1 Overview of Fission Yeast Septation

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1 ABSTRACT

Cytokinesis is the final process of the vegetative cycle, which divides a cell into two independent daughter cells once mitosis is completed. In fungi, as in animal cells, cytokinesis requires the formation of a cleavage furrow originated by constriction of an actomyosin ring which is connected to the plasma membrane and causes its invagination. Additionally, since fungal cells have a polysaccharide cell wall outside the plasma membrane, cytokinesis requires the formation of a septum coincident with the membrane ingression. Fission yeast *Schizosaccharomyces pombe* is a unicellular, rod-shaped fungus that has become a popular model organism for the study of actomyosin ring formation and constriction during cell division. Here we review the current knowledge of the septation and separation processes in this fungus, as well as recent advances in understanding the functional interaction between the transmembrane enzymes that build the septum and the actomyosin ring proteins.

1 INTRODUCTION

Cytokinesis is the final stage of the eukaryotic cell cycle during which, after mitotic 2 exit, the formation of a cleavage furrow separates the cell giving rise to two new cells. 3 Cell division in fungal and animal cells is well conserved. Cleavage furrow formation 4 always requires the establishment and closure of a cytokinetic actomyosin ring (AR). A 5 6 major difference between fungal and animal cells is that fungi are surrounded by a rigid 7 cell wall; therefore, in fungal cells AR contraction occurs simultaneously with the biosynthesis of a cell wall structure known as a septum (Willet et al., 2015b; Rincón 8 9 and Paoletti, 2016) (Figure 1A). In unicellular fungi such as yeasts, at the end of cytokinesis there is a controlled septum degradation that separates the two daughter 10 11 cells. The fission yeast Schizosaccharomyces pombe has been used as a model organism 12 to study the eukaryotic cytokinesis because of the high degree of conservation among AR components throughout evolution. S. pombe is a simple, genetically tractable 13 14 organism with highly regular rod-shaped, stable growth patterns (Mitchison et al., 1985). Additionally, S. pombe divides symmetrically, giving rise to two daughter cells of 15 the same size (Mitchison, 1957). In contrast, Saccharomyces cerevisiae and other yeasts 16 grow asymmetrically forming a bud that will give rise to a daughter cell. They also 17 18 divide asymmetrically, with the mother cell larger than the daughter cell. Therefore, the positioning of the division plane in the geometrical center of S. pombe cells is different 19 20 from S. cerevisiae and similar to the majority of animal cells (Balasubramanian et al., 21 2004). Several recent reviews discuss the regulatory mechanisms that control division 22 plane positioning in S. pombe, which proceeds through the assembly of cytokinetic 23 precursors on the medial cortex into nodes that coalesce into AR (Pollard and Wu, 2010; 24 Lee et al., 2012; Willet et al., 2015b; Rincón and Paoletti, 2016). Here we review the 25 current knowledge of septation in the fission yeast, emphasizing the importance of

- 1 correct septum formation for cell integrity and survival especially during cell separation.
- 2 Additionally, we discuss recent advances on the cooperation between the AR and
- 3 septum during the cleavage furrow ingression.

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Cell wall and septum composition in fission yeast

All fungi contain a polysaccharide cell wall which includes glucans and mannoproteins 6 as major components. Two types of glucans are the major structural components of 7 8 fission yeast cell wall: branched $\beta(1,3)$ -D-glucan with 14% of $\beta(1,6)$ branches constitutes 48-54% of total cell wall polysaccharides, and $\alpha(1,3)$ -D-glucan which 9 10 constitutes 28-32% (reviewed in Durán et al., 2004). Additionally, there is a small amount of linear $\beta(1,3)$ -D-glucan, mainly present in the primary septum and cell tips 11 (Cortés et al., 2007). This glucan might play a role similar to chitin, present in most 12 13 fungi but not found in S. pombe (Kreger, 1954; Horisberger et al., 1978). There is also a 14 small proportion of $\beta(1,6)$ -D-glucan that might be important for cross-linking different polysaccharides (Magnelli et al., 2005), and of galactomannan linked to the cell wall 15 glycoproteins (Ballou et al., 1994). 16 Analysis by transmission electron microscopy (TEM) has found the septum to be a 17 three-layered structure with a central primary septum (PS) flanked by two layers of 18 19 secondary septum (SS) (Johnson et al., 1973) (Figure 1B). The SS deposition is simultaneous to the PS growth (Cortés et al., 2007). The PS is a special layer of the cell 20

wall that in fission yeast is rich in linear $\beta(1,3)$ -D-glucan. In S. cerevisiae and other

fungi the PS is mainly made of chitin (Cabib et al., 2005); the SS contains the same

polymers of the cell wall (Humbel et al., 2001).

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Fungal wall $\beta(1,3)$ -D-glucan is synthesized by the enzymatic complex $\beta(1,3)$ -D-glucan 1 2 synthase (EC 2.4.1.34, UDP-glucose:1,3-β-D-glucan 3-β-D-glucosyltransferase). This complex, conserved in all fungi, includes at least two proteins: a catalytic subunit that is 3 a large protein with several transmembrane domains, and the Rho1 GTPase acting as 4 regulatory subunit that activates the catalytic subunit when bound to GTP (Arellano et 5 al., 1996; Drgonova et al., 1996). Different paralog genes coding for the catalytic 6 7 subunit, named Fks or Bgs, are present in fungal cells (Free, 2013). In budding yeast 8 and other fungi, these subunits have partially redundant roles (Mazur et al., 1995) while each of the four subunits present in fission yeast is essential. Bgs1, 3, and 4 function 9 10 during vegetative growth and Bgs2 functions during sporulation. Bgs1,3 and 4 localize to growing poles, division area, and sites of wall synthesis during sexual differentiation 11 12 (Roncero et al., 2010). Ultrastructural analysis of the septa formed in $bgs1\Delta$ 13 germinating spores established that Bgs1 is responsible for the linear $\beta(1,3)$ -D-glucan synthesis and PS formation (Cortés et al., 2007) (Figure 1B). The function of Bgs3 is 14 15 not yet known. Bgs4 is responsible for the synthesis of the major cell wall β-glucan, and is essential for the maintenance of cell integrity especially during cell separation, SS 16 formation, and for correct PS completion (Cortés et al., 2005; Muñoz et al., 2013) 17 18 (Figure 1B). The only enzyme identified as a putative α -glucan synthase is Ags1, also named Mok1. 19 Like the Bgs enzymes, Ags1 is a membrane protein essential for cell integrity and is 20 21 detected at the growing poles and the septum (Katayama et al., 1999). Ags1 is required

for SS formation (Figure 1B) and for gradual and balanced cell separation (Cortés et al.

2012). Ags1 orthologs are not found in budding yeasts but are widely extended in other

fungi although they are not always essential (Edwards et al., 2011; Henry et al., 2012).

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Establishment of the septum position

The position of the septum depends on the AR formed at the middle of the cell cortex. 2 3 In fission yeast, the nucleus and the anillin Mid1 mark the position of AR assembly which is initiated by the maturation of medial cortical nodes (reviewed in Willet et al., 4 5 2015b; Rincón and Paoletti, 2016). Once the ring is formed, it is necessary to keep it in 6 position until constriction starts (Wu et al., 2003; Arasada and Pollard 2014; McDonald 7 et al., 2016). In spherical protoplasts deprived of the wall, the AR slides to the poles (Mishra et al., 2012) suggesting that the AR needs to be anchored through the 8 9 membrane to the extracellular cell wall, and that the cylindrical shape of fission yeasts might also play a role in the AR stability (Mishra et al., 2012). However, it has been 10 shown in cylindrical cells that the β-glucan synthesized by Bgs4 plays a main role in 11 12 maintaining the AR in the cell middle before septum formation begins (Muñoz et al., 13 2013). Bgs1 has also been implicated in the maintaining of AR position. Accordingly, the AR of cells carrying cps1-191, a temperature-sensitive allele of $bgs1^+$, slides along 14 15 the plasma membrane (Arasada et al., 2014).

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Coupling AR contraction with septum synthesis.

The septation initiation network (SIN) is a kinase cascade that activates Sid2, a kinase from the NDR (Nuclear Dbf-2-related) family. Sid2 is essential in the regulation of cytokinesis (reviewed in Simanis, 2015). The SIN is orthologous to the mitotic exit network (MEN) in budding yeast, and the Hippo pathway in animal cells (Hergovich *et al.*, 2006). The SIN cooperates in the regulation of mitotic entry, spindle elongation and checkpoint inactivation, telophase nuclear positioning, assembly of AR, and, importantly, it is essential for the AR contraction and concomitant synthesis of the

septum (reviewed in Simanis, 2015). Additionally the SIN inhibits a second NDR 1 2 kinase pathway called the morphogenesis Orb6 (MOR) network (Ray et al., 2010), which is orthologous to the network called regulation of Ace2 and morphogenesis 3 4 (RAM) in S. cerevisiae and other fungi (Saputo et al., 2012). The MOR network is required for cell separation and apical growth (Gupta et al., 2014). Most SIN 5 6 components are essential, and temperature-sensitive SIN mutants form defective AR but 7 do not initiate septum synthesis, leading to the formation of elongated multinucleated cells at the restrictive temperature (reviewed in Krapp et al., 2008; Simanis, 2015). 8 9 Because Rho1 GTPase can rescue some SIN mutants, it has been proposed that SIN 10 activates Rho1, which in turn activates the Bgs enzymes, (Jin et al., 2006), but it has not 11 been proved. Additionally, it has been proposed that there is a feedback loop were Rho1 12 activates the SIN to ensure SIN activity while septation is progressing (Alcaide-Gavilan 13 et al., 2014). 14 In animal cells AR constriction is dependent on myosin type II and exerts the force needed to pull the plasma membrane and form the cleavage furrow. In fungi AR 15 constriction is also myosin II dependent (Mishra et al., 2013) and is required for the 16 initiation of septum formation, but does not provide the pulling force for the furrow 17 ingression (Proctor et al., 2012). Since AR constriction and septum synthesis occur 18 simultaneously (**Figure 1A**), it has been proposed that linear $\beta(1,3)$ -D-glucan synthesis 19 20 provides the major force for the furrow closure (Proctor et al., 2012). On the other hand, 21 the AR is dispensable when the septum is already forming but ingression is much 22 slower in its absence (Proctor et al., 2012). It seems therefore that ring constriction 23 activates septum synthesis. Supporting this hypothesis, two recent works propose that 24 septum synthesis is mechanosensitive and somehow coupled to contractile AR tension 25 (Thiyagarajan et al., 2015; Zhou et al., 2015). By manipulating the curvature of the

- 1 cleavage furrow it was shown that the AR promotes local septum growth in a curvature-
- dependent manner suggesting that Bgs1 is regulated by AR tension (Zhou et al., 2015).
- 3 Conversely, as mentioned above, when Bgs1 is defective the AR slides (Arasada et al.,
- 4 2014), is disorganized (Cortés et al., 2015), and does not contract (Liu et al., 1999).
- 5 Whether and how the AR contractile force stimulates the cell wall machinery, and how
- 6 the cell wall maintains the AR and stimulates its contraction are currently major
- 7 questions in fungal septation.
- 8 Focal adhesions in animal cells connect the extracellular matrix to the cytoskeleton and 9 transmits signals in both directions. Similarly, during fission yeast cytokinesis several proteins might form a complex to connect the cell wall with the AR through the plasma 10 11 membrane. Major candidates to organize this complex are the F-BAR proteins (Roberts-12 Galbraith and Gould, 2010), which can bridge the plasma membrane and the cytoskeleton. Cdc15 is the founding member of the PCH family (Carnahan and Gould, 13 14 2003; Wu et al, 2003) and contributes to AR formation via the direct binding of its F-BAR domain to the formin Cdc12 that nucleates the AR actin filaments (Willet et al., 15 2015a). Additionally, Cdc15 contains an SH3 domain that binds proteins required for 16 17 septation such as Px11, Fic1, or Rgf3 (Roberts-Galbraith et al., 2009; Ren et al., 2015). A second F-BAR protein named Imp2 (Demeter et al., 1998) also has an SH3 domain 18 functionally interchangeable with that of Cdc15 and both collaborate in the recruitment 19 20 of the mentioned proteins (Roberts-Galbraith et al., 2009; Ren et al., 2015). It has been 21 recently described that Cdc15 oligomerization is critical for fission yeast cytokinesis 22 (McDonald et al., 2015). Pxl1 is the fission yeast ortholog of animal cells paxillin, it binds to myosin II and stabilizes the AR (Ge et al., 2008; Pinar et al., 2008). Pxl1 23 24 collaborates with Bgs1 to maintain the AR and form the septum (Cortés et al., 2015). 25 Bgs1 depletion generates abnormal septa made of SS layers, which somehow are guided

by the AR (Figure 1B). When Bgs1 depletion occurs in the absence of Px11 there is no 1 2 septum synthesis initiation (Figure 1B). It is possible that Px11, by binding to myosin II and Cdc15 (Cortés et al., 2015; Ren et al., 2015), transmits the AR tension to the 3 membrane in order to concentrate the Ags1 and Bgs4 synthases and narrow the area of 4 septum synthesis (Figure 2). Fic1 is a C2-domain containing protein whose role is not 5 6 defined in S. pombe but which becomes essential in the absence of the Pxl1 (Roberts-7 Galbraith et al., 2009, Ren et al., 2015). Rgf3 is an essential GEF that activates Rho1 (Arellano et al., 1996; Tajadura et al., 2004; Morrell-Falvey et al., 2005; Mutoh et al., 8 2005). In turn, this GTPase directly activates the Bgs enzymes and glucan synthesis 9 10 (Arellano et al., 1996; Tajadura et al., 2004; Morrell-Falvey et al., 2005; Mutoh et al., 2005). Whether or how Rgf3 and Pxl1 functionally interact to transform the AR 11 12 contraction into an activation signal for the biosynthetic enzymes that form the septum 13 remains to be discovered. 14 Another F-BAR protein, Rga7, forms a complex with Cdc15 and Imp2 (Martín-Garcia et al., 2014). Rga7 also contains a Rho GTPase activating protein (GAP) domain and 15 acts as negative regulator of Rho2 (Martin-Garcia et al., 2014). These complex 16 17 networks of proteins might contribute to the coordination of contractile ring constriction and septum formation (Figure 2). 18 19 F-BAR proteins may also have a role in the traffic of Bgs enzymes. Two recent works 20 propose that Cdc15 participates in the transport of Bgs1 from late Golgi to the 21 membrane at the division area, and similarly, Rga7 contributes to the transfer of the Bgs4 (Arasada et al., 2014, Arasada et al., 2015). The role of exocytosis, endocytosis, 22 23 and membrane traffic during the formation of the septum is only beginning to be

uncovered.

Cell separation and cell integrity

Separation is the most critical process of the cell cycle for the cell integrity. In animal 3 4 cells, the terminal step of cytokinesis is called abscission and includes microtubule 5 severing and membrane splitting that is mediated by the endosomal sorting complex required for transport (ESCRT) proteins (Bhutta et al., 2014). In filamentous fungi cell 6 separation does not occur and they form hyphae composed of cell compartments 7 8 delimited by septa with a small central pore. These septa are important for maintaining 9 hyphal integrity. Thus, they are sealed immediately upon injury impeding an extensive loss of cytoplasm (Mourino-Perez et al., 2013). 10 Cell separation does not seem to be essential for the vegetative cycle of the fission 11 yeast. A broad range of viable mutants defective in cell separation have been described 12 13 in the past (reviewed in Sipiczki, 2007), and recently a systematic visual screening of 14 the deletion collection of S. pombe haploids identified new genes based on the "long 15 branched" phenotype of viable cells (Hayles et al., 2013). Not all of these genes participate directly in the process of cell separation but play a role in earlier steps of 16 cytokinesis. Thus, most cells lacking Pxl1, or temperature-sensitive SIN mutants grown 17 18 at semi-restrictive temperature are septated even if they do not have a separation defect. 19 It is possible that the separation machinery, which at least includes the septins, the exocyst, Rho GTPases, and glucanases (reviewed in Martín-García and Santos 2016), is 20 21 set at the beginning of septation, and if the AR constriction/septation is delayed, 22 separation can no longer take place. The Sep1-Ace2 transcription-factor cascade 23 regulates the periodic expression of many genes encoding proteins required for AR 24 constriction and for cell separation (Rustici et al., 2004; Alonso-Nuñez et al., 2005).

- 1 Additionally, a posttranslational control on cell separation might be exerted during
- 2 cytokinesis by the SIN, which inhibits the MOR pathway (Gupta et al., 2014). This
- 3 pathway regulates septum degradation although the mechanism is not yet known (Gupta
- 4 *et al.*, 2014).
- 5 To prevent cell lysis during separation, a precisely controlled degradation of the lateral
- 6 cell wall at the division area and the PS is required. The remaining SS gradually curves
- 7 concavely due to the internal cell pressure, and forms the new end in the daughter cells.
- 8 (Cortés et al., 2012; Atilgan et al., 2015). Bgs4 depletion induces unopposed cell wall
- 9 degradation leaving the plasma membrane without cell wall and consequently, the
- internal turgor pressure causes the cell lysis (Cortés et al., 2005; Muñoz et al., 2013). In
- 11 Ags1-depleted cells lysis also occurs during separation suggesting that a correct SS
- assembly is essential for cell viability (Cortés *et al.*, 2012).
- Degradation of the lateral cell wall requires the Agn1 1,3- α -glucanase (Dekker *et al.*,
- 2004; Garcia et al., 2005), and Eng1 1,3-β-glucanase is necessary to digest the PS
- 15 (Alonso-Nuñez et al., 2005). Precise secretion of these enzymes involves the formation
- of septin rings and the directed activity of the exocyst complex (Martin-Cuadrado et al.,
- 17 2005). Rho4 GTPase is also required for glucanase secretion (Santos *et al.*, 2005), likely
- through exocyst regulation (Perez et al., 2015). This GTPase is activated by Gef3,
- which interacts with and is localized by the septins (Muñoz et al., 2014; Wang et al.,
- 20 2015). In this way, a precise spatio-temporal regulation of glucanase secretion preserves
- 21 cell integrity.

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Concluding remarks

Although this review focuses on fission yeast, similar mechanisms of septation exist in 1 2 other fungi. There are still a number of open questions on the septation process that need to be further addressed: the targets of the SIN that activate septation; the different 3 4 functions of F-BAR proteins during septum formation; the role of Rho GTPases and other molecules that regulate cell wall synthesis; etc. In all these questions the 5 6 connection between the cell wall and the AR through the plasma membrane is emerging 7 as an important condition for a successful cytokinesis and for the maintenance of cell integrity. Although it is still unknown how this connection is accomplished, some of the 8 main players such as Bgs enzymes, F-BAR proteins, Pxl1, and other cytoskeleton 9 10 binding proteins have been already identified. The characterization of new double conditional mutants, proteomics, and high-resolution microscopy techniques will help to 11 12 further characterize this connection.

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FIGURE LEGENDS

- 21 Figure 1. A) Time-lapse fluorescence micrographs showing simultaneously AR
- constriction and septum synthesis by using cells expressing GFP-cdc15 to label the AR,
- 23 RFP-Bgs1 to label the membrane and calcofluor (CW) staining to label the cell wall

- 1 glucan. Bar 5 μm. B) The ultrastructure of the *Schizosaccharomyces pombe* septum.
- 2 Transmission electron micrographs of wild type septum showing the middle layer of PS
- flanked by the SS; septum of cells with repressed $bgs4^+$ showing no SS and floppy PS
- 4 (adapted from Muñoz et al., 2013); septum of cells with repressed ags1⁺ showing
- 5 defective SS and floppy PS (adapted from Cortés et al., 2012); with repressed bgs1⁺
- 6 showing parallel SS depositions in wild type cells and absence of septum in cells
- 7 lacking Pxl1 (adapted from Cortés *et al.*, 2015). Bar 1 μm.
- 8 Figure 2. Model of the protein complexes connecting AR, septum membrane, and
- 9 septum wall.
- 10 Figure 3. A) Model of cell separation at the end of cytokinesis. B, C) Transmission
- electron micrograph showing how degradation of the lateral cell wall (B) and PS (C)
- leaves the SS that, upon separation, changes from flat to round shape (adapted from
- 13 Cortés et al., 2012).

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