

## **Role of quorum sensing and chemical communication in fungal biotechnology and pathogenesis**

**One sentence summary:** This review focuses on the complexity of quorum sensing mechanisms in fungi and its potential applications.

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### **Abstract**

Microbial cells do not live in isolation in their environment, but rather they communicate with each other using chemical signals. This sophisticated mode of cell-to-cell signalling, known as quorum sensing, was first discovered in bacteria, and coordinates the behaviour of microbial population behaviour in a cell-density dependent manner. More recently, these mechanisms have been described in eukaryotes, particularly in fungi, where they regulate processes such as

pathogenesis, morphological differentiation, secondary metabolite production and biofilm formation.

In this manuscript, we review the information available to date on these processes in yeast, dimorphic fungi and filamentous fungi. We analyse the diverse chemical “languages” used by different groups of fungi, their possible cross-talk and interkingdom interactions with other organisms. We discuss the existence of these mechanisms in multicellular organisms, the ecophysiological role of QS in fungal colonisation, and the potential applications of these mechanisms in biotechnology and pathogenesis.

**Key words:** cell-to-cell signalling, yeast, dimorphic, filamentous fungi, farnesol, consortia

## INTRODUCTION

### What is quorum sensing (QS)?

Since the 1960's, it has been known that unicellular microorganisms do not live in isolation in their environment, but there is communication among cells that allows the population to coordinate its behaviour (Nealson *et al.* 1970). This sophisticated cell-to-cell signalling system, known as quorum sensing (QS), is mediated by small molecules that diffuse into the environment and accumulate during the growth of the microbial population. Each individual cell is able to sense the amount of quorum sensing molecules (QSMs) present in the medium which, upon reaching a threshold concentration, coordinate specific gene expression (Bandara *et al.* 2012). Microbes monitor their population density through QS-mediated mechanisms that regulate a number of population density related processes (Miller and Bassler 2001). Moreover, the QSMs are able to stimulate their own production, acting as “autoinducers” (Bandara *et al.* 2012). QS confers a great evolutionary advantage, allowing microbial populations to adapt to changing environments. Some authors suggest that these processes are neo-Darwinian mechanisms of evolution that may have

played an important role in the appearance of the first multicellular organisms (Miller and Bassler 2001).

### **QS in prokaryotes**

The QS phenomenon was first studied in bacteria, where it is known to be involved in the regulation of processes such as pathogenesis, symbiosis, competence, conjugation, nutrient uptake, morphological differentiation, secondary metabolite production, and biofilms formation (Bandara *et al.* 2012). The first organised response dependent on population density was reported in the luminous marine bacterium *Aliivibrio fischeri* (Nealson *et al.* 1970). This pioneer study on this Gram-negative bacterium showed that bioluminescence was regulated by a QS mechanism controlled by a two-component system integrated by a synthase (LuxI), responsible for the production of the QSMs *N*-acyl-homoserine-lactones (AHLs), and a sensor receptor (LuxR) whose activity is dependent on the presence of the QSM. Subsequently, other forms of homoserine lactones and other molecules including quinolones, the Diffusible Signal Factor (DSF), and the Cholera Autoinducer 1, were identified as QSMs in Gram-negative bacteria (Papenfort and Bassler 2016).

QS-mediated signalling in Gram-positive bacteria is often regulated by small peptides after intracellular processing of certain polypeptides and secretion of the final molecule, which is detected by a sensor histidine-kinase (Hawver *et al.* 2016). However, Gram-positive *Streptomyces*, use  $\gamma$ -butyrolactones as QSMs (Biarnes-Carrera *et al.* 2015). Finally, the autoinducer 2 (AI-2) has been described as a QS system common to both Gram-positive and Gram-negative bacteria, although its role as a true QSM has been questioned for some microorganisms (Chen *et al.* 2002; Winzer *et al.* 2002; Bandara *et al.* 2012; Papenfort and Bassler 2016). This system gives rise to a family of molecules that are thought to serve as a common “language” for most bacteria. Many bacteria possess more than one QS system. They can communicate with their own species using their particular QS system (e.g. DSF-type molecules), and also communicate with bacteria from

other genera using AHLs or the AI-2 system, establishing a cross-talk among different bacterial species (Miller *et al.* 2004; Ryan *et al.* 2009; Dow *et al.* 2016).

The study of QS in natural populations in environmentally complex settings is often complicated. This is because there are a variety of cell-intrinsic and cell-extrinsic factors that modulate the kinetics of QS signal production, detection and response (Fig. 1). Intrinsic factors include the roles of other regulators outside of the direct QS circuit that respond to environmental stimuli, such as phosphate availability (Blus-Kadosh *et al.* 2013), metabolic pools that may influence signal synthesis (Bjarnsholt and Givskov, 2007), and genetic differences between strains that alter the dynamics of QS activation. Extrinsic factors include QS interference or “quorum quenching” in which QSMs are inactivated or degraded (Zhu and Kaufmann 2013), environmental factors that influence the stability and diffusion of QSM (Connell *et al.*, 2010), diffusion kinetics, and population size. Furthermore, in some species, there are members of the population that lack certain components of the QS system. These cells may have lost the ability to produce exoproducts beneficial to the population and are referred to as “cheaters” (Diggle *et al.* 2007). Others may have lost their ability to synthesise QSMs (e.g. LuxI defective strains) but are still able to respond to QSMs, this is the case of bacterial strains harbouring LuxR “solos” (Venturi and Ahmer, 2015) that may be referred to as “listeners”. The “cheaters” may have some fitness benefits in the presence of cells that are making quorum-sensing regulated public goods (Diggle *et al.* 2007), while the “listeners” may respond to external QSMs and express favourable traits induced by the population (Venturi and Ahmer, 2015). On the other hand, QS-defective endemic strains are also present in the environment suggesting that there may be specific niches in which QS is not required for fitness (Hammond *et al.*, 2016).

## **QS in eukaryotes**

It has long been known that eukaryotic cells are able to secrete and sense signalling molecules (Youk and Lim 2014). However, now we know that certain types of eukaryotic

microorganisms monitor their population density through QS mechanisms (Hogan 2006a). This would be expected as many bacteria and eukaryotic microbes occupy similar ecological niches and face common challenges. QS in microscopic eukaryotes was first described in the pathogenic dimorphic fungus *Candida albicans* (Hornby *et al.* 2001). Most of the published works on QS in eukaryotes relate to fungi, but it is unclear if this is because QS regulation is more common in fungi or merely easier to detect and study. Nevertheless, the existence of QS mechanisms in the Protista kingdom has been recently claimed, particularly in the parasite *Trypanosoma brucei* (Mony *et al.* 2014) and in the social amoeba *Dictyostelium discoideum* (Tarnita *et al.* 2015).

In fungi, QS mechanisms have been described to regulate processes such as sporulation, secondary metabolite production, morphological transition and enzyme secretion. In this developing field, there are a range of fungal genera displaying these mechanisms and using variety of signalling molecules with different structures. For instance, the inoculum size effect has been well studied in many fungi such as *Ophiostoma (Ceratocystis) ulmi*, *Ophiostoma novo-ulmi*, *Ophiostoma piceae*, *Ophiostoma floccosum*, *Histoplasma capsulatum*, *Mucor rouxii*, *Aureobasidium pullulans*, *Candida albicans*, *Cladosporium werneckii*, *Aspergillus flavus*, *Penicillium isariaeforme*, *Neurospora crassa* and *Schizophyllum commune* (Nickerson *et al.* 2006; Horowitz Brown *et al.* 2008; Berrocal *et al.* 2014; de Salas *et al.* 2015; Wedge *et al.* 2016). Furthermore, fungi can communicate with bacteria, and even their plants or mammals hosts.

QS mechanisms have emerged among the most important aspects in the study of bacterial behaviour, and are often well characterised at the genetic and molecular levels due to their implication in biofilm formation and their potential as future drug targets to be used alone or in conjunction with antibiotics (Camara *et al.* 2002). In contrast, knowledge on QS mechanisms in fungi at the molecular, physiological and ecological levels is scarce. Here we aim to review the role of QS mechanisms in fungi due to their relevance in pathogenesis, ecology and biotechnology. In this context, we will discuss the intrinsic and extrinsic factors that modulate QS (Fig. 1). We will

establish a correlation between the QSMs utilised in different fungal taxa or habitats to decipher the different “languages” used by fungi, their evolution, and their potential for cross-talk. We will discuss the prominence of these mechanisms in filamentous (multicellular) fungi, speculate on interkingdom communication and the formation of bacterial-fungal consortia, and propose future directions for the application of these mechanisms.

### **QS mechanisms in yeast and dimorphic fungi**

Some evidence suggests that the yeast *Saccharomyces cerevisiae* exhibits a QS type of regulation mediated by aromatic alcohols derived from amino acids (Hogan 2006b). This budding yeast produces tryptophol and phenylethyl alcohol to control the formation of pseudohyphae, biofilm formation (Chen and Fink 2006) and perhaps virulence towards the grapevine *Vitis vinifera* (Gognies *et al.* 2006). Some QSMs that were discovered first in other fungal species, such as *Candida albicans*, have been subsequently described to have roles in *S. cerevisiae*. For example, *S. cerevisiae* can produce tyrosol (Avbelj *et al.* 2015) and respond to farnesol (Egbe *et al.* 2017), two QSMs first characterised in *C. albicans* as discussed in detail below.

On the other hand, the only fungus from the group of the basidiomycetes with described QS mechanisms is *Cryptococcus neoformans*, a human pathogen. This fungus, the etiologic agent of cryptococcosis, undergoes cellular changes that result in the appearance of different cellular types during infection. After capsule enlargement and growth of the whole cell “titan cells” are formed (Albuquerque *et al.* 2014). The main virulence factors in this fungus are the formation of the polysaccharide capsule and melanin accumulation, which are regulated by QS mechanisms. Madhani (2011) described QS regulation via peptides (QSP1-4) that are secreted outside the cell prior to uptake via the transporter Opt1. QSP1 is a central signalling molecule that regulates pathogenesis in *C. neoformans*, being a direct target of three transcription factors required for virulence. It is known that QSP1 regulates secreted protease activity and promotes cell wall function at high cell densities. Here, similar to Gram-positive bacteria, the signal production

requires extracellular processing of the pro-peptide proQsp1 by a cell-associated protease (Homer *et al.* 2016).

### **QS in *Candida albicans***

QS mechanisms are best described in the dimorphic yeast *C. albicans*. This species is the causal agent of most common mycosis and is also a member of the resident flora in humans and animals (Nucci *et al.* 2010). Under certain conditions, this organism undergoes a controlled, reversible interconversion of cell morphology, switching from yeast to hyphal form. This transformation is crucial to its pathogenicity and adaptation to the environment (Madhani 2011; Sudbery 2011). The morphological switch in dimorphic fungi is governed by many environmental signals, including temperature, pH, nutrients composition and concentration, carbon dioxide levels, transition metals, chelating agents, and cell population density (Romano 1966; Gauthier 2015). These stimuli act through several well-established signalling cascades (Lo *et al.* 1997). In *C. albicans* two signalling compounds with opposite effects have been described as QSMs: farnesol and tyrosol. Farnesol is a 12-carbon backbone sesquiterpene alcohol ((2E,6E)-3,7,11-trimethyldodeca-2,6,10-trien-1-ol) made up of three isoprene units (Nickerson *et al.* 2006). It is an intermediate in sterol biosynthesis that prevents differentiation from yeast to hyphae (Hornby *et al.* 2001) and induces the switch from hypha back to yeast growth (Lindsay *et al.* 2012). Tyrosol (2-(4-hydroxyphenyl) ethanol) stimulates the exit from lag phase such that germ-tube formation and hyphal development is accelerated in its presence (Chen *et al.* 2004). *C. albicans* QSMs also influence the formation and structure of biofilms (Ramage *et al.* 2002; Cordeiro *et al.* 2015) as well as the dispersal of cells from a biofilm; hence, these molecules play important roles in pathogenesis.

The regulation of morphological changes mediated by *E,E*-farnesol in *C. albicans* is complicated, but well studied. This molecule regulates hyphal development through the repression of the Ras1-cAMP/protein kinase A (PKA) signalling pathway (Davis-Hanna *et al.* 2008). Evidence indicates that farnesol can affect filamentation by suppressing the cAMP pathway through

inhibition of adenylyl cyclase (Cyr1) activity (Hall *et al.* 2011). Activation of the cAMP-PKA pathway down-regulates the expression of Nrg1, the major repressor of hyphal development. Farnesol is able to repress the expression of the kinase required for Nrg1 degradation, Sok1 (Lu *et al.* 2014). Moreover, the transcription repressor Cup9 has been found to be responsible for the regulation of *SOK1* expression in response to farnesol inhibition. Interestingly, farnesol inhibits Cup9 degradation mediated by the N-end rule E3 ubiquitin ligase, Ubr1 (Lu *et al.* 2014). The histidine kinase (Chk1) and the zinc finger 1 (Czf1) from *C. albicans* (Kebaara *et al.* 2008) have also been identified as important response factors to farnesol (Langford *et al.* 2013) Farnesol also modulates the transcription levels of *TUPI1*, a major repressor of the morphological transition (Kebaara *et al.* 2008). Nevertheless, the *E,E*-farnesol export pathways in the cell are unknown, but its secretion has been shown to vary across different strains (Weber *et al.* 2010).

### **QS in other dimorphic fungi**

Over the years, QS mechanisms have been identified in few other dimorphic fungi. Many of these are pathogens of plants or animals, and many have applications in biotechnology. This is the case for fungi from the genus *Ophiostoma*, which includes plant pathogenic species such as *O. ulmi* and its aggressive sister species *O. novo-ulmi*, causal agents of Dutch elm disease. Both species belong phylogenetically to the so called “*O. piceae*-*O. ulmi* complex” (Haridas *et al.* 2013; Forgetta *et al.* 2013; Khoshraftar *et al.* 2013; De Beer and Wingfield 2013). The effect of inoculum size on the morphology of *O. ulmi* as a manifestation of QS regulation has been studied. In 2004 Hornby *et al.* showed that in medium inoculated with  $>10^6$  spores per mL, cells initially developed as budding yeasts, while when inoculated at  $<10^6$  spores per mL, cells developed as mycelia. This inoculum size-dependent effect was related to the presence of QSMs that accumulated throughout the growth of the fungus. When the QSMs were extracted from spent medium with organic solvent and subsequently added back to fresh cultures, the QS activity demonstrated a dose-dependent effect specific for *O. ulmi*, without an appreciable effect on *C. albicans*. Similarly, *O. ulmi* did not respond



to farnesol addition. Further studies indicated that the QSM mediating these processes could be 2-methyl-1-butanol (Berrocal *et al.* 2012). Later, it was found that the *O. ulmi* strain used in this study actually belongs to *O. novo-ulmi* species (Naruzawa *et al.* 2014). Furthermore, inoculum size effects on yeast-mycelium dimorphism have been reported in several strains of *O. novo-ulmi* (Wedge *et al.* 2016). This report revealed that different strains of this species respond differently to inoculum size, confirming a certain level of intra-specific variation. In addition, transcriptome analysis of *O. novo-ulmi* has confirmed that several genes in the cAMP and PKA pathways are differentially expressed during the morphological transition from yeast to hyphae, suggesting that some of these genes might be targets for QSMs, as is the case for *C. albicans* (Nigg and Bernier 2016).

Other species from the genus *Ophiostoma* are plant saprophytes, as is the dimorphic fungus *O. piceae*. QS mechanisms mediated by farnesol have been described in the strain *O. piceae* CECT 20416. The accumulation of farnesol in the culture medium of this dimorphic fungus during the exponential growth phase coordinates the morphological transition from yeast to hyphae, and this is likely modulated by the transcription repressor Tup1 (de Salas *et al.* 2015). Furthermore, the addition of exogenous *E,E*-farnesol or spent medium to fresh cultures of *O. piceae* had a significant effect on fungal morphology; however, its effect was opposite to that produced by the same compounds in *C. albicans*, where it induced filamentation of the fungus (de Salas *et al.* 2015). Experiments with another saprophytic fungus, the albino strain *O. floccosum* FIF1A55 (Berrocal *et al.* 2014) showed an inoculum size effect similar to that reported in *C. albicans* (Hornby *et al.* 2001). In *O. floccosum*, three cyclic sesquiterpenes were described as QSMs in a defined culture medium with L-proline as a sole nitrogen source (Berrocal *et al.* 2014). These studies have shown the diversity of QSMs in the phylogenetically related species *O. piceae*, *O. floccosum* and *O. novo-ulmi*. Further studies are needed to establish if the ability of these fungi to colonise wood is influenced by the morphological switch and whether QSMs play a role in this process.

The presence of QS mechanisms was recently reported in another dimorphic fungus, the insect pathogen *Metarhizium rileyi* (Boucias *et al.* 2016). The study revealed that the yeast-like hyphal bodies-to-mycelia transition, which occurs in the insect host at the late infection stage, is regulated by QS mechanisms. A QSM is produced when the density of hyphal bodies in the hemolymph reaches a threshold, triggering the synchronous switch in the tissue-invasive mycelia. In this case, QSM is not identified yet, but lack of QS activity from of *M. rileyi* spent media suggests that the chemical signal mediating this response is produced by the host tissues in response to a high density of hyphal bodies (Boucias *et al.* 2016).

### **QS in filamentous fungi**

Filamentous fungi, developing mycelial growth, are not unicellular; hence the necessity of exporting a QSM to the environment in these organisms is controversial. However, QS mechanisms have been reported in this group, particularly in species from the genera *Aspergillus* and *Penicillium* (Raina *et al.* 2010; Guo *et al.* 2011; Sorrentino *et al.* 2010; Raina *et al.* 2012; Williams *et al.* 2012). In these studies, fungi seem to induce secondary metabolism as a competition strategy. *Penicillium sclerotiorum* produces sclerotiorin, a secondary metabolite with antibiotic properties. In this fungus,  $\gamma$ -butyrolactone-containing molecules such as multicolic acid function as QSMs. The exogenous addition of extracts containing  $\gamma$ -butyrolactones produced by *P. sclerotiorum* enhances sclerotiorin production in *P. sclerotiorum* cultures (Raina *et al.* 2010). Similarly, the effect of farnesol on fungal morphology and cellulase production in *Penicillium decumbens* has been investigated. In this case, the addition of a high concentration of exogenous farnesol (1 mM) promoted hyphal growth facilitating higher cellulase secretion (Guo *et al.* 2011). Further kinetic studies monitoring QSM production during the exponential growth phase are needed to determine the role of endogenous QS in the control of enzyme production and to corroborate the population density-dependent nature of this phenomenon.

In *Aspergillus* species, it has become apparent that QS mechanisms influence population-dependent behaviours, such as morphogenesis and secondary metabolite production. Oxylipins in *Aspergillus flavus* act as QSMs that regulate morphological differentiation and the production of either asexual spores or sclerotia. The specific structure of these oxylipins is still unknown (Horowitz Brown *et al.* 2008). Similarly, oxylipins regulate a QS-dependent pathway in *A. flavus* controlling development and mycotoxin production (Affeldt *et al.* 2012). However, in this case, the production of oxylipins has not been demonstrated as a population-dependent phenomenon. Furthermore, the addition of linoleic acid, an oxylipin precursor, to cultures of *O. novo-ulmi* affected fungal morphology, but this effect was not regulated by QS mechanisms (Naruzawa *et al.* 2016). In *Aspergillus terreus*, the addition of linoleic acid improved production of the secondary metabolite lovastatin (a cholesterol-lowering drug) in stirred tank fermenters (Sorrentino *et al.* 2010). Later, a butyrolactone was proposed as a QSM in *A. terreus* for its ability to affect both lovastatin and its own production. Exogenous addition of 100 nM butyrolactone I resulted in a 2.5-fold increase in lovastatin production, regulating the transcription of genes involved in the lovastatin biosynthesis. Exogenous addition of butyrolactone I also increased the levels of endogenous butyrolactone I (2.5-fold), suggesting an auto-stimulatory function (Raina *et al.* 2012). The strain AXTII9 of *A. nidulans* secretes a small diffusible molecule during its growth cycle, which accumulates at high population densities and alters growth profile and secondary metabolite production (Williams *et al.* 2012). The addition of ethyl acetate extracts from stationary phase culture supernatants to low cell-density cultures of *A. nidulans* abolished the lag phase, decreased the final cell dry weight and increased penicillin production. The bioactive molecule present in the stationary phase extract was purified and identified as  $\gamma$ -heptalactone (Williams *et al.* 2012).

The empirical evidence indicates the existence of QS in filamentous fungi, but the biological meaning is still unclear. Further studies are needed to elucidate whether these mechanisms are a form of communication among fungal microcolonies (de Bekker *et al.* 2011) and/or are involved in the formation of biofilms in these fungi (Harding *et al.* 2009).

## FAMILIES OF QS MOLECULES

The signalling molecules described so far (Table I) belong to four families of compounds: i) Lactones, which are cyclic esters of hydroxycarboxylic acids (Safari et al. 2014), including, N-acyl-homoserine-lactones from Gram-negative bacteria; and butyrolactone I (Raina *et al.* 2012),  $\gamma$ -heptalactone (Williams *et al.* 2012) and  $\gamma$ -butyrolactones (Raina *et al.* 2010), exclusively from filamentous fungi. Interestingly,  $\gamma$ -butyrolactones exist in the filamentous bacterium *Streptomyces* sp. (Biarnes-Carrera *et al.* 2015), suggesting convergent evolution or horizontal gene transfer; ii) Terpenes are derived biosynthetically from isoprene units, such as farnesol produced by the dimorphic fungi *C. albicans* (Hornby *et al.* 2001) and *O. piceae* (de Salas *et al.* 2015), or the cyclic sesquiterpenes from the dimorphic fungus *O. floccosum* (Berrocal *et al.* 2014); iii) Alcohols derived from amino acids, and are produced by yeast and dimorphic fungi. QS alcohols include tryptophol and phenylethyl alcohol in the baker's yeast *S. cerevisiae* (Hogan 2006b), tyrosol in *S. cerevisiae* and *C. albicans* (Chen *et al.* 2004; Avbelj *et al.* 2015), and 2-methyl-1-butanol in *O. novo-ulmi* (Berrocal *et al.* 2012); and iv) Peptides, produced by one basidiomycete, *C. neoformans* (May 2016), as well as by several Gram-positive bacteria (Hawver *et al.* 2016).

In this context, it could be hypothesised that fungi use a similar "language" to communicate with related species, and a "common language" to communicate to species from other phyla. However, this plausible scenario might prove to be an over-simplification; as new QSMs are discovered, a more detailed picture could emerge (Table I).

In general, as QSMs are produced in small concentrations (nM- $\mu$ M) researchers face an experimental problem when larger amounts are needed, e.g. for purification, identification and the discovery of new molecular structures. In this context, ways to increase their production (including molecular biology tools or scale up fermentations) will prove useful.

## INTERKINGDOM COMMUNICATION

While bacterial species can sense different families of QSMs, there is no evidence of inter-species communication among fungi. However, this cross-talk has been described between bacteria and fungi, and even bacteria and fungi with their plants or mammalian hosts (Barriuso *et al.* 2008; Pérez-Montaña *et al.* 2013; Camara *et al.* 2002; Hargarten *et al.* 2015). There are only a few reports dealing with interkingdom communication involving fungi, and most are focused on *C. albicans*. It is known that opaque cells of *C. albicans* are able to attract leukocytes to the site of infection by using the QSM farnesol (Langford *et al.* 2013; Hargarten *et al.* 2015). Moreover, farnesol can influence the behaviour of bacteria, inhibiting growth and virulence factors in some cases (Jabra-Rizk *et al.* 2006). It has also been reported that co-cultivation of *C. albicans* and the nonpathogenic filamentous fungus *A. nidulans* results in farnesol-dependent inhibition of *A. nidulans* growth (Semighini *et al.* 2006). *C. albicans* may use farnesol to reduce competition from other microbes. However, it is not clear to what extent this is a population density-mediated process. The hydrophobic nature of farnesol and the high concentrations used in these examples may favour its accumulation in the membrane and its leakage.

Interkingdom communication through QS systems has been reported in polymicrobial biofilms between the caries-promoting bacterium *Streptococcus mutans* and *C. albicans*, which are of medical importance (Sztajer *et al.* 2014). In this case, dual-species biofilms reached higher biomass and cell numbers than mono-species biofilms. Spent media from mixed biofilms but not from *C. albicans* or *S. mutans* alone activated alternative sigma factor SigX of *S. mutans*, which is regulated by the bacterium's QSMs and influences its virulence properties (Sztajer *et al.* 2014) (Fig. 2A). However, in this case, the signalling molecule involved has not been fully identified yet. In another example, the Gram-positive bacterium *Enterococcus faecalis* inhibits hyphal morphogenesis and virulence of *C. albicans* in the context of *in vitro* biofilm and in a *Caenorhabditis elegans* infection model (Cruz *et al.* 2013). Both microbes are commensals in the

same niche and frequently induce polymicrobial infections. The inhibition is partially dependent on the Fsr QS system of *E. faecalis*, specifically, two proteases regulated by Fsr, GelE and SerE, are partially required. Preliminary characterisation of the inhibitory signal has revealed that it is heat resistant with a molecular weight between 3 and 10 kDa; later, this factor has been identified as the EntV secreted bacteriocin (Graham *et al.* 2017) (Fig. 2B).

In addition, the association between *Candida* and other pathogens, such as the bacterium *Pseudomonas aeruginosa*, increases the complexity of infections. In this case, the 12-carbon backbone QSM 3-oxo-C12-HSL secreted by *P. aeruginosa* inhibits the adenylyl cyclase (Cyr1) from the yeast by binding directly to the catalytic site, and prevents *C. albicans* filamentation (Hogan *et al.* 2004; Hall *et al.* 2011). The fungal adenylyl cyclase (Cyr1) contains multiple domains acting as sensors for a diverse range of signals, including serum, peptidoglycan (PGN), and elevated CO<sub>2</sub> concentration (Klengel *et al.* 2005). QSMs also bind to the catalytic domain of Cyr1p at a site distal to that of the CO<sub>2</sub> binding site. Biochemical studies suggest that 3-oxo-C12-HSL is unique in its ability to mimic the actions of farnesol by binding to the farnesol inhibitory site in Cyr1p. Due to the importance of this signalling pathway in both fungal pathogenesis and interkingdom communication, the fungal soluble adenylyl cyclase has been proposed as a hub of signal sensing and integration (Hogan and Muhlschlegel 2011). On the other hand, farnesol is able to induce the production of the QSMs quinolone and C4-HSL in *P. aeruginosa* biofilms (Cugini *et al.* 2010) (Fig. 2C).

In addition to investigations on interkingdom cross-talk with *C. albicans*, there is an increasing number of publications describing the interactions of other fungi with bacteria. For example, *P. aeruginosa*, which causes biofilm lung infections in cystic fibrosis patients, is able to inhibit *Aspergillus fumigatus* biofilm formation by means of a secreted heat-stable soluble factor in a concentration-dependent manner (Ramage *et al.* 2011). Co-culture of *P. aeruginosa* mutants in the master QS system LasI/R did not significantly inhibit *A. fumigatus* biofilms formation and filamentation to the same extent as did wild type. Subsequently, the secreted factor was determined

to be the siderophore pyoverdine, whose production is regulated by the QS systems in *P. aeruginosa* (Sass *et al.* 2017) (Fig. 2D). The diffusible lipopeptide ralsolamycin, produced by the ubiquitous soil-borne plant pathogenic bacterium *Ralstonia solanacearum*, is a novel interkingdom interaction signal that induces chlamydospore development in a wide range of fungal taxa (Ascomycetes, Basidiomycetes and Zygomycetes) (Spraker *et al.* 2016). Interestingly, the production of ralsolamycin is controlled in a bacterial population density-dependent manner by the PhcBSR QS system (Li *et al.* 2017). It is thought that ralsolamycin contributes to the invasion of fungal hyphae, providing a specific niche for bacterial colonisation, but also diminishing survival of the fungus by preventing the production of protective agents against oxidative stress (Spraker *et al.* 2016; Khalid *et al.* 2018). Finally, it has been recently found that the ComX pheromone from *Bacillus licheniformis* (NCIMB-8874) inhibits growth of *Aspergillus flavus* strains (NRRL 3357 and ESP 15). This can lead to biotechnology-based strategies for the control of aflatoxin damage in agriculture, avoiding pre- and post-harvest aflatoxin contamination (Esmailshirazifard 2016; Esmailshirazifard *et al.* 2017).

These examples show how microorganisms use their QS systems to sense their population density as well as to directly interact with other microbes, or indirectly regulate the production of molecules that affect the survival of their neighbouring populations; and highlight the importance and timeliness of research to investigate how microorganisms communicate with each other and with their environment. Therefore, it is essential to examine other microbes in each fungal habitat and identify potential interactions among the microbial community (Johnston *et al.* 2016).

## **ECOPHYSIOLOGICAL ROLE AND PATHOGENESIS**

As it has been shown, QS mechanisms play a central role in the ability of fungi to colonise their habitat, form biofilms, and cause infections. In order to understand the clinical implications of QS and to exploit it as a therapeutic target, further studies of QS mechanisms in fungi are needed (Fig. 3).

*C. albicans* is a successful coloniser of the human host. Important virulence-associated traits of the fungus, such as the dimorphic switch and biofilm formation, are controlled by the quorum sensing molecule farnesol. *C. albicans* also simultaneously produces and tolerates high farnesol concentrations that are lethal to other microbes (Polke *et al.* 2017). The formation of biofilm structures in *C. albicans* has been documented as a form of resistance against antimycotic drugs (Nobile and Johnson, 2015). Since traditional antimicrobial therapies are often ineffective against *C. albicans* biofilms, different approaches have been tested to enhance the susceptibility of biofilms. In this context, the clinical relevance of farnesol and tyrosol was demonstrated by the finding that the exogenous addition of these molecules inhibits biofilm formation in *Candida* species, enhancing their susceptibility to antifungals (Cordeiro *et al.* 2015; Ramage *et al.* 2002).

The study of QS processes in fungi may uncover strategies to prevent colonisation of hosts by not only human and animal pathogens, but also plant pathogens (Fig. 3). For example, *O. ulmi* and *O. novo-ulmi*, causal agents of two successive Dutch elm disease pandemics, are transported to elm trees by an intermediate vector, the elm bark beetle (Santini and Faccoli 2015). Fungal spores accumulate in the xylem vessels of the plant and disperse by translocation. The ability of these organisms to undergo morphological changes is thought to be a crucial factor for their virulence. Since QS regulation in *O. novo-ulmi* dimorphism has been identified (Wedge *et al.* 2016), the pathogenesis of this fungus may be regulated by this mechanism, and thus may be interrupted by targeting this pathway. Other genera from the family *Ophiostomataceae*, such as *Grosmannia* and the anamorphic genus *Sporothrix*, contain saprophytic and pathogenic species of relevance that are dimorphic or polymorphic. QS mechanisms have not been described in these fungi but are worthy of investigation since the anamorphic genus *Sporothrix*, phylogenetically related to the teleomorphic genus *Ophiostoma*, includes several species that are pathogenic to humans or animals. In this sense, it has been described that spent medium from *Sporothrix schenckii* increased the growth of *C. neoformans* (Albuquerque *et al.* 2013). On the other hand, *Grosmannia clavigera* is able to infect pine trees (DiGuistini *et al.* 2011). The yeast is the infective form, while usually



grows as mycelia in the environment (López-Romero *et al.* 2011). Population-level signalling, such as QS, that modulates morphology is yet to be described in these species; and warrants future studies.

Little is known in regards to the role of QS in filamentous fungi, although there is an example of population density dependent phenomena in *Plectosphaerella cucumerina* regulating the infection strategy decision; however, the molecule regulating this process remains unknown (Pétriaccq *et al.* 2016). In the case of *A. flavus* the role of oxylipins as QS molecules has to do with the reproductive cycle that the fungus adopts to produce asexual spores or sclerotia (Horowitz Brown *et al.* 2008). In other fungi, e.g. *Aspergillus* and *Penicillium* species, QS mechanisms are used to induce secondary metabolisms as a competition strategy (Raina *et al.* 2010; Sorrentino *et al.* 2010; Guo *et al.* 2011; Raina *et al.* 2012; Williams *et al.* 2012).

## **BIOENGINEERING OF FUNGAL QS SYSTEMS**

The exploitation of QS mechanisms as biotechnological tools is a developing field in bacteriology (Choudhary and Schmidt-Dannert 2010); however there are few examples in mycology despite the fact that many fungi have biotechnological applications. For example, species such as *O. piceae* and *O. floccosum* produce the pigment melanin which is responsible for sapstain conifers. Colonisation of wood by these fungi results in substantial economic losses in the forest industry products, making biological control of these fungi essential regarding both environmental and economic considerations (Krokene and Solheim 1998). *Ophiostoma* albino strains have been developed and are used as biocontrol agents (Held *et al.* 2003), but the exploration of the QS systems of these fungi (Berrocal *et al.* 2014; de Salas *et al.* 2015) may represent an alternative approach to avoid colonisation and hence the blue-staining (Fig. 3). Moreover, this review has addressed fungal species able to produce important secondary metabolites including antibiotics, organic acids and vitamins, and species capable of producing enzymes for decomposition of organic matter. For example, *O. piceae* CECT 20416 is able to produce a versatile lipase/sterol esterase of

biotechnological relevance that acts on triglycerides, esters of *p*-nitrophenol, and cholesterol (Barriuso *et al.* 2016). The modification of the QS parameters in fermentations with this fungus resulted in a 2-fold increased secretion of the enzyme (de Salas *et al.* 2015). This strategy has also been explored in *P. sclerotiorum* and *A. terreus* for the production of metabolites (Raina *et al.* 2010; Williams *et al.* 2012) (Fig. 3).

Furthermore, some fungi take part in the production of foods; the most illustrative example is *S. cerevisiae*. It has been shown in this yeast that QS during fermentations may influence the production and quality of wine. In particular, ethanol stress during wine fermentation reduces the production of 2-phenylethanol, tryptophol, and tyrosol, which opens new challenges in the control of food fermentations (Avbelj *et al.* 2015; Zupan *et al.* 2013) (Fig. 3).

QS systems from different microorganisms have been artificially modified by synthetic and systems biology techniques to engineer the communication among different species. Using the principles of engineering and standardisation, systems biology is taking advantage of microbial QS to create new regulatory circuits with new features and diverse biotechnological applications, allowing complex behaviours, and the division of labour within a community (Purnick and Weiss 2009). In bacteria there are many examples of artificial QS circuits, biosensors based on QS mechanisms, and even bacteria that are able to release a drug in tumour tissues *in vivo* using QS mechanisms (Choudhary and Schmidt-Dannert 2010). In fungi, there are some examples of artificial QS circuits depicting artificial intra- or interspecific communications. In this respect, Khakhar *et al.* (2016) developed a modified *S. cerevisiae* strain with an orthologous QS system, based on the transcription factor CRISPR and the plant hormone auxin. This engineered cell-to-cell communication system was composed of a sender strain able to produce auxin and a receiver strain with a broad range of auxin sensitivities based on a library of auxin-degradable CRISPR transcription factors (Fig. 3).

Oscillatory bacterial communities have been created already using QS mechanisms and systems with a bi-stable behaviour to increase the robustness of the system (Chen *et al.* 2015), but

the construction of new biological systems taking advantage of the QS systems of bacteria and fungi has never been explored.

One of the current challenges in biocatalytic processes is to improve their efficiency by using microbial consortia to carry out complex biotransformations. This strategy exploits the biotechnological potentials of different microorganisms that are put to work together. The interactions between fungi and bacteria would be of particular interest because both organisms have different, and in many cases complementary, biotechnological abilities such as the production of extracellular degrading enzymes or high added-value compounds (Fig. 3). In this context, it is worthwhile to study the pool of bacteria that surround and effectively interact with fungi in nature; in other words, the microbiome (Johnston *et al.* 2016).

## CONCLUSIONS

In this paper we have presented a broad overview of a novel field of research, QS mechanisms in fungi. This review tackles the fundamental issue of how microorganisms relate to each other and their environment. Since this is a maturing area of research, there is much to be discovered. It is now clear that there is no universal communicating “language” among fungi, as QSMs are not generic across genera or species. Over the coming years, we will be able to elucidate whether fungal communication is common to taxonomic groups or is strain specific. In addition, an understanding of the molecular components of QS systems will be required to determine the extent of cross-talk among fungi, to decipher the evolutionary origin of the fungal “languages,” and to unravel the way fungi communicate with other organisms, such as bacteria, plants and animals. As our knowledge of the sophisticated microbial communication process increases and distils, we can determine if fungal QS is modulated in response to environmental conditions through extrinsic and intrinsic factors, and whether microevolution of QS within fungal populations occurs. This knowledge will enhance the potential for targeting fungal QS processes for biotechnological and clinical purposes.

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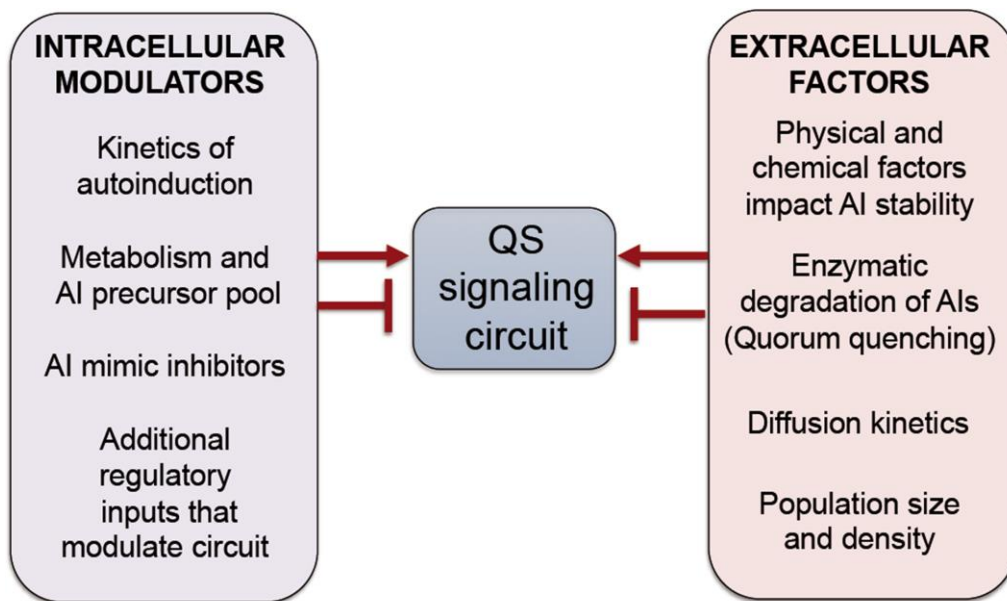
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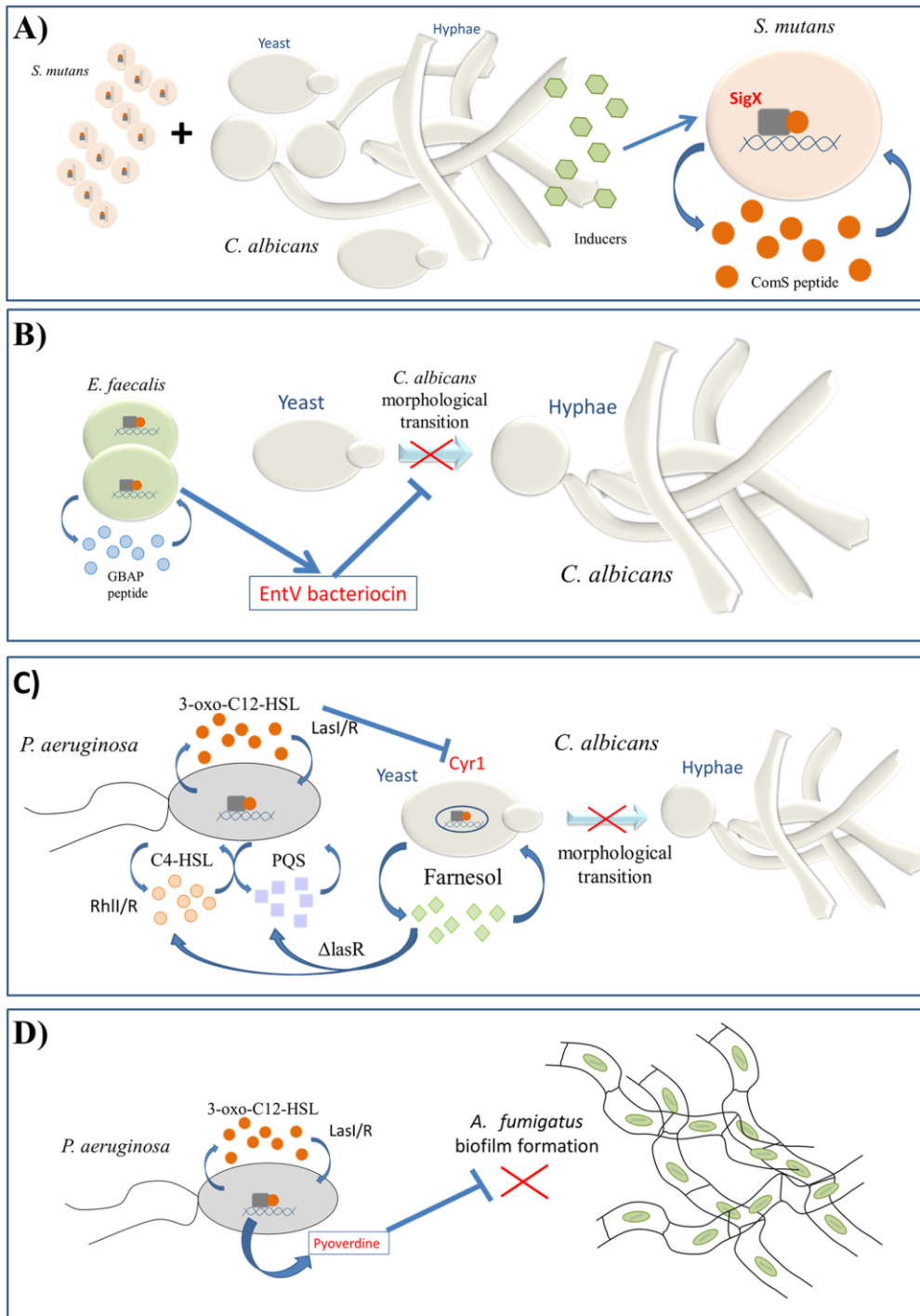
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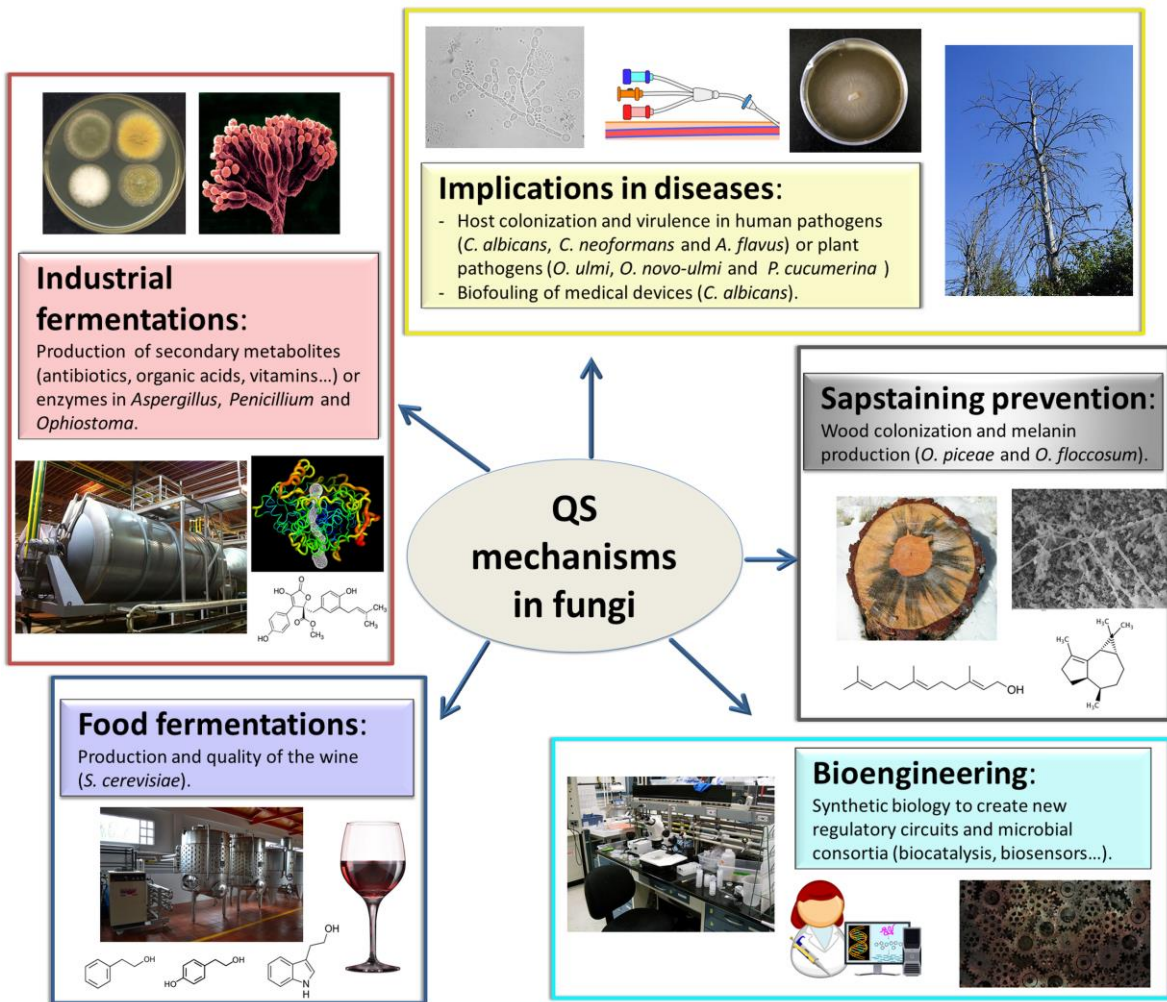
**Figure 1.** Summary of the intrinsic and extrinsic factors that modulate QS signaling in microbial cells. The intracellular modulators include the dynamics of signal detection and the activity of the positive feedback loop that characterizes signal production, and this can vary between species or strains within a species. Metabolic state, and the effects of metabolism of autoinducer (AI) production, can also impact AI production. AI mimics that inhibit AI detection have been described. In bacteria, other environment-responsive regulatory elements, such as transcription factors and small RNAs, can positively or negatively tune the QS circuit. The extracellular factors that modulate QS signaling include physical and chemical elements that impact the stability of secreted AI molecules and the action of enzymes that can degrade AIs. In addition, the population size and factors that influence the diffusion and availability of the AI will influence the QS circuit.



**Figure 2.** Communication between bacteria and fungi has co-evolved into polymicrobial biofilms. There are several examples of interkingdom cross-talk where bacteria and fungi exchange signal molecules depending on their population densities within a biofilm. A) Spent medium from dual-

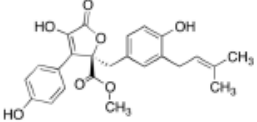
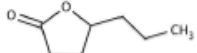

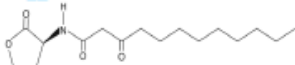
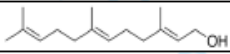

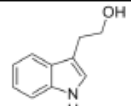
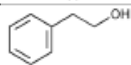
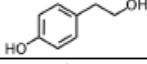
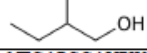
species biofilms between *S. mutans* and *C. albicans* (not from *C. albicans* or *S. mutans* alone) activates alternative sigma factor SigX of *S. mutans*, which in turn is regulated by the bacterium's QSM ComS, resulting in a biofilm with higher cell number and biomass. B) The bacterium *E. faecalis* inhibits hyphal morphogenesis of *C. albicans* by secreting the EntV bacteriocin in the context of *in vitro* biofilm. This inhibition is partly dependent on the Fsr QS system of *E. faecalis*. C) The QSM from *P. aeruginosa* 3-oxo-C12-HSL inhibits the adenylyl cyclase (Cyr1) from *C. albicans* and prevents the morphological transition from yeast to hyphae in this fungus. On the other hand, the QSM from *C. albicans* farnesol is able to induce the production of the QSMs C4-HSL and quinolones (PQS) in biofilms of *P. aeruginosa*  $\Delta$ lasR mutants. D) *P. aeruginosa* inhibits *A. fumigatus* biofilm formation by producing the siderophore pyoverdine, whose production is regulated by the QS systems in *P. aeruginosa*.





**Figure 3.** Potential biotechnological applications of the fungal QS mechanisms.

**Table 1.** QS molecules identified or active in fungi.

Family	Name	Structure	Producer species	References
Lactones	Butyrolactone I		<i>A. terreus</i>	Raina <i>et al.</i> 2012
	$\gamma$ -heptalactone		<i>A. nidulans</i>	Williams <i>et al.</i> 2012
	$\gamma$ -butyrolactone		<i>P. sclerotiorum</i> and Gram-positive bacterium <i>Streptomyces</i> sp.	Raina <i>et al.</i> 2010; Biarnes-Carrera <i>et al.</i> 2015
	3-oxo-C12-homoserine-lactone		Gram-negative bacterium <i>P. aeruginosa</i>	Hall <i>et al.</i> 2011
Terpenes	Farnesol		<i>C. albicans</i> , other <i>Candida</i> spp. and <i>O. piceae</i>	Hornby <i>et al.</i> 2001; de Salas <i>et al.</i> 2015
	Cyclic sesquiterpenes		<i>O. floccosum</i>	Berrocal <i>et al.</i> 2014
Alcohols	Tryptophol		<i>S. cerevisiae</i>	Hogan 2006
	Phenylethyl alcohol		<i>S. cerevisiae</i>	Chen and Fink 2006
	Tyrosol		<i>S. cerevisiae</i> and <i>C. albicans</i>	Chen <i>et al.</i> 2004; Avbelj <i>et al.</i> 2015
	2-methyl-1-butanol		<i>O. ulmi</i>	Berrocal <i>et al.</i> 2012
Peptides	QSP1	NFGAPGGAYPW	<i>Cryptococcus neoformans</i>	May 2016
	ComX	ADPITRQW(Farnesyl)GD	Gram-positive bacterium <i>Bacillus subtilis</i> 168	Hawver <i>et al.</i> 2016

Interestingly, the signaling using peptides among fungi have only been described in a basidiomycete, lactones have only been identified in filamentous fungi, while alcohols have only been described in yeast. However, dimorphic fungi such as *Ophiostoma*, and *Candida* have been shown to use terpenes and alcohols.

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