., Boyden, E. S., Tsai, L.-H., & Bacskaï, B. J. (2017). Optogenetics shed light on Alzheimer's disease. *EC Neurology*, ECO.01, 27–29.
- Kastanenka, K. V., Hou, S. S., Shakerdige, N., Logan, R., Feng, D., Wegmann, S., ... Bacskaï, B. J. (2017). Optogenetic restoration of disrupted slow oscillations halts amyloid deposition and restores calcium homeostasis in an animal model of Alzheimer's disease. *PLoS One*, 12(1), e0170275. <https://doi.org/10.1371/journal.pone.0170275>
- Kayser, C., Logothetis, N. K., & Panzeri, S. (2010). Millisecond encoding precision of auditory cortex neurons. *Proceedings of the National Academy of Sciences of the United States of America*, 107(39), 16976–16981. <https://doi.org/10.1073/pnas.1012656107>
- Kelly, S. C., He, B., Perez, S. E., Ginsberg, S. D., Mufson, E. J., & Counts, S. E. (2017). Locus coeruleus cellular and molecular pathology during the progression of Alzheimer's disease. *Acta Neuropathologica Communications*, 5(1), 8. <https://doi.org/10.1186/s40478-017-0411-2>
- Khani, A., & Rainer, G. (2016). Neural and neurochemical basis of reinforcement-guided decision making. *Journal of Neurophysiology*, 116(2), 724–741. <https://doi.org/10.1152/jn.01113.2015>
- Kiani, R., Cueva, C. J., Reppas, J. B., & Newsome, W. T. (2014). Dynamics of neural population responses in prefrontal cortex indicate changes of mind on single trials. *Current Biology*, 24(13), 1542–1547. <https://doi.org/10.1016/j.cub.2014.05.049>
- Kiebel, S. J., Daunizeau, J., & Friston, K. J. (2008). A hierarchy of time-scales and the brain. *PLoS Computational Biology*, 4(11), e1000209. <https://doi.org/10.1371/journal.pcbi.1000209>
- Kok, P., & de Lange, F. P. (2014). Shape perception simultaneously up- and downregulates neural activity in the primary visual cortex. *Current Biology*, 24(13), 1531–1535. <https://doi.org/10.1016/j.cub.2014.05.042>
- Korogod, N., Petersen, C. C., & Knott, G. W. (2015). Ultrastructural analysis of adult mouse neocortex comparing aldehyde perfusion with cryo fixation. *Elife*, 4. <https://doi.org/10.7554/eLife.05793>
- Lau, B., Monteiro, T., & Paton, J. J. (2017). The many worlds hypothesis of dopamine prediction error: Implications of a parallel circuit architecture in the basal ganglia. *Current Opinion in Neurobiology*, 46, 241–247. <https://doi.org/10.1016/j.conb.2017.08.015>
- Laughlin, S. B. (2001). Energy as a constraint on the coding and processing of sensory information. *Current Opinion in Neurobiology*, 11(4), 475–480.
- Ledoux, E., & Brunel, N. (2011). Dynamics of networks of excitatory and inhibitory neurons in response to time-dependent inputs. *Frontiers in Computational Neuroscience*, 5, 25. <https://doi.org/10.3389/fncom.2011.00025>
- Lee, S. H., & Dan, Y. (2012). Neuromodulation of brain states. *Neuron*, 76(1), 209–222. <https://doi.org/10.1016/j.neuron.2012.09.012>
- Leinweber, M., Ward, D. R., Sobczak, J. M., Attinger, A., & Keller, G. B. (2017). A sensorimotor circuit in mouse cortex for visual flow predictions. *Neuron*, 96(5), 1204. <https://doi.org/10.1016/j.neuron.2017.11.009>
- Li, X., Gutierrez, D. V., Hanson, M. G., Han, J., Mark, M. D., Chiel, H., ... Herlitze, S. (2005). Fast noninvasive activation and inhibition of neural and network activity by vertebrate rhodopsin and green algae channelrhodopsin. *Proceedings of the National Academy of Sciences of the United States of America*, 102(49), 17816–17821. <https://doi.org/10.1073/pnas.0509030102>
- Litwin-Kumar, A., & Doiron, B. (2012). Slow dynamics and high variability in balanced cortical networks with clustered connections. *Nature Neuroscience*, 15(11), 1498–1505. <https://doi.org/10.1038/nn.3220>
- Liu, A. K., Chang, R. C., Pearce, R. K., & Gentleman, S. M. (2015). Nucleus basalis of Meynert revisited: Anatomy, history and differential involvement in Alzheimer's and Parkinson's disease. *Acta Neuropathologica*, 129(4), 527–540. <https://doi.org/10.1007/s00401-015-1392-5>
- London, M., & Hausser, M. (2005). Dendritic computation. *Annual Review of Neuroscience*, 28, 503–532. <https://doi.org/10.1146/annurev.neuro.28.061604.135703>
- Lundstrom, K. (2018). Viral vectors in gene therapy. *Diseases*, 6(2), 42. <https://doi.org/10.3390/diseases6020042>
- Lutcke, H., Gerhard, F., Zenke, F., Gerstner, W., & Helmchen, F. (2013). Inference of neuronal network spike dynamics and topology from calcium imaging data. *Frontiers in Neural Circuits*, 7, 201. <https://doi.org/10.3389/fncir.2013.00201>
- Maass, W., Natschlag, T., & Markram, H. (2002). Real-time computing without stable states: A new framework for neural computation based on perturbations. *Neural Computation*, 14(11), 2531–2560. <https://doi.org/10.1162/089976602760407955>
- Magistretti, P. J., & Allaman, I. (2015). A cellular perspective on brain energy metabolism and functional imaging. *Neuron*, 86(4), 883–901. <https://doi.org/10.1016/j.neuron.2015.03.035>



- Magistretti, P. J., & Morrison, J. H. (1988). Noradrenergic- and vasoactive intestinal peptide-containing neuronal systems in neocortex: Functional convergence with contrasting morphology. *Neuroscience*, 24(2), 367–378.
- Manninen, T., Havela, R., & Linne, M. L. (2018). Computational models for calcium-mediated astrocyte functions. *Frontiers in Computational Neuroscience*, 12, 14. <https://doi.org/10.3389/fncom.2018.00014>
- Marder, E. (2015). Understanding brains: Details, intuition, and big data. *PLoS Biology*, 13(5), e1002147. <https://doi.org/10.1371/journal.pbio.1002147>
- Markov, N. T., Vezoli, J., Chameau, P., Falchier, A., Quilodran, R., Huissoud, C., ... Kennedy, H. (2014). Anatomy of hierarchy: Feedforward and feedback pathways in macaque visual cortex. *The Journal of Comparative Neurology*, 522(1), 225–259. <https://doi.org/10.1002/cne.23458>
- Markram, H., Müller, E., Ramaswamy, S., Reimann, M. W., Abdellah, M., Sanchez, C. A., ... Schürmann, F. (2015). Reconstruction and simulation of neocortical microcircuitry. *Cell*, 163(2), 456–492. <https://doi.org/10.1016/j.cell.2015.09.029>
- Marr, D., & Poggio, T. (1976). *From understanding computation to understanding neural circuitry*. Cambridge, MA: Computer Science and Artificial Intelligence Lab, Massachusetts Institute of Technology.
- Martin-Fernandez, M., Jamison, S., Robin, L. M., Zhao, Z., Martin, E. D., Aguilar, J., ... Araque, A. (2017). Synapse-specific astrocyte gating of amygdala-related behavior. *Nature Neuroscience*, 20(11), 1540–1548. <https://doi.org/10.1038/nn.4694>
- Martin, R., Bajo-Graneras, R., Moratalla, R., Perea, G., & Araque, A. (2015). Circuit-specific signaling in astrocyte-neuron networks in basal ganglia pathways. *Science*, 349(6249), 730–734. <https://doi.org/10.1126/science.aaa7945>
- Masgrau, R., Guaza, C., Ransohoff, R. M., & Galea, E. (2017). Should we stop saying 'GliA' and 'Neuroinflammation'? *Trends in Molecular Medicine*, 23(6), 486–500. <https://doi.org/10.1016/j.molmed.2017.04.005>
- Mastrogiuseppe, F., & Ostojic, S. (2018). Linking connectivity, dynamics, and computations in low-rank recurrent neural networks. *Neuron*, 99, 609–623.e29. <https://doi.org/10.1016/j.neuron.2018.07.003>
- Mazurek, M. E., Roitman, J. D., Ditterich, J., & Shadlen, M. N. (2003). A role for neural integrators in perceptual decision making. *Cerebral Cortex*, 13(11), 1257–1269. <https://doi.org/10.1093/cercor/bhg097>
- Mesulam, M. M. (1998). From sensation to cognition. *Brain*, 121(Pt 6), 1013–1052.
- Meunier, C. N., Chameau, P., & Fossier, P. M. (2017). Modulation of synaptic plasticity in the cortex needs to understand all the players. *Frontiers in Synthetic Neuroscience*, 9, 2. <https://doi.org/10.3389/fnsyn.2017.00002>
- Meunier, D., Lambiotte, F., Fornito, A., Ersche, K. D., & Bullmore, E. T. (2009). Hierarchical modularity in human brain functional networks. *Frontiers in Neuroinformatics*, 3, 37. <https://doi.org/10.3389/neuro.11.037.2009>
- Micheva, K. D., O'Rourke, N., Busse, B., & Smith, S. J. (2010). Array tomography: high-resolution three-dimensional immunofluorescence. *Cold Spring Harbor Protocols*, 2010(11), pdb top89. <https://doi.org/10.1101/pdb.top89>
- Mongillo, G., Barak, O., & Tsodyks, M. (2008). Synaptic theory of working memory. *Science*, 319(5869), 1543–1546. <https://doi.org/10.1126/science.1150769>
- Murray, J. D., Bernacchia, A., Freedman, D. J., Romo, R., Wallis, J. D., Cai, X., ... Wang, X. J. (2014). A hierarchy of intrinsic timescales across primate cortex. *Nature Neuroscience*, 17(12), 1661–1663. <https://doi.org/10.1038/nn.3862>
- Nagel, G., Szellas, T., Huhn, W., Kateriya, S., Adeishvili, N., Berthold, P., ... Bamberg, E. (2003). Channelrhodopsin-2, a directly light-gated cation-selective membrane channel. *Proceedings of the National Academy of Sciences of the United States of America*, 100(24), 13940–13945. <https://doi.org/10.1073/pnas.1936192100>
- Nasser, H. M., Calu, D. J., Schoenbaum, G., & Sharpe, M. J. (2017). The dopamine prediction error: Contributions to associative models of reward learning. *Frontiers in Psychology*, 8, 244. <https://doi.org/10.3389/fpsyg.2017.00244>
- Navarrete, M., Perea, G., Fernandez de Sevilla, D., Gomez-Gonzalo, M., Nunez, A., Martin, E. D., & Araque, A. (2012). Astrocytes mediate in vivo cholinergic-induced synaptic plasticity. *PLoS Biology*, 10(2), e1001259. <https://doi.org/10.1371/journal.pbio.1001259>
- Nogueira, R., Abolafia, J. M., Drugowitsch, J., Balaguer-Ballester, E., Sanchez-Vives, M. V., & Moreno-Bote, R. (2017). Lateral orbitofrontal cortex anticipates choices and integrates prior with current information. *Nature Communications*, 8, 14823. <https://doi.org/10.1038/ncomms14823>
- Oberheim, N. A., Takano, T., Han, X., He, W., Lin, J. H., Wang, F., ... Nedergaard, M. (2009). Uniquely hominid features of adult human astrocytes. *Journal of Neuroscience*, 29(10), 3276–3287. <https://doi.org/10.1523/JNEUROSCI.4707-08.2009>
- Octeau, J. C., Chai, H., Jiang, R. T., Bonanno, S. L., Martin, K. C., & Khakh, B. S. (2018). An optical neuron-astrocyte proximity assay at synaptic distance scales. *Neuron*, 98(1), 49. <https://doi.org/10.1016/j.neuron.2018.03.003>
- O'Doherty, J. P., Cockburn, J., & Pauli, W. M. (2017). Learning, reward, and decision making. *Annual Review of Psychology*, 68, 73–100. <https://doi.org/10.1146/annurev-psych-010416-044216>
- Oliveira, J. F., Sardinha, V. M., Guerra-Gomes, S., Araque, A., & Sousa, N. (2015). Do stars govern our actions? Astrocyte involvement in rodent behavior. *Trends in Neurosciences*, 38(9), 535–549. <https://doi.org/10.1016/j.tins.2015.07.006>
- Otsu, Y., Couchman, K., Lyons, D. G., Collot, M., Agarwal, A., Mallet, J. M., ... Charpak, S. (2015). Calcium dynamics in astrocyte processes during neurovascular coupling. *Nature Neuroscience*, 18(2), 210–218. <https://doi.org/10.1038/nn.3906>
- Pachitariu, M., Stringer, C., Dipoppa, M., Schröder, S., Rossi, F., Dalgleish, H., ... Harris, K. D. (2016). Suite2p: Beyond 10,000 neurons with standard two-photon microscopy. *bioRxiv*, 061507. <https://doi.org/10.1101/061507>
- Panatier, A., Vallee, J., Haber, M., Murai, K. K., Lacaille, J. C., & Robitaille, R. (2011). Astrocytes are endogenous regulators of basal transmission at central synapses. *Cell*, 146(5), 785–798. <https://doi.org/10.1016/j.cell.2011.07.022>
- Paninski, L., & Cunningham, J. P. (2018). Neural data science: Accelerating the experiment-analysis-theory cycle in large-scale neuroscience. *Current Opinion in Neurobiology*, 50, 232–241. <https://doi.org/10.1016/j.conb.2018.04.007>
- Panzeri, S., Macke, J. H., Gross, J., & Kayser, C. (2015). Neural population coding: Combining insights from microscopic and mass signals. *Trends in Cognitive Sciences*, 19(3), 162–172. <https://doi.org/10.1016/j.tics.2015.01.002>
- Panzeri, S., Petersen, R. S., Schultz, S. R., Lebedev, M., & Diamond, M. E. (2001). The role of spike timing in the coding of stimulus location in rat somatosensory cortex. *Neuron*, 29(3), 769–777.
- Paukert, M., Agarwal, A., Cha, J., Doze, V. A., Kang, J. U., & Bergles, D. E. (2014). Norepinephrine controls astroglial responsiveness to local circuit activity. *Neuron*, 82(6), 1263–1270. <https://doi.org/10.1016/j.neuron.2014.04.038>
- Pellerin, L., & Magistretti, P. J. (1997). Glutamate uptake stimulates Na⁺,K⁺-ATPase activity in astrocytes via activation of a distinct subunit highly sensitive to ouabain. *Journal of Neurochemistry*, 69(5), 2132–2137.
- Pelluru, D., Konadhode, R. R., Bhat, N. R., & Shiromani, P. J. (2016). Optogenetic stimulation of astrocytes in the posterior hypothalamus increases sleep at night in C57BL/6J mice. *The European Journal of Neuroscience*, 43(10), 1298–1306. <https://doi.org/10.1111/ejn.13074>
- Perea, G., & Araque, A. (2005). Properties of synaptically evoked astrocyte calcium signal reveal synaptic information processing by astrocytes. *The Journal of Neuroscience*, 25(9), 2192–2203. <https://doi.org/10.1523/JNEUROSCI.3965-04.2005>

- Perea, G., Gomez, R., Mederos, S., Covelo, A., Ballesteros, J. J., Schlosser, L., ... Araque, A. (2016). Activity-dependent switch of GABAergic inhibition into glutamatergic excitation in astrocyte-neuron networks. *eLife*, 5, e20362. <https://doi.org/10.7554/eLife.20362>
- Perea, G., Yang, A., Boyden, E. S., & Sur, M. (2014). Optogenetic astrocyte activation modulates response selectivity of visual cortex neurons in vivo. *Nature Communications*, 5, 3262. <https://doi.org/10.1038/ncomms4262>
- Pnevmatikakis, E. A., Soudry, D., Gao, Y., Machado, T. A., Merel, J., Pfau, D., ... Paninski, L. (2016). Simultaneous Denoising, deconvolution, and Demixing of calcium imaging data. *Neuron*, 89(2), 285–299. <https://doi.org/10.1016/j.neuron.2015.11.037>
- Porto-Pazos, A. B., Veiguela, N., Mesejo, P., Navarrete, M., Alvarellos, A., Ibanez, O., ... Araque, A. (2011). Artificial astrocytes improve neural network performance. *PLoS One*, 6(4), e19109. <https://doi.org/10.1371/journal.pone.0019109>
- Poskanzer, K. E., & Molofsky, A. V. (2018). Dynamism of an astrocyte in vivo: Perspectives on identity and function. *Annual Review of Physiology*, 80, 143–157. <https://doi.org/10.1146/annurev-physiol-021317-121125>
- Poskanzer, K. E., & Yuste, R. (2016). Astrocytes regulate cortical state switching in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 113(19), E2675–E2684. <https://doi.org/10.1073/pnas.1520759113>
- Quiñ Quiroga, R., & Panzeri, S. (2009). Extracting information from neuronal populations: Information theory and decoding approaches. *Nature Reviews Neuroscience*, 10(3), 173–185. <https://doi.org/10.1038/nrn2578>
- Reimann, M. W., Nolte, M., Scolamiero, M., Turner, K., Perin, R., Chindemi, G., ... Markram, H. (2017). Cliques of neurons bound into cavities provide a missing link between structure and function. *Frontiers in Computational Neuroscience*, 11, 48. <https://doi.org/10.3389/fncom.2017.00048>
- Renart, A., Moreno-Bote, R., Wang, X. J., & Parga, N. (2007). Mean-driven and fluctuation-driven persistent activity in recurrent networks. *Neural Computation*, 19(1), 1–46. <https://doi.org/10.1162/neco.2007.19.1.1>
- Reynolds, J. P., Zheng, K., & Rusakov, D. A. (2019). Multiplexed calcium imaging of single-synapse activity and astroglial responses in the intact brain. *Neuroscience Letters*, 689, 26–32. <https://doi.org/10.1016/j.neulet.2018.06.024>
- Reynolds, J. H., & Heeger, D. J. (2009). The normalization model of attention. *Neuron*, 61(2), 168–185. <https://doi.org/10.1016/j.neuron.2009.01.002>
- Rolls, E. T., Critchley, H. D., Mason, R., & Wakeman, E. A. (1996). Orbitofrontal cortex neurons: Role in olfactory and visual association learning. *Journal of Neurophysiology*, 75(5), 1970–1981. <https://doi.org/10.1152/jn.1996.75.5.1970>
- Rose, C. R., & Verkhratsky, A. (2016). Principles of sodium homeostasis and sodium signalling in astroglia. *Glia*, 64(10), 1611–1627. <https://doi.org/10.1002/glia.22964>
- Rosenberg, A., Patterson, J. S., & Angelaki, D. E. (2015). A computational perspective on autism. *Proceedings of the National Academy of Sciences of the United States of America*, 112(30), 9158–9165. <https://doi.org/10.1073/pnas.1510583112>
- Roth, B. L. (2016). DREADDs for neuroscientists. *Neuron*, 89(4), 683–694. <https://doi.org/10.1016/j.neuron.2016.01.040>
- Saez, I., Lin, J., Stolk, A., Chang, E., Parvizi, J., Schalk, G., ... Hsu, M. (2018). Encoding of multiple reward-related computations in transient and sustained high-frequency activity in human OFC. *Current Biology*, 28(18), 2889–2899 e2883. <https://doi.org/10.1016/j.cub.2018.07.045>
- Sanchez-Vives, M. V., Massimini, M., & Mattia, M. (2017). Shaping the default activity pattern of the cortical network. *Neuron*, 94(5), 993–1001. <https://doi.org/10.1016/j.neuron.2017.05.015>
- Santello, M., Toni, N., & Volterra, A. (2019). Astrocyte function from information processing to cognition and cognitive impairment. *Nature Neuroscience*, 22(2), 154–166. <https://doi.org/10.1038/s41593-018-0325-8>
- Sara, S. J. (2009). The locus coeruleus and noradrenergic modulation of cognition. *Nature Reviews Neuroscience*, 10(3), 211–223. <https://doi.org/10.1038/nrn2573>
- Sardinha, V. M., Guerra-Gomes, S., Caetano, I., Tavares, G., Martins, M., Reis, J. S., ... Oliveira, J. F. (2017). Astrocytic signaling supports hippocampal-prefrontal theta synchronization and cognitive function. *Glia*, 65(12), 1944–1960. <https://doi.org/10.1002/glia.23205>
- Sasaki, T., Beppu, K., Tanaka, K. F., Fukazawa, Y., Shigemoto, R., & Matsui, K. (2012). Application of an optogenetic byway for perturbing neuronal activity via glial photostimulation. *Proceedings of the National Academy of Sciences of the United States of America*, 109(50), 20720–20725. <https://doi.org/10.1073/pnas.1213458109>
- Savtchouk, I., & Volterra, A. (2018). Gliotransmission: Beyond black-and-white. *The Journal of Neuroscience*, 38(1), 14–25. <https://doi.org/10.1523/JNEUROSCI.0017-17.2017>
- Schmidt-Hieber, C., Toleikyte, G., Aitchison, L., Roth, A., Clark, B. A., Branco, T., & Hausser, M. (2017). Active dendritic integration as a mechanism for robust and precise grid cell firing. *Nature Neuroscience*, 20(8), 1114–1121. <https://doi.org/10.1038/nn.4582>
- Schoenbaum, G., Nugent, S. L., Saddoris, M. P., & Setlow, B. (2002). Orbitofrontal lesions in rats impair reversal but not acquisition of go, no-go odor discriminations. *Neuroreport*, 13(6), 885–890.
- Schroter, M., Paulsen, O., & Bullmore, E. T. (2017). Micro-connectomics: Probing the organization of neuronal networks at the cellular scale. *Nature Reviews Neuroscience*, 18(3), 131–146. <https://doi.org/10.1038/nrn.2016.182>
- Schummers, J., Yu, H., & Sur, M. (2008). Tuned responses of astrocytes and their influence on hemodynamic signals in the visual cortex. *Science*, 320(5883), 1638–1643. <https://doi.org/10.1126/science.1156120>
- Sejnowski, T. J., Koch, C., & Churchland, P. S. (1988). Computational neuroscience. *Science*, 241(4871), 1299–1306.
- Seung, H. S. (1996). How the brain keeps the eyes still. *Proceedings of the National Academy of Sciences of the United States of America*, 93(23), 13339–13344.
- Sevigny, J., Suhy, J., Chiao, P., Chen, T., Klein, G., Purcell, D., ... Barakos, J. (2016). Amyloid PET screening for enrichment of early-stage Alzheimer disease clinical trials: Experience in a phase 1b clinical trial. *Alzheimer Disease and Associated Disorders*, 30(1), 1–7. <https://doi.org/10.1097/WAD.0000000000000144>
- Seymour, K., Clifford, C. W., Logothetis, N. K., & Bartels, A. (2010). Coding and binding of color and form in visual cortex. *Cerebral Cortex*, 20(8), 1946–1954. <https://doi.org/10.1093/cercor/bhp265>
- Shenoy, K. V., Sahani, M., & Churchland, M. M. (2013). Cortical control of arm movements: A dynamical systems perspective. *Annual Review of Neuroscience*, 36, 337–359. <https://doi.org/10.1146/annurev-neuro-062111-150509>
- Shigetomi, E., Tong, X., Kwan, K. Y., Corey, D. P., & Khakh, B. S. (2011). TRPA1 channels regulate astrocyte resting calcium and inhibitory synapse efficacy through GAT-3. *Nature Neuroscience*, 15(1), 70–80. <https://doi.org/10.1038/nn.3000>
- Sofroniew, N. J., Flickinger, D., King, J., & Svoboda, K. (2016). A large field of view two-photon mesoscope with subcellular resolution for in vivo imaging. *eLife*, 5, e14472. <https://doi.org/10.7554/eLife.14472>
- Song, T., Pan Zheng, M. L., Wong, D., & Wang, X. (2016). Design of logic gates using spiking neural P systems with homogeneous neurons and astrocytes-like control. *Information Sciences*, 372, 380–391. <https://doi.org/10.1016/j.ins.2016.08.055>
- Soudry, D., Keshri, S., Stinson, P., Oh, M. H., Iyengar, G., & Paninski, L. (2015). Efficient "shotgun" inference of neural connectivity from highly sub-sampled activity data. *PLoS Computational Biology*, 11(10), e1004464. <https://doi.org/10.1371/journal.pcbi.1004464>
- Sporns, O. (2013). Structure and function of complex brain networks. *Dia-logues in Clinical Neuroscience*, 15(3), 247–262.
- Srinivasan, R., Huang, B. S., Venugopal, S., Johnston, A. D., Chai, H., Zeng, H., ... Khakh, B. S. (2015). Ca²⁺ signaling in astrocytes from *Ip3r2(-/-)* mice in brain slices and during startle responses in vivo. *Nature Neuroscience*, 18(5), 708–717. <https://doi.org/10.1038/nn.4001>



- Stellwagen, D., & Malenka, R. C. (2006). Synaptic scaling mediated by glial TNF- α . *Nature*, 440(7087), 1054–1059. <https://doi.org/10.1038/nature04671>
- Stephan, K. E., Iglesias, S., Heinzle, J., & Diaconescu, A. O. (2015). Translational perspectives for computational neuroimaging. *Neuron*, 87(4), 716–732. <https://doi.org/10.1016/j.neuron.2015.07.008>
- Stobart, J. L., Ferrari, K. D., Barrett, M. J. P., Gluck, C., Stobart, M. J., Zuend, M., & Weber, B. (2018). Cortical circuit activity evokes rapid astrocyte calcium signals on a similar timescale to neurons. *Neuron*, 98(4), 726–735.e724. <https://doi.org/10.1016/j.neuron.2018.03.050>
- Stringer, C., & Pachitariu, M. (2018). Computational processing of neural recordings from calcium imaging data. *Current Opinion in Neurobiology*, 55, 22–31. <https://doi.org/10.1016/j.conb.2018.11.005>
- Stuart, G., Spruston, N., Sakmann, B., & Häusser, M. (1997). Action potential initiation and backpropagation in neurons of the mammalian CNS. *Trends in Neurosciences*, 20(3), 125–131.
- Summerfield, C., Trittschuh, E. H., Monti, J. M., Mesulam, M. M., & Egner, T. (2008). Neural repetition suppression reflects fulfilled perceptual expectations. *Nature Neuroscience*, 11(9), 1004–1006. <https://doi.org/10.1038/nn.2163>
- Suzuki, A., Stern, S. A., Bozdagi, O., Huntley, G. W., Walker, R. H., Magistretti, P. J., & Alberini, C. M. (2011). Astrocyte-neuron lactate transport is required for long-term memory formation. *Cell*, 144(5), 810–823. <https://doi.org/10.1016/j.cell.2011.02.018>
- Sweatt, J. D. (2016). Neural plasticity and behavior—Sixty years of conceptual advances. *Journal of Neurochemistry*, 139 Suppl 2, 179–199. <https://doi.org/10.1111/jnc.13580>
- Sweeney, P., Qi, Y., Xu, Z., & Yang, Y. (2016). Activation of hypothalamic astrocytes suppresses feeding without altering emotional states. *Glia*, 64(12), 2263–2273. <https://doi.org/10.1002/glia.23073>
- Takata, N., Mishima, T., Hisatsune, C., Nagai, T., Ebisui, E., Mikoshiba, K., & Hirase, H. (2011). Astrocyte calcium signaling transforms cholinergic modulation to cortical plasticity in vivo. *The Journal of Neuroscience*, 31(49), 18155–18165. <https://doi.org/10.1523/JNEUROSCI.5289-11.2011>
- Tang, F., Lane, S., Korsak, A., Paton, J. F., Gourine, A. V., Kasparov, S., & Teschemacher, A. G. (2014). Lactate-mediated glia-neuronal signalling in the mammalian brain. *Nature Communications*, 5, 3284. <https://doi.org/10.1038/ncomms4284>
- Tani, H., Dulla, C. G., Farzampour, Z., Taylor-Weiner, A., Huguenard, J. R., & Reimer, R. J. (2014). A local glutamate-glutamine cycle sustains synaptic excitatory transmitter release. *Neuron*, 81(4), 888–900. <https://doi.org/10.1016/j.neuron.2013.12.026>
- Thiele, A., & Bellgrove, M. A. (2018). Neuromodulation of attention. *Neuron*, 97(4), 769–785. <https://doi.org/10.1016/j.neuron.2018.01.008>
- Thunemann, M., Lu, Y., Liu, X., Kilic, K., Desjardins, M., Vandenbergh, M., ... Kuzum, D. (2018). Deep 2-photon imaging and artifact-free optogenetics through transparent graphene microelectrode arrays. *Nature Communications*, 9(1), 2035. <https://doi.org/10.1038/s41467-018-04457-5>
- Torkildsen, O., Myhr, K. M., & Bo, L. (2016). Disease-modifying treatments for multiple sclerosis—A review of approved medications. *European Journal of Neurology*, 23(Suppl 1), 18–27. <https://doi.org/10.1111/ene.12883>
- Tsunada, J., & Cohen, Y. E. (2014). Neural mechanisms of auditory categorization: From across brain areas to within local microcircuits. *Frontiers in Neuroscience*, 8, 161. <https://doi.org/10.3389/fnins.2014.00161>
- Vanmarcke, S., Calders, F., & Wagemans, J. (2016). The time-course of ultrarapid categorization: The influence of scene congruency and top-down processing. *Iperception*, 7(5), 204166951667338. <https://doi.org/10.1177/2041669516673384>
- Vidaurre, D., Smith, S. M., & Woolrich, M. W. (2017). Brain network dynamics are hierarchically organized in time. *Proceedings of the National Academy of Sciences of the United States of America*, 114(48), 12827–12832. <https://doi.org/10.1073/pnas.1705120114>
- Vishwanathan, A., Daie, K., Ramirez, A. D., Lichtman, J. W., Aksay, E. R. F., & Seung, H. S. (2017). Electron microscopic reconstruction of functionally identified cells in a neural integrator. *Current Biology*, 27(14), 2137–2147.e2133. <https://doi.org/10.1016/j.cub.2017.06.028>
- Vogelstein, J. T., Packer, A. M., Machado, T. A., Sippy, T., Babadi, B., Yuste, R., & Paninski, L. (2010). Fast nonnegative deconvolution for spike train inference from population calcium imaging. *Journal of Neurophysiology*, 104(6), 3691–3704. <https://doi.org/10.1152/jn.01073.2009>
- Wacongne, C., Labyt, E., van Wassenhove, V., Bekinschtein, T., Naccache, L., & Dehaene, S. (2011). Evidence for a hierarchy of predictions and prediction errors in human cortex. *Proceedings of the National Academy of Sciences of the United States of America*, 108(51), 20754–20759. <https://doi.org/10.1073/pnas.1117807108>
- Wang, X., Lou, N., Xu, Q., Tian, G. F., Peng, W. G., Han, X., ... Nedergaard, M. (2006). Astrocytic Ca²⁺ signaling evoked by sensory stimulation in vivo. *Nature Neuroscience*, 9(6), 816–823. <https://doi.org/10.1038/nn1703>
- Wang, X. J. (1999). Synaptic basis of cortical persistent activity: The importance of NMDA receptors to working memory. *The Journal of Neuroscience*, 19(21), 9587–9603.
- Westerberg, C. E., Mander, B. A., Florczak, S. M., Weintraub, S., Mesulam, M. M., Zee, P. C., & Paller, K. A. (2012). Concurrent impairments in sleep and memory in amnesic mild cognitive impairment. *Journal of the International Neuropsychological Society*, 18(3), 490–500. <https://doi.org/10.1017/S135561771200001X>
- Xu, Y., Zou, P., & Cohen, A. E. (2017). Voltage imaging with genetically encoded indicators. *Current Opinion in Chemical Biology*, 39, 1–10. <https://doi.org/10.1016/j.cbpa.2017.04.005>
- Yamins, D. L., & DiCarlo, J. J. (2016). Using goal-driven deep learning models to understand sensory cortex. *Nature Neuroscience*, 19(3), 356–365. <https://doi.org/10.1038/nn.4244>
- Yang, J., Yu, H., Zhou, D., Zhu, K., Lou, H., Duan, S., & Wang, H. (2015). Na⁺/Ca²⁺ exchanger mediates ChR2-induced [Ca²⁺]_i elevation in astrocytes. *Cell Calcium*, 58(3), 307–316. <https://doi.org/10.1016/j.ceca.2015.06.008>
- Yang, L., Qi, Y., & Yang, Y. (2015). Astrocytes control food intake by inhibiting AGRP neuron activity via adenosine A1 receptors. *Cell Reports*, 11(5), 798–807. <https://doi.org/10.1016/j.celrep.2015.04.002>
- Yang, W., & Yuste, R. (2017). In vivo imaging of neural activity. *Nature Methods*, 14(4), 349–359. <https://doi.org/10.1038/nmeth.4230>
- Yizhar, O., Fenno, L. E., Davidson, T. J., Mogri, M., & Deisseroth, K. (2011). Optogenetics in neural systems. *Neuron*, 71(1), 9–34. <https://doi.org/10.1016/j.neuron.2011.06.004>
- Yuste, R. (2013). Electrical compartmentalization in dendritic spines. *Annual Review of Neuroscience*, 36, 429–449. <https://doi.org/10.1146/annurev-neuro-062111-150455>
- Yuste, R. (2015). From the neuron doctrine to neural networks. *Nature Reviews Neuroscience*, 16(8), 487–497. <https://doi.org/10.1038/nrn3962>
- Zeisel, A., Muñoz-Manchado, A. B., Codeluppi, S., Lönnerberg, P., La Manno, G., Jureus, A., ... Linnarsson, S. (2015). Brain structure. Cell types in the mouse cortex and hippocampus revealed by single-cell RNA-seq. *Science*, 347(6226), 1138–1142. <https://doi.org/10.1126/science.aaa1934>
- Zheng, K., Bard, L., Reynolds, J. P., King, C., Jensen, T. P., Gourine, A. V., & Rusakov, D. A. (2015). Time-resolved imaging reveals heterogeneous landscapes of Nanomolar Ca²⁺ in neurons and Astroglia. *Neuron*, 88(2), 277–288. <https://doi.org/10.1016/j.neuron.2015.09.043>

How to cite this article: Kastanenka KV, Moreno-Bote R, De Pittà M, et al. A roadmap to integrate astrocytes into Systems Neuroscience. *Glia*. 2020;68:5–26. <https://doi.org/10.1002/glia.23632>