Is the anthelmintic effect of heather supplementation to grazing goats always accompanied by antinutritional effects?

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To test the hypothesis that the beneficial anthelmintic effect of consuming moderate amounts of tannins may not always be accompanied by anti-nutritional effects in goats, two experiments were conducted. In the first, 48 Cashmere goats were randomly assigned to two treatments: supplementation with tannin-containing heather (6.4% total tannins) or non-supplementation. All goats grazed continuously from May to September under farm conditions in a mountainous area of northern Spain. The mean percentage of heather incorporated into the diet of the supplemented animals was 29.1%. Supplementation reduced the mean number of nematode eggs in faeces (P < 0.001) and the goat mortality rate (P < 0.05). The rumen ammonia concentration was markedly reduced in the goats receiving the heather supplement (160 vs. 209 mg/l; P < 0.01), while volatile fatty acid (VFA) concentrations were significantly greater (63.0 vs. 53.6 mmol total VFA/l; P < 0.05). The heather-supplemented goats also showed a lower loss of live weight (P < 0.01) and body condition score (P < 0.001). In the second experiment, batch cultures of rumen microorganisms with rumen fluid from nine goats whose diet included 29% heather – or not, were used to incubate three substrates (pasture, pasture + heather and pasture + heather + polyethylene glycol) to investigate in vitro ruminal fermentation. Differences (P < 0.01) among substrates were observed in terms of dry matter disappearance (DMD), true substrate digestibility (ivTSD), gas production and ammonia concentration, the greatest values always associated with the pasture substrate. Cultures involving rumen inoculum derived from goats receiving the heather-containing diet showed slightly lower DMD (46.9 vs. 48.5 g/100 g; P < 0.05), ivTSD (64.6 vs. 65.9 g/100 g; P < 0.10) and gas production (105 vs 118 ml/g; P < 0.001) values, but much greater total VFA concentrations (48.5 vs. 39.3 mmol/l; P<0.05), and suggest the efficiency of ruminal fermentation in these animals was probably improved. Together, the results support the absence of a clear nutritional cost counteracting the
beneficial anthelmintic effect of supplementing the diet of grazing goats with tannin-containing heather.

**Keywords:** gastrointestinal nematode, rumen fermentation, tannin, body weight

**Introduction**

A growing number of studies relates the consumption of tannin-rich plants by small ruminants with the regulation of their gastrointestinal (GI) nematode populations. This could reduce dependence on conventional chemotherapy, to which many parasite species have developed resistance, and facilitate the sustainable control of gastrointestinal nematode parasitism (Githiori *et al.*, 2006; Hoste *et al.*, 2006). Parasite-infected goats that consume moderate quantities of tannin-containing plants show improved resistance and resilience to parasites, and significant decreases in faecal egg counts (FEC) and parasite burden (Min *et al.*, 2005; Paolini *et al.*, 2005; Osoro *et al.*, 2007a and 2007b). Goats can, however, also experience the anti-nutritional effects of tannins (e.g., Silanikove *et al.*, 1997; Ben Salem *et al.*, 2003).

It is widely accepted that the consumption of tannins by parasitized animals may result in either favourable or detrimental net effects depending on whether or not the positive anthelmintic action of these plant secondary metabolites outweighs their negative nutritional cost to the host (Houdijk and Athanasiadou, 2003). Nevertheless, some tannins have also been reported as capable of producing a positive nutritional effect when consumed in moderate concentrations, which is associated with the reduction of dietary protein degradation in the rumen (Barry and McNabb, 1999; Mueller-Harvey, 2006).

Earlier studies (Osoro *et al.*, 2007a and 2007b) showed that grazing goats on a diet supplemented with tannin-containing heather had a lower FEC and a better performance, in comparison with animals on non-supplemented diets. On the basis of those studies, we
hypothesized that the beneficial anthelmintic effect of the consumption of moderate amounts of tannins might not always be accompanied by marked anti-nutritional effects in goats. Two experiments were conducted to test this hypothesis. The first was performed under farm conditions in a less-favoured mountainous region of Spain where grazing animals are raised extensively and form part of the dynamics of the natural ecosystem. The second experiment involved an in vitro assay, with batch cultures of rumen microorganisms, to further investigate the nutritional effects of the inclusion of heather in the diet observed in the first.

**Materials and methods**

Experiments 1 and 2 were performed in accordance with Spanish Royal Decree 1201/2005 for the protection of animals used for experimental and other scientific purposes.

**Experiment 1 (in vivo)**

**Experimental site**

This experiment was conducted in a mountainous area (altitude 1,000 m) of northwestern Spain (6º 53’ W, 43º 21´ N; Sierra de San Isidro, Illano, Asturias) dominated by shrubby heather-gorse vegetation. Improved pastures were established in 2001 by removing the shrubs present and sowing perennial ryegrass (*Lolium perenne* L.) and white clover (*Trifolium repens* L.). A plot of approximately 20,000 m² previously managed homogeneously was divided into two identical plots of approx. 10,000 m² to accommodate all the animals used in this experiment.

During the experimental grazing season (May-September), the mean monthly rainfall ranged from 48 mm in August to 133 mm in September. The maximum and minimum temperatures were recorded in September (31°C) and May (2°C) respectively; the mean average monthly values ranging between 11.8°C in May and 17.8°C in July.
Animals and experimental design

Forty-eight lactating Cashmere goats (weighing 36.8 ± 1.13 kg at the beginning of the trial), balanced for live body weight (BW) and body condition score (BCS), were randomly assigned, together with their single kids, to one of two treatments: supplementation with heather (+H) and non-supplementation (−H). Each group was confined to one of the two plots of pasture (containing no heather, as explained above) of approximately 10,000 m² for the whole grazing season. The +H goats were offered, in the morning and once every 3 days, freshly cut heather ad libitum; the −H goats received no supplement. All animals were reared outdoors under farm conditions (i.e., not preventing nematode infection).

Two weeks before the experiment started, all animals were orally treated against gastrointestinal nematodes with ivermectin (Oramec, Merial, Lyon, France) at about twice the recommended dose (2 mg/kg of BW). The animals were turned out on the pastures on April 19th and left grazing until September 10th.

Sampling procedures and analyses

Pasture. To monitor grass availability, the sward surface height was measured every 10-15 days using an HFRO swardstick (Barthram, 1986), taking 200 measurements at random in each plot. The botanical composition of the pasture was assessed in August using a point quadrat (Grant, 1981) and recording 200 vertical hits per plot. Pasture samples collected in August were analyzed for dry matter (DM; ISO 6496:1999), organic matter (OM; ISO 5984:2002) and crude protein (CP; ISO 5983-2:2005). Neutral and acid detergent fibre (NDF and ADF) and acid detergent lignin (ADL) were determined by the method of Goering and Van Soest (1970) and Van Soest et al. (1991).
Heather. The species composition of the heather offered to the goats was assessed in August by recording 1,000 random contacts with the HFRO swardstick over the area where the heather was cut. Green shoots of this heather (less than 3 mm in diameter) were collected for chemical analysis. The nutritive quality (DM, OM, CP, NDF, ADF, and ADL) was analyzed following the same procedures as used to analyze the pasture. The total phenol (TP) and total tannin (TT) contents of the heather offered to the goats were determined using the Folin-Ciocalteu assay in combination with polyvinyl-polypyrrolidone, using tannic acid (Merck, Damstadt, Germany) as the reference standard (Makkar, 2003).

Diet selection. In August, half of the +H animals were randomly chosen and subjected to estimations of the percentage of heather incorporated into the diet using n-alkane markers (Ferreira et al., 2005). Samples of pasture, heather, and faeces were collected and calculations performed using a least squares optimization procedure, that minimizes the discrepancies between the actual concentration of alkanes in faeces (adjusted for incomplete faecal recoveries using the recovery values obtained by Ferreira et al., 2005) and the estimated proportion in the diet.

Parasitological procedures. Approximately once per month between May and September, spot samples of goat faeces were individually collected by rectal grab to assess gastrointestinal nematode egg excretion. The number of eggs per gram of faeces (faecal egg count; FEC) were estimated using the modified McMaster technique (MAFF, 1978) with sodium chloride as the flotation medium, in which every egg is regarded as equivalent to 15 eggs per gram (epg) fresh faeces. Faecal cultures were performed for each group by pooling samples from the different individuals as described by the MAFF (1978) and the genus of third-stage nematode larvae identified according to Van Wyk et al. (2004).
**Body weight and body condition score.** All animals were weighed and their BCS assessed on a scale of 1 to 5 (1 = emaciated, 5 = fat; Russel, 1990), at the beginning and end of the experiment and at monthly intervals.

**Ruminal fermentation variables.** In August, after an overnight fast, a sample of ruminal fluid was individually collected from each goat via a stomach tube and visually checked to ensure that it did not contain saliva. Immediately afterwards each fluid was strained through two layers of gauze and 4 ml were acidified with 4 ml 0.2 N HCl for ammonia determination. A further aliquot of 0.8 ml was added to 0.5 ml of a deproteinizing solution (2% metaphosphoric and 0.4% crotonic acids (wt/vol) in 0.5 N HCl) for determination of volatile fatty acid (VFA). All samples (two per animal and parameter to be analyzed) were stored at −30°C until analysis. The ammonia concentration was determined by colorimetry and VFA by gas chromatography, using crotonic acid as the internal standard, both in centrifuged samples (Frutos et al., 2004).

**Mortality rate.** Goats were daily checked for clinical signs of illness. When an animal was considered terminally ill, it was humanely euthanized and complete post-mortem examination was performed to discard mortality for other causes.

**Statistical analysis**

Sward height, FEC, BW and BCS data were analyzed by repeated measures analysis using the MIXED procedure of the Statistical Analysis System program (SAS, 1999). The first measurement in May was used as the covariate. For FEC, BW and BCS, the animals were nested within the treatment to provide the error term to contrast the effect of heather...
supplementation. Data on FEC were log transformed \((\log_{10} x + 1)\) to normalize their distribution.

The rumen variables were analyzed by one-way analysis of variance, using the GLM procedure. The mortality rate was analyzed using the Chi-squared test (SAS, 1999).

Significant differences were declared at \(P < 0.05\) and tendencies at \(P < 0.10\).

**Experiment 2 (in vivo - in vitro)**

*Animals and diets*

Nine non-lactating Cashmere goats (28.3 ± 2.30 kg BW) were used in this experiment, which was performed at an experimental research station. Animals were individually fed a mixture of alfalfa and grass hays (AGH; 18 g DM/kg of BW) once a day over a 14-day period. They were then switched to a diet containing 71% of the same mixture of hays and 29% heather (AGH+H; 18 g DM/kg of BW) for 12 more days. The heather was collected from the same area as in Experiment 1. These diets (treatments) were designed on the basis of the outcome of diet selection (see results from Experiment 1), to experimentally simulate the \(\sim H\) and \(+H\) treatments.

Table 1 shows the chemical composition of the mixture of grass and alfalfa hays and of the heather used in this experiment.

**In vitro ruminal fermentation**

The effect of the diet on *in vitro* ruminal fermentation was studied in four runs of incubation using a modification of the gas production technique described by Theodorou et al. (1994). On day 12 of the experiment, after an overnight fast, ruminal fluid was collected from each animal by stomach tube (as in Experiment 1). The collected samples were transferred to the laboratory in pre-warmed thermos flasks, strained through a double layer of muslin, and kept
under CO₂ flushing. The nine inocula were mixed to form a single inoculum (run 1). The whole process was repeated on day 14 to obtain the replicate (run 2). When the goats received the heather-containing diet, rumen fluids were again withdrawn on days 24 (run 3) and 26 (run 4).

Samples of the pasture and heather collected in August during Experiment 1, ground through a 1-mm screen, were used to provide two substrates: pasture (P; 100% pasture) and pasture + heather (P+H; 71% pasture and 29% heather). A third substrate (P+H+PEG) was prepared by adding one gram per flask of a tannin-binding agent (polyethylene glycol, PEG, MW 6000; Fluka Chemie GmbH, Buchs, Switzerland) - which inhibits the effect of tannins (Makkar, 2003) - to the P+H substrate. The large quantity of PEG was chosen because the efficiency of this agent is believed to depend on the tannins/PEG ratio, and to avoid the possibility that too small an amount of this inert compound might be associated with a lack of effect.

Each incubation run included a total of twelve 125 ml sealed serum flasks. Three samples of each substrate (≈ 500 mg) and three blanks (i.e., buffered rumen fluid without substrate) were incubated at 39°C with 10 ml rumen fluid and 40 ml phosphate-bicarbonate buffer. The buffer solution was prepared as described by Goering and Van Soest (1970), with the exception that no trypticase was added.

Gas production was determined by measuring head space gas pressure at 24 h post-inoculation. Pressure values, corrected for the quantity of substrate OM incubated and gas released from the blanks, were used to generate gas volume estimates using a predictive equation derived from earlier simultaneous pressure and volume measurements (Hervás et al., 2005). Immediately afterwards, the fermentations were ended by swirling the bottles on ice. The samples were then centrifuged at 976 × g for 10 min and aliquots of the supernatant collected for ammonia and VFA determinations as in Experiment 1.
In vitro DM disappearance (DMD; g/kg) and true substrate digestibility (ivTSD) after 24 h of incubation were estimated by filtering the residues using pre-weighed sintered glass crucibles (100–160 µm; Pyrex, Stone, UK) and determining the NDF content, as reported in Frutos et al. (2004).

Chemical analyses

All chemical analyses were performed as in Experiment 1.

Statistical analysis

Data for in vitro ruminal variables were analyzed by one-way analysis of variance, using the GLM procedure of the Statistical Analysis System program (SAS, 1999) to examine the effects of the two main sources of variation, goat diets and substrates, as well as their interaction. The mean value of each set of three flasks per substrate and run was considered as the experimental unit. Significance was set at $P < 0.05$; trends were declared at $P < 0.10$. 


Results

Experiment 1

Sward height, and pasture and heather botanical and chemical composition

As shown in Figure 1, the available sward height differed significantly between treatments, with greater values being recorded in the pasture used by the +H animals ($P < 0.001$). Mean sward height gradually fell from 10.2 cm at the beginning of the experiment (May) to 3.7 cm at the end (September) ($P < 0.001$).

The botanical composition of the available pasture in August was on average 6.8% white clover, 21.9% perennial ryegrass, 43.1% other grasses (mainly *Agrostis capillaris* L.), and 27.9% dead matter, with no differences between treatments. The chemical composition of the pasture samples taken in August showed a CP content of 143 g/kg of DM, and values for NDF, ADF and ADL of 590, 269 and 25 g/kg of DM, respectively (Table 1).

The heather offered was mostly composed of *Calluna vulgaris* (L.) Hull (61%), with 25% *Erica umbellata* L., 12% *E. cinerea* L., 1% *E. tetralix* L., and 1% *Daboecia cantabrica* (Hudson) C. Koch. Chemical composition analysis showed a lower CP content (61 g/kg of DM) and greater cell wall content (557, 436 and 224 g/kg of DM for NDF, ADF and ADL, respectively), as expected for shrub species. Total phenol and total tannin contents were estimated to be 94 and 64 g tannic acid equivalents/kg of DM, respectively (Table 1).

Diet composition

The mean percentage of heather incorporated into the diet of the +H animals was 29.1% ($\pm$ 2.06) in August. The remaining 70.9% corresponded to herbaceous species.

Faecal egg counts and goat mortality
Supplementation with heather reduced the mean FEC ($P < 0.001$; see Figure 1); values ranged from 15 epg at the beginning of the trial to a maximum of 2653 at the end of August. The $−H$ goats with the greatest FEC in the two samplings performed in August died before the September sampling (seven in total), which explains the reduction seen in Figure 1. Supplementation with heather significantly reduced the mortality rate ($P < 0.05$): at the end of the grazing period, two $+H$ and eight $−H$ does had to be euthanized.

The nematode genera identified were *Trichostrongylus*, *Teladorsagia*, and *Oesophagostomum*. *Trichostrongylus* was the predominant genus (60–95%) over the entire experimental period; *Teladorsagia* and *Oesophagostomum* were detected in lower percentages (5–25%).

**Live weight and body condition changes**

Table 2 shows that heather supplementation was responsible for the smaller losses of live body weight ($P < 0.01$) and body condition score ($P < 0.01$) recorded over the experimental grazing season in the $+H$ animals.

**Ruminal fermentation variables**

Table 3 shows the ammonia and VFA concentrations in the ruminal fluid of the experimental animals. The rumen ammonia concentration was markedly reduced in the goats receiving tannin-containing heather ($P < 0.01$). On the contrary, total VFA concentration was significantly greater in these animals ($P < 0.05$), mainly due to an increase in acetic acid (approx. 26%) but also in propionic acid (approx. 16%). The concentrations of valerate and branched-VFAs (referred to as “others” in Table 3), however, decreased. The molar proportions of VFA (data not shown) were also modified by supplementation with heather ($P < 0.001$), with increases in acetate and reductions in butyrate, valerate, and branched-VFAs.
Experiment 2

As shown in Table 4, significant differences in gas production, dry matter disappearance, *in vitro* true substrate digestibility and ammonia and volatile fatty acid concentrations were observed due to both goat diet (i.e., including or not heather: AGH vs. AGH+H) and substrates incubated. However, no significant interaction (diet x substrate) was found for any *in vitro* rumen fermentation variables (*P* > 0.10).

Cultures with rumen inoculum derived from goats receiving the heather containing diet (AGH+H) showed slightly lower values of DMD (-3%), ivTSD (-2%) and gas production (-11%) compared to cultures from AGH animals. On the contrary, those incubations (AGH+H) returned greater values of total VFA (23%), mainly accounted for by greater concentrations of acetate (34%) and butyrate (21%). The molar proportions of VFA (data not shown) tended to be modified by the incorporation of heather into the diet (*P* < 0.10), with increases in acetate and reductions in propionate, and valerate and branched-VFAs (referred to as “others” in Table 4).

Differences among substrates (*P* < 0.01) were observed in terms of gas production, DMD, ivTSD and ammonia concentrations, the greatest values always being found for the pasture (P) substrate and the lowest for the P+H substrate. When PEG was added to the mixture (P+H+PEG), gas production and ivTSD were significantly increased. However, the addition of PEG did not significantly inhibit the negative effect of heather on DMD and ammonia concentration.

No significant differences among substrates were observed for the concentration of any particular VFA.
Discussion

The control of GI parasites with anthelmintics is becoming increasingly difficult worldwide due to drug resistance and concerns over pesticide residues in the food chain and the general environment (Van Houtert and Sykes, 1996). Thus, the greater variety of control methods used in combination, the longer we can expect to have effective worm control. The use of tannin-containing plants has proven effective in the control of GI parasite infections, as confirmed in this study where supplementing the diets of grazing goats with tannin-containing heather significantly reduced faecal egg counts (see Figure 1). This is in line with previous reports for the same area by Osoro et al. (2007a and 2007b). However, the mean FEC values (as high as 2652 epg) were notably different from those found in the above papers, which is almost certainly related to the effect of climatic conditions on parasite infections (Stromberg, 1997).

Several authors have reported that the intake of low to moderate quantities of condensed tannins (CTs) may have beneficial effects on parasitized ruminants. For example, we reported reductions in FEC of up to 75%, compared to control goats, in an experiment that included supplementation with a heather containing between 7 and 8.6% tannin (expressed in tannic acid equivalents; Osoro et al., 2007b). Butter et al. (2000) observed a reduction in FEC when 5% of a commercial extract of quebracho (Schinopsis spp.) CTs was included in a low protein diet. Min et al. (2005) reported a great reduction in FEC when does were fed Sericea lespedeza (Lespedeza cuneata) containing 15.2% tannins (using an internal standard from the plant itself as the reference).

In the present study, FEC reduction was achieved with the dietary incorporation of an estimated 29% heather containing 6.4% tannin (which may be considered a low-to-moderate intake of tannins). The lowest dietary tannin threshold needed for anthelmintic effects remains unclear, however. Moreover, it should be remembered that different plants contain
tannins with different structures and hence reactivities, so the notion of a “low-to-moderate” content could be misleading. In addition, different research groups use different methods to analyse this large group of phenolic compounds, but the equivalents in which the tannin content are expressed (e.g., tannic acid, internal standards extracted from the plants themselves, etc.) are not always given, which means that comparisons cannot be made (Makkar, 2003; Álvarez del Pino et al., 2005).

A number of investigations have focused on the potential use of CTs to control GI parasitism because these are relatively stable in the digestive tract of ruminants and rarely have toxic effects (Mueller-Harvey, 2006). CTs may affect protein nutrition by reducing dietary protein degradation in the rumen, thus enhancing the amount of protein available for digestion in the small intestine (Barry and McNabb, 1999; Frutos et al., 2000; Mueller-Harvey, 2006). According to Coop and Kyriazakis (2001), protein nutrition, and therefore CTs, may influence the development and consequences of parasitism by increasing the resilience and resistance of the host or by directly affecting the parasite populations.

Given the low content of protein in the offered heather (61 g/kg of DM), it seems very unlikely that the anthelmintic effect of its supplementation would be due to a greater availability of protein, although there might be a greater availability of non-degradable protein due to the effect of the tannin (Barry and McNabb, 1999). A common characteristic of GI parasitism is an increased loss of endogenous protein to the GI tract (Coop and Kyriazakis, 2001) and some authors have suggested that the inclusion of CTs might be an alternative to providing high protein rations as a means of reducing the consequences of parasitism (Van Houtert and Sykes, 1996; Butter et al., 2000; Coop and Kyriazakis, 2001).

The supplementation with tannin-containing heather significantly reduced the goat mortality rate, which is consistent with previous studies reporting the positive effect of tannin consumption on the resilience of goats (Paolini et al., 2005).
In addition to the effect of heather supplementation and due to the differences observed between treatments in the sward height, it cannot be completely ruled out that the –H goats consumed more contaminated pasture than +H goats. A shorter sward may have larger parasite burdens because it is easier for the parasites to migrate to the top (Niezen et al., 1998), so the non-supplemented animals might be at a higher risk of ingestion of nematode infective larvae. Nevertheless, as both experimental plots had the same previous management, they presumably had a similar parasitic status at the start of the trial.

As observed in earlier work (Osoro et al., 2007a and 2007b), the negative BW gains agree with the relationship established between green sward height and BW change (Merchant and Riach, 1994). Considerable BW losses were observed after July, when the sward height fell below 6–7 cm (see Figure 1), probably due to the limited availability of pasture and the increase in parasite burden during the grazing season (Coop and Kyriazakis, 2001). However, losses were mitigated by the effect of heather intake. Houdijk and Athanasiadou (2003) reported evidence suggesting that the consumption of CT-rich plants can result in reduced parasitism and improved performance in parasitized small ruminants, which would probably be the outcome of indirect nutritional benefits (e.g., extra protein availability for the parasitized grazing hosts). On the other hand, those authors also suggest that the net benefit in the performance of parasitized hosts might be the result of the anti-parasitic effects of CTs outweighing their anti-nutritional effects.

The performance results found in this study were supported by the nutritional outcomes. Although the results on ruminal fermentation obtained in vitro and in vivo were not exactly the same, as expected, they followed a very similar pattern and support the general conclusion that the dose of tannins ingested with the heather diet was insufficient to adversely affect the majority of the rumen microbial populations in goats adapted to its consumption.
Several authors have indicated that dietary concentrations of <50 g CT/kg DM are nutritionally beneficial (these recommendations originated mainly from feeding trials with *Lotus* species and cannot be directly applied to other feeds; Mueller-Harvey, 2006). However, erroneous generalizations have persisted since the first reviews in the 1960s and 1970s stated that tannins are harmful or toxic to mammals (Mueller-Harvey, 2006). The main benefit of tannins in ruminant nutrition stems from their negative effect on proteolysis and the first consequence is a lower concentration of ammonia in the ruminal fluid, as observed in our *in vivo* and *in vitro* approaches. The differences between assays 1 and 2 may be due to the fact that, in *in vitro* studies, lower rumen fluid ammonia concentrations can only be attributed to lower concentrations of protein in the substrate, a reduced deamination of amino acids or a greater rate of uptake by bacteria. In addition, differences may also be explained because in the *in vitro* experiment 29% of the pasture was replaced by exactly the same amount of heather (while substitution in the *in vivo* grazing trial was probably incomplete), and because the response of the ruminal microorganisms to tannins may not be exactly the same *in vivo* and *in vitro*. The fact that no significant differences in ammonia concentration were observed between substrates P+H and P+H+PEG suggests other factors, further to tannins, may also be at work (e.g., the lower nutritive value of P+H compared to P). In the grazing experiment, a significant and negative correlation was found between the percentage of heather incorporated into the diet and the rumen ammonia concentration ($r = -0.60; P < 0.05$).

As well known, VFAs are the most important by-products of fermentation and the major energy source of ruminants. The higher concentrations of VFAs for the supplemented goats in both trials (17% and 23%, respectively) is in line with the better performance (BW and BCS) of these animals. The fact that no significant differences were found in total VFA concentration between substrates, but were found when comparing goat diets, suggests an
adaptation of rumen microorganisms to the consumption of these phenolic compounds (Smith et al., 2005) and a subsequently better digestive utilization of the diet.

The direct effects reported for tannins on rumen VFAs are not consistent and different authors have indicated different results (e.g., Makkar et al., 1995; Hervás et al., 2003); these discrepancies probably depend on the amount and type of tannins and on the animal species that ingest them. Results from experiments 1 and 2 revealed a significant increase in acetate production when heather was present, as well as differences in the fermentation pathways, reflected in the different molar proportions. The increase in acetate and the reduction in gas production may be accounted for by an effect of tannins on acetate-utilizing rumen bacteria, which is probably related to the beneficial and known inhibition of methane-producing microorganisms by these secondary compounds (Tavendale et al., 2005). These results support the improved efficiency of ruminal fermentation in animals consuming tannin-containing heather and confirm those reported in a previous grazing experiment performed under similar conditions (Osoro et al., 2007b).

Comparisons of the results for goats adapted versus non-adapted to the consumption of heather (AGH vs. AGH+H; Table 4) show the latter to be associated with reductions in gas production, DMD and ivTSD (in accordance with that widely reported for diets containing tannins; Frutos et al., 2004; Mueller- Harvey, 2006) of a lower magnitude than the improvements in the mean indicators for the utilization of nitrogen or energy (ammonia and VFA concentrations).

In conclusion, the present results suggest there is no substantial nutritional cost counteracting the anthelmintic effect of supplementing the diets of grazing goats with heather, and they therefore support the hypothesis under test. Further research, including the estimation of total pasture and heather intake, would help explain the effects of heather supplementation on nutrition and performance in grazing goats. Eventually, it would also
contribute to the development of a sustainable method for controlling GI nematode parasitism in extensive goat production systems.

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performance in Cashmere and local Celtiberic goats on pasture. Small Ruminant Research 67, 184-191.


Table 1 Chemical composition (g/kg of DM, except for DM itself [g/kg of fresh matter]) of feeds and diets used in Experiments 1 and 2

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<tr>
<td><strong>Experiment 2 (diets)(^4)</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AGH</td>
<td>865</td>
<td>899</td>
<td>154</td>
<td>591</td>
<td>347</td>
<td>61</td>
<td>nd</td>
<td>nd</td>
</tr>
<tr>
<td>AGH+H</td>
<td>776</td>
<td>921</td>
<td>127</td>
<td>581</td>
<td>364</td>
<td>101</td>
<td>23</td>
<td>33</td>
</tr>
</tbody>
</table>

1 g tannic acid equivalents/kg of DM.

2 Samples collected in August.

3 nd = not determined.

4 AGH = alfalfa and grass hays; AGH+H = alfalfa and grass hays (71%) + heather (29%).

DM = dry matter; OM = organic matter; CP = crude protein; NDF = neutral detergent fibre; ADF = acid detergent fibre; ADL = acid detergent lignin; TT = total tannins; TP = total phenols.
Table 2  Body weight (BW, kg) and body condition score (BCS) during the experimental grazing period (from May to September) in goats supplemented (+H) or not (−H) with heather

<table>
<thead>
<tr>
<th></th>
<th>BW</th>
<th></th>
<th></th>
<th>BCS</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>−H</td>
<td>+H</td>
<td>SEM</td>
<td>−H</td>
<td>+H</td>
</tr>
<tr>
<td>May</td>
<td>37.0</td>
<td>36.3</td>
<td>1.12</td>
<td>2.57</td>
<td>2.56</td>
</tr>
<tr>
<td>June</td>
<td>34.5</td>
<td>34.2</td>
<td>0.89</td>
<td>2.57</td>
<td>2.59</td>
</tr>
<tr>
<td>July</td>
<td>36.2b</td>
<td>37.7a</td>
<td>0.92</td>
<td>2.53b</td>
<td>2.85a</td>
</tr>
<tr>
<td>August</td>
<td>30.3b</td>
<td>32.4a</td>
<td>0.83</td>
<td>2.17b</td>
<td>2.38a</td>
</tr>
<tr>
<td>September</td>
<td>29.7b</td>
<td>30.7a</td>
<td>0.87</td>
<td>1.45b</td>
<td>2.00a</td>
</tr>
</tbody>
</table>

P - value$^2$

- Treatment: $< 0.01$  
- Date of sampling: $< 0.01$  
- Treatment x date: $< 0.01$

$^1$ SEM = standard error of the mean.

$^2$ Probability value.

$^{a,b}$ For each variable, means in a row with a different superscript differ significantly ($P<0.05$).
Table 3 Ammonia (mg/l) and volatile fatty acid (VFA, mmol/l) concentrations in the ruminal fluid of grazing goats whose diets were supplemented (+H) or not supplemented (−H) with heather

<table>
<thead>
<tr>
<th></th>
<th>−H</th>
<th>+H</th>
<th>SEM</th>
<th>P – value²</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ammonia</td>
<td>209.4a</td>
<td>159.7b</td>
<td>9.95</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>Total VFA</td>
<td>53.61b</td>
<td>62.98a</td>
<td>2.340</td>
<td>0.02</td>
</tr>
<tr>
<td>Acetate</td>
<td>36.11b</td>
<td>45.47a</td>
<td>1.845</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>Propionate</td>
<td>8.80b</td>
<td>10.25a</td>
<td>0.500</td>
<td>0.07</td>
</tr>
<tr>
<td>Butyrate</td>
<td>5.48</td>
<td>5.08</td>
<td>0.270</td>
<td>0.34</td>
</tr>
<tr>
<td>Others³</td>
<td>3.22a</td>
<td>2.17b</td>
<td>0.137</td>
<td>&lt; 0.01</td>
</tr>
</tbody>
</table>

¹ SEM = standard error of the mean.

² Probability value.

³ Calculated as the sum of valerate, isovalerate, isobutyrate and caproate.

a,b Means in a row with a different superscript differ significantly (P<0.05).
Table 4  Gas production (ml/g OM), dry matter disappearance (DMD; g/100 g), in vitro true substrate digestibility (ivTSD; g/100 g), ammonia (mg/l) and VFA concentration (mmol/l) in batch cultures after 24 h incubation

<table>
<thead>
<tr>
<th>Goat diet¹</th>
<th>Substrate²</th>
<th>P – value³</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AGH</td>
<td>AGH+H</td>
</tr>
<tr>
<td>Gas production</td>
<td>118.8ᵃ</td>
<td>105.2ᵇ</td>
</tr>
<tr>
<td>DMD</td>
<td>48.48ᵃ</td>
<td>46.94ᵇ</td>
</tr>
<tr>
<td>ivTSD</td>
<td>65.95ᵃ</td>
<td>64.61ᵇ</td>
</tr>
<tr>
<td>Ammonia</td>
<td>287.7</td>
<td>284.7</td>
</tr>
<tr>
<td>Total VFA</td>
<td>39.2ᵃ</td>
<td>48.45ᵃ</td>
</tr>
<tr>
<td>Acetate</td>
<td>23.6ᵇ</td>
<td>31.8ᵃ</td>
</tr>
<tr>
<td>Propionate</td>
<td>10.44</td>
<td>10.82</td>
</tr>
<tr>
<td>Butyrate</td>
<td>3.01ᵇ</td>
<td>3.65ᵃ</td>
</tr>
<tr>
<td>Others</td>
<td>2.17</td>
<td>2.18</td>
</tr>
</tbody>
</table>

¹ AGH = alfalfa and grass hays; AGH+H = alfalfa and grass hays (71%) + heather (29%).
² P = pasture; P+H = pasture (71%) + heather (29%); PEG = polyethylene glycol.
³ SEM = standard error of the mean.
⁴ Probability value.
⁵ Calculated as the sum of valerate, isovalerate, isobutyrate and caproate.
⁶ For each source of variation (goat diet or substrate), means in a row with a different superscript differ significantly (P<0.05).
Faecal gastrointestinal nematode egg counts (FEC; eggs/g; solid lines —) and sward height (cm; short-dash lines ---) over the experimental season in grazing goats whose diets were supplemented - or not - with heather. Mean log transformed FEC values = 2.20 vs. 2.66 eggs/g for supplemented vs. non-supplemented animals (sed = 0.115; $P < 0.001$), and mean sward heights = 6.8 vs. 5.7 cm for supplemented vs non-supplemented animals (sed = 0.07; $P < 0.001$). (sed = standard error of the differences of means)
FIGURE 1

Sward height

May June July August September

FEC

non-supplemented with heather
supplemented with heather