Regional genetic differentiation among populations of *Cladocora caespitosa* in the Western Mediterranean

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*Keywords. Microsatellites, genetic differentiation, broadcast spawning coral, self-recruitment, oceanographic barriers*
Abstract

Cladocora caespitosa is the only reef-forming zooxanthellate scleractinian in the Mediterranean Sea. This endemic coral has suffered severe mortality events at different Mediterranean sites, due to anomalous summer heat waves related to global climate change. In this study, we assessed the genetic structure and gene flow among four populations of this species in the Western Mediterranean Sea: Cape Palos (SE Spain), Cala Galdana (Balearic Islands), Columbretes Islands and L’Ametlla (NE Spain). The results obtained, from Bayesian approaches, FST statistics and Bayesian analysis of migration rates, suggest certain levels of genetic differentiation driven by high levels of self-recruitment, a fact that is enhanced by egg retention mechanisms. On the other hand, genetic connectivity among distant populations, even if generally low, seems to be related to sporadic dispersal events through regional surface currents linked to the spawning period which has been described to occurs during the end of the summer-beginning of autumn. These features, together with a certain isolation of the Columbretes Islands, could explain the regional genetic differentiation found among populations. These results will help to better understand population structure and connectivity of the species, and will serve as an approach for further studies on different aspects of the biology and ecology of C. caespitosa.

Introduction

The Mediterranean Sea is a semi-enclosed sea that has been characterized as a “miniature ocean” responding quickly to changes in temperature and increases in extreme events (Lejeusne et al. 2009). Global climate change, in synergy with other disturbances such as water pollution, coastal development, massive algal blooms, pathogenic organisms and invasive species, among others, is expected to have significant effects on Mediterranean biodiversity in the near future (Lejeusne et al. 2009; Templado 2014). For over a decade, mass mortality events have recurrently impacted Mediterranean benthic communities due to prolonged heat waves, affecting some emblematic sessile invertebrates (Perez et al. 2000; Cerrano et al. 2009; Garrabou et al. 2009; Lejeusne et al. 2009). The scleractinian coral Cladocora caespitosa is among these affected species, and mortalities have been noted in some of its most remarkable reefs, such as those in the Adriatic Sea (Kružić et al. 2012), Gulf of La
Spezia (Rodolfo-Metalpa et al. 2005) and Colombretes Islands, where over 50 % of the coral cover has been affected by necrosis during the last decade (Kersting et al. 2013a).

*Cladocora caespitosa* (Linnaeus, 1767) is the sole Mediterranean colonial and zooxanthellate scleractinian with reef-forming capacity (Morri et al. 1994). It is considered a relict species from the subtropical late Pliocene and Quaternary periods (Kühlman et al. 1991; Peirano et al. 2009) according to the fossil record (e.g., Fornós et al. 1996; Bernasconi et al. 1997; Aguirre and Jiménez 1998). Nowadays, the total abundance of *C. caespitosa* is reduced overall in the Mediterranean. The remaining populations of this coral are patchily distributed across the entire Mediterranean Basin and only few living banks (large colonies more than 1 m high and covering several square meters in surface area) and/or beds (groups of small globose to hemispherical colonies) of this coral been recorded in some spread locations (Laborel 1961; Schiller 1993; Morri et al. 1994; Peirano et al. 1999; Kružic and Benković 2008; Özalp and Alparslan 2011; Kersting and Linares 2012). The decrease of the geographical range of *C. caespitosa* reefs in the Mediterranean with respect to its fossil distribution prompted Augier (1982) to include this coral in a list of marine species under extinction risk. Furthermore, this regression is still in progress enhanced by the mortality events that the coral has suffered in recent decades due to heat waves and other threats such as the proliferation of invasive algal species (Kružic et al. 2008b, Kersting et al. 2014a).

The reproductive ecology of *C. caespitosa* was described in populations from the Adriatic Sea (Kružic et al. 2008) and Western Mediterranean Sea (Kersting et al. 2013b). In the Adriatic Sea, the colonies of *C. caespitosa* have been described as hermaphroditic and the spawning time was observed at the beginning of the summer, in coincidence with rising seawater temperatures (Kružic et al. 2008). Contrary to the Adriatic Sea, in the Western Mediterranean this coral has been regarded as gonochoric
and the spawning period seems to occur at the end of summer (August-October) coinciding with the beginning of the cooling of the seawater (Kersting et al. 2013b). *In situ* observations in the Adriatic Sea indicated that the release of male and female gametes was not synchronous for each hermaphroditic colony. Eggs were released by the polyps in a mucus coating, also called “clumps”, that bound the eggs together, while sperm were freely released in sperm bundles (Kružic et al. 2008a). According to the authors, fertilization may be enhanced by synchronous spawning and eggs retention on the colony surface.

The ability of marine populations to resist and recover from environmental disturbances depends in part on population connectivity and recruitment processes, which contribute to the addition of new individuals to populations and therefore to the population resilience of marine organisms (Caley et al. 1996), particularly corals (Adjeroud et al. 2007) including *C. caespitosa* in the Mediterranean (Kersting et al. 2014b). In order to improve our knowledge on the recovery potential of this bioconstructive species, *C. caespitosa*, a better understanding of the resilience of its populations is needed. Population genetics studies enable the estimation of the structure and connectivity of populations and identify the associated processes, detecting populations self-recruitment, barriers to gene flow, introgression, isolation and fragmentation of the populations (Baums 2008). It should be noted that these features are linked to species life history traits, such as species dispersal capacity at each stage of its life cycle, and their interaction with associated biotic and abiotic factors (DiBacco et al. 2006). Moreover, dispersal among populations can be affected by physical barriers (hydrographic fronts, upwelling systems, eddies or counter currents), or enhanced by oceanographic features such as global and local water currents or rafting events (Pineda et al. 2007).
The present study aims to examine the genetic structure and connectivity patterns of some populations of the Mediterranean reef-forming coral *C. caespitosa* from the Western Mediterranean Basin. The egg retention mechanisms described for this coral may increase self-recruitment and evidences of it can be found in the distribution and recruitment patterns of its populations (Kersting and Linares 2012, Kersting et al. 2014b). Therefore, it is hypothesized that populations of *C. caespitosa* in the Western Mediterranean basin are isolated showing different levels of genetic differentiation between them.

**Material and Methods**

**Study area and sample collection**

Colonies of *C. caespitosa* were sampled at four localities in the Western Mediterranean Sea: Cape Palos (Murcia, SE Spain), Cala Galdana (Menorca, Balearic Islands), Columbretes Islands (at the edge of the continental shelf 60 km off the nearest coast of E Spain) and L´Ametlla (Tarragona, NE Spain) (Table 1, Fig. 1).

At each sampling locality, individual polyps from 13 to 37 colonies were collected at depths between 10-15 m, using SCUBA diving. Sampled colonies were randomly chosen within a given area of 300-500 m², considering a minimum distance between colonies of 1-2 m to avoid sampling the same colony twice. To minimize the damage to the sampled colonies, 1-2 polyp tips were carefully detached from one of the edges of each colony with a hammer and chisel and placed in labeled bags. Samples were stored in vials of absolute ethanol until laboratory analyses. It should be noted that the small number of samples from Cape Palos is due to the low number of colonies found in the area.
**DNA extraction, microsatellite amplification and genotyping**

Total DNA was extracted from a total of 108 polyps using a Qiagen BioSprint IT 15DNA Blood Kit (Table 1). Eight microsatellite loci specifically developed for *C. caespitosa* (Casado-Amezua et al. 2011) were amplified with eight primer pairs with a fluorescently-labeled forward primer, following the PCR conditions described in Casado-Amezua et al. (2011). PCR products were visualized with an automated sequencer (ABI PRISM 3730 DNA Sequencer, Applied Biosystems) with the GeneScan-500 (LIZ) internal size standard. Electropherograms were analyzed for allele scoring with GeneMapper software 3.0 (Applied Biosystems).

**Data analysis**

The possibility of the presence of clone mates (i.e., individuals with the same genotypes) was estimated from the allele frequencies observed in the multilocus genotypes in each population using GenAlEx (Peakal and Smouse 2006). In addition, the probability that the two randomly chosen individuals in a population had identical genotypes by chance was calculated with the Probability of Identity (P_ID) index using GeneAlEx software. This index resulted in a low average value across populations of $3.04 \times 10^{-5}$. Therefore, since individuals that showed identical genotypes could be considered as clones, each distinct eight-locus genotype in the data set was included for statistical analysis.

Linkage disequilibrium (LD) was tested among all pairs of loci at each of the sampled locations with a permutation test using GenePop version 3.4 (Raymond and Rousset 1995). Analysis of significance was tested with Markov Chain Monte Carlo (MCMC that was run using 1,000 dememorizations with 100 batches and 1,000 iterations per batch). MICRO-CHECKER v.2.2.3 software (van Oosterhout et al. 2004)
was used to check for scoring errors due to stuttering, large allele dropout and to estimate null allele frequencies.

**Genetic diversity**

Parameters of genetic diversity for each population and for the global sample were estimated. Allelic diversity ($N_a$) was quantified as the number of alleles per locus over all loci and localities using GenAlEx 6.0 software (Peakall and Smouse 2006). Allelic richness and private allelic richness were estimated with a rarefaction procedure using HP-RARE software (Kalinowsky 2005), with the minimum number of genes set at 24 ($N_{S(24)}$; $P_{(24)}$). Analyses of departures from Hardy-Weinberg equilibrium (HWE) within populations for each locus and over all loci were quantified as the observed ($H_o$) and expected ($H_e$) heterozygosities using GenAlEx 6.0 software (Peakall and Smouse 2006). Estimations of the inbreeding coefficient, $F_{IS}$, an estimate of the deficit or excess of heterozygotes, within each population for each locus and over all loci were computed with Genetix (Belkhir et al. 2004). Significance of the estimation analysis was tested with 10,000 permutations.

Step-down Bonferroni (Holm 1979) correction was applied to p-values in all the statistical analyses that included multiple comparisons.

**Inference of population differentiation**

POWSIM v4.0. (Ryman and Palm 2006) was used to estimate the statistical power provided by the microsatellite dataset for correctly testing for population genetic differentiation. The four localities sampled were used for testing allele frequency homogeneity at the eight loci and combining information for the multiple loci using Fisher’s exact and chi-squared tests. Simulations were run using various combinations of $N_e$ (effective population size) and $t$ (time since divergence), leading to $F_{ST}$ values of
0.001 to 0.05, reproducing the magnitude of $F_{ST}$ values estimated from the data. Simulations were carried out for effective population sizes $N_e = 2,000$, $N_e = 5,000$ and $N_e = 10,000$ to yield $F_{ST}$ values of 0.001, 0.025 and 0.05. One thousand replicates were run and the power of the analysis was indicated by the proportion of tests that were significant at $p < 0.05$ using the respective allele frequencies at the eight loci studied.

For the measures of genetic differentiation, sample sites were used as a population unit. $F_{ST}$ values between sampling localities were estimated using ARLEQUIN 3.1.1. software (Excoffier et al. 2005), and their statistical significance calculated 10,000 bootstrap replicates (Weir and Cockerham 1984). As null alleles can induce overestimation of genetic distances (Chapuis and Estoup 2007), pairwise estimates were computed with and without correction for null alleles (Brookfield 1996).

Population genetic structure was also inferred using a Bayesian approach. The number of genetically differentiated $C. caespitosa$ populations, $K$, with the highest posterior probability given the data was estimated with STRUCTURE 2.3.3 (Pritchard et al. 2000). The software was run using the admixture model under the “location prior” function (LOCPRIOR) with correlated allele frequencies. LOCPRIOR was considered as the most accurate model because even while it learns from a priori assignment of individuals into populations, it does not tend to infer structure when none exists (Pritchard et al. 2000; Hubisz et al. 2009). Simulations included 20 replicated runs for $K$ values, 1 to 10 and a mean log probability of the data ($\ln P(K)$) was calculated. MCMC of each run consisted of 100,000 burn-in iterations followed by 1,000,000 sampled iterations. $\Delta K$ values are an index for deciding the probable number of genetically clustering populations. $\Delta K$ values were calculated under the Evanno et al. (2005) method as implemented on STRUCTURE Harvester (Earl and von Holdt 2012)
The influence of geographic distance on population genetic differentiation was calculated with two models: Rousset (1997) model for two-dimensional habitats with the correlation between pairwise population linearized $F_{ST}$ ($F_{ST}/ (1-F_{ST})$) and the logarithm of the geographical distance (in meters); and Slatkin (1993), which used the logarithm of $\hat{M}$ calculated separately for pairs of populations, (where $\hat{M} = (1-F_{ST})/4F_{ST}$) as a measure of similarity, versus the logarithm of the geographical distance. This parameter corresponds to the number of migrants required to account for observed genetic differences if migrants could move directly between populations. In both cases, Mantel tests (10,000 permutations) were used to assess the statistical significance. The strength of the isolation by distance (IBD) relationship was quantified with the slope and intercept of genetic similarity ($\hat{M}$) or distance (linearized $F_{ST}$) against geographic distance. Both parameters were calculated using reduced major-axis regression (RMA) (Sokal and Rohlf 1981). Asymmetric 95% confidence intervals around the RMA regression coefficient were calculated with 10,000 bootstraps around individual population pairs. All the analyses were done with IBDWS software (Jensen et al. 2005). Geographical distances were measured by means of dead-reckoning distances using MatLab software.

**Estimation of migration rates**

We used the Bayesian model implemented in BayesAss+ v1.3 (Wilson and Rannala 2003), to estimate recent migration rates, $m$, among populations. The model was run under default parameters. As suggested by Baums et al. (2005) and Cibrián-Jaramillo et al. (2010), a jackknife procedure was used sequentially omitting each of the sampling localities from the migration matrix. This approach is based on the fact that particular population features, such as demographic factors (Cornuet et al. 1999; Wilson and Rannala 2003), may affect migration rates of the other localities. Thus, if a
population contributes migrants, its exclusion will increase the other population's self-recruitment rates. If a population contribution is minimal for the rest of the populations, its exclusion might decrease mean self-recruitment rates. Self-recruitment migration matrices were obtained following the procedure in Baums et al. (2005), by obtaining four matrices sequentially omitting each of the sampling localities and one matrix for all the populations. The diagonals present in the resulting matrices were summarized and compared with the overall population’s matrix.

Results

A total of 108 individuals were genotyped for 8 microsatellite loci. After removing identical genotypes, from all populations except Cape Palos were no identical genotypes were found, a total of 101 unique genotypes were considered for the analyses (33 in Cala Galdana, 27 in Columbretes Islands, 28 in L’Ametlla and 13 from Cape Palos (Table 1)). Linkage disequilibrium (LD) among loci was found only in three of the twenty-eight pairwise comparisons per sampling locality (P < 0.05 after step-down Bonferroni correction). None of the analyses were significant at the same time for all loci in all populations. Therefore physical linkage can be discarded.

Across localities, allelic diversity (\(N_a\)) ranged from 4.1±1.1 (L’Ametlla) to 5.9±1.5 (Columbretes Islands) (mean value ± standard error (SE) here and hereafter) and private allelic richness from 0.1±0.1 (L’Ametlla) to 1.5±0.7 (Columbretes Islands) (Table 1). After rarefaction, allelic diversity parameters ranged from 3.5±0.9 (L’Ametlla) to 4.9±1.3 (Cape Palos) and private allelic richness from 0.1±0.1 (L’Ametlla) to 1.0±0.5 (Columbretes Islands) (Table 1). Departures from HWE, measured as \(F_{IS}\), were not generalized over all loci in each population. In cases of heterozygote deficiencies, evidence for null alleles was checked and their frequencies
were computed at each locus for each sampled site. No evidence of allelic dropout or scoring errors due to stuttering was found. The analysis detected the possibility of null alleles in locus CcL5 in Cala Galdana, Columbretes Islands and L’Ametlla and in CcL16 in L’Ametlla, due to an excess of homozygotes in most allele size classes. Null allele frequencies for locus CcL5 ranged from 0.09 (Columbretes Islands) to 0.27 (L’Ametlla). However, applying the corresponding correction for null alleles (Brookfield 1996, in all cases) did not qualitatively affect the results (Table S1). Overall loci, FIS values ranged from -0.035 (Cala Galdana) to 0.087 (L’Ametlla) (Table 1). After null allele correction, the overall loci FIS value from L’Ametlla decreased, however the value remained significant for the Columbretes Islands.

*Population genetic structure and gene flow*

Simulations performed with POWSIM v4.0 suggested that the statistical power of the microsatellite was high for detecting FST values of 0.01 (91% and 88% probabilities according to the chi-square test and Fisher, respectively) and 0.05 (98.2% probability according to chi-square and Fisher tests). The α error (type I, proportion of false significances) was also estimated by performing simulations of no divergence among populations (i.e., setting t=0). This resulted in a proportion of 1% significances, which is lower that the intended value of 5%.

The Bayesian clustering analysis detected the highest likelihood (LnP(K) = -1760.83±46.83) with K=5. Also, the modal value of ΔK (Evanno et al. 2005) was shown at K=5 (Fig.1 supplementary material), showing genetic differentiation among the studied population nevertheless with certain levels of admixture. The highest proportion of membership of each sampled population within each cluster defined by STRUCTURE corresponded to (Fig. 2): Cluster 1- Columbretes Islands, Cluster 2- L’Ametlla, Cluster 3-Cala Galdana, Cluster 4- Cape Palos. The fifth cluster (Cluster 5)
was shown to be formed on a lesser or greater extent from individuals of the four sampled localities, however being the population of L’Ametlla the one contributing more to the cluster (Fig. 2).

Pairwise $F_{ST}$ values (Table 2) ranged from 0.012 (Cape Palos-L’Ametlla) to 0.051 (Cala Galdana-Columbretes Islands). After correction for null alleles, no significant differences between pairwise $F_{ST}$ values and pairwise $F_{ST}$ values corrected for null alleles were observed (T-test, $p = 0.560$). Therefore, it is possible to assume that the presence of null alleles did not affect the analyses. The highest $F_{ST}$ values were found in relation to pairwise comparisons concerning Columbretes Islands and the other localities (Table 2), that is in concordance with the clustering analysis which shows two groups, one formed by the populations of Cape Palos, Cala Galdana and L’Ametlla and the other with Columbretes Islands.

$F_{ST}$ analyses were also performed considering the clustering approach in which, even if the optimal K value was 2, three different groups of individuals would be considered: a group formed mainly by individuals from L’Ametlla, another one of individuals from Columbretes Islands and a third one admixed between the remaining two groups of populations (Cape Palos-Cala Galdana). The results indicate levels of genetic differentiation from 0.013 (Cape Palos-Cala Galdana vs L’Ametlla) to 0.040 (Cape Palos-Cala Galdana vs Columbretes Islands) (Table 3). Neither of the two isolation by distance models, Slatkin or Rousset, showed significant relationships between genetic and geographic distances: (Slatkin model: $F_{ST}/(1-F_{ST}) = 0.025-0.037\log(\text{dist})$, $R^2 = 0.20$, $p = 0.80$; Rousset model: $\log(M) = 1.35-6.24\log(\text{dist})$, $R^2 = 0.28$, $p = 0.95$).

Self-recruitment rates showed values of 0.69±0.02 in Cape Palos to 0.90 ±0.02 in L’Ametlla. Recent migration rates were high only between the population of
L’Ametlla and the others, as indicated with immigration rates of ca. 0.3. Migration rates between the other populations were very low (≤ 0.04). Despite the overall lack of differentiation between localities, the jackknife procedure showed higher rates of self-recruitment in all the populations when excluding L’Ametlla (Table 4).

**Discussion**

The population structure of marine organisms reflects the historical and contemporary interaction at different spatial and temporal scales among a complex set of ecological, demographic, behavioral, oceanographic, climatic and geological processes (Grosberg and Cunningham 2001). Past geological and ecological events, together with the corals' sexual reproductive traits and their relation to oceanographic and geomorphologic processes may explain the present genetic structure of *Cladocora caespitosa*. Our results suggest that the studied populations are highly dependent on self-recruitment, likely enhanced by the egg retention mechanisms shown by this species (Kružić et al. 2008); all of which is reflected in the distribution and recruitment patterns described for this species (Kersting and Linares 2012, Kersting et al. 2014b).

The clustering analysis shows that *C. caespitosa* in the Western Mediterranean Sea forms five clusters and denotes genetic structure between the studied populations. It should be noted that the studied clusters share alleles and allelic frequencies, and therefore, are not fully isolated from each other. According to the Bayesian analysis, the individuals from the sampled populations are divided in 4 different clusters and the gene flow of individuals from not sampled populations might affect the obtainment of a fifth cluster (Slatkin 2005; Hellberg 2007). Even so, this analysis should be considered with caution as the performance of STRUCTURE algorithms decreases with sample size (Latch et al. 2006), such as might have occurred for the population of Cape Palos (N= 13).
Small but highly significant $F_{ST}$ values were found between the Columbretes Islands population and the others, while slight but significant or non-significant values were found among the other populations. Migration rates showed the prevalence of self-recruitment processes within each population (from 0.69±0.02 in Cape Palos to 0.90±0.02 in L’Ametlla). These features might be influenced by the characteristic spawning mechanism of the coral (“clumps” of eggs, Kružic et al. 2008) and may suggest that genetic connectivity is somehow maintained by sporadic events of gene flow among the studied populations (Hellberg 2002). There is a general lack of studies regarding the behavioral mechanisms of *C. caespitosa* planulae and its relation to environmental factors. However, with the obtained results we could assume that sporadic gene flow events might be the consequence of variable hydrodynamic conditions and of structure and continuity of rocky substrates along the coastline along together with other external factors. Another explanation of the small values of $F_{ST}$ together with a certain degree of structure could be due the occurrence of recruitment pulses, i.e. sporadic events of successful recruitment from external sources which might be influenced by the reproductive biology of the species coupled with extrinsic factors such as the variable current system, which in some anthozoans and marine invertebrates have been shown to have a great impact on the increase in population densities (Yoshioka 1996; Connell et al. 1997; Sams and Keough 2012).

The sampled area is located in the Balearic Basin. This area is dominated by the Northern Current, which flows southwards along the Iberian Peninsula until it reaches the Ibiza Channel, where it both continues southwards and re-circulates cyclonically, to a lesser or greater extent, over the Balearic Islands forming the Balearic Current (Font et al. 1990). Both currents, the Northern and the Balearic, have been regarded as the motor for the flow surface circulation of the region (Font et al. 1995). The surface layer
circulation is seasonal in character. During the spring and summer, the formation of a gyre partially deviates the Northern current towards the Balearic Sea, thus lowering water flow through the Ibiza Channel. During the autumn-winter season, this gyre is less evident due to the weaker character of the Balearic Front and the increase in intensity of the Northern Current (LaViolette et al. 1990; Font et al. 1995). The relative seasonal intensities of the Northern Current and the mentioned gyre play an important role in the gene flow in this region. For example, in the red gorgonian \textit{Paramuricea clavata}, the seasonal gyre that occurs before the Ibiza Channel has been thought to reduce southward gene flow between populations, due to the coincidence in the reproductive timing of the species (between June and July) and the intensification of the gyre (Mokthar-Jamaï et al. 2011). In contrast, in the case of \textit{C. caespitosa}, our results showed that southwards gene flow is not impeded as most of the immigrants were determined to come from the area of L’Ametlla. In the fish \textit{Serranus cabrilla}, it has been suggested that this barrier is directional, as gene flow moves from north to south and from the southern localities of this channel towards the Balearic Islands (Schunter et al. 2011). According to the data obtained here, this barrier also seems to act as a directional barrier for \textit{C. caespitosa}, as the highest level of gene flow was found southwards. Therefore, north to south gene fluxes are much more important in magnitude than south to north fluxes via recirculation over the Balearic Islands. This is in concordance with the spawning period of \textit{C. caespitosa} in the western Mediterranean, which occurs at the end of summer-beginning of autumn (Kersting et al. 2013b), also in coincidence with the increase in intensity of the Northern Current.

The population from Columbretes Islands was shown to have a certain genetic differentiation. This might also explain the excess of homozygotes found in the Columbretes Islands even after the correction analyses of the null alleles, as
heterozygosity deficiencies may be caused by historical and demographic events such as selection, population mixing and non-random mating (Luikart et al. 2003). Colonies from this population were sampled in the Illa Grossa Bay, which is a semi-enclosed C-shaped volcanic caldera, open to the direction of winter storm waves, in which the coral colonies form banks and beds in a mixed manner (Kersting and Linares 2012). The colonies of *C. caespitosa* in the Columbretes Islands present a highly aggregated distribution, since most of the colonies are concentrated inside the volcanic caldera. This distribution has been associated with sea bottom morphology and hydrodynamic protection together with the reproductive strategies of the species (Kersting and Linares 2012). The isolated location of the Columbretes Islands (60 km off the nearest coast) together with the protection of the bay homing the coral banks and its egg-retention mechanisms, which may reduce their dispersal, may be the causes of the differentiation of the Columbretes Islands population from the other sampled populations, as previously suggested by Kersting and Linares (2012).

This is the first study dealing with population genetics features of *C. caespitosa*, a relict species in the Mediterranean Sea. The high self-recruitment levels and the low and probably intermittent connectivity found, together with the slow dynamics of the species (Kersting et al. 2014b), may indicate a reduced recovery potential from the recurrent heat wave-induced mortalities that are affecting the species (Kersting et al. 2013a). The results obtained will promote conservation plans for the populations of this coral. An adequate marine reserve network be designed considering source populations of the coral, such as L’ Ametlla, and differentiated populations, such as those in the Columbretes Islands. This will assure connectivity among populations and genetic diversity, as has been suggested for other anthozoans in which populations are highly maintained by self-recruitment processes in the Mediterranean, such as the red
gorgonian *Paramuricea clavata* (Mokhtar-Jamaï et al. 2011) and the orange coral *Astroides calycularis* (Casado-Amezúa et al. 2012). Moreover, studies on modular growth, planulae behavior and dispersal abilities, and recruitment processes are recommended in order to better understand the population dynamics and resilience of *C. caespitosa* in the face of a changing environment, thus informing management and conservation strategies.

**Acknowledgements**

We want to acknowledge A Luque, M Taviani, P Montagna, J Fayos, E Serrano and A Addamo for their help either in the field work or providing samples, and MH Nielsen for help on data analysis. We thank S Young and K Nielsen for English revision of grammar and style. C Papetti helped us to improve the manuscript with suggestions on the analyses. Two anonymous reviewers help to improve this manuscript. We thank the Secretaría General de Pesca Maritima (MAGRAMA) and the Columbretes Islands Marine Reserve staff for their logistical support. We thank the Spanish authorities for Environment and Fisheries, Valencia Autonomous Community and Balearic Islands Regional Government the permits for collecting the samples. This study was supported by the Autonomous Organism of National Parks (Spanish Ministry of Environment) and projects CGL2011-23306 from the Spanish Ministry of Science and Innovation, and COCONET-- “Towards COast to COast NETworks of marine protected areas (from the shore to the high and deep sea), coupled with sea-based wind energy potential”-- from the VII FP of the European Commission. P. Casado- Amezua is currently funded by a postdoctoral internship from the Alfred Wegener Institute for Polar and Marine Research (AWI).

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Figures

**Figure 1** Left panel, schematic map showing the distribution range of *Cladocora caespitosa* in the Mediterranean as well as the main known living reefs of the species. CL, Columbretes Islands (Kersting and Linares 2012); MA, Port- Cross, Gulf of Marseille (Zibrowius 1980); SP, Gulf of La Spezia (Morri et al. 1994); PI, Bay of Piran (Schiller 1993); IZ, Iz Island; PG, Pag Island; PR, Prvic; ML, Mjlet (Kružić and Benkovic 2008); EU, Eubée, Gulf of Atalante (Laborel 1961); MR, Marmara Sea (Özalp and Alparslan 2011). Right panel: sampled populations of *Cladocora caespitosa*. A schematic of sea-surface currents and main oceanographic barriers is shown. BF: Balearic Front; IC: Ibiza Chanel; MC: Mallorca Chanel. Continuous arrows indicate main currents; discontinuous arrows indicate mesoscale currents throughout the year (modified from Ruiz et al. 2009).

![Schematic map showing the distribution range of *Cladocora caespitosa*](image1.png)

**Figure 2.**
Bar plot of the Bayesian clustering analyses performed with STRUCTURE 2.3.3. software. Cluster 1 is represented by green, cluster 2 by violet, cluster 3 by pink cluster 4 by yellow and cluster 5 by red. The values above the figure indicate the proportion of membership of each pre-defined population (i.e. sampled locality) in each of the 5 clusters.

![Bar plot of the Bayesian clustering analyses](image2.png)
Tables

**Table 1** Data of sampled populations of *Cladocora caespitosa*. N: number of collected and analyzed individuals. Ng: number of unique multilocus genotypes. Na: average number of alleles per population. \( N_{S(24)} \): mean allelic richness standardized to the smallest sample size. \( P_{S(24)} \): number of private alleles per population standardized to the smallest sample size. \( H_o \) and \( H_e \): mean observed and expected heterozygosities. \( F_{IS} \): inbreeding coefficient. Bold \( F_{IS} \): significant values of probability estimates. * Significant \( F_{IS} \) values after Brookfield (1996) null allele correction.

<table>
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<tr>
<th>Basin</th>
<th>Population</th>
<th>Location</th>
<th>N</th>
<th>Ng</th>
<th>Na</th>
<th>( P_a )</th>
<th>( N_{S(24)} )</th>
<th>( P_{S(24)} )</th>
<th>( H_o )</th>
<th>( H_e )</th>
<th>( F_{IS} )</th>
</tr>
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<tbody>
<tr>
<td>Algerian Basin</td>
<td>Cape Palos</td>
<td>37°34′34.98″N 0°46′32.64″W</td>
<td>13</td>
<td>13</td>
<td>5.0</td>
<td>0.5</td>
<td>4.9</td>
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<td>39°52′52.62″N 3°59′31.20″E</td>
<td>37</td>
<td>33</td>
<td>5.8</td>
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<td>(0.3)</td>
<td>(1.1)</td>
<td>(0.1)</td>
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<tr>
<td></td>
<td>Columbretes Islands</td>
<td>39°53′38.55″N 0°41′10.37″E</td>
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<td>27</td>
<td>5.9</td>
<td>1.5</td>
<td>4.4</td>
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<td>0.444</td>
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<td>(0.7)</td>
<td>(1.1)</td>
<td>(0.5)</td>
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<td>(0.057)*</td>
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<tr>
<td></td>
<td>L’ Ametlla</td>
<td>40°50′27.20″N 0°44′58.90″E</td>
<td>30</td>
<td>28</td>
<td>4.1</td>
<td>0.1</td>
<td>3.5</td>
<td>0.1</td>
<td>0.413</td>
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<td>0.087</td>
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<td>(0.1)</td>
<td>(0.9)</td>
<td>(0.1)</td>
<td></td>
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<td>(-0.046)</td>
</tr>
</tbody>
</table>

**Table 2**. Pairwise \( F_{ST} \) values between populations. Significant values after step-down Bonferroni correction are highlighted in bold (p < 0.01).

<table>
<thead>
<tr>
<th></th>
<th>Cape Palos</th>
<th>Cala Galdana</th>
<th>Columbretes Islands</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cala Galdana</td>
<td>0.016</td>
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<td></td>
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<tr>
<td>Columbretes Islands</td>
<td>0.028</td>
<td>0.051</td>
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</tr>
<tr>
<td>L’ Ametlla</td>
<td>0.012</td>
<td>0.020</td>
<td>0.027</td>
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</table>
Table 4. Migration rates of *Cladocora caespitosa* (means ± SD) among sampled populations estimated by BayesAss+. Source populations are given in columns, recipient localities in rows. Values along the diagonal are populations' self-recruitment rates.

<table>
<thead>
<tr>
<th>From</th>
<th>Cape Palos</th>
<th>Cala Galdana</th>
<th>Columbretes Islands</th>
<th>L’Ametlla</th>
</tr>
</thead>
<tbody>
<tr>
<td>Into Cape Palos</td>
<td>0.69±0.02</td>
<td>0.02±0.01</td>
<td>0.02±0.02</td>
<td>0.27±0.03</td>
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<tr>
<td>Cala Galdana</td>
<td>0.01±0.01</td>
<td>0.70±0.01</td>
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<td>0.29±0.02</td>
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<td>Columbretes Islands</td>
<td>0.01±0.01</td>
<td>0.01±0.02</td>
<td>0.75±0.02</td>
<td>0.23±0.02</td>
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<tr>
<td>L’Ametlla</td>
<td>0.04±0.01</td>
<td>0.04±0.01</td>
<td>0.01±0.01</td>
<td>0.90±0.02</td>
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</tbody>
</table>